

## NOVEL APPROACHES AND CUTTING EDGE IMMUNOTHERAPIES IN MULTIPLE SCLEROSIS

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### 1. ABSTRACT

MS is a chronic inflammatory disease of the central nervous system (CNS). MS is a predominantly CD4<sup>+</sup> T cell mediated autoimmune disorder. Recent studies have challenged this existing paradigm by supporting the role of other immune cells and factors (even

non-immune) including CNS antigen-driven clonally expanded B cells, autoantibodies, complement and mediators of the innate immune responses in MS lesions. Further expansion of this global CNS dysfunction includes oligodendroglial cell (OGC) loss, attenuated remyelination,

axonopathy, and gliosis. The recognition of new "players" directing effector and regulatory functions and further insight into reparative mechanisms occurring at various stages of the disease within a given individual will influence ongoing and future therapeutic trials. The following discussion will encompass evolving concepts in the pathogenesis of MS with a focus on novel immunotherapies. These new approaches reflect targeting of a multifaceted spectrum of immune activity. The immunotherapies will be characterized by their intervening role of specific and/or multiple pathogenic steps including initiation, peripheral activation, molecular co-stimulation and immune effector responses during early, transitional and late phases of disease. Emerging strategies for the enhancement of neuroprotection and reparative mechanisms will also be reviewed. Classification of novel approaches will include the following main types of immunotherapies: (1) targeting of myelin specific T cells: antigen-specific therapies (2) targeting of B cell and autoantibody responses (3) targeting of immunologic steps of disease pathology (4) targeting of reparative stages of disease: neurotrophic and neuroprotective, (5) global therapies: broad-based polydirectional strategies.

## 2. GENERAL INTRODUCTION

### 2.1. Overview

The present review of current novel immune trials is intended to assist the reader in the understanding of evolving MS pathogenic paradigms and provide the reader with an update of cutting edge therapies in multiple sclerosis (MS), and potential future strategies toward the advancement of care of MS patients.

### 2.2. Introduction

Multiple sclerosis (MS) is a clinical neurological disease characterized by chronic inflammation, demyelination, variable axonopathy and gliosis of the central nervous system (CNS) (1-6). It is the principal (excluding trauma) neurological disease of individuals in early to middle adulthood and has been estimated to include 350,000 people in this country alone although the actual incidence including the undiagnosed may be considerably higher (7, 8). Immune mediated tissue injury in MS appears to develop in genetically susceptible individuals after exposure to a causal environmental agent that is yet undefined (9-12). The disease is clinically and histopathologically heterogeneous, with several clinical types of MS. (8, 13). Four histopathologic subtypes of MS have been described with the demonstration of both CD4+ and CD8+ cells in MS lesions (3, 4, 14-17). The majority of MS cases present as a relapsing-remitting (RRMS) condition lasting approximately five to ten years that transitions into a secondary chronic progressive state. Here patients have less frequent relapses and radiographic evidence of blood brain barrier break down but continue to develop progressive neurologic deficits. Approximately 10-20% of patients begin with a primary progressive (PPMS) course characterized by a continuous progression without acute relapses (13, 14). Recent studies suggest the role of both regulatory and effector dysregulation in MS and have raised the possibility of an intrinsic perturbation of CNS

factors and neurodegenerative mechanisms contributing to MS especially in the late chronic phases of disease. (18-20). Demonstration that both axonopathy and cerebral atrophy occur early in disease argue for early treatment which may limit long term disability (2-4, 6).

Recently reported CHAMPS and ETOMS trials have provided more direct evidence of the benefit of early initiation of therapy (21, 22). There are currently three FDA approved therapies for the treatment of RRMS and secondary progressive MS. This includes interferon-beta, glatiramer acetate and mitoxantrone. (23-26). These medications have modest effect on disease activity but do demonstrate the ability to reduce attack rate and frequency of active lesions as determined by gadolinium-enhanced magnetic resonance imaging (MRI) (24, 27). The limited efficacy, side effect profile of approved treatments for RRMS along with the failure to treat PPMS and chronic progressive phases of disease has highlighted the need for the development of new therapies to be utilized as mono or combination therapy addressing the heterogeneity of this disease (28, 29).

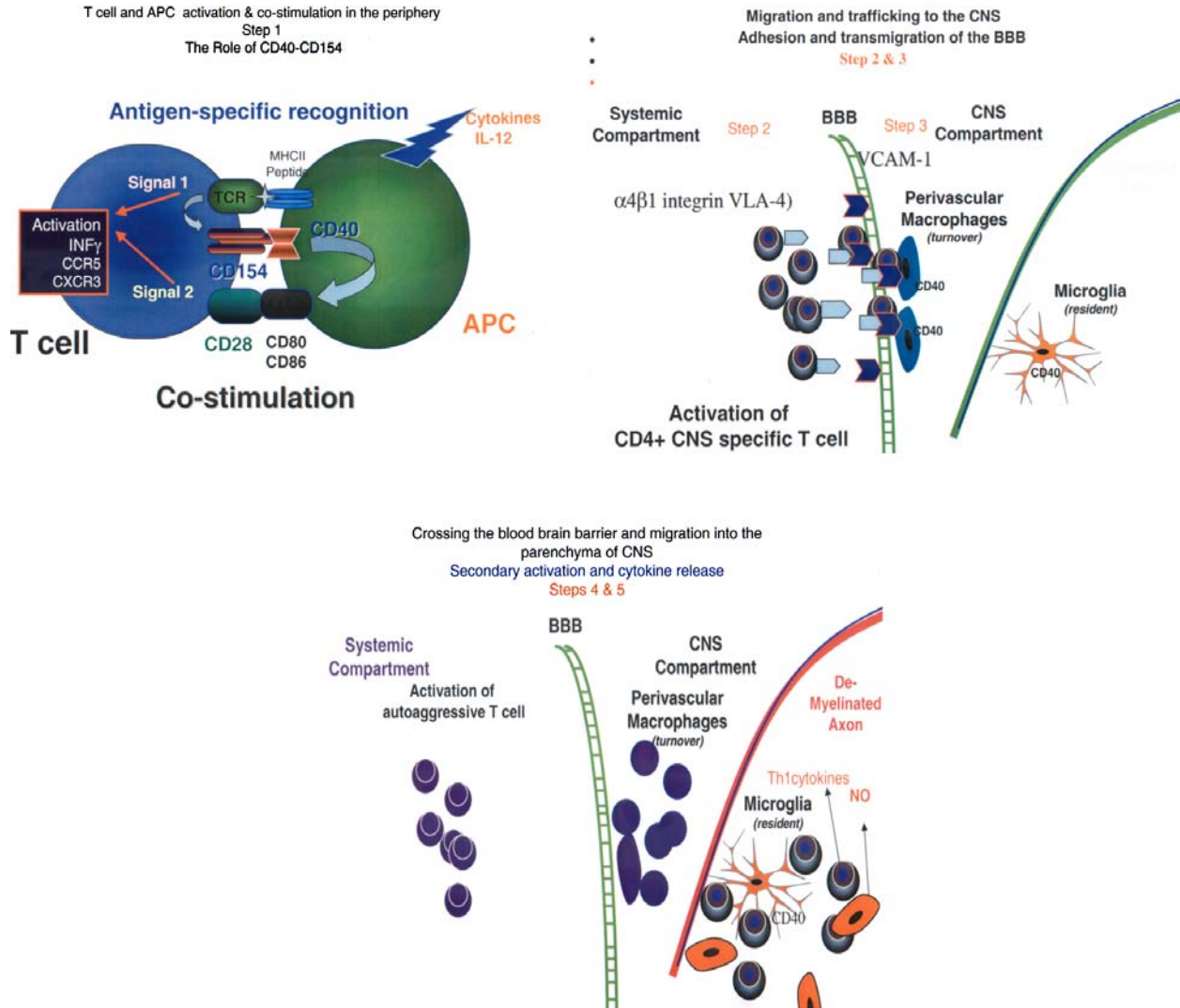
## 3. THE IMMUNOLOGICAL BASIS FOR DISEASE INTERVENTION AND RATIONALE FOR NOVEL IMMUNOTHERAPIES

### 3.1 Disease induction

MS pathogenesis involves a complex sequence of cascading immune events that culminates in chronic disease (Figure 1<sup>step 1</sup>) MS is presumed an autoimmune condition mediated at least in part by T cells (18, 30-34). The precise initiating trigger and induction of the acute phases of disease is unknown but occurs when immune tolerance is disrupted. This process may occur due to molecular mimicry whereby our immune system may attack "self" if a microbe and human share a common gene sequence encoding for a conserved structural peptide/protein(s) such as CNS immunogenic antigens such as, myelin basic protein (MBP), myelin oligodendrocyte glycoprotein (MOG), proteolipid protein (PLP) (5, 35-37). Initiation may occur from other mechanisms such as microbial superantigens, or a self protein. It is possible that the release of CNS antigens to the periphery following a CNS insult such as, the introduction of non-self proteins (CNS viral infection), or acute brain injury (trauma, stroke) may be a mechanism of initiation (19). This process may be perpetuated and expanded by "epitope spreading" that has been defined in EAE, the experimental model of human MS (38). Once initiated, neural antigens are processed by antigen-presenting cells (APC) for presentation to sensitized T cells (CD4+) with resultant activation and co-stimulation in the periphery (33, 39). B cells and CD8+ T responses primed and clonally expanded in the periphery may also be contributory to disease pathology (17, 40-43).

### 3.2. Peripheral co-stimulation

Paramount in MS immunopathogenesis is the activation, expansion, and differentiation of T cells that are dependent on a coordinated series of signals exchanged between the antigen-presenting cell (APC), the T cell, and the environment. (Figure 1<sup>step 1</sup>) The maturity of the APC is



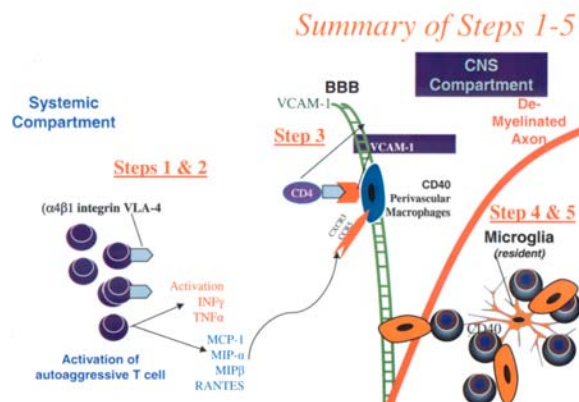
**Figure 1.** Schematic depiction of the pathogenic steps resulting in tissue injury in MS: (1-5) (1) Activation and co-stimulation of autoreactive T cells in the periphery (2) Migration and trafficking to the CNS (upregulation of homing chemokines & adhesion molecules) (3) Adhesion of proinflammatory autoreactive T cells to the endothelium, release of proteases and MMPs degrading ECM & affording transmigration of the BBB of T, B, monocytes/macrophages into the parenchyma of CNS (4) Secondary activation and amplification of the CNS intraparenchymal cellular and humoral responses with autoantibodies, induction of cytotoxic T lymphocyte (CTL) and cytokine/neurotoxin (NO) mediated CNS damage. (5) Effector stage of disease with injury to the myelin sheath.

controlled by the inflammatory milieu and expression of APC maturation factors (CD154, TNF, RANKL) by the cognate T cells. Initial recognition of peptide-MHC class II by the T cell leads to TCR stimulation delivering the "first signal" which then induces expression of CD154, which in turn binds to CD40 expressed on the APC. CD40 stimulation results in the upregulation of a series of co-stimulatory molecules, including CD80 and CD86. CD80 and CD86 engage the T-cell co-receptor CD28, thereby delivering a "second signal" that culminates in T-cell activation, proliferation, and proinflammatory cytokine production (INF $\gamma$ , IL-2, IL12, TNF $\alpha$ ). Evidence indicates that there are multiple interactions between the T cell and the APC leading to fulminant T-cell activation and expansion. Interruption of these co-stimulatory interactions (such as CD154-CD40) incapacitates APC maturation, thereby impairing the activation, proliferation, and differentiation of

antigen-specific T cells becoming potential targets of novel therapies (39, 44, 45).

### 3.3. CNS migration and trafficking

After priming, highly coordinated sequential interactions involving cellular homing (most likely dependent on preferential expression of chemokine receptors, CXCR3, CCR5) (46, 47), and trafficking into the CNS compartment proceed. Specifically a number of proinflammatory (Th1)-type cytokines are elicited that induce upregulation of adhesion molecules, and alter permeability of the blood-brain barrier (30, 48). This subsequently promotes the adhesion of proinflammatory autoreactive T cells (integrins VLA-4 also known as  $\alpha 4 \beta 1$ , LFA-1 on T cells) to the endothelium (via corresponding VCAM, ICAM receptors) allowing T, B, monocytes, macrophages to the extravasate into the



**Figure 2.** Schematic depiction of the pathogenic steps resulting in tissue injury in MS: (1-5) (1) Activation and co-stimulation of autoreactive T cells in the periphery (2) Migration and trafficking to the CNS (upregulation of homing chemokines & adhesion molecules) (3) Adhesion of proinflammatory autoreactive T cells to the endothelium, release of proteases and MMPs degrading ECM & affording transmigration of the BBB of T, B, monocytes/macrophages into the parenchyma of CNS (4) Secondary activation and amplification of the CNS intraparenchymal cellular and humoral responses with autoantibodies, induction of cytotoxic T lymphocyte (CTL) and cytokine/neurotoxin (NO) mediated CNS damage. (5) Effector stage of disease with injury to the myelin sheath, oligodendrocytes, and axons.

extracellular matrix (ECM) of the BBB (49, 50). In order to then effectively breach the BBB and traffic into the CNS compartment, activated cells must next pass through the ECM which is comprised of type IV collagen. Upon direct contact with collagen, T cells produce matrix metalloproteinases (MMPs) 2 and 9 which specifically degrade collagen type IV, as well as contribute to the proteolysis of myelin components in MS (51-53). Alpha 1 integrin on activate T cells may play a role in the initial binding to collagen type IV (54). MMPs are inhibited by tissue inhibitors of matrix metalloproteinases (TIMPs) (52). In MS brain tissue and spinal cord fluid patients MMP-2, MMP-7, MMP-9, MMP-12 and TIMP-1 have been reported (52, 55, 56). Adhesive molecules, chemokines and MMPs therefore represent another potential site of targeted intervention (Figure 1<sup>step 2&3</sup>).

### 3.4. Effector mechanisms in the CNS compartment

Immune cells including B cells and sensitized T cells (CD4+ helper and CD8+ cytotoxic) that successfully traffic into the CNS are reactivated by antigen-presenting macrophages and resident microglial cells with a subsequent amplification of CNS cellular, humoral and complement effector responses leading to the production of antibodies, complement, toxic cytokines, apoptosis-mediating molecules, and release of other neurotoxic mediators such as the oxygen and nitrogen free radicals, glutamate and osteopontin (57-59). This complex and inter-related process reflects both the innate and acquired immune systems which target and remove the antigenic source from the CNS tissue (19). For instance, nitric oxide (NO) secreted by activated microglia is a potent mediator of oligodendroglial cell loss. Expression of induced nitric oxide synthetase (iNOS) catalyzes NO and transcription

of iNOS is upregulated by macrophage/ microglial elaborated (TNF $\alpha$ ) and T cell elaborated (IFN $\gamma$ ). In the setting of this proinflammatory milieu, NO and TNF $\alpha$  along with autoantibodies and complement produce demyelination. Macrophages and microglia not only present antigen but in this setting phagocytose myelin debris, and capture antibody-antigen complexes by their Fc receptors. Th1 cytokines (INF $\gamma$ , IL-12) are further perpetuated by macrophage/T cell elaborated osteopontin while downregulating Th2 cytokines (53). This concerted attack of varied immune cells and their relative effector mechanisms results in damage to oligodendrocytes, myelin sheaths and even invariable degrees of axons and neurons as well as, cerebral atrophy during early acute inflammatory phases of disease. (Figure 1<sup>step 3&4</sup>) If these effector mechanisms are of significant severity or persistence resultant irreversible axonopathy and neuronopathy may occur (60). The contribution of axonopathy appears to correlate to irreversible disability (2, 6, 61). Clinically this stage is reflected early by exacerbations during the relapsing of disease which may be antagonized by immunoregulatory and reparative mechanisms (20, 62-70).

### 3.5. Protective and reparative mechanisms operative in MS

Neuroprotective and homeostatic regulatory processes appear concurrent with the immune attack down-regulating inflammation such as upregulation of regulatory cell populations. Even proinflammatory mediators such as TNF $\alpha$  may be involved in a dual role of tissue repair (71). Oligodendrocyte progenitor cells (OCPs) are recruited to areas of demyelination, expand and differentiate into remyelinating cells that repair local tissue damage (72, 73).

Remyelination appears most efficient in the early inflammatory phases of disease and within acute MS lesions. In chronic lesions and progressive phases of disease oligodendroglial cells may still be observed but with little remyelination present (74, 75).

The debate regarding the cause of inadequate repair is ongoing and may be due to a neurodegenerative processes such as intrinsic abnormalities in recruitment and/ or differentiation of OCPs or possibly secondary to chronic severe demyelination (74, 75). Likewise axonal damage may be a consequence of direct effector mechanisms of the immune attack (macrophages, CD8 cytotoxic cells) (15, 17), or may arise secondarily to chronic severe demyelination and membrane destabilization. This may lead to increased Ca<sup>2+</sup> influx and disrupted axonal transport, and/or primary axonal dysfunction (76). Enhancement of neuroprotective and regenerative factors have become attractive targets of novel therapeutic strategies (62-65, 67, 77).

## 4. EMERGING NOVEL THERAPEUTICS

(Figure 2, Tables 1).

### 4.1. Immunotherapies targeting myelin specific T cells: antigen-specific therapies

Induction of tolerance by targeting immunoregulation and the modification of autoreactive T cells or antigenic epitopes is currently being explored.

## Novel Immunotherapies in Multiple Sclerosis

**Table 1**

Target BBB	Agent	Human Study	Animal Study	I.	II 1.	II 2.	II 3.	II 4.	II 5.	III	IV
	Anti- $\alpha_4$ integrin (VLA-4) mAb-natalizumab Antegren	Phase III	x			x					
	*MMPs inhibitors		x			x					
	*TIMPs		x								
	Chemokine blockade		x		x	x					
MHC-Class II	Anti-MHC mAbs		x	x							
	iNOS inhibitor		x						x		
	Hypervariable Peptide vaccines		x	x							
Antigen	Altered peptide ligands	Phase I/II topped	x	x							
	Myelin specific T cell Immunization MBP298	Multiple		x							
	DNA encoding Autoantigen vaccine			x	x						
TCR	TCR V $\beta$ immuni-zation/NeuroVax	Multiple		x							
	TCR peptides		x								
Co-Stim Ulatory Ligands	Anti-CD154 mAb IDEC-131	Phase I/II DB, PC	x		x						
	Anti-IL12 (p40) mAb CNTO 1275	PhaseI/II DB, PC	x		x						
	Anti-IL2 $\alpha$ mAb Daclizumab	Phase I/II OL, CO, R	x		x						
	Anti-CTLA4 mAb	Phase II DB, PC			x						
	Recombinant IL-1 receptor Antagonist Anakinra	Phase I/II DB, PC	x				x				
	IL-10		x		x						
PPAR $\gamma$ receptor	Agonist Rosiglitazone maleate Avandia	PhaseII DB, PC	x								x
Cell Recruitment	Anti-cytokines Chemokines (i.e MIP-1)		x		x	x					
Macrophage	Inhibitors of exocytotoxin Products (free radicals, PAF,		x						x		
Astrocyte	Cytokines TNF $\alpha$ inhibitors		x				x	x			
	Treosulfan Apoptosis APC and T cell		x					x			
	Protein kinase inhibitors		x								
	Caspase signaling inhibitors		x					x			
CD8 cell	Chemotherapy Anti-CD8+ mAbs		x					x			
B cell	Chemotherapy Anti-CD20+ mAbs/Rituxamab	Phase II/III	x					x			
Complement	Anticomplement mAbs		x					x			
	Inhibitors of complement MAC Membrane attack Complex interactions		x					x			
Transplantation	Autologous bone marrow/peripheral stem cell	Multiple OL	x								
	Oligodendrocytes (fetal, adult)		x							x	
	Schwann cells		x							x	
Hormone	Estrilol	Phase I/II	x								x
	AndroGel Testosterone	Phase II CO	x								x
Regeneration	Riluzole Inhibitor of glutamate receptor (AMPA/kainite)	OL	x						x	x	
	iNOS2 inhibitor in glial cells/Ansamysins		x						x		
Neuroprotection	Ginko Biloba	Phase I								x	
	A-lipoic acid, VitamenE/Selenium	Phase II								x	
	Inosine/precursor of Antioxidant uric acid	Phase II								x	
Neurotrophic Growth factors	CNTF, NGF, IGF-1	IGF-1 PhaseI/II								x	

Table modified from the National Sclerosis Society at <http://www.nationalmssociety.org/pdf/research/clinicaltrials.pdf> Subtypes of immune therapy; (I) Antigen Specific Therapy, (II) Directed Immunotherapy see figure 1 and 2, [1. Activation and co-stimulation of autoreactive T cells in the periphery 2. Migration and trafficking to the CNS 3. Adhesion of proinflammatory cells to the endothelium 4.Secondary activation and amplification of the CNS ] (III) Repair Therapy, (IV) Global-polydirectional Therapy DB, double blinded; OL, open labeled; PC, placebo controlled, CO, crossover

Antigen-specific therapies target the autoreactive myelin-specific T cell, a major player of MS pathology. The therapeutic goal is to enhance homeostasis and restore immune tolerance. Suppression of clonally expanded effector or helper T-cells and/or enhancement of regulatory

populations are potential therapeutic strategies. These strategies include (1) induction of T cell anergy (2) activation induced T cell apoptosis whereby an activated T cell upon exposure to an antigen undergoes cell death and deletion (3) induction of bystander suppression whereby an

activated T cell upon exposure to the same or modified version of the autoantigen induces T cells with immunoregulatory function.

To accomplish the above goal of immune tolerance several novel therapies for the reinduction of peripheral tolerance through immunization of the putative pathogenic T cell, TCR receptor, and autoantigens have been explored in animal and human studies. Efficacy of T-cell immunizations may be less beneficial than those reported in animal models due to the diversity of potential antigenic epitopes, pathogenic T cells and TCR repertoire in the outbred human population. Additionally, intact immunoregulatory mechanisms present in experimental allergic encephalomyelitis (EAE) an animal model for MS may be dysregulated in the MS population. The challenge is to identify the "key" autoantigens or encephalopathic T cell populations and TCR repertoire that contribute to disease and maybe overcome to provide a safe and viable therapy in MS. The following are few examples of these currently pursued approaches.

### 4.1.1. Immunoregulatory mechanisms

Induction of regulatory cell populations such as CD4<sup>+</sup>CD25<sup>+</sup>CD45RB<sup>lo</sup> and invariant Cd1 NKT cells have been shown to restore peripheral tolerance and attenuate in EAE (20, 78-80). These regulatory cells may delete and/or suppress pathogenic T cells by a variety of mechanisms including the induction of anergy or a shift in polarity from a Th1 to a Th2 cytokine milieu. Analogous human regulatory populations have been identified and appear to play a critical role in balancing the need for autoimmunity for protection and risk for non-controlled autoimmunity offering potential future strategies (69, 81, 82). Although the specific neural peptides involved in the induction of these regulatory cells have not yet been characterized, this would represent a novel approach for the induction of antigen-specific regulatory response in active MS.

### 4.1.2. T-Cell and T-Cell receptor immunization

One novel strategy has utilized immunization with putative pathogenic T cells (MBP-specific T cells) reporting effectiveness in MBP induced EAE model (83). In EAE and several small pilot T cell vaccination trials in MS short term depletion of T cells reactive against different myelin antigens was demonstrated. Peripheral tolerance appeared to be mediated through deletion of myelin specific cells by CD8<sup>+</sup> lysis in a MHC Class I restricted manner, or by CD4<sup>+</sup> lysis MHC Class II restricted cytolytic activity (84-88). A more recent phase II open label trial of MBP-specific T cell immunization of RRMS and SPMS demonstrated depletion of MBP specific T cells following vaccination (43). Another T cell mediated approach is to utilize T cell receptor V $\beta$  (TCR) peptides derived from the complementarity-determining region 2 and 3 (CDR2) and (CDR3) of autoreactive T cells. These T cells have been shown to ameliorate disease in EAE models (89, 90). MS patients may have an overrepresentation of TCR V $\beta$  subsets. Early pilot TCR V $\beta$  peptide vaccination trials in MS patients reported safety, induction of Th2 IL-10 cytokine and some depletion of the complimentary targeted T cells (91). A later phase II trial did not show a clinically

significant reduction gadolinium-enhanced MRI lesion, their primary outcome (92). These modalities require larger, randomized, double blind, controls studies to demonstrate clinical efficacy in MS patients.

### 4.1.3. Modification and targeting of antigenic epitopes

Other antigen-specific therapies underway have attempted to downregulate autoreactive T cells by the development of auto-antigenic peptides designed to resemble the antigenic epitopes such as, altered peptide ligands (APL), new modified copolymers, and vaccination with DNA-encoding auto-antigen. Altered peptide ligand (APL) is an auto-antigenic peptide with modifications (amino acid substitution) in TCR contact positions. Promising animal studies demonstrated APL could alter pathogenic T cell responses to native peptide by T cell anergy, TCR antagonism, partial agonism, or bystander suppression (93-95). An APL of the human immunodominant MBP (83-99) peptide was utilized in two separate phase II clinical trials in MS but stopped due to adverse events (96, 97). These preliminary trials however highlighted important immunologic findings. APL induce hypersensitivity reactions in several patients (97) induced APL-specific Th1 cells (98). Recently, a study confirmed that myelin antigens induce classic anaphylactic responses but can be easily treated with antihistamine prophylaxis (99). This trial although clinically disappointing linked disease exacerbation with expansion of activated MBP (83-99). This gave support that disease induction and tissue injury in MS is driven by myelin specific CD4<sup>+</sup> Th1 cell population against CNS myelin components. Interestingly lower doses of APL showed a trend towards clinical benefit associated with a reduction of inflammation demonstrated by MRI and a polarity skewing of APL-specific cells with a Th2 immunoregulatory phenotype, suggesting APL bystander suppression (96, 100). To establish optimal dose, timing and administration for safe and efficacious therapy larger, randomized, double blind, controls studies are planned to address these issues.

Glatiramer acetate (GA) (Copaxone®, Teva Industries) and FDA approved drug appears to employ several mechanisms including bystander suppression with the induction of GA-specific T cells that cross react with native MBP autoantigen (101). New copolymers and peptides are presently in investigational development to find molecules with optimal benefit. One recent report has shown that a four amino acid copolymer based on MBP (85-99) but distinct from GA (aa size and content) had greater affinity for the HLA-Dr2- restricted T cell clones and more effectively attenuated EAE (102).

### 4.1.4. Modification of gene DNA encoding autoantigen

Another novel antigen-specific approach being explored is DNA vaccination. This approach exploits recent advances in gene therapy with the goal of preventing the generation of encephalogenic T cells (103-106). A recent study of DNA vaccine under development (106) involves covaccination with DNA encoded myelin autoantigen(s) alone or given with DNA encoded immunoregulatory cytokines such as IL-4 in EAE. This model of proteolipid

protein (PLP) induced EAE was attenuated after the covaccination with IL-4 DNA and naked DNA encoding (PLP139-151). PLP-specific T cells demonstrated a polarity skewing towards a Th2 phenotype (106) DNA vaccination in the presence of statins may also be explored. Although preclinical methods offer potential site-directed efficacy further investigation are essential to establish safety and efficacy in humans.

### 4.2. Targeting of B cells

Autoreactive B cells and humoral responses appear to contribute to EAE and MS pathology. The functions of these B cells is not yet fully understood and the functions may be quite diverse (107). Autoreactive B cells can produce antibody that can directly mediate effector mechanisms responsible for some of the pathologies associated with autoimmune disease. Immune complex deposition in the kidney and the resulting tissue damage is a hallmark of systemic lupus erythematosus (SLE). Antibody-mediated demyelination in the CNS in primates is also characteristic of MS and MOG induced EAE induced in rats, mice and marmosets (5, 108-110). Secretion of effector antibodies may provoke direct CNS damage or may also play less obvious roles in redirecting T cell activities. This may be through mechanisms which generate inflammatory mediators and chemokines that alter the recruitment of T cells into an inflammatory site, as has been shown in a model of oophoritis. In addition, antibodies may facilitate the cross-presentation of antigens via immune complex binding to FcR on dendritic cells and facilitate the generation of self-reactive CD8<sup>+</sup> T cells. Beyond the pathological effects of autoantibodies, B cells themselves can also influence the development of autoimmunity through their antigen presentation functions and regulatory capacities. B cells, through their membrane immunoglobulin can capture autoantigens and present these antigens to self-reactive T cells. Under such circumstances B cells can also function in the context of antigen uptake, processing and presentation resulting in the expansion of self-reactive CD4<sup>+</sup> T cells. In model systems where B cells cannot secrete autoantibody but can express autoreactive surface immunoglobulin, these B cells have been shown to be able to mediate a systemic autoimmune syndrome, clearly showing that the secretion of autoantibody is not critical for autoimmunity to develop. In summary increasing evidence supports the role of B cell and humoral responses in MS (3, 12, 42, 111-116).

#### 4.2.1. Anti-human CD20 mab (Rituximab) B cell deletion

Rituximab is a chimeric anti-human CD20 mab used in the treatment of B cell lymphoma that maybe of potential importance in the treatment of both RRMS and primary progressive disease. The anti-CD20 monoclonal antibody (mAb) is a genetically engineered chimeric murine human monoclonal antibody that depletes circulating and tissue based B-cells. CD20 Rituximab has been granted FDA approval for relapsed or refractory B cell lymphoma with delineated safety and toxicity guidelines. Rituximab (anti-CD20) has additionally been administered in a number of B cell mediated autoimmune diseases such as rheumatoid arthritis (RA) (117-122), cold

agglutinin, disease (123, 124), warm antibody hemolytic anemia (125), idiopathic thrombocytopenic purpura (126), paraproteinemic polyneuropathy, (127) and myasthenia gravis (128). Studies in RA suggest that the immunologic benefit of rituximab is related more closely to decreases in circulating autoantibodies than to B-cells (121). Like MS, both B and T cell activation and proliferation may mediate the pathogenesis of progressive RA, and several trials support the targeting of B-cell immunomodulation in autoimmune diseases such as MS (129). Both a Phase II randomized trial in RRMS as well as a Phase II/III randomized, multicenter, double blind, parallel group, placebo controlled study of Rituximab are currently under development.

### 4.3. Immunotherapies targeting directed immunologic steps of disease pathology

#### 4.3.1. Targeting peripheral co-stimulatory and proinflammatory cytokines

##### 4.3.1.1. CD40L-CD40 co-stimulatory blockade

Treatment with a humanized monoclonal antibody CD154 is a potential new immunomodulatory treatment for RRMS. Preclinical studies have suggested the importance of CD40L-CD40 costimulatory interaction in the pathogenesis of MS. In specific the engagement of CD40 and it's ligand CD154 are critical in eliciting the activation of T cells and cell-mediated immunity (CMI) responses (130-133). Blockade of CD154/CD40 interactions prevents the development of CMI and a variety of autoimmune disease models in mice. Importantly it has been shown to prevent the progression of both monophasic and relapsing remitting EAE models (39). Animal studies also support that CD40-CD154 interactions are ongoing in human MS plaques (132, 134). It is hypothesized that the effects of CD154 blockade on CMI are due to a central impairment of APC maturation (130, 132, 135, 136). It is clear that CD40 signaling is critical for the "maturation" of APC via the induction of a wide spectrum of APC activities and that blockade in this pathway may lead to T-cell tolerance or T-cell skewing. A phase I clinical trial of the treatment of RRMS with anti-CD154 showed no evidence of either systemic or neurological toxicity, which were the primary outcomes for the study. Although the number of subjects was insufficient to draw significance, several of the secondary outcomes including MRI and EDSS suggest that therapy may stabilize the disease process. These results provide a rationale for a larger scale study and a phase II masked, placebo-controlled, randomized partial crossover study in 46 subjects with relapsing-remitting multiple sclerosis (RRMS) is underway to test safety and efficacy (Figure 2, Table 1).

##### 4.3.1.2. Interleukin - 12p40 (IL-12p40) blockade

Another novel therapeutic approach involves the inhibition of a proinflammatory IL-12, a heterodimeric predominant cytokine in immune mediated inflammatory disorders (137-140). Accumulating evidence indicates that IL12 plays a pivotal role in the pathogenesis of EAE mediating both cellular (Th1) and humoral responses. (141). IL-12 antagonists and neutralization of IL-12 can prevent EAE in both rodent and marmoset models, as well as, being implicated in the pathogenesis of MS (142-150).



The IL-12p40 subunit appears most important and other monokines such as IL-23 that express this subunit may also participate in the development of EAE (151). Local expression of IL-12 within the CNS has been demonstrated in EAE animal models (149, 152). Furthermore local expression of IL-12 within the CNS of MS patients as well as increased levels of IL-12 in CSF, plasma, serum and PBMCs during active disease has been demonstrated (153-156). A Phase I/II clinical trial for the treatment of psoriasis another Th1 mediated disorder with a humanized monoclonal anti-IL-12p40 antibody evidenced statistical clinical benefit. These studies provided theoretical evidence that therapy directed against IL-12p40 may be another new effective MS treatment. A multi-center Phase I double blind, placebo-controlled trial in patients with relapsing forms of MS, evaluating the safety of a single administration of monoclonal antibody to IL-12p40 (CNT01275) is presently in progress. A phase II trial is planned later this year to examine drug efficacy in RRMS patients (Figure 2, Table 1).

### 4.3.1.3. IL-2/IL-2 receptor $\alpha$ blockade

Daclizumab (Zenapax) is a humanized monoclonal antibody against the IL-2 receptor alpha subunit. IL-2 is a autocrine growth factor necessary for T cell growth. The rationale for this novel therapy is that the interaction of IL-2 and its receptor IL-2 on the surface on T cells leads to the activation and expansion of autoreactive T cells and blockade may result in the downregulation of encephalogenic T cells. This agent has been FDA approved in renal transplant recipients to induce immunosuppression for optimization of graft acceptance (157) patients who had responded incompletely to interferon beta therapy was pursued.

They reported during the past three years the tolerability and safety of monthly administration of daclizumab, as well as a reduction of disease activity by MRI imaging. A further amendment of this study demonstrated persistent efficacy of daclizumab after the interferon beta therapy was discontinued. A current Phase II (ZAP) open-label, baseline to treatment cross over trial is testing the efficacy of daclizumab alone in RRMS patients.

### 4.3.1.4. Targeting B7-CD28-CTLA-4 co-stimulatory pathway with CTLA-4Ig agonist

Cytotoxic T lymphocyte-associated antigen 4 Ig [CTLA4Ig] is a soluble chimeric protein. Costimulatory blockade using CTLA-4Ig has recently been explored as a novel therapeutic in human studies. A phase I clinical trial of the treatment of psoriasis vulgaris, with CTLA-4Ig improved clinical outcomes and was associated with reduced cellular activation of lesional T cells, keratinocytes, dendritic cells (DCs), and vascular endothelium (158). Another pilot clinical trial of the treatment rheumatoid arthritis evaluating CTLA-4Ig also demonstrated safety, tolerance and dose-dependent effectiveness (159). A current phase I clinical trial is testing the safety of CTLA4Ig (BMS-188667) and CTLA4Ig (Repligen-RG2077) in MS patients.

### 4.3.2. Targeting adhesion molecules and cell trafficking across the blood brain barrier (BBB)

Cellular homing, adhesion and transmigration of a number of activated immune cells including autoreactive

encephalopathic T cells across the BBB into the CNS compartment is an early and critical step involved in the immune pathology of MS. This is supported by *in vitro* and *in vivo* animal and human studies (160). Targeting these steps are integral in a number of current and novel MS therapies. The following are a few examples of these novel strategies.

### 4.3.2.1. Inhibition of $\alpha 4 \beta 1$ integrin-VCAM-1 mediated adhesion

Natalizumab (Antegren) is a humanized monoclonal antibody against the  $\alpha 4$  chain of the  $\alpha 4 \beta 1$  integrin (VLA 4) expressed on the surface of activated lymphocytes and monocytes.  $\alpha 4 \beta 1$  integrin (VLA 4) and its associated receptor VCAM-1 on endothelial cells are important prerequisites of adhesion, transendothelial migration and enhanced cellular activation within inflamed tissue. In an EAE model natalizumab evidenced inhibition of T cells trafficking across the BBB with subsequent amelioration of disease (160). In a Phase II clinical trial, two doses of natalizumab were administered over a 8 week course. The results of this novel study showed a reduction of gadolinium-enhancing MRI lesions at 12 weeks but not at the follow-up second 12 weeks which were the primary outcomes for the study. Secondary outcome measures showed a significant increase in relapse rate in the follow up period (161). This suggested a rebound effect with drug cessation and the possible need for long term drug therapy. (161). In a multicenter, randomized, double-blind, placebo controlled phase II trial, 213 patients with RRMS or relapsing SPMS were assigned 3mg/kg, 6mg/kg or placebo every 28 days for 6 months. This extended both treatment and follow-up period (162). In both treatment arms a 90% reduction in the number of new gadolinium lesions, the primary outcome. A secondary outcome showed a significant reduction of clinical relapses compared to placebo but was not powered for clinical outcomes (162). Patients returned to pre-treatment relapse rate following discontinuation of therapy. Currently two larger multicenter Phase III trials are in progress in RRMS. One trial is a monotherapy trial of 1200 patients (AFFIRM) and the other is a combined trial of 900 patients with interferon beta-1a (Avonex) (SENTINAL). A smaller phase III trial of 110 patients has combined natalizumab with glatiramer acetate. Another Phase III trial utilizing small-molecule antagonists has been initiated. These agents may offer advantages over monoclonal antibodies such as oral availability and lack antigenicity in the future. (163).

### 4.3.2.2. Inhibition of matrix metalloproteinases (MMPs): treatment of minocycline

Preclinical evidence supports the pathologic role in MS for several of 23 MMP family members. In normal CNS low to non detectable levels of MMPs are found whereas upregulation and high levels are found in several neurologic diseases including MS (55, 56, 164, 165). A recent study extended the identified members of the MMP family in MS and showed a distinctive pattern of expression in T, B, and monocytes (166). Several EAE animal studies support the promotion of disease activity and course (52, 165). Synthetic inhibitors of MMPs (TIMP-1 like) have been shown to ameliorate or prevent



EAE and genetically deficient MMP-9 mice are more resistant to EAE induction compared with wild-types (167, 168). A pivotal role of MMPs in MS and EAE is the facilitation of leukocyte transmigration across the BBB, reversed by inhibitors of MMP activity (169-171). However many other effector functions of MMPs may contribute to MS pathology including breakdown of the BBB, promotion of CNS inflammation, and direct neurotoxicity (172-174). Based on the above data, identification and targeting of MMP members is a promising area of future treatments. Interferon beta IFN- $\beta$  has been shown to inhibit the activity of MMPs. Minocycline, an antibiotic has been shown to attenuate EAE (175) and inhibit MMP activity and production (MMP-9). A phase II open label cross over trial of minocycline in RRMS is underway.

### 4.3.3. Targeting secondary activation and amplification within the CNS

As previously discussed immune cells including B cells and sensitized T cells (CD4<sup>+</sup> helper and CD8<sup>+</sup> cytotoxic) that successfully traffic into the CNS are reactivated by interactions with resident APCs with the subsequent amplification of CNS cellular and humoral effector responses. These complex molecular interactions in the CNS may lead to inflammation, demyelination, oligodendrocyte loss and axonal and neuronal dysfunction. Several animal and human studies are currently exploring this class of immunomodulators and immune independent neurodegenerative mechanisms operative in the CNS compartment which may provide effective future MS therapeutic interventions. The following are a few examples of preliminary animal studies elucidating data that support pivotal roles of several novel CNS targets but which require further research to determine therapeutic possibilities in MS patients.

#### 4.3.3.1. Targeting of a brain proinflammatory mediator: osteopontin

In recent animal studies performing genetic sequencing of MS brain libraries revealed a role of a pleiotrophic brain binding protein osteopontin produced by glial cells may have a role in MS progression (59). In transgenic mice deficient for osteopontin (OPN) the progression of EAE was inhibited and severity of disease was reduced. (59, 169, 176, 177). Furthermore, inhibition of CD44, a ligand of OPN prevented EAE and elevated a non-inflammatory Th2 cytokine IL-10 (169). This study suggests that the proinflammatory effect of OPN may be mediated by CD44 and provide a new CNS target. (169). Genetic osteopontin polymorphisms appear to correlate with disease course in MS. (178, 179). In addition, elevated osteopontin levels have recently been shown to be associated with disease activity in relapsing remitting MS patients (10, 180) supporting their future role in the arsenal of novel strategies.

#### 4.3.3.2. Suppressive effects of ansamycins on inducible nitric oxide synthetase 2 (iNOS2)

Another central target of immune effector mechanisms in the CNS is the suppression of inducible nitric oxide synthetase (181). The inducible form of nitric

oxide synthetase (iNOS2) by brain glial cells is thought to contribute to the production of neurotoxic mediators and expression of proinflammatory cytokines such as diseases as Alzheimer and MS (181). Reduction of iNOS2 by a heat shock response (HSR) attenuated the histologic and clinical symptoms of EAE (182). Recently ansamycins, a class of antibiotics demonstrated a suppressive effect on iNOS2 exerting a potent anti-inflammatory response on brain glial cells in EAE (181). Other novel nitric oxide scavengers such as (NOX-100) have been shown to reduce the severity or ameliorate EAE progression in mice (183).

#### 4.3.3.3. Targeting of intracellular cyclic amino monophosphate (cAMP) in the CNS

Human and animal studies examining phosphodiesterases (PDE) which critical enzymes expressed in the immune system and brain have been shown to be responsible for the degradation of cAMP and/or cGMP (184). PDE enzymes exist as 11 distinct families (135, 185, 186). Recent data has shown that inflammatory cells predominantly express PDE4 followed by PDE3 and, to a lesser extent, PDE7 with isoforms of PDE4 being preferentially expressed in the brain (184). Inhibitors of cAMP-specific PDE4 have been shown to inhibit T cell proliferation, proinflammatory mediator release with modulation of T cell-polarization (Th1 to Th2 skewing) contributing to the cytokine milieu and influencing the upregulation of distinct costimulatory signals. They recently have been shown to alter DC capacity and cytokine production (187-190). PDE4-specific inhibitors, such as Rolipram, Cilomilast and Mesopram, (191), have been demonstrated to elevate cAMP levels, and inhibit proliferation, cytokine production and mediator release of several cells, including T cells, monocytes and eosinophils (96, 192). Other animal studies have shown PDE4 inhibitors demonstrate the ability to abrogate acute and chronic-relapsing EAE in mice, rats and marmosets (188, 193, 194). Human *in vitro* studies have demonstrated similar immunomodulating effects of PDE4 inhibitors with a preferential inhibition of Th1 type cytokines elaborated by autoreactive T cells in MS patients versus healthy controls (188, 193-195). These studies lead to the support of PDE inhibitors as candidate therapies for T<sub>H</sub>1-mediated human diseases such as MS. Presently a phase II, open label, crossover trial with the treatment of Rolipram (PDE4 inhibitor) for RRMS and SPMS patients is in progress. The trial will use MRI measures as the primary outcome and the treatment phase will be for 8 months.

Salbutamol also inhibits cAMP albeit by distinct mechanisms versus PDE inhibitors and demonstrated disease suppression in EAE models (196). The administration of oral salbutamol evidence a shift in T cell polarity toward a Th2 phenotype in peripheral blood monocyte cells (PBMCs) of MS patients (196, 197). Presently phase II trials of this agent are underway to examine drug efficacy in MS patients.

### 4.4. Immunotherapies targeting reparative stages of disease

Recent evidence supports that axonal and neuronal degeneration occur as the disease progresses (2-6,

60). These MS lesions predominantly involve the white matter however, recently demyelination and neuronal pathology have also been demonstrated in the (gray matter) cerebral cortex of MS patients. Novel approaches inhibiting brain neurotoxicity and/or promotion of repair and recovery affording ODG, axonal and neuronal protection represent a rapidly evolving area of research potentially offering protective and beneficial treatment for MS patients at various stages of disease. Interestingly, even immune cells associated with inflammatory responses in the CNS can produce a variety of neurotrophic factors of different molecular families supporting their potential for not only detrimental but beneficial effects in MS. Indeed some of the most potent members of the neurotrophin family such as nerve growth factor, (NGF), brain-derived neurotrophic factor (BDNF) act on or are endogenously produced by immune cells in MS lesions (62-65, 77). Resident CNS cells appear also to have immuno-regulatory properties (66). The following are only a few examples of the investigations underway exploring the use of neuroprotective-reparative agents.

### 4.4.1. Neurotrophic factor

Neurotrophic factors are proteins that direct differentiation and survival/apoptosis acting through specific neurotrophin receptors (65, 77, 198, 199) that have been shown to shift Th1-Th2 and may promote neuroprotection (65, 200). Potential neurotrophic therapeutic candidates include brain-derived neurotrophic factor (BDNF), glial growth factor, ciliary neurotrophic factor (CNTF), neuroimmunophilin ligand (FK506) and NGF (77, 201-203). CNTF elaborated by activated astrocytes induced growth and trophic factors such as FGF-1 and IGF-1 which indirectly protected neurons from cell death and promoted oligodendrocyte generation. CNTF was also shown to inhibit neuronal and glial degeneration resulting from microglial cytotoxins (202). A phase I/II trial examining insulin-like growth factor is currently being explored. There is evidence from other neurodegenerative systems to support the use of gene delivery of growth factors for the promotion of remyelination (204).

### 4.4.2. Inhibition of the glutamate receptor $\alpha$ -amino-3-5-methyl-4-isoxazolepropionic acid/kainite (AMPA/kainite) receptor

Preliminary studies suggest the glutamate neurotoxicity is an important contributing factor in MS pathogenesis (69, 70, 205). In EAE and MS lymphocytes, brain microglia and macrophages release excessive levels of glutamate which activate AMPA ( $\alpha$ -amino-3-5methyl-4-isoxazolepropionic acid)/kainate receptors on oligodendrocytes (OGCs) and neurons. OGCs are especially vulnerable to glutamate-AMPA/kainate excitotoxicity (206, 207). Blockade with AMPA/kainate antagonists have been shown to ameliorate EAE (58, 76, 205). AMPA/kainate antagonists also appear to protect OGCs and axons from immune-mediated damage. A recent study investigated riluzole affect in MOG-induced EAE a chronic model and demonstrated this agent attenuated the clinical severity of disease and reduced inflammation, demyelination and axonal damage in the CNS (205) of MS patients versus controls. MS patients also demonstrated

increased CSF glutamate levels correlating with disease severity (70). An anti-glutamatergic agent riluzole (2-amino-6-trifluoromethoxy benzothiazole) found to be protective in several models of neurodegenerative disease including ALS (208), Parkinson's (209) and ischemia (210). Based on riluzole's neuroprotective properties, safety and tolerability the FDA approved this agent for the treatment of ALS (211). Riluzole is presently being explored as a potential neuroprotector in MS. An open label clinical trial in primary progressive MS patients to test the neuroprotection of riluzole (Rilutek®) is currently in progress.

### 4.4.3. Targeting of neurotoxic and nitrogen free radical mediators

Other studies suggest a role of oxygen and nitrogen free radicals in the immunopathogenesis of EAE and MS. In animal models of MS, these chemical reactions have been associated with break-down of the BBB and CNS tissue injury. Additionally, increased levels of iNOS have been evidenced in active demyelinating lesions, as well as, showing increased CNS and CSF levels of reactive nitrogen oxide species (RNOS) in MS patients versus matched controls (164, 212). Uric acid (UA) is a RNOS scavenger and natural inhibitor of chemical interactions associated with peroxynitrite (213). In several mouse EAE models uric acid administration attenuated disease severity and was associated with alterations in BBB permeability, inhibition of CNS inflammation, and tissue injury (213-215). The above data and observations that MS patients have serum uric acid levels that are lower than age and sex matched healthy controls (216-218). This data provided the rationale for a novel treatment of MS aimed at raising levels of the natural antioxidant UA or its precursor inosine (219).

A small preliminary clinical trial of oral administration of inosine in 11 MS patients showed clinical stability in 9 patients and improvement in 3 patients. Further in two patients who had notable pretreatment gadolinium-enhanced lesions none were detectable following inosine treatment. Currently a phase II, double blind, placebo controlled trial is underway to determine whether oral treatment with inosine has an effect on cumulative number of newly active lesions on MRI and to evaluate safety and tolerability of inosine in 30 RRMS and SPMS patients. Upcoming phase I/II trials examining three other natural antioxidants (ginkgo biloba,  $\alpha$ -lipoic/essential fatty acids and selenium) for MS treatment is planned and sponsored by National Center for Complimentary and Alternative Medicine (NCCAM).

### 4.5. Global Therapies: broad based polygenic mechanisms

The following class of novel therapies have diverse properties but share the characteristics of exhibiting polygenic mechanisms and broad based immunomodulatory targets. Below are a few examples of this category of agents.

#### 4.5.1. Depletion of multiple immune cellular subsets: treatment with anti-CD52, alemtuzumab

Alemtuzumab (Campath) directed at CD52 appears to have polygenic mechanisms including the

depletion of leukocytes (T & B) as well as, monocytes and macrophages. This agent was initially approved by the FDA (2001) to treat patients with B-cell chronic lymphocytic leukemia and revealed immune suppressing actions. A phase II clinical trial in SPMS demonstrated that alemtuzumab had pronounced effects the immune system reducing relapse rate and brain inflammation as shown by serial MRI. Specifically, during the 18 month follow phase gadolinium enhanced lesions were significantly reduced (220). Safety concerns were raised however, due to the observation that about 30% of patients in these early studies developed Grave's autoimmune thyroiditis (221). Currently there are two clinical studies extending the investigation of Campath® affect on immune function including a retrospective study of 58 RRMS and SPMS patients and an open labeled multicentered clinical trial of Campath®/MABCampath® versus Rebif (interferon-beta-1a) in early active RRMS.

### 4.5.2. Statins

A class of orally administered cholesterol (lipid) lowering drugs, the statins or (HMG-CoA) reductase inhibitors are safe and appear to have biological effects independent of their cholesterol reducing properties (222). Neuhaus and colleagues reported that cells (PBMCs) of RRMS exposed *in vitro* to several forms of statins including mevastatin, simvastatin (Zocar®) and lovastatin (Mevastatin®) inhibited several different immune responses involved in MS. This study along with other early laboratory studies demonstrated that statins have several immunomodulatory effects by varied polygenic mechanisms including; (i) suppression of T and B cell proliferation, (ii) reduced expression of activation-induced adhesion molecules on T cells, (iii) skewed the polarity of Th1 to Th2 cytokines, (iv) downregulated chemokine receptors of T and B cells and (v) reduced the secretion of MMP-9 protease, inhibition of potential neurotoxins such as TNF $\alpha$  and inducible nitric oxide synthetase (iNOS) (223-229). Furthermore, statins have been shown to ameliorate in several EAE models modifying the balance of Th1 and Th2 cells to a proinflammatory Th2 phenotype (230, 231). A small pilot open labeled proof of concept/Phase II was the first human study of simvastatin (Zocar®) in 30 RRMS. The preliminary analysis of the study results were recently reported demonstrating that Zocar® safely reduced the number of new lesions. Analysis of immune responses suggested a shift away from inflammation however no differences were observed in neurologic disability in this short study (232) controlled trials of simvastatin and atorvastatin (Lipitor®) are in the planning and will begin this year.

### 4.5.3. Pregnancy-induced hormonal therapy: treatment with oral estradiol (E<sub>3</sub>) hormone

Pregnancy induced hormones and their potential as therapeutic agents has been entertained. This has evolved from longstanding clinical observations that sex-specific differences and hormonal influences exist in MS patients. Patients with MS have fewer relapses during pregnancy especially during the third trimester and disease exacerbations during first few months after birth (233). In several EAE models exogenous estrogens (estradiol/estradiol)

ameliorated disease by altering multiple immune responses (234-236). These polygenic mechanisms included; (i) suppression of Th1 proinflammatory cytokines, (ii) reduced the secretion of metalloproteases, (iii) downregulated of chemokines and chemokine receptors of T and B cells with inhibition of T cell, B and macrophage recruitment (234-236). A small open label, cross-over, proof of concept/Phase II trial with the treatment of estradiol in nonpregnant women with RRMS and SPMS was performed. Compared with pretreatment baseline, patients with RRMS but not SPMS showed a significant reduction of number and volume of gadolinium enhancing lesions. Furthermore, when the estradiol was stopped during the posttreatment phase the number and volume of gadolinium enhancing lesions returned to pretreatment values and then decreased when treatment was reinstated (237). Correlating with these clinical findings were decreases in proinflammatory cytokines including INF- $\gamma$  (237). The above studies are the rationale for a larger placebo-controlled trials to examine long term efficacy with an acceptable side effect profile. One such trial is currently in the planning. The potential long term benefits must be weighed against side effects and toxicities including carcinogenesis and thrombosis.

### 4.5.4. Targeting with oral peroxisome proliferator-activated receptor gamma (PPAR $\gamma$ ) agonists: treatment with avandia

Potential targeting of EAE and MS with oral peroxisome proliferator-activated receptor gamma (PPAR $\gamma$ ) agonists are currently being investigated. The ligand agonists of the peroxisome proliferator-activated receptor (PPAR $\gamma$ ) exert anti-inflammatory effects on a number of inflammatory cells including glial cells. This molecular interaction results in reduced proliferation and activation of T cells, and induction of myelin gene expression (238-241). Several of these oral agents have been FDA approved to treat diabetes. PPAR $\gamma$ -deficient heterozygous mice evidence myelin oligodendrocyte glycoprotein (MOG) induced EAE exacerbation and Th1 response (239). Several other models of EAE have shown that orally administered PPAR gamma ligands ameliorate the severity of monophasic, chronic disease and of relapsing disease in mice immunized with myelin oligodendrocyte glycoprotein (MOG) and/or myelin basic protein (MBP). Attenuation of clinical signs appear to correlate with decreased CNS inflammation and a reported reduction in lymphocyte infiltration, inflammatory chemokine and cytokine expression, and increased inhibitor of kappa B (IkB) expression in the brain (238-240). A clinical phase II double blinded placebo controlled trial of an oral diabetic agent (Avandia ®) is underway to examine safety and effectiveness in RRMS patients.

## 5. SUMMARY

Advancing knowledge of disease mechanisms and a deeper understanding of the immune pathogenesis in MS will refine current drug-specific therapies and shape the rationale for future immunotherapeutic strategies. Exploding technologies in areas such as neuroimaging, genomic cDNA microarray and proteomic analysis of MS

will further transform and delineate the heterogeneity and mechanisms underlying MS, yielding new targets for therapeutic strategies. Currently a more complex and intriguing picture of MS immune dysregulation is evolving. For example, immune cells and CNS inflammatory reactions may have dual roles inducing both proinflammation and neurotoxicity as well as, anti-inflammatory and protective immunity. Insight into this interrelated and aberrant neuroimmune imbalance, reflecting variable components such as encephalogenic effector cells, dysregulated regulatory populations and possible neurodegenerative processes will yield more specific and complimentary therapies. These mechanisms of disease pathology may impact differently upon subpopulations of patients and even vary within different stages of MS in a given individual. Future treatment options therefore will most likely utilize a combination of synergistic therapies with distinct mechanisms of action to address the treatment of a diverse MS patient population in all stages of disease offering greater efficacy and long term benefit. Tomorrow's arsenal of novel immune agents will reflect a patient-tailored treatment regimen of immune modulatory, suppressive and protective strategies for the treatment of MS shaped by different disease types and stages.

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## 7. REFERENCES

1. Steinman, L: Multiple sclerosis: a coordinated immunological attack against myelin in the central nervous system. *Cell* 85, 299 (1996)
2. De Staphano, N., P.M. Matthews and Narayanan: Axonal dysfunction and disability in a relapse of multiple study of a patient. *Neurology* 49, 1138-41 (1997)
3. Lucchinetti, C., W. Brück, J. Parisi, B. Scheithauer, M. Rodriguez and H. Lassmann: Heterogeneity of multiple sclerosis lesions: implications for the pathogenesis of demyelination. *Ann Neurol* 47, 707 (2000)
4. Lucchinetti, C. B., W. and J Noseworthy: Multiple sclerosis: recent developments in neuropathology, pathogenesis, magnetic resonance imaging studies and treatment. *Curr Opin Neurol* 14, 259 (2001)
5. Storch, M. K. S., A. U. Brehm, R. Weissert, E. Wallstrom, M. Kerschensteiner, T. Olsson, C. Linington, and H. Lassman: Autoimmunity to myelin oligodendrocyte glycoprotein in rats mimics the spectrum of multiple sclerosis pathology. *Brain Pathol* (2001)
6. Trapp, B. D. P., J. R.M. Ransohoff, R. Rudick, S. Mork and B. Lo: Axonal transection in the MS lesion. *N Engl J Med* 338, 278 (1998)
7. Dean: How many people in the world have multiple sclerosis? *Neuroepidemiology* 13, 1 (1994)
8. Noseworthy, J. L., C., M. Rodriguez and B.G. Weinshenker: Multiple sclerosis. *N Engl J Med* 343, 983 (2000)
9. Baranzini, S. E. O., J. R and S. L. Hauser: New insights into the genetics of multiple sclerosis. *Journal of Rehabilitation Research and Development* 39, 201 (2002)
10. Abdul-Majid, K. B. S., A. C. Bourquin, H. Lassmann, C. Linington, T. Olsson, S. Kleinau and R.A. Harris: Fc receptors are critical for autoimmune inflammatory damage to the central nervous system in experimental autoimmune encephalomyelitis. *Scand J Immunol* 55, 70 (2002)
11. Kenealy, S. J. P.-V., M.A. and J.L. Haines: The genetic epidemiology of multiple sclerosis. 143, 7 (2003)
12. Robinson, W. H. U., P and L. Steinman: Genomic and proteomic analysis of multiple sclerosis. *Current Opinion in Immunology* 15, 660 (2003)
13. Lublin, F. D. and S.C. Reingold: Defining the clinical course of multiple sclerosis: results of an international survey. *Neurology* 46, 907 (1996)
14. Weinshenker, BG. B. Bass, GPA Rice, J. Noseworthy, W. Carriere and J Baskerville: The natural history of multiple sclerosis: a geographically based study. *Brain* 112, 133 (1989)
15. Raine, C. S: The Dale E. McFarlin Memorial Lecture: the immunology of the multiple sclerosis lesion. *Ann Neurol* 36 suppl S61-S72 (1994)
16. Lucchinetti, C. M., RN. D. McGavern, W. Bruck, G. Gleich, RM. Ransohoff, C. Trebst, B. Weinshenker, D. Wingerchuk, JE. Parisi and H. Lassmann: A role for humoral mechanisms in the pathogenesis of Devic's neuromyelitis optica. *Brain* 125, 1450 (2002)
17. Babbe, H. E. A: Clonal expansion of CD8+ T cells dominate the T cell infiltrate in active multiple sclerosis lesions as shown by micromanipulation and single cell polymerase chain reaction. *J Clin Invest* 192, 393 (2002)
18. Steinman, L: Multiple sclerosis: a two-stage disease. *Nature Immunology* 2, 762 (2001)
19. Hemmer, B. A., J. and H.P. Hartnung: New concepts in the immunopathogenesis of multiple sclerosis. *Nature Reviews* 3, 291 (2002)
20. Kohm, A. P. C. P. A. S. D. M: Regulation of experimental autoimmune encephalomyelitis (EAE) by

CD4+CD25<sup>+</sup> regulatory T cells. *Novartis Found Symposium* 252, 45 (2003)

21. Jacobs, L. D. B., R.W J.H. Simon, R.P. Kinkel, C.M. Brownsheildle, T.J. Murray and the CHAMPS Study Group: Intramuscular inteferon beta-1a therapy initiated during the first demyelinating event in multiple sclerosis. *New England Journal of Medicine* 343, 898 (2000)

22. Comi, G. F., M. and F. Barkof: Interferon beta 1a (Rebif) in patients with acute syndromes suggestive of multiple sclerosis: a multi-center, randomized, double-blind, placebo-controlled study. *Neurology* 54 (2000)

23. Hartung, H. G., R. and MIMS Study Group: Mitoxantrone in progressive multiple sclerosis: a placebo-controlled, randomised, observer-blind phase III trial: clinical results and 3-year follow up. *Neurology* 52 (1999)

24. Goodin, D. S. F., E.M. and G.P. Garmany: Disease modifying in multiple sclerosis. Report of the Therapeutics and Technology Assessment Subcommittee of the American Academy of Neurology and the MS Council for Clinical Practice Guidelines. *Neurology* 58, 169 (2002)

25. Calabresi, P. A: Considerations in the treatment of relapsing-remitting multiple sclerosis. *Neurology Volume* 58(8), S10 (2002)

26. Edan, G. B., B. and D. Brassat: Safety profile of mitoxantrone in a cohort of 802 multiple sclerosis patients. *Neurology* 58, A168 (2002)

27. O'Connor, P: Key issues in the diagnosis and treatment of multiple sclerosis. *Neurology* 59, S1 (2002)

28. Tullman, M. L., F. and A. Miller: Immunotherapy of multiple sclerosis-Current practice and future directions. *Journal of Rehabilitation Research and Development* 39, 273 (2002)

29. Noseworthy, J. H: Management of multiple sclerosis: current trials and future options. *Current Opinion Neurol* (2003)

30. Martin, R. M., H.F. and D.E. McFarland: Immunologic aspects of demyeinating disease. *Annual Reviews in Immunology* 10, 153 (2003)

31. Zhang, J. W., H.L. and D.A. Hafler: Autoreactive T cells in multiple sclerosis. *Int Rev Immunol* 9, 183 (1992)

32. Van Gool, S. V., P. M. de Boer and JL Ceuppens: CD80, CD86 and CD40 provide accessory signals in a multiple-step T-cell activation model. *Immunol Rev* 153, 47 (1996)

33. Martin, R. S., C.S. and H.F. McFarland: Immunotherapy of multiple sclerosis: Where are we? Where should we go? *Nature Immunology* 2, 785 (2001)

34. Prat, E. R. M: The immunopathogenesis of multiple sclerosis. *Journal of Rehabilitation Research and Development* 39, 187 (2002)

35. Zamvil, S. S. N., P.A. D.J.RL Mitchell, Knobler, R.B. Fritz and L. Steinman: Encephalitogenic T cell clones specific for myelin basic protein. An unusual bias in antigen recognition. *J Exp Med* 162, 2107 (1985)

36. Wucherpfennig, K. W., and J.L. Strominger: Molecular mimicry in T cell mediated autoimmunity: viral peptides activate human T cell clones specific for myelin basic protein. *Cell* 80, 695 (1995)

37. Wucherpfennig, K. C., I. S. Stefan Hausmann, L. Jack J.L. Strominger, L. Steinman and K. Warren: Recognition of the immunodominant myelin basic protein peptide by autoantibodies and hla-dr2-restricted t cell clones from multiple sclerosis patients identity of key contact residues in the B-cell and T-cell epitopes. *Clinical Investigation Volume* 100, 1114 (1997)

38. Vanderlugt, C. L. N., K.L K.M Nikceovich, T.N. Eagar, J.A. Bluestone and SD Miller: Pathologic role and temporal appearance of newly emerging autoepitopes in relapsing experimental autoimmune encephalomyelitis. *J Immunology* (2000)

39. Gerritse, K. L., J.D. R.J. Noelle, A. Aruffo, J.A. Ledbetter, W.J. Boersma and E. Claassen: CD40-CD40 ligand interactions in experimental allergic encephalomyelitis and multiple sclerosis. *Proc Natl Acad Sci* 93, 2499 (1996)

40. Baranzini, S. E. E. A: B-cell repertoire diversity and clonal expansion in multiple sclerosis brain lesions. *J Immunology* 163, 5133 (1999)

41. Columbo, M. E. A: Accumulation of clonally related B lymphocytes in the cerebral spinal of multiple sclerosis. *J Immunology* 164, 2782 (2000)

42. O'Connor, K. C. B.-O., A and D.A. Hafler: The neuroimmunology of multiple sclerosis: possible roles of T and B cells in immunopathogenesis. *J Clin Immunol* 21, 81 (2001)

43. Zhang, J. Z. R., V.M. and M.V. Tehada-Simon: T cell vaccination in MS: results of a preliminary study. *J Neurol* 249, 212 (2002)

44. Seguin, R., and L. H. Kasper: Sensitized lymphocytes and CD40 ligation augment interleukin-12 production by human dendritic cells in response to Toxoplasma gondii. *J Infect Dis* 179, 467 (1999)

45. Chitnis, T. S. J. K: Role of costimulatory pathways in the pathogenesis of multiple sclerosis and experimental autoimmune encephalomyelitis. *Journal of Allergy and Clinical Immunology* 112, 837 (2003)

46. Balas, K. E. R., J.B. H.L. Weiner and W.W. Hancock: CCR5<sup>+</sup> and CCR3<sup>+</sup> T cells are increased in multiple sclerosis and their lignads MIP-1alpha and IP-10 are expressed in demyelinating brains. *Proc Natl Acad Science* 96, 6873 (1999)

47. Weiner, H. L: Multiple sclerosis: a disease of immune dysregulation. *Actrims* 2003, 3 (2003)
48. Flugel, A. B., T. T. Ritter, M. Labeur, D.E. Jenne and Z. Li: Migratory activity and functional changes of green fluorescent effector cells before and during experimental autoimmune encephalomyelitis. *Immunity* 14, 547 (2001)
49. McCarron, R. M. K., O. M. Spatz and D.E. McFarlin: Interaction between myelin basic protein-sensitized T lymphocytes and murine cerebral vascular endothelial cells. *J Immunol* 137, 3428 (1986)
50. Washington, R. B., J. R.F. Todd, W. Newman, L. Dragovic and P. Dore-Duffy: Expression of immunologically relevant endothelial cell activation antigens on isolated central nervous microvessels: from patients with multiple sclerosis. *Ann Neurol* 35, 89 (1996)
51. Archelos, J. J. H. P. H: The role of adhesion molecules in multiple sclerosis: biology, pathogenesis, and therapeutic implications. *Mol. Med Today* 3, 310 (1997)
52. Kieseier, B. C. S., T. Giovannoni, G. and H.P. Hartung: Matrix metalloproteinases in inflammatory demyelination: targets for treatment. *Neurology* 53, 20 (1999)
53. Steinman, L. M., R. C. Bernard, P. Conlan and J.R. Okensberg: Multiple sclerosis: deeper understanding of its pathogenesis reveals new targets for therapy. *Annual Rev Neuroscience* 25, 491 (2002)
54. Kern, A. B., R. I. Bank and E.E. Marcantonio: The role of the I domain in ligand binding of the human integrin  $\alpha 1 \beta 1$ . *J of Biol Chem* 269, 22811 (1994)
55. Cossins, J. A. C., J.M. J. Ford, K.M. Miller, R. Pigott and W. Vos: Enhanced expression of MMP-7 and MMP-9 in demyelinating multiple sclerosis lesions. *Acta Neuropathology* 94, 590 (1997)
56. Vos, C. M. P. V. H., E.S. C.J.A. de Groot, P. van der Valk and H.E. de Vries: Matrix metalloproteinase-12 is expressed in phagocytic macrophage lesions in active multiple lesions. *J Neurommunology* 138, 106 (2003)
57. Werner, P. A. D. P. E. A: Multiple sclerosis: altered glutamate homeostasis in lesions correlates with oligodendrocytes and axonal damage. *Ann Neurol* 50, 169 (2000)
58. Pitt, D. W., P. and C.S. Raine: Glutamate excitotoxicity in a model in multiple sclerosis. *Nature Medicine* 6, 67 (2000)
59. Chabas, D. B., S. D. Mitchell, C.C.A. Bernard and S. Rittling: The influence of the pro-inflammatory cytokine, osteopontin, on autoimmune demyelinating disease. *Science* 294, 1734 (2001)
60. Trapp, B. D. E. A: Axonal pathology in multiple sclerosis: relationship to neurologic disability. *Curr Opin Neurol* 12, 295 (1999)
61. Losseff, N. A. E. A: Progressive cerebral atrophy in multiple sclerosis. A serial MRI study. *Brain* (1999)
62. Kerschensteiner, M. G., E. L. Behrens, V. Vargas, L. Misgeld, E.F. Wolfgang, K. Klinkert, R. Kolbeck, E. Hoppe, R.L. Oropesa-Wekerle, I. Bartke, C Stadelmann, H. Lassmann, H. Wekerle and R. Hohlfeld: Activated Human T Cells, B Cells, and Monocytes Produce Brain-derived Neurotrophic Factor *In Vitro* and in Inflammatory Brain Lesions: A Neuroprotective Role of Inflammation? *Journal of Experimental Medicine* 189, 865 (1999)
63. Kerschensteiner, M. S., C. G. Dechant, H. Wekerle and R. Hohlfeld: Neurotrophic cross-talk between the nervous and immune systems: Implications for neurological diseases. *Annals of Neurology Volume* 53, 292 (2003)
64. Kerschensteiner, M., and R. Hohlfeld: Neurotrophic factors protect myelin from attack. *International MS Journal* 10, 2 (2003)
65. Hohlfeld, R. K., M. C. Stadelmann, H. Lassmann and H. Wekerle: The neuroprotective effect of inflammation: implications for the therapy of multiple sclerosis. *Journal of Neuroimmunology* 107, 161 (2000)
66. Becher, B. P., A. and J.P. Antel: Brain-immune connection: immuno-regulatory properties of CNS-resident cells. *Glia* 29, 293 (2000)
67. Graumann, U. R., R. A.J. Steck and N. Schaeren-Wiemers: Molecular changes in normal appearing white matter in multiple sclerosis are characteristic of neuroprotective mechanisms against hypoxic insult. *Brain Pathology* 13, 554 (2003)
68. Kapoor, R. D., M. P.A. Blaker, S.M. Hall and K.J. Smith: Protection of axons from the degeneration caused by nitric oxide can be achieved using blockers of the sodium channel and sodium-calcium exchanger. *Ann Neurol* 53, 174 (2003)
69. Schwartz, M: The neuroprotective effect of T cells. *Actrims* 2003
70. Schwartz, M. S., I. J. Fisher, T. Mizrahi and H. Schori: Protective immunity against the enemy within: fighting glutamate toxicity. *Trends in Neuroscience* 26, 297 (2003)
71. Moalem, G. L.-A., R. E. Yoles, F. Mor, I.R. Cohen and M. Schwartz: Autoimmune T cells protect neurons from secondary degeneration after central nervous system axotomy. *Nat Med* 5, 49 (1999)
72. Scolding, N. J. R. J. F: Remyelination in demyelinating disease. *Belleras Clin Neurology* 6, 525 (1997)
73. Levine, J. M. R., R. and J.W. Fawcett: The oligodendrocyte precursor in health and disease. *Trends in Neuroscience* 24 (2001)
74. Wolswijk, G: Chronic stage multiple sclerosis lesions contain a relatively quiescent population of oligodendrocyte. *J Neuroscience* 16, 601 (2001)

75. Chang, A. T., W.W. R Rudick and B.D. Trapp: Premyelinating oligodendrocytes in chronic lesions of multiple sclerosis. *J Exp Med* 346, 165 (2002)
76. Smith, K. J: Axonal protection in inflammatory demyelinating disease. Multiple Sclerosis: Clinical and Laboratory Research 9, S3 (2003)
77. Stadelmann, C. K., M., T Misgeld, Wolfgang Brück, R Hohlfeld and H. Lassmann: BDNF and gp145trkB in multiple sclerosis brain lesions: neuroprotective interactions between immune and neuronal cells? *Brain Vol* 125, 75 (2002)
78. Pal, E. T. K., T. M. Taniguchi, S. Miyake and T. Yamamura: Co-stimulation -dependent modulation of experimental autoimmune encephalomyelitis by ligand stimulation of Va14 NK T cells. *J of Immunol* 166, 662 (2001)
79. Singh, A. K. W., M.T. and L. Van Kaer: Natural killer T cell activation protects mice against experimental autoimmune encephalomyelitis. *J Exp Med* 12, 1801 (2001)
80. Furlan, R. B., A. D. Cantarella, E. Bramilla, M. Taniguchi, P. Dellabona, G. Casorati and G. Martino: Activation of invariant NKT cells by aGalCer administration protects mice from MOG35-55 induced EAE: critical roles of administration route and INF $\gamma$ . *Eur J Immunol* 33, 1830 (2003)
81. Jonuleit, H. S., E. M. Stassenb, A. Tuettenberg, J. Knopa and A.H. Enka: Identification and functional characterization of human CD4<sup>+</sup>CD25<sup>+</sup> T cells with regulatory properties isolated from peripheral blood. *Journal of Experimental Medicine* 193, 1285 (2001)
82. Araki, M. K., T., J.E. Gumperz, M.B. Brenner and Y. Miyake: Th2 bias of CD4<sup>+</sup> NKT cells derived from multiple sclerosis in remission. *Int Immunol* 15, 279 (2003)
83. Ben-Nun, A. W., H. and I.R. Cohen: Vaccination against autoimmune encephalomyelitis with T- lymphocyte line cells reactive against myelin basic protein. *Nature* 292, 60 (1981)
84. Lider, O. R., T. E Beraund, A. Ben-Nun and I.R. Cohen: Anti-idiotypic network induced by T cell vaccination against experimental autoimmune encephalomyelitis. *Science* 239, 181 (1988)
85. Lohse, A. W. M., F. N. Karin and I.R. Cohen: Control of experimental autoimmune encephalomyelitis by T cells responding to activated T cells. *Science* 244, 820 (1989)
86. Zhang, J. M., R. P. Stinissen, D. Hafler and J. Raus: MHC-restricted depletion of human myelin basic protein-reactive T cells by T cell vaccination. *Science* 261, 1451 (1989)
87. Medaer, R. S., P. L. Truyen, J. Raus and J. Zhan G: Depletion of myelin-basic-protein autoreactive cells by T cell vaccination: pilot trial in multiple sclerosis. *Lancet* 346, 807 (1995)
88. Correale, J. L., B. M. McMillian, D.Y. Ko, K. McCarthy and L.P. Weiner: T cell vaccination in secondary progressive multiple sclerosis. *J Neuroimmunol* 107, 130 (2000)
89. Kumar, V. T., R. H.M. Geysen and E. Sercarz: Immunodominant framework region 3 peptide from TCR V beta 8.2 chain controls murine experimental autoimmune encephalomyelitis. *J Immunology* 154 (1995)
90. Howell, M. D. W., S.T. T. Olee, H.C. Powell, D.J. Carlo and S.W. Brostoff: Vaccination against experimental allergic encephalomyelitis with T cell receptor peptides. *Science* 246, 668 (1989)
91. Morgan, E. N., C, J, J.P. Dively: Vaccination with a CDR2 BV6S2/6S5 peptide in adjuvant induced peptide-specific T cell responses in patients with multiple sclerosis. *J Neuroscience Res* 64, 298 (2001)
92. Killestein, J. O., T. and E. Wallenstrom: Antibody-mediated suppression of Vbeta5.2/5.3(+) T cells in multiple sclerosis results from an MRI-monitored phase II clinical trial. *Ann Neurology* 51, 467 (2002)
93. Nicholson, L. B., J.M. Greer, R.A Sobel, M.B. Lees and V.K. Kuchroo: An altered peptide ligand mediates immune deviation and prevents autoimmune encephalomyelitis. *Immunity* 3, 397 (1995)
94. Nicholson, L. B. M., A. B.P. Hafler, A. Sette and V.K. Kuchroo: A T cell receptor antagonist peptide induces T cells that mediate bystander suppression and prevent autoimmune encephalomyelitis induced with multiple myelin antigens. *Proc Natl Acad Science* 94, 9279 (1997)
95. Young, D. A. L., L.D. and S.S. Booth: IL-4, IL-10, IL-13 and TGB- $\beta$  from an altered peptide ligand Th2 cell clone down regulation adoptive transfer of experimental autoimmune encephalomyelitis. *J Immunology* 164, 3563 (2000)
96. Bielekova, B. L., A. H. McFarland and R. Martin: Therapeutic Potential of Phosphodiesterase-4 and -3 Inhibitors in Th1-Mediated Autoimmune Diseases. *The Journal of Immunology* 164, 1117 (2000)
97. Kappos, L. C., G. and H. Panitch: Induction of a nonencephalitogenic Th2 helper cell autoimmune response after administration of an altered peptide in multiple sclerosis ligand in a placebo controlled, randomized phase II trial. The Altered Peptide Ligand in Relapsing MS Study Group. *Nature Medicine* (2000)
98. Beilekova, B. G., B. and N. Richert: Encephalitogenic potential of the myelin basic protein peptide( amino acids 83-99) in multiple sclerosis: results of a phase II clinical trial with an altered peptide ligand. *Nature Medicine* 6, 1167 (2000)



99. Pedotti, R. M., D. J. Wedemeyer, M. Karpuj and D. Chabas: An unexpected version of horror autotoxicus: anaphylactic shock to a self-peptide. *Nat Immunol* 216 (2001)
100. Crowe, P. D. Q., Y. P.J. Conlon and J.P. Antel: NBI-5788, an altered peptide MBP83-99 peptide, induces a T-helper 2-like immune response in multiple sclerosis patients. *Ann Neurol* 48, 758 (2000)
101. Dhib-Jalbut, S: Mechanisms of action of interferons and glatiramer acetate in multiple sclerosis. *Neurology* 58, S3 (2002)
102. Fridkis-Hareli, M. S., L. J.N. Stern, L. Fugger, C. Brosman and J.L.Strominger: Novel synthetic amino acid copolymers that inhibit autoantigen-specific T cell responses and suppress experimental autoimmune encephalomyelitis. *J Clin Invest* 109, 1635 (2002)
103. Piccirillo, C.A. and G.J. Prud'homme: Prevention of experimental allergic encephalomyelitis by intramuscular gene transfer with cytokine-encoding plasmid vectors. *Hum Gene Ther* 10, 1915 (1990)
104. Waisman, A. E. A: Suppressive vaccination with DNA encoding a variable region gene of the T-cell receptor prevents autoimmune encephalomyelitis and activates Th2 immunity. *Nature Med* 2, 899 (1996)
105. Weissert, R: Protective DNA vaccination against organ-specific autoimmunity is highly specific and discriminates between single amino acid substitutions in the peptide autoantigen. *Proc Natl Acad Sci USA* 97, 1689 (2000)
106. Garren, H. R., P.J. T.A. Watkins, P. Fontoura, L.T. Nguyen, E.R. Estline, D.L. Hirschberg and L. Steinman: Combination of Gene Delivery and DNA Vaccination to Protect from and Reverse Th1 Autoimmune Disease via Deviation to the Th2 Pathway. *Immunity* 15, 15 (2001)
107. Qin, Y. D., P. Y. Zhang, M. Olek, J. Da, R.R. Richardson, J.P. Antel, J. Talbot, N.R. Cashman, W.W. Tourtellotte, H. Wekerle and S. van den Noort: Intrathecal B-Cell Clonal Expansion, an Early Sign of Humoral Immunity, in the Cerebrospinal Fluid of Patients with Clinically Isolated Syndrome Suggestive of Multiple Sclerosis. *Lab Investigation* 83, 1081 (2003)
108. Raine, C. S. C., B. S. L. Hauser and C.P. Genain: Demyelination in primate autoimmune encephalomyelitis and acute multiple sclerosis lesions: a case for antigen-specific antibody mediation. *Ann Neurol* 46 (1999)
109. Genain, C. P. C., B. S.L. Hauser and C.S. Raine: Identification of autoantibodies associated with myelin damage in multiple sclerosis. *Nat Med* 5, 170 (1999)
110. Svensson, L. A.-M., K.B. J. Bauer, H. Lassmann, R.A. Harris and R. Holmdahl: A comparative analysis of B cell-mediated myelin oligodendrocyte glycoprotein-experimental autoimmune encephalomyelitis pathogenesis in B cell-deficient mice reveals an effect on demyelination. *Eu J Immunol* 32, 1939 (2002)
111. Linington, C. B., M H. Lassmann, C. Brummer and K. Vass: Augmentation of demyelination in rat acute allergic encephalomyelitis by circulating mouse monoclonal antibodies directed against a myelin/oligodendrocyte glycoprotein. *Am J Pathol* 130, 443 (1998)
112. Myers, K. J. S., J. J. P. Dougherty and Y. Ron: Synergy between encephalitogenic T cells and myelin basic protein specific antibodies in the induction of experimental autoimmune encephalomyelitis. *J Neuroimmunol* 41, 1 (1992)
113. Lyons, J. A. S., M. P. Happ and A. H. Cross: B cells are critical for induction of experimental allergic encephalomyelitis by protein but not by a short encephalitogenic peptide. *Eur J Immunol* 29, 3432 (1999)
114. Reindl, M. E. A: Antibodies against myelin oligodendrocyte glycoprotein and the myelin basic protein in multiple sclerosis and other neurologic diseases. *Brain* 122, 2047 (1999)
115. Archelos, J. J. S., M.K. and H.P. Hartung: The role of B cells and autoantibodies in multiple sclerosis. *Ann Neurol* 47, 694 (2000)
116. Cross, A. H. T., J.L. and J. Lyons: B cells and antibodies in CNS demyelinating disease. *J Neuroimmunology* 112, 1 (2001)
117. Davis, T. A. W., C.A. A.J. Grillo-Lopez W.S. Velasquez, B. Link and D. G. Maloney: Single-agent monoclonal antibody efficacy in bulky non-Hodgkin's lymphoma: Results of the a phase II trial of Rituximab. *J Clin Oncol* 17, 1851 (1999)
118. Edwards, J.C.W. L. Szczepanski, J. Szechinski, A. Filiowicz-Sosnowska, D. Close and R. M.Stevens: B-lymphocyte depletion therapy in rheumatoid arthritis and other autoimmune disorders. *Biochem. Soc Trans* 30, 824 (2001)
119. Edwards, J. C. W. S., L. J. Szechinski, A. Filiowicz-Sosnowska, D. Close and R. M.Stevens: Efficacy and safety of rituximab, a B-cell targeted chimeric monoclonal antibody: a randomized, placebo-controlled trial in patients with rheumatoid arthritis. *Arthritis and Rheum* 46, S197 (2002)
120. DeVita, S. Z., F. S. Sacco, A. DeCandia, R. Ranin and G. Ferraccioli: Efficacy of selective B cell blockade in the treatment of rheumatoid arthritis. *Arthritis and Rheum* 46 (2002)
121. Cambridge, G. L., M.J. J.C.W. Edwards, M.R. Ehrenstien, M. Salden and D.B. Webster: B lymphocyte depletion in patients with rheumatoid arthritis: serial studies of immunological parameter. *Arthritis and Rheum* 46, S508 (2002)

122. Boyle, J. E., T. and A. Engert: An overview of the current clinical use of the anti-CD20 monoclonal antibody rituximab. *Ann of Oncology* 14, 520 (2003)
123. Berntsen, S. T., G.E. B.T. Gjertsen, J. Hammerstrom, R. Langh and J.H. Sorbo: Rituxan (rituximab) therapy for chronic cold agglutinin disease [abstract]. *Blood* 93, 315a (2000)
124. Zaja, F. R., D. G. Fuga, T. Muchelutti, A. Sperotto and R. Fanin: Rituximab in case of cold agglutinin disease. *Br J Hematol* 115, 232 (2001)
125. Lee, E. Z., K.W. T.C. Gentile and A. Zimrin: Rituxan in the treatment of auto-immune Hemolytic anemia (AIHA) [abstract]. *Blood* 96, 2560a (2000)
126. Salah, M. N., J. Gutiheil, M. Moore, B. Feinberg, P. Bunch and J. Butler: A pilot study of anti-CD20 MoAb rituximab in patients with refractory immune thrombocytopenic purpura (ITP). *Blood* 96, 1086a (2000)
127. Levine, T.D. and A. Pestronk: IgM antibody-related polyneuropathies: B-Cell depletion chemotherapy using rituximab. *Neurology* 52, 1701 (1999)
128. Zaja, F. R., D. G. Fuga, G. Perella and M. Baccarani: Rituximab for myasthenia gravis developing after bone marrow transplant. *Neurology* 55, 1062a (2000)
129. Takemura, D. K., P.A. A. Braun, J.J. Goronzy and C.M. Weyland: T cell activation in rheumatoid synovium is B cell dependent. *J Immunol* 167, 4710 (2001)
130. Grewal, I.S. and R.A. Flavell: CD40 and CD154 in cell-mediated immunity. *Annu Rev Immunol* 16 (1998)
131. Banchereau, J. B., F. D. Blanchard, F. Briere, J.P. Galizzi and C. van Kooten: 1994. The CD40 antigen and its ligand. *Immunol Today* 12, 881 (1998)
132. Durie, F. H. F., T.M. S.R. Masters, J.D. Laman and R. J. Noelle: The role of CD40 in the regulation of humoral and cell-mediated immunity. *Immunol Today* 5, 404 (1994)
133. Hollenbaugh, D. O., H.D. R.J. Noelle, J.A. Ledbetter and A. Aruffo: The role of CD40 and its ligand in the regulation of the immune response. *Immunol Rev* 138, 24 (1994)
134. Vogel, L. A. and R. J. Noelle: CD40 and its crucial role as a member of the TNFR family. *Semin Immunol* 10, 435 (1998)
135. Dyke, H.J. and J.G. Montana: Update on the therapeutic potential of PDE4 inhibitors. *Expert Opin Investig Drugs* 11, 1 (2002)
136. Noelle, R. J: CD40 and its ligand in cell-mediated immunity. *Agents Actions Suppl* 49, 17 (1998)
137. Segal, B. M. D., B.K. and E.M. Shevach: An interleukin (IL)-10/IL-12 immunoregulatory circuit controls susceptibility to autoimmune disease. *J Exp Med* 187, 537 (1998)
138. Chang, J. T., R. M. Segal, E. M. Shevach: Role of costimulation in the induction of the IL-12/IL-12 receptor pathway and the development of autoimmunity. *J Immunol* 164, 100 (2000)
139. Stafford, E.A. and N.R. Rose: Newer insights into the pathogenesis of experimental autoimmune thyroiditis. *Int Rev* 6, 501 (2001)
140. Schimdt, C. M., T. B.M. A. Wittig, H. Hombach, H. Abken and A. Stallmach: Interleukin-12 antagonists as new therapeutic agents in inflammatory bowel disease. *Pathobiology* 70, 177 (2002)
141. Zhang, G. X. S. Y., B. J. Gran, I. Li, X. Siglienti, D. Chen, E. Calida, M. Ventura, Kamoun and A. Rostami: Role of IL-12 Receptor  $\beta$ 1 in Regulation of T Cell Response by APC in Experimental Autoimmune Encephalomyelitis. *J Immunol* 171, 4485 (2003)
142. Bright, J. J. D., C. M. Coon, S. Sriram and S.J. Klaus: Prevention of experimental autoimmune encephalomyelitis via inhibition of IL-12 signaling and IL-12-mediated Th1 differentiation: an effect of the novel anti-inflammatory drug lisofylline. *J Immunol* 161, 7015 (1998)
143. Segal, B. M. and E. M. Shevach: IL-12 unmasks latent autoimmune disease in resistant mice. *J Exp Med* 184, 771 (1997)
144. Tuohy, V. K., M. M. Yu, L. Yin, P.M. Mathisen, J. M. Johnson and J. A. Kawczak: Modulation of the IL-10/IL-12 cytokine circuit by interferon- $\gamma$  inhibits the development of epitope spreading and disease progression in murine autoimmune encephalomyelitis. *J Neuroimmunol* 111, 55 (2000)
145. Ichikawa, M. K., C.S. A. Inoue, J. Tsuyusaki, M. Yamazaki, Y. Inaba, Y. Sekiguchi, M. Itoh, H. Yagita and A. Komiyama: Anti-IL-12 antibody prevents the development and progression of multiple sclerosis-like relapsing-remitting demyelinating disease in NOD mice induced with myelin oligodendrocyte glycoprotein peptide. *J Neuroimmunol* 102, 56 (2000)
146. Leonard, J. P., K. E. Waldburger and S. J. Goldman: Prevention of experimental autoimmune encephalomyelitis by antibodies against interleukin 12. *J Exp Med* 181, 381 (1995)
147. Constantinescu, C. S. H., B. E.S. Ventura, M. Wysocka, L. Showe, E. Lavi, T. Fujioka, P. Scott, Trinchieri and A.M. Rostami: Modulation of susceptibility and resistance to an autoimmune model of multiple sclerosis in prototypically susceptible and resistant strains by neutralization of interleukin-12 and interleukin-4, respectively. *Clin Immunol* 98, 23 (2001)

148. Ahmed, Z. G., D. G. D. Pryce, D. Baker, J.P. Leonard and M.L. Cuzner: Myelin/axonal pathology in interleukin-12 induced serial relapses of experimental allergic encephalomyelitis in the Lewis rat. *Am J Pathol* 158 (2001)
149. Brok, H. V. M., M. E. Blezer, A. Schantz, D. Peritt, G. Treacy, J.D. Laman, J. Bauer and B.A.'t Hart: Prevention of experimental autoimmune encephalomyelitis in common marmosets using an anti-IL-12p40 monoclonal antibody. *Journal of Immunology* 169, 6554 (2002)
150. Ludmila, B. V. W., L.P. and B. M. Segala: IL-12 dependent/IFN independent expression of CCR5 by myelin-reactive T cells correlates with encephalitogenicity. *Journal of Neuroimmunology* 137, 109 (2003)
151. Becher, B. D., B.G. and R.J. Noelle: Experimental autoimmune encephalitis and inflammation in the absence of interleukin-12. *Clin Invest* 493 (2002)
152. Bright, J. J. M., B.F. C. Du and S. Sriram: Expression of IL-12 in CNS and lymphoid organs of mice with experimental allergic encephalomyelitis. *J Immunol* 82, 22 (1998)
153. Windhagen, A. N., J. and F. Dangond: Expression of co-stimulatory molecules B7-1 (CD80), B7-2 (CD86) and interleukin 12 cytokine in multiplesclerosis. *J Exp Med* 182, 1985 (1995)
154. Nicoletti, F. P., F. and C. Cocuzza: Elevated serum levels of interleukin-12 in chronic progressive multiple sclerosis. *J Neuroimmunology* 70, 87 (1996)
155. van Boxel-Dezaire, A. H., S. C. Hoff, B. W. van Oosten, C. L. Verweij, A. M. Drager, H. J. Ader, J. C. van Houwelingen, F. Barkhof, C. H. Polman, and L. Nagelkerken: Decreased interleukin-10 and increased interleukin-12p40 mRNA are associated with disease activity and characterize different disease stages in multiple sclerosis. *Ann Neurol* 45, 695 (1999)
156. Balashov, K. E. C., M. T. Ohashi, S.J. Khoury and H. L. Weiner: Defective regulation of IFN and IL-12 by endogenous IL-10 in progressive MS. *Neurology* 55, 192 (2000)
157. Webster, A. C. P., E.G. Higgins, G. Chapman and J.R. Craig: Interleukin 2 receptor antagonists for renal transplant recipients: a meta-analysis of randomized trials. *Transplantation* 77, 166 (2004)
158. Abrams, J. R. K., S.L. E. Hayes, T. Kikuchi, M.J. Brown, and S. Kang: Blockade of T lymphocyte costimulation with cytotoxic T lymphocyte-associated antigen 4-immunoglobulin (CTLA4Ig) reverses the cellular pathology of psoriatic plaques, including the activation of keratinocytes, dendritic cells, and endothelial cells. *J Exp Med* 681 (2000)
159. Moreland, L. W. A., R. F. Van den Bosch, T. Appelboom, M. Leon and P. Emery: Costimulatory blockade in patients with rheumatoid arthritis: a pilot, dose-finding, double-blind, placebo-controlled clinical trial evaluating CTLA-4Ig and LEA29Y eighty-five days after the first infusion. *Rheum* 46, 1470 (2002)
160. von Andrian, U.H. and B. Engelhardt: Alpha4 integrins as therapeutic targets in autoimmune disease. *N Engl J Med* 348, 68 (2003)
161. Tubridy, N. B., P.O. R. Capildeo, A. Chaudhuri, R. Forbes, C. P. Hawkins, R. A. C. Hughes, J. Palace, B. Sharrack, R. Swingle, C. Young, I. F. Moseley, D. G. MacManus, S. Donoghue and D. H. Miller: The effect of anti-[alpha]4 integrin antibody on brain lesion activity in MS. *The UK Antegren Study Group: Neurology Volume* 53, 466 (1999)
162. Miller, D. H. K., O.A. and W.A. Sheramata: A controlled trial of natalizumab for relapsing multiple sclerosis. *N Engl J Med* 348, 15 (2003)
163. Jackson, D.Y: Alpha 4 integrin antagonists. *Curr Opin Neurol* 8, 1229 (2002)
164. Cross, A. H. E. A. Peroxynitrite formation within the central nervous system in active multiple sclerosis. *J Neuroimmunology* 88, 45 (1998)
165. Yong, V. W. P., C. P. Forsyth and D.R. Edwards: Metalloproteinases in biology and pathology of the nervous system. *Nature Review Neuroscience* 2, 502 (2001)
166. Bar-Or, A. N., R.K. M. Duddy, A. Alter, H.J. Kim, I. Ifergan, C. Pennington, P. Bourgoin, D.R. Edwards and V.W. Yong: Analysis of all matrix metalloproteinase members in leukocytes emphasize monocytes as major inflammatory mediators in multiple sclerosis. *Brain* 126, 2738 (2003)
167. Hewson, A. K. S., T. J.P. Leonard and M.L. Cuzner: Suppression of experimental allergic encephalomyelitis in the Lewis rat by the matrix metalloproteinase inhibitor Ro31-9790. *Inflamm Res* 44, 345 (1995)
168. Dubois, B. M., S. U. Hurtenbach, L. Paeman, H. Heremans and J. van den Oord: Resistance of young gelatinase B-deficient mice to experimental autoimmune encephalomyelitis andnecrotizing tail lesions. *J Clin Invest* 104, 1507 (1999)
169. Brocke, S. P., C. L. Stinman, I.L. Wissman and T. Veromma: Antibodies to CD44 and integrin alpha 4, but not L-selectin, prevent CNS inflammation and experimental encephalomyelitis by blocking secondary leukocyte recruitment. *Proc Natl Acad Sci USA* 96, 6896 (1999)
170. Lanone, S. Z., T. Z. Zhu, W. Liu, C. G. Lee and B. Ma: Overlapping and enzymes contributions of matrix-metalloproteinases-9 and -12 in IL-13 induced inflammation and remodeling. *J Clin Invest* 110, 463 (2002)

171. Opdenakker, G. I. Nelissen and Van Damme: Functional roles and therapeutic targeting of gelatinase B and chemokines in multiple sclerosis. *Lancet Neurology* 12, 747 (2003)
172. Opdenakker, G. and J. Van Damme: Cytokine-regulated proteases in autoimmune diseases. *Immunol Today* 15, 103 (1994)
173. Gu, Z. K., M. B. Yan, S. J. Kridel, J. Cui and A. Strongin: S-nitrosylation of matrix metalloproteinases: signaling pathway to neuronal cell death. *Science* 297, 1186–90 (2002)
174. Rosenberg, G: Matrix metalloproteinases and neuroinflammation in multiple sclerosis. *Neuroscientist* 8, 586 (2002)
175. Brundula, V. R., N.B. L. M. Metz, C. C. Bernard and W.W. Yong: Targeting leukocytes MMPs and transmigration: minocycline as a potential therapy for multiple sclerosis. *Brain* 125, 1297 (2002)
176. Jansson, M. V. Panoutsakopoulou, J. Baker, L. Klein and Cantor: Cutting edge: attenuated experimental autoimmune encephalomyelitis in eta-1/osteopontin-deficient mice. *J Immunology* 168, 2096 (2002)
177. Steinman, L. Y., S. W. J. van Venrooij, D. Chabas, S. E. Baranzini, S. Rittling, D.T. Denhardt, R.A. Sobel, C. Lock and R. Pedotti: Response to comment on "The influence of the proinflammatory cytokine, osteopontin, on autoimmune demyelinating disease. *Science* 299, 1845 (2003)
178. Callier, S. B., L. S. E. Barazani, A. Swerdlow, R. Lincoln, L. Steinman, and J. R. Oskenberg: Osteopontin polymorphisms and disease course in MS. *Genes Immun* (2003)
179. Niino, M. K., S. T. Fukazawa, I. Yabe and K. Tashiro: Genetic polymorphisms of osteopontin in association with multiple sclerosis in Japanese patients. *J Neuroimmunology* 136, 125 (2003)
180. Vogt, M: Elevated osteopontin levels are associated with disease activity in relapsing remitting MS patients. *Ann Neurol* (2003)
181. Murphy, P. S., A. J. Shin, V. Gavriluk, C. Dello Russo, G. Weinberg, F. Sharp, A. Lu, M. T. Heneka, and D.L. Feinstein: Suppressive effects of ansamycins on inducible nitric oxide synthase expression and the development of experimental autoimmune encephalomyelitis. *J of Neuroscience Research* 67, 461 (2002)
182. Heneka, M. T. S., A. P. Murphy, J. A. Lyons, L. Dumitrescu and D. L. Feinstein: The heat shock response reduces myelin oligodendrocyte glycoprotein-induced experimental autoimmune encephalomyelitis in mice. *J Neurochem* 77, 568 (2001)
183. Jolivalta, C. G. H., R.B. S. Long, Chena, A.P. Mizisinb and C.S. Lai: A novel nitric oxide scavenger in combination with cyclosporine A ameliorates experimental autoimmune encephalomyelitis progression in mice. *Journal of Neuroimmunology* 138, 56 (2003)
184. Essayan, D. M: Cyclic nucleotide phosphodiesterases. *J Allergy and Clinical Immunology* 108, 671 (2001)
185. Bloom, T. J. and J. A. Beavo: Identification and tissue-specific expression of PDE7 phosphodiesterase splice variants. *Proc. Natl Acad Sci USA* 93, 14188 (1996)
186. Ekholm, D. H., B. G. Gao, M. Vergelli, R. Martin and V. Manganiello: Differential expression of cAMP cyclic nucleotide phosphodiesterase PDE3 and PDE4 activities in human T cell clones specific for myelin basic protein. *J Immunol* 159, 1520 (1997)
187. Lacour, M. A., J.F. K.M. Muller, C. Carlberg, J.H. Saurat and C. Hauser: cAMP up-regulates IL-4 and IL-5 production from activated CD4+ T cells while decreasing IL-2 release and NF-AT induction. *Int Immunol* 6, 1333 (1994)
188. Sommer, N. M., R. and H.F. McFarland: Therapeutic potential of phosphodiesterase type 4 inhibition in chronic autoimmune demyelinating disease. *J Neuroimmunology* 79, 54 (1997)
189. Eigler, A. S., B. U. Emmerich, K.H. Baumann, G. Hartmann, and S. Endres: Anti-inflammatory activities of cAMP-elevating agents: enhancement of IL-10 synthesis and concurrent suppression of TNF production. *J Leuk Biol* 63, 101 (1998)
190. Heystek, H. C. T. A. C. S., P. and C. Moulon: Phosphodiesterase 4 inhibitors reduce human dendritic cell inflammatory cytokine production and Th1-polarizing capacity. *International Immunology* 15, 827 (2003)
191. Griswold, D. E., E. F. Webb, A. M. Badger, P. D. Gorycki, P. A. Levandoski, M. A. Barnette, M. Grous, S. Christensen and T. J. Torphy: SB 207499 (Ariflo), a second generation phosphodiesterase 4 inhibitor, reduces tumor necrosis factor alpha and interleukin-4 production *in vivo*. *J Pharmacol Exp Ther* 287, 705 (1998)
192. Layseca-Espinosa, E. B., L. B. Alvarado-Sanchez, D. Portales-Perez, H. Portillo-Salazar and R. Gonzalez-Amaro: Rolipram Inhibits Polarization and Migration of Human T Lymphocytes. *J Invest Dermatol* 121, 81 (2003)
193. Genain, C. P. R., T. R.L. Davis: Prevention of autoimmune demyelination in non-human primates by cAMP-specific phosphodiesterase inhibitor. *Proc Natl Acad Sci USA* 92, 3601 (1995)
194. Sommer, N. L., P.A. and G.H. Nortoff: The antidepressant rolipram suppresses cytokine production and prevents autoimmune encephalomyelitis. *Nat Med* 1, 244 (1995)

195. Rott, O. C., E. and B. Fleischer: Phosphodiesterase inhibitor pentoxifylline, a selective suppressor of T- helper 1 but not type-2-associated lymphokine production, prevents induction of experimental autoimmune encephalomyelitis. *Eur J Immunol* 23 (1993)
196. Muthyla, S. W., K. D. H. Kim, B. G. Arnson and E. Chelmicka-Schorr: Experimental allergic encephalomyelitis, beta-adrenergic receptors and interferon gamma-secreting in beta-adrenergic agonist treated rats. *Int Immunol* 17, 895 (1995)
197. Makhoulouf, K. W., H.L. and S.J. Khoury: Potential of beta2-adrenergic agonist as add-on therapy for multiple sclerosis: focus on salbutamol (albuterol). *CNS Drugs* 16, 1 (2002)
198. Bothwell, M: Functional interactions of neurotrophins and neurotrophin receptors. *Ann Rev Neuroscience* 18, 223 (1995)
199. Gao, Y. N., E. W. Mellado and M.T. Filbin: Neurotrophins elevate cAMP to reach a threshold required to overcome inhibition by MAG through extracellular signal-regulated kinase-dependent inhibition of phosphodiesterase. *J Neuroscience* 23, 117770 (2003)
200. Villoslada, P. E. A: Human nerve growth factor protects common marmosets against autoimmune encephalomyelitis by switching the balance of T helper cell type 1 and 2 cytokines within the central nervous system. *J Exp Med* 191, 1799 (2000)
201. Linker, R. A. E. A: CNTF is a major protective factor in demyelinating CNS disease: a neurotrophic cytokine as modulator in neuroinflammation. *Nat Med* 8, 620 (2002)
202. Levison, S. W. A., P.J. A. Basu and J.K. Krady: Roles of ciliary neurotrophic factor in recovery from demyelinating diseases. *Actrims* 2003, 9 (2003)
203. Gold, B. G. E. A: Neuroimmunophilin ligands promote neuroprotection in a model of MS: effects of FK506 and nonimmuno-suppressant FK506 derivative (FR131706) in EAE. *ACTRIMS* (2003)
204. Stankoff, B. E. A: Ciliary neurotrophic factor (CNTF) enhances myelin formation: a novel role for CNTF and CNTF-related molecules. *J Neuroscience* 22, 9221 (2002)
205. Gilgun-Sherki, Y. P., H. E. Melamed, and D. Offen: Riluzole suppresses experimental autoimmune encephalomyelitis: implications for the treatment of multiple sclerosis. *Brain Research* 989, 196 (2003)
206. Matute, C: Properties of acute and chronic kainate excitotoxic damage to the optic nerve. *Proc Natl Acad Sci USA* 95, 10229 (1998)
207. McDonald, J. W. A., S. and K. Hyre: Oligodendrocytes from forebrain are highly vulnerable to AMPA/kainate receptor mediated excitotoxicity. *Nat Med* 4, 291 (1998)
208. Gurney, M. E. F., T.J. and C.S. Himes: Riluzole preserves motor in transgenic model of familial amyotrophic lateral sclerosis. *Neurology* 50, 62 (1998)
209. Anaki, T. K., T. and M. Matsubara: Protective effect of riluzole on MPTP-induced depletion of dopamine and its metabolite content in mice. *Metab Brain Dis* 15, 193 (2000)
210. Pratt, J. R., J. and F. Bardot: Effect of riluzole on focal cerebral ischemia in rats. *Neuroscience Lett* 140, 225 (1992)
211. Bensimon, L. L. and V. Meninger: A controlled trial of riluzole in amyotrophic lateral sclerosis. *N Engl J Med* 330, 585 (1994)
212. Bagasara, O. E. A: Activation of the inducible form of nitric oxide synthetase in the brains of the patients with multiple sclerosis. *Proc Natl Acad Sci USA* 92, 12041 (1995)
213. Ghafourifar, P. E. A: Mitochondrial nitric oxide synthetase stimulation causes cytochrome c release from isolated mitochondria. Evidence for intramitochondrial peroxynitrite. *J Biol Chem* 274, 31185 (1999)
214. Hooper, D. C. S., S. R.B. Kean, J.M. Champion, G.M. Dickson, I. Chaudhry and H. Korowski: Uric acid, a natural scavenger of peroxynitrite, in experimental allergic encephalomyelitis and multiple sclerosis. *Proc Natl Acad Sci USA* 95, 675 (1998)
215. Hooper, D. C. S., G.S. A. Zborek, T. Mikheeva, R.B. Kean, H. Korowski and S.V. Spitsin: Uric acid, a peroxynitrite scavenger, inhibits CNS inflammation, blood-CNS barrier permeability changes, and tissue damage in a mouse model of multiple sclerosis. *J FASEB* 5, 691 (2000)
216. Koprowski, H. S., S.V. and D.C. Hopper: Prospects for the treatment of multiple sclerosis by raising serum levels of uric acid, a scavenger of peroxynitrite. *Ann Neurol* 49, 139 (2001)
217. Spitsin, S. D.C. Hooper, T. Leist, L.J. Streletz, T. Mikeheeva and H. Koprowski: Inactivation of peroxynitrite in multiple sclerosis patients after oral administration of inosine may suggest possible approaches to therapy of the disease. *Multiple Sclerosis* 5, 313 (2003)
218. Sotgiu, S. E. A: Serum uric acid and multiple sclerosis. *Neurol Science* 23, 183 (2002)
219. Kazem, M. A.R. Dehpour, A. Minager and P. Ghafourifar: Uric acid: a novel treatment strategy for multiple sclerosis. *Trends in Pharmacological Sciences* 24, 563 (2003)

220. Paolillo, A. C., A.J. and P.D. Molyneux: Quantitative MRI in patients with secondary progressive MS treated with monoclonal antibody Campath 1H. *Neurology* 53, 751 (1999)
221. Coles, A. J. W., M. and S Smith: Pulsed monoclonal antibody treatment and autoimmune thyroid disease in multiple sclerosis. *Lancet* 169, 95 (1999)
222. Kobashigawa, J. A. K. and S. Laks: Effect of pravastatin on outcomes after cardiac transplantation. *N Engl J Med* 333, 621 (1995)
223. Stanislaus, R. P., K. A.K. Singh and I. Singh: Amelioration of experimental allergic encephalomyelitis in Lewis rats by lovastatin. *Neuroscience Lett* 269, 71 (1999)
224. Pahan, K. S., F.G. A.M. Namboodiri and Singh: Lovastatin and phenylacetate inhibit the induction of nitric oxide synthetase and cytokines in rat astrocytes, microglia, and macrophages. *J Clin Invest* 100, 2671 (1997)
225. Romano, M. D., L. and M. Sironi: Inhibition of monocyte chemotactic protein-1 synthesis by statins. *Lab. Investigation* 80, 1095 (2000)
226. Wong, B. L., W.C. A.M. Smith, J.T. Sosko, S.D. Wright and T.Q. Cai: Statins suppress Th-1 cell migration and secretion of matrix metalloproteinases 9 by inhibiting gerylation. *J Leuko Biol* 6, 959 (2001)
227. Weitz-Schidt, G. W., K. and V. Brinkmann: Statins selectively inhibit leukocyte function antigen-1 by binding to a novel regulatory integrin site. *Nat Med* 7, 687 (2001)
228. Zamvil, S. S. L. S. E. A: Cholesterol-lowering statins possess anti-inflammatory activity that might be useful for treatment of MS. *Neurology* 59, 970 (2002)
229. Neuhaus, S. S.-F. and F. Fazekas: Statins as immunomodulators: comparison with interferon- $\beta$  in MS. *Neurology* 59, 990 (2002)
230. Youseff, S. S., O. and J.C. Patarroyo: The HMG-CoA reductase inhibitor, atorvastatin, promotes a Th2 bias and reverses paralysis in central nervous system autoimmune disease. *Nature* 420, 78 (2002)
231. Aktas, O. W., S. and A. Smorodchenko: Treatment of relapsing paralysis in experimental encephalomyelitis by targeting Th1 cells through atorvastatin. *J Exp Med* 197, 725 (2003)
232. Vollmer, T. D., V. and W. Taylor: An open label, single arm study of simvastatin as a therapy for multiple sclerosis (MS). *Neurology* 60, A84 (2003)
233. Confavreux, C. H., M. M.M. Hours, P. Cortinovos-Touraniaire and T. Moreau: Rate of pregnancy-related relapse in multiple sclerosis. Pregnancy in Multiple Sclerosis Group. *N Engl J Med* 339, 285 (1998)
234. Kim, S. L., S.M. M.A. Dala, M.A. Verity and R.R. Voskuhl: Estriol ameliorates autoimmune demyelinating disease: implications for multiple sclerosis. *Neurology* 52, 1230 (1999)
235. Bebo, B. F., A. Fyre-Johnson, K. Adlard, A.G. Beam, A.A. Vanderbark and H. Offner: Low-dose estrogen therapy ameliorates experimental autoimmune encephalomyelitis in two different inbred mouse strains. *J Immunology* 166, 2080 (2001)
236. Subramanian, S. M., A. A. Zamora, A.A. Vandenberg and H. Offner: Oral feeding with ethinyl estradiol suppresses and treats experimental autoimmune encephalomyelitis in SJL mice and inhibits recruitment of inflammatory cells in the central nervous system. *J Immunology* 170, 1548 (2003)
237. Sicotte, N. L. L., S.M. and R. Klutch: Treatment of multiple sclerosis with pregnancy hormone estriol. *Ann Neurol* 52, 421 (2002)
238. Zhang, Y. C., L.W. C.J. Sinal, F.J. Gonzalez and P.A. Edwards: Peroxisome proliferator-activated receptor- $\gamma$  coactivator 1 $\alpha$  (PGC-1 $\alpha$ ) regulates triglyceride metabolism by activation of the nuclear receptor FXR. *Genes Development* (2004)
239. Bright, J. J. C., N. G. Muthian, Y. Barak and R.M. Evans: Peroxisome proliferator-activated receptor-g-deficient heterozygous mice develop and exacerbated neural antigen-induced Th1 response and experimental allergic encephalomyelitis. *J of Immunology* 171, 5743 (2003)
240. Feinstein, D. L. G., E. V. Gavriluk, C. Brosnan, C.C. Whitacre, L. Dumitrescu-Ozimek, G. Landreth, H. Pershad Singh, G. Weinberg and M.T. Heneka: Peroxisome proliferator-activated receptor-g-agonists prevent experimental autoimmune encephalomyelitis. *Ann Neurol* 51, 694 (2002)
241. Diab, C. D., J. D. R.Z. Smith, B.P. Hussain, A.E. Lovett-Racke, P.D. Drew and M.K. Racke: Peroxisome Proliferator-Activated Receptor- Agonist 15-Deoxy-12, 14-Prostaglandin J2 Ameliorates Experimental Autoimmune Encephalomyelitis. *J Immunology* 168, 2508 (2002)

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