

## INTRACELLULAR SIGNALING MECHANISMS OF SEX HORMONES IN ACUTE MYOCARDIAL INFLAMMATION AND INJURY

Daniel R. Meldrum<sup>1,2</sup>, Meijing Wang<sup>1</sup>, Ben M. Tsai<sup>1</sup>, Ajay Kher<sup>1</sup>, Jeffrey M. Pitcher<sup>1</sup>, John W. Brown<sup>1</sup>, Kirstan K. Meldrum<sup>1</sup>

*Departments of Surgery<sup>1</sup> and Cellular and Integrative Physiology<sup>2</sup>, Indiana University Medical Center, IB 461, Indianapolis, Indiana 46202*

### TABLE OF CONTENTS

1. Abstract
2. Introduction
3. Clinical Outcomes
4. Myocardial Inflammation
5. Sex Differences in Myocardial Injury
6. Estrogen and Testosterone
7. Summary and Perspective
8. Acknowledgement
9. References

### 1. ABSTRACT

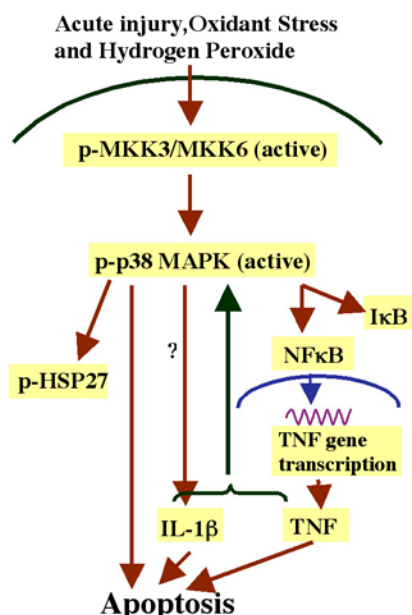
Sex hormones are important modifiers of the acute inflammatory response to injury, an important aspect of myocardial depression and apoptosis following ischemia or endotoxemia. Hemorrhage, trauma, ischemia/reperfusion, burn and sepsis each lead to cardiac dysfunction. Gender has been shown to influence the inflammatory response as well as outcomes following acute injury. The mechanisms by which sex affects the inflammatory response and the outcome to acute injury are being actively investigated. It is now recognized that myocardial inflammation plays a crucial role in I/R-induced myocardial dysfunction. Inflammatory mediators, such as TNF-alpha are produced by cardiomyocytes and contribute to myocardial functional depression and apoptosis. Gender differences in the inflammatory response following burn injury have been demonstrated. However, gender differences in the setting of acute I/R-induced inflammation are unclear. In addition, a critical component of the signal transduction pathway leading to myocardial inflammation is the activation of p38 mitogen-activated protein kinase (MAPK). In other systems, it appears that gender differences exist in the p38 MAPK signaling pathway. The inflammatory response, including the p38 MAPK signaling cascade and expression of proinflammatory cytokines such as TNF-alpha and IL-1beta, may precipitate cardiomyocyte apoptosis following I/R injury. Apoptosis may be an essential component in the pathogenesis of heart failure, and there is evidence that myocyte apoptosis in the failing human heart is markedly lower in women than in men. The prevention of cell death attenuates I/R-induced injury on myocardial anatomy and performance. This review will: 1) examine evidence for gender differences in the outcome to acute injury; 2) explain the myocardial inflammatory response to acute injury; and 3) elucidate the various mechanisms by which gender and sex hormones affect the myocardial response to acute injury.

### 2. INTRODUCTION

Estrogen and testosterone have emerged as important modifiers of the acute inflammatory response to injury, which plays an important role in the development of acute myocardial injury (1-42). Cardiovascular disease is the number one cause of death among women, accounting for nearly fifty percent of female deaths (43). Statistics show that women on average develop cardiovascular disease ten to fifteen years later in life than men, and that the risk may increase after the menopause (44). This observation has led to much speculation as to what physiological change(s) associated with the menopause is responsible for the higher risk of atherosclerosis. Estrogen and its potential as a cardioprotective agent and as an immunomodulator of the inflammatory response in atherosclerosis has received the most attention from researchers. The inflammatory response to injury is a double-edged sword playing an important part in the damage produced by the injury as well as in the process of repair. In the heart, it has been shown that the inflammatory response is produced not only by resident macrophages but also by the cardiomyocytes (5, 45). The cytokines produced during an inflammatory response cause depression of cardiac function (46-48) and hence methods that block the inflammatory response may be protective (49-51). Sex differences have been noted in these responses and the potential reasons for the differences have been the subject of extensive research (1-4, 52, 53). This review will: 1) examine evidence for gender differences in the outcome to acute injury; 2) explain the myocardial inflammatory response to acute injury; and 3) elucidate the various mechanisms by which sex affects the myocardial response to acute injury.

### 3. CLINICAL OUTCOMES

Gender differences have been noted in outcome to acute injuries like myocardial infarction, burns, trauma



**Figure 1.** p38 mitogen-activated protein kinase (MAPK) signaling pathway.

and sepsis. Hospital based clinical studies have shown that females have a higher mortality rate after myocardial infarction compared to males (54-56). In general, the females in many of these studies were older, had higher risk factors (diabetes, hypertension and congestive heart failure), more complications, and lower likelihood of receiving treatment (57). Importantly, more males died from myocardial infarction before reaching the hospital and the 28 day mortality for males and females was the same (58-60). This actually suggests that females are relatively protected in the immediate aftermath of a myocardial infarction but are similar to males at the end of a month.

Sepsis and trauma are two other inflammatory conditions associated with sex dependent outcomes. For mortality in trauma, there is either no sex difference (61-63) or sex difference in blunt trauma but not in penetrating trauma (64). Studies that have found sex differences are inconsistent. Some studies showed benefit only in females > 50 years (64) while others showed benefit only in females < 50 years old (65, 66). Females however have lower incidence of pneumonia, sepsis and multi organ failure after trauma (62, 66-68). In sepsis, some studies have found a higher mortality rate in females > 80 years old (69), while others have found lower mortality rates for females (70). In a study by Schroeder and colleagues (71) involving septic patients, females demonstrated lower mortality, higher interleukin-10 (IL-10) and lower tumor necrosis factor alpha (TNF) levels. Fewer female patients in intensive care units developed sepsis, although once sepsis developed the mortality rate was the same (72). Clinical studies on sex differences in mortality after burns present inconsistent evidence. Some showed females (73) or only females aged 30-59 years (74) to have higher mortality while others showed that females have lower incidence of multi-organ dysfunction and sepsis after burns (75).

In contrast to the clinical studies, animal studies have consistently found that females do better. Protective effects of acute administration of estrogen, in an *in vivo* left anterior descending (LAD) coronary artery ischemia/reperfusion (I/R) model, have been shown in different animals (76-79). Chronic administration of estrogen provides protection from I/R injury in isolated hearts undergoing global ischemia and in hearts undergoing *in vivo* LAD obstruction (80, 81). Estrogen also protected against reperfusion induced arrhythmias after LAD I/R injury (78, 81, 82). Ovariectomized females have worse cardiac functional recovery after global I/R, in an isolated heart, than sham ovariectomized females or ovariectomized females with estradiol replacement (83, 84). After burn injury, females have lower cytokine production and better cardiac function (85). Trauma-hemorrhage leads to depressed immune function and this depression is more severe in males (86-88). The immune depression is in part caused by testosterone (89, 90) since both castration (87) and receptor blockade (91-94) attenuated this depression. Estrogen also prevented the immune depression caused by trauma-hemorrhage (95, 96).

Animal studies have consistently shown that females are protected against acute injury while clinical studies appear inconsistent. A possible reason is that in animal studies the female population is well controlled and only proestrous females are used while clinical studies have a heterogeneous population. Furthermore, the underlying condition of humans is less uniform. Indeed, the few animal studies that used diestrous females showed that diestrous females had functional recovery equivalent to males, but lower than proestrous females (85, 97). This has been borne out by a few clinical studies which showed that cardiac function fluctuates with the hormonal changes of menstrual cycle (98-100). Thus, it is important to know the hormonal status of females and future clinical studies that take this into account may produce more consistent results. The remainder of this review will focus on gender differences in the myocardial inflammatory response to acute injury.

#### 4. MYOCARDIAL INFLAMMATION

Acute injuries lead to the production of an inflammatory cascade. The inflammatory cascade is triggered through many pathways but it may converge onto a few key regulatory proteins. Perhaps important among these are p38 mitogen-activated protein kinase (MAPK) and nuclear factor kappa B (NFκB). Ischemia reperfusion (I/R), sepsis, trauma and burn injury lead to oxidative stress (5, 101) which activates p38 MAPK (24, 102, 103) and NFκB (104, 105) (Figure 1). p38 MAPK is regulated by upstream kinases referred to as MAP kinase kinase (MAPKK) which themselves are regulated by MAP kinase kinase kinases (MAPKKK). This sequence of phosphorylation causes amplification of the signal. p38 MAPK is crucial in the cascade leading to TNF gene induction and its inhibition is protective (106-110) (Figure 1). p38 MAPK is also involved in the production of IL-1, IL-4, IL-6 and IL-8 (111-114). Wang and colleagues (108) showed that the increase in TNF, IL-1 and IL-6

production, after endotoxin infusion in an isolated heart, could be inhibited by a p38 MAPK inhibitor. TNF sequestration alone led to a decrease in IL-1 and IL-6. This suggests that TNF is upstream to IL-1 and IL-6 and that the effect of p38 MAPK on other cytokines might be mediated through TNF.

NFkB is involved in the regulation of many processes including apoptosis, cell growth, stress responses, innate and acquired immunity and sepsis. NFkB is bound to inhibitory kappa B (IkB) in the cytoplasm, this prevents its nuclear localization and DNA binding. Phosphorylation of IkB by inhibitory kappa B kinase (IKK) results in dissociation of IkB from NFkB, allowing NFkB to translocate to the nucleus. MAPK, protein kinase C (PKC) and phosphatidylinositol 3 kinase (PI3K)/Akt converge on IKK for NFkB activation (115, 116). p38 MAPK plays an important role in activation of NFkB and expression of NFkB-dependent genes (109, 115, 117, 118) (Figure 1). NFkB activation leads to production of TNF and IL-1 (36, 119, 120).

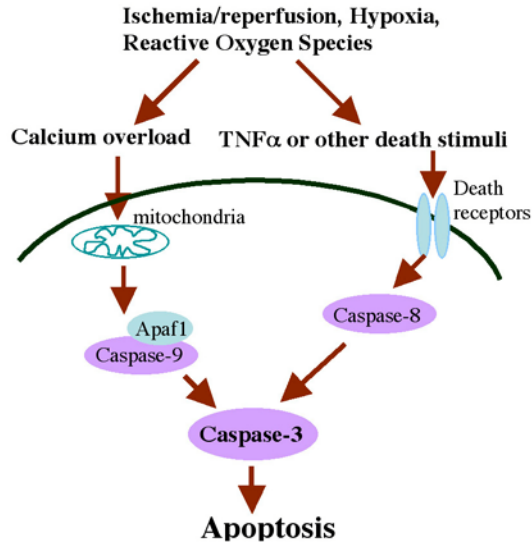
MAPK and NFkB activation also occur through other mechanisms. TNF and IL-1 activate p38 MAPK and NFkB (Figure 1). IL-1 leads to formation of a complex between IL-1 receptor-associated kinase (IRAK) and myeloid differentiation factor 88 (MyD88) (121, 122). IRAK is released and binds to TNF-receptor associated factor (TRAF). TRAF activates a MAPKKK, which activates p38 MAPK and NFkB (123). TNF also activates them through TRAF. TNF and IL-1 activate p38 MAPK and NFkB, they increase the production of TNF and IL-1, thus forming a feed-forward mechanism and amplifying the inflammatory response. Lipopolysaccharide (LPS) through its interaction with CD14 provokes rapid activation of protein tyrosine kinase which activates a pathway involving Ras/Raf-1/MEK/MAPK/NFkB (5). Recently, LPS has also been shown to use the MyD88/IRAK pathway (116, 124). These pathways have been delineated but their role in myocardial inflammatory response to sepsis is not known and should be the subject of further research.

TNF causes decreased myocardial contractile efficiency and reduced ejection fraction, hypotension, decreased systemic vascular resistance and biventricular dilation. These effects are produced through calcium dyshomeostasis, direct cytotoxicity, oxidant stress, disruption of excitation-contraction coupling, myocyte apoptosis and induction of other cardiac depressant cytokines such as IL-1 and IL-6 (5). TNF, through sphingosine, disrupts L-type channel-induced calcium release and thereby depresses calcium transients (125-127). NO appears to mediate TNF-induced desensitization of myofilaments to intracellular calcium (128, 129). Anti-TNF measures have been protective however a study using TNF receptor knockout mice showed increased infarcts in the knockout mice after LAD occlusion (130). This suggests that TNF leads to activation of both protective and damaging responses and a decrease in the excessive TNF production after acute injury may be protective but a complete absence is harmful. This might be why clinical studies of anti-TNF measures have not shown the benefit expected (131).

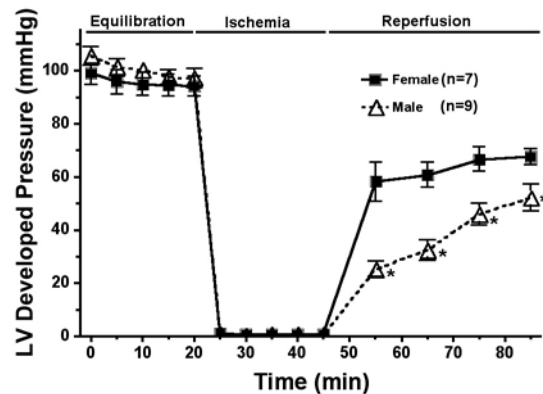
TNF, IL-1 and IL-6 lead to increased expression of adhesion molecules like intercellular adhesion molecule-1 (ICAM-1) (132-134). Neutrophil adhesion occurs leading to a respiratory burst which produces reactive oxygen species (135). The importance of neutrophils in cardiac injury is made clear by the cardioprotection obtained with neutrophil depletion (136-138). The reactive oxygen species produced during acute injury can also cause leukocyte chemotaxis (139) and adhesion (140) possibly through complement activation (141), upregulating P selectin expression (142), by inducing ICAM-1 expression (143) or by increasing the ability of ICAM-1 to bind neutrophils (144). Protective effects of antioxidant enzymes were shown in left circumflex coronary artery I/R by infusing antioxidant enzymes (145) and in isolated hearts of transgenic mice overexpressing antioxidant enzymes (146, 147). Disappointingly, clinical studies using them have found no significant benefit (148, 149).

Accumulating evidence indicates that cytokines are important mediators of cardiovascular disease. Acute injury in the form of ischemia, endotoxemia, or burn trauma results in myocardial functional suppression, in part via the local production of inflammatory mediators such as TNF, IL-1, and IL-6. Ischemia-reperfusion injury induces the local production of TNF, IL-1 and IL-6. Indeed, locally produced inflammatory mediators may be an important contributor to postischemic myocardial dysfunction and cardiomyocyte apoptosis. TNF, IL-1, or IL-6-induced depression of myocardial function in an ex vivo model has been observed. The mechanisms by which TNF causes myocardial dysfunction include calcium dyshomeostasis, direct cytotoxicity, oxidant stress, disruption of excitation-contraction coupling, and myocyte apoptosis, as well as the induction of other cardiac depressants such as IL-1 and IL-6. This study examines the mechanisms by which the heart produces these substances and uses targeted therapies to determine if blocking inflammatory cytokine production helps the heart. This information may help save heart cells during conditions of low blood flow such as heart attacks or heart surgery.

One of the signaling enzymes involved in proinflammatory cytokine production is p38 mitogen-activated protein kinase (MAPK). Ischemia-reperfusion, oxidant stress, and hydrogen peroxide directly activate p38 MAPK. Activation of p38 MAPK is required for TNF and IL-1 production in cardiomyocytes. Recently, p38 MAPK activation has been correlated with proinflammatory cytokine production in myocardium after endotoxemia and burn trauma. In addition, myocardial I/R injury in animal and human studies results in activation of myocardial p38 MAPK, and p38 MAPK inhibition results in improved myocardial function following I/R injury. Therefore, p38 MAPK may mediate post-ischemic myocardial dysfunction by upregulating the inflammatory response. However, it remains unclear how activation of p38 MAPK induces postischemic myocardial inflammatory cytokine production. Active p38 MAPK may lead to inhibitory  $\kappa$ B (IkB) phosphorylation, which subsequently results in disruption of the NFkB-IkB complex and activates NFkB.



**Figure 2.** Myocardial apoptosis signaling pathway.



**Figure 3.** Changes in left ventricular developed pressure (LVDP) following ischemia and reperfusion in normal age matched male and female rat hearts perfused with modified Krebs-Henseleit solution. Results are mean  $\pm$  SEM, \* $p$ <0.01 vs. female at the corresponding time points).

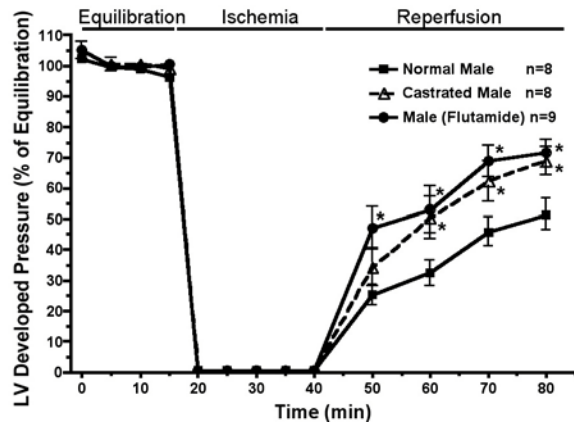
Activated NF $\kappa$ B translocates from the cytoplasm to the nucleus, where it docks to TNF promoter and activates TNF gene transcription. Recently, caspase-1 and caspase-11 were shown to function upstream of IL-1 maturation. IL-1 is initially synthesized as an inactive precursor requiring the IL-1-converting enzyme (ICE or caspase-1) for cleavage to the mature, biologically active molecule. Caspase-11 is crucial for the activation of caspase-1. ICE is required for IL-1 activation in the postischemic heart. It has been demonstrated that caspase-11 induction by LPS and hypoxia in microglia was mediated by p38 MAPK (150, 151). However, it is still unknown whether active p38 MAPK may directly induce myocardial IL-1 after acute ischemic injury. Thus, understanding the role of the p38 MAPK signaling pathway in I/R injury may differentiate p38 MAPK, NF $\kappa$ B, caspase-1, or caspase-11 as putative therapeutic targets.

Apoptosis may be an essential component of myocardial dysfunction following I/R (Figure 2). Myocyte apoptosis in heart failure differs between women and men. Apoptosis can be mediated by either extrinsic death receptor signaling or intrinsic mitochondrial control pathway. The extrinsic pathway involves TNF or other death signaling binding to death receptor TNFR1, which recruits procaspase-8 and activates it. Caspase-8 then activates downstream caspase-3 and induces apoptosis. The release of cytochrome *c* from mitochondria into the cytoplasm initiates the intrinsic signaling of apoptosis. Cytochrome *c* binds to apoptotic protease-activating factor-1 (Apaf-1) and then leads to recruitment and activation of procaspase-9, which results in the activation of procaspase-3 and apoptosis. The activation of intrinsic apoptotic pathway (the release of cytochrome *c* and activation of caspase-9) during reperfusion, but not ischemia has been observed in chick cardiomyocytes and activation of caspase-8, caspase-3 exists in cardiomyocytes response to hypoxia and ischemia. However, no information exists regarding the influence of gender and sex hormones in myocardial proapoptotic signaling cascades to I/R.

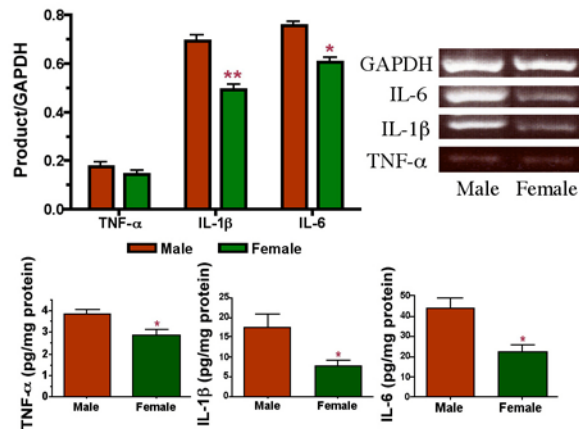
## 5. SEX DIFFERENCES IN MYOCARDIAL INJURY

Sex differences in cytokine production after acute injury have been shown in many studies (1, 2) and appears to correlate with myocardial function (Figures 3-5). Our recent studies showed that normal female myocardium produces less TNF and IL-1 when compared to male myocardium undergoing the same insult (1). In clinical studies, Schroeder and colleagues (71) showed that females had lower TNF and higher IL-10 levels in sepsis and Oberholzer and associates (62) showed that male trauma patients had higher IL-6 levels. In animal studies, females had lower cardiomyocyte secretion and serum concentrations of TNF, IL-1, IL-6 and IL-10 after burn injury (85). Deshpande and colleagues (152) showed that estradiol attenuated the LPS-induced production of IL-1, IL-6 and TNF by macrophages and also decreased NF $\kappa$ B binding activity. Estradiol also attenuated the increase in IL-6 after trauma-hemorrhage (153). As stated earlier, trauma-hemorrhage leads to depressed immune function and this depression is more severe in males (86-88). The immune depression is in part caused by testosterone (89, 90) since both castration (87) and receptor blockade (91-94) attenuated this depression. Estrogen also prevented the immune depression caused by trauma-hemorrhage (95, 96). Even though gender difference in the cytokine production after acute injury has been noted, the mechanisms by which these differences are mediated are not known.

As p38 MAPK is an important regulatory protein in the inflammatory cascade, recent evidence showing sex differences in p38 MAPK point towards a possible role for it in mediating sex differences in response to acute injury. Sex differences in p38 MAPK activation have been shown after trauma-hemorrhage and I/R. Wang and colleagues (154) found that males had higher levels of phosphorylated p38 MAPK (the activated form of p38 MAPK) in isolated hearts after global I/R. They also showed that



**Figure 4.** Changes in left ventricular developed pressure (LVDP) following ischemia and reperfusion in normal male, castrated male and male with androgen receptor blocker rat hearts. Results are mean  $\pm$  SEM, \* $p$ <0.01 vs. normal male at the corresponding time points).



**Figure 5.** Cardiac TNF- $\alpha$ , IL-1 $\beta$  and IL-6 in normal age matched male and female rat hearts after I/R. Right one in the top shows relative optical density of target PCR signals, normalized to GAPDH PCR signal. Left one is representative gel photograph. Protein levels are shown in the bottom (Mean  $\pm$  SEM,  $n$ =5/group. \* $p$ <0.05, \*\* $p$ <0.01 vs. male).

ovariectomized females had phosphorylated p38 MAPK equivalent to males and that treatment of males and ovariectomized females with estradiol prevented the increase in phosphorylated p38 MAPK caused by I/R. This suggests that the decreased cytokine production and cardioprotection caused by estrogen might be mediated through p38 MAPK. Indeed, our data indicates decreased p38 MAPK activation in females following ischemia (Figure 6).

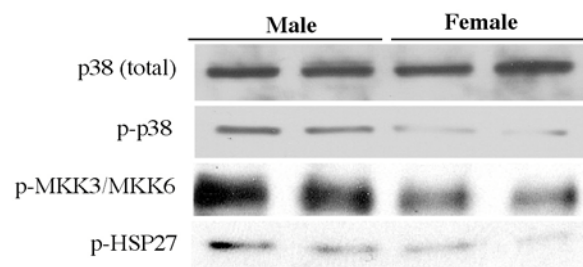
Chaudry's group (155) have presented interesting findings on p38 MAPK activation after trauma-hemorrhage. They showed that sex, after trauma-hemorrhage, did not alter the expression of the nonphosphorylated p38 MAPK in macrophages but altered the expression of phosphorylated p38 MAPK. Macrophages from female shams had increased

phosphorylated p38 MAPK compared with males. Trauma-hemorrhage increased phosphorylated p38 MAPK in males but decreased it in females. They also showed that castration attenuated the increase in phosphorylated p38 MAPK caused by trauma-hemorrhage and that supplementation with 5 $\alpha$  Dihydrotestosterone restored the ability of trauma-hemorrhage to activate p38 MAPK. This suggests that testosterone leads to p38 MAPK activation after trauma-hemorrhage. The significance of the gender difference in p38 MAPK in shams is not known and it will be interesting to see if it is valid in humans and other animals and whether it leads to differences in cytokine production. Both studies show higher p38 MAPK activity after acute injury in males suggesting that acute injuries lead to activation of similar pathways and p38 MAPK is involved in them (154, 155). Previous studies indicate that after trauma-hemorrhage there is a decreased capability of macrophages to release cytokines and that this is more severe in males. This raises the question as to whether increased p38 MAPK activation in males is the cause for the immune depression and if it is why does it lead to immune depression instead of activation. A hypothesis proposed by Chaudry's group (155) that integrates all the evidence is that the increased p38 MAPK activation leads to such an increase in the inflammatory cytokine response that it leads to an exhaustion of the capacity of the cells to respond to any further stimulus. These studies have shown that estradiol and testosterone modify p38 MAPK and that this may be responsible for gender differences in response to acute injury though further studies are needed to clarify the issue.

As oxidant stress is one of the stimuli for inflammatory cytokine production, any gender difference in antioxidants could lead to differences in the inflammatory response to acute injury. The sudden increase in production of hydroxyl radicals that occurs during reperfusion was reduced by estrogen in a canine model of cardiac ischemia and reperfusion (156). Using a similar I/R model it was found that estrogen decreased lipid peroxidation and in an in-vitro study estrogen decreased superoxide anion production from coronary artery segments undergoing hypoxia/reoxygenation (81). A possible mechanism for the antioxidant effect of estrogen is increased GSH (reduced glutathione) in the myocardium. This was supported by using a GSH synthesis antagonist, which reduced GSH and partially reversed the beneficial effects of estrogen on left ventricular diastolic pressure, systolic shortening and lipid peroxidation after LAD I/R (157). As there was only a partial reversal by using a GSH synthesis antagonist other mechanisms must also play a role in the protective effects of estrogen.

Another possible antioxidant mechanism is increased levels of superoxide dismutase in females. Barp and colleagues (158) showed decreased lipid peroxidation and increased superoxide dismutase in female hearts compared with males. Ovariectomized females had significant decrease in superoxide dismutase and an increase in lipid peroxidation. However, this study did not induce acute injury and hence the role of these changes in the protective effects seen in females after acute injury is





**Figure 6.** The expression of activated p38 MAPK signaling pathway was increased after I/R in male rat hearts compared to females. Shown are representative immunoblots. n=4/group.

unknown. Oxygen radicals have also been shown to upregulate expression of adhesion molecules. Squadrito and colleagues (159) showed that estrogen decreased serum and macrophage TNF levels and decreased ICAM-1 expression in the myocardium after left main coronary artery I/R. This led to decreased leukocyte accumulation and smaller infarcts in the estradiol treated group. Other studies have also shown that estrogen decreases neutrophil accumulation after LAD I/R (76, 77). The mechanism by which estrogen decreases the leukocyte accumulation is not well defined though there is evidence for a role of antioxidant mechanisms, nitric oxide (NO) and decreased TNF production. Many physiological and pharmacological actions have been attributed to NO and several of these are cardio-protective. The protective effects appear dependent on endothelial NO synthase (eNOS) (160) and inducible NO synthase (iNOS) (161). Estrogen increases NO production through increased expression of iNOS and eNOS in cardiomyocytes (78, 162-165). NO provides protection by reducing the expression of adhesion molecules, especially P selectin, and decreasing neutrophil accumulation (166, 167). Increased infarct size occurred when a NOS inhibitor was used (168) NO also modulates calcium channels and affects myocardial contractility.

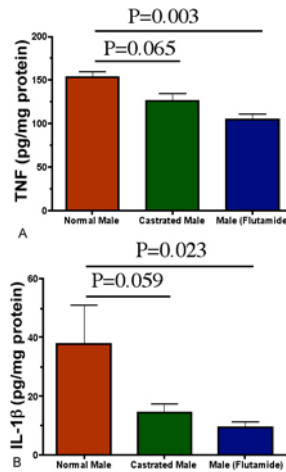
Cardiac contraction is triggered by calcium release through L-type calcium channels and sarcoplasmic reticulum (SR). During relaxation sodium-calcium exchanger (NCX) and sarcoplasmic reticulum calcium ATPase (SRCA) act to remove intracellular free calcium. Increase in intracellular calcium occurs with myocardial ischemia and inhibition of this is protective (169, 170). NO decreases free intracellular calcium by inhibiting L-type calcium channels (171, 172) and inhibiting the calcium release from SR (173-175). The inhibition of calcium release from SR may be mediated by inactivation of ryanodine receptor calcium release channel by NO (174). NO may also modulate calcium by activating or potentiating the effects of ATP sensitive potassium ( $K_{ATP}$ ) channels (176, 177). Thus, estrogen leads to increased production of NO and through it decreases neutrophil accumulation and free intracellular calcium and provides cardioprotection.

The sympathetic stimulation that occurs after acute injury acts through beta adrenoreceptors to increase

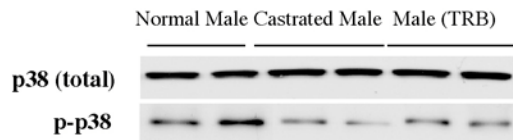
cAMP and cAMP-dependent protein kinase A. This kinase phosphorylates many effector proteins, one of them being Phospholamban (PLB). PLB is a sarcoplasmic reticulum (SR) protein that when phosphorylated dissociates from sarcoplasmic reticulum calcium ATPase (SRCA) and increases the affinity of SRCA for calcium (178). This leads to increased SR calcium, increased SR calcium release and increased myocardial contractility (179-181). Golden and coworkers (182) have shown that protein kinase A also increases expression of NCX. Thus, betaadrenergic stimulation not only increases calcium release from SR but also increases NCX expression. These pathways lead to increased intracellular calcium and increased myocardial contractility but after acute injury these same pathway lead to increased damage.

Estrogen and progesterone depletion causes upregulation of beta<sub>1</sub> adrenergic receptors in the heart and this was reversed by supplementation of either estrogen or progesterone (183). Estrogen deficiency led to increased density of beta adrenoreceptors in the heart though the affinity remained the same. Isoproterenol (selective beta adrenoreceptor agonist) increases infarct size in isolated hearts undergoing I/R but estrogen protects against the injury produced by it (184). The increased damage caused by beta adrenoreceptors is probably due to increased accumulation of intracellular calcium after injury. Using Phospholamban-knockout mice, Cross and colleagues (185), found that the knockout mice had greater baseline myocardial contractility but worse myocardial recovery after I/R. This study confirmed the fact that increased calcium through SRCA leads to increased baseline myocardial contractility but it also leads to increased damage after acute injury. They also found that female knockout mice recovered better from I/R compared to male knockout mice. Use of L-NAME (a NOS inhibitor) blocked the protective effects in females while giving SNAP (a NO donor) to males provided protection equivalent to female knockouts. In another study, when isolated hearts were treated with Isoproterenol or high  $Ca^{2+}$  increased I/R injury was found in males (186). Females were protected against this injury but lost the protection when treated with L-NAME. The probable mechanism by which NO provides this protection is by inhibiting the ryanodine receptor calcium release channel and thereby inhibiting calcium release from cardiac sarcoplasmic reticulum.

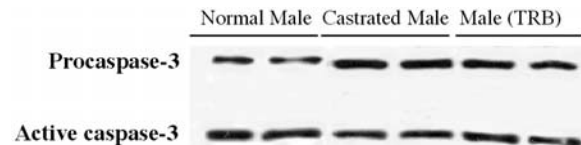
Castration causes a reduction in the myocardial expression of beta adrenergic receptor, L-type calcium channel and sodium-calcium exchanger (NCX) and supplementation with androgens reversed these effects (187). This led to reduced contractility in cardiomyocytes from castrated males. Indeed, our recent studies demonstrated that endogenous testosterone contributes to the increased myocardial TNF and IL-1, and activation of p38 MAPK and caspase 3 (Figures 7-9). The impact of these changes on myocardial damage, after acute injury, has not been studied but it can be postulated that the increased calcium influx produced by them would lead to increased myocardial injury.



**Figure 7.** The expression of myocardial TNF and IL-1 $\beta$  following ischemia and reperfusion in normal males, castrated males and males with testosterone receptor blockade. TNF and IL-1 $\beta$  protein levels are lower in castrated males and males with testosterone receptor blockade than in normal males (n=5-7/group).



**Figure 8.** Myocardial p38 MAPK activation following acute ischemia and reperfusion in normal males, castrated males and males with testosterone receptor blockade. More activation of p38 MAPK exists in normal male rat hearts compare to castrated males and males with testosterone receptor blockade (n=4/group).



**Figure 9.** Myocardial caspase-3 after acute ischemia and reperfusion in normal males, castrated males and males with testosterone receptor blockade. More active caspase-3 and less procaspase-3 in normal male rat hearts was observed compared to castrated males and males with testosterone receptor blockade (n=4/group).

Horton and colleagues (85) found that the cardiomyocyte secretion of inflammatory cytokines after burns by diestrous rats was similar to males but the sodium/calcium ( $\text{Na}^+/\text{Ca}^{2+}$ ) accumulation was significantly less and diestrous rats had better cardiac function. This suggested that differences in  $\text{Na}^+/\text{Ca}^{2+}$  accumulation are instrumental in mediating sex differences in cardiac function after burns. Sodium – calcium exchanger (NCX) is present on the sarcolemmal membrane and countertransports three  $\text{Na}^+$  ions for one  $\text{Ca}^{2+}$  ion. NCX can function in either direction depending on the transmembrane gradients of the ions and the membrane potential. A role for this protein in myocardial injury was suspected because manipulations of intracellular sodium

led to corresponding changes in intracellular calcium and that affected the myocardial injury produced (188, 189). NCX normally works in the calcium removal mode but in ischemia there is an increase in intracellular  $\text{Na}^+$  and change in the membrane potential, which leads to reversing of the NCX (190). Isolated hearts of transgenic male mice overexpressing NCX had greater I/R injury than wild type mice (191). Female transgenic mice were protected from the increased I/R injury compared to male transgenics and this protection was lost partially when female transgenic mice were ovariectomized. The mechanism of the protective effect in these transgenic females is unknown as male and female transgenic hearts did not have a difference in the overexpression of NCX. The possible mechanisms are that females are better able to withstand the higher intracellular calcium produced by NCX overexpression or that even with similar NCX overexpression females have lower intracellular calcium due to lower intracellular sodium. Evidence in support of the latter has been provided by Sugishita and colleagues (192). They studied myocytes isolated from NCX overexpressing mice and found lower intracellular calcium after metabolic inhibition in females. They also found lower intracellular sodium in females suggesting that the decreased calcium in females might be due to this. Due to the stoichiometry of NCX, exchanges 3 sodium ions for 1 calcium ion, even small differences in intracellular sodium would lead to larger differences in calcium. This suggests that females may be protected due to lower intracellular sodium after acute injury though the reason for the lower intracellular sodium in females remains unanswered.

ATP sensitive potassium ( $\text{K}_{\text{ATP}}$ ) channels have been shown to be present in cardiac mitochondria and to mediate protection against ischemic injury. These channels are activated by many stimuli including adenosine (193), NO (177) and free radicals and these then activate Protein Kinase C (PKC) which links to mitochondrial  $\text{K}_{\text{ATP}}$  channels (194, 195). Mitochondrial calcium overload plays an important part in I/R injury (196, 197). Opening of mitochondrial  $\text{K}_{\text{ATP}}$  channels results in potassium influx, which decreases the driving force for calcium uptake (198). Use of  $\text{K}_{\text{ATP}}$  channel agonists has shown the decreased mitochondrial calcium concentration caused by opening of  $\text{K}_{\text{ATP}}$  channels (199). Estrogen has been shown to decrease infarct size in canine LAD I/R through mitochondrial  $\text{K}_{\text{ATP}}$  channels (200) and decrease reperfusion-induced arrhythmias through sarcolemmal  $\text{K}_{\text{ATP}}$  channels (201). The mechanism by which estrogen activates these channels is unknown. The evidence that estrogen increases NO production and that NO can activate these channels indicates that this might be a possible mediator. Estrogen has been used in coronary angioplasty patients and has been shown to reduce myocardial ischemia caused by balloon inflation, possibly through  $\text{K}_{\text{ATP}}$  channels (202).

Apoptosis is a genetically controlled process by which cells undergo nonnecrotic cellular death. Testosterone has been shown to promote apoptosis in vascular endothelial cells (203) and renal tubular cells (204) while anabolic-androgenic steroids do so in

cardiomyocytes (205). Verzola and colleagues (204) showed a dose dependent effect of testosterone on apoptosis in renal tubular cells. They also showed that testosterone upregulated Fas, Fas ligand and Fas associated death domain. The use of caspase-3 inhibitor, caspase-8 inhibitor or caspase-9 inhibitor reduced the apoptosis produced by testosterone. Testosterone was shown to decrease Bcl-2 expression (203). These studies indicate the possible role of testosterone in promoting apoptosis though further research is needed to delineate the mechanisms.

Estrogen has been shown to decrease apoptosis of cardiac myocytes induced by staurosporine and this was associated with decreased caspase 3 activity and decreased NFkB (206). Wang and colleagues (154) showed that females had lower active caspase 3, and active caspase 8 but higher Bcl-2 after global I/R, in isolated hearts. They also showed that ovariectomized females had increased active caspase-3 while estradiol administration to males and ovariectomized females reduced the level of active caspase-3. Camper-Kirby and colleagues (207) found that females have higher nuclear localization of phospho-Akt in myocardium. Akt kinase activity in female nuclei was higher and they had higher phospho-forkhead levels (which is a downstream target of Akt). Administration of estradiol increased nuclear localization of phospho-Akt. Akt has previously been shown to inhibit apoptosis in cardiomyocytes in I/R (208, 209). It has also been shown to mediate the anti-apoptotic effects of insulin growth factor-1 (IGF-1) (210). The possible mechanisms of the antiapoptotic effects of Akt are phosphorylation of BAD, (211) phosphorylation of caspase 9 (212) and phosphorylation of FKHL1 which leads to blocking of Fas ligand expression (213). This suggests that Akt might be a mediator for the anti-apoptotic effect of estrogen after acute injury.

TNF is also known to produce apoptosis. TNF induces apoptosis by binding to either TNF type 1 receptor or Fas (5, 124). Both are associated with death domains, TNF receptor-associated death domain (TRADD) and Fas-associated death domain (FADD). These death domains interact with receptor-interaction protein and activate endonucleases. The endonucleases destroy the nuclear DNA leading to apoptosis. As stated earlier, estrogen decreases TNF production after acute injury and hence may decrease the apoptosis produced by TNF.

## 6. ESTROGEN AND TESTOSTERONE

In recent studies (1-3) we have provided evidence that inherent gender differences affect the otherwise normal myocardial response to acute ischemic injury. These results suggest that differences between the female and male response to I/R are related to the correlation between myocardial function and inflammatory signaling. Understanding the mechanisms that lead to these differences may allow low side effect therapeutics. The main findings of those studies were that after I/R, compared to normal males, castrated males and males treated with testosterone receptor blockade (TRB=flutamide): 1)

exhibited cardiac functional protection; 2) had decreased proinflammatory cytokine production (TNF- $\alpha$ , IL-1 $\beta$  and IL-6), and active p38 MAPK and caspase-1; 3) had decreased expression of apoptotic-related proteins caspase-3 and caspase-11; and 4) had increased expression of anti-apoptotic protein Bcl-2.

Sex differences exist in the response of myocardium to acute injury (85, 227, 228). Most studies have focused on the role of estrogen in cardiac protection, whereas little data exists regarding the effect of testosterone on cardiovascular disease. The role of testosterone on cardiac injury may be important because the heart can accumulate testosterone at higher concentrations than other androgen target organs (229), and functional androgen receptors are present in isolated cardiac myocytes (230). Furthermore, testosterone can modulate nuclear transcription by membrane receptor second messenger cascades (231); therefore, testosterone may be involved in mediating L-type calcium channel activity, coronary vasomotion by NO-dependent (232) and NO-independent mechanisms (233), and apoptotic cell death of cardiomyocytes (205, 234). In this study, rat hearts from castrated males and flutamide-treated males exhibited better functional recovery after acute I/R compared to normal males. This observation is consistent with other reports of protected cardiac performance in animals with testosterone depletion or testosterone receptor blockade after trauma-hemorrhage (93, 235), and suggests that endogenous testosterone may have a negative effect on the heart subjected to acute I/R.

Accumulating evidence indicates that cytokines are important mediators of injury-induced cardiovascular dysfunction. Acute injury in the form of ischemia, endotoxemia, or burn trauma results in myocardial functional suppression, in part via the local production of inflammatory mediators such as TNF and IL-1. Ischemia-reperfusion injury induces the local production of TNF, IL-1, and IL-6 (5, 214, 215). Indeed, locally produced proinflammatory mediators may be important contributors to postischemic myocardial dysfunction and cardiomyocyte apoptosis. p38 MAPK has been shown to be an important mediator of TNF production. In our study, castrated males and TRB-treated males, both of which had improved post-ischemic functional recovery compared to normal males, also had less myocardial TNF, IL-1 and IL-6 production, and decreased p38 MAPK activation.

It remains unclear how I/R induces myocardial inflammatory cytokine production. One possible mechanism may involve myocardial p38 MAPK signaling. Stimuli such as ischemia-reperfusion, oxidant stress, and hydrogen peroxide directly activate p38 MAPK (5). Activation of p38 MAPK is required for TNF and IL-1 production in cardiomyocytes (220, 225, 226). TNF production following I/R is dependent on NFkB translocation, which may occur via p38 MAPK activation, and regulation of this process occurs pretranscriptionally (5). On the other hand, caspase-1 and caspase-11 are shown to function upstream of IL-1 maturation (92). Caspase-1 is recognized as the IL-1-converting enzyme



(ICE) for cleavage of IL-1 precursor to the mature form (236, 237). Precursor caspase-1 results in the generation of mature p10/p20 and p10/p20 form ICE (238). ICE is required for IL-1 activation in the postischemic heart (239). Furthermore, the activation of caspase-1 is dependent on caspase-11 (240). Caspase-11 is thought to activate downstream signals caspase-1 and caspase-3, and thus, it may be important in both inflammation and apoptosis (241). It has been demonstrated that p38 MAPK mediates caspase-11 induction in microglia subjected to hypoxia and endotoxin exposure (150, 151). In our study, we observed increased procaspase-1, but decreased caspase-1 p20 and caspase-11 in castrated males and TRB-treated males compared to untreated males.

Myocardial apoptosis is responsible for the loss of cardiomyocytes and depression of myocardial function following I/R. Myocyte apoptosis in heart failure is reduced in women compared to men (242). Apoptosis may be mediated by either the extrinsic death receptor signaling pathway or the intrinsic mitochondrial control pathway (243). Caspases play a crucial role in each of these pathways. The activation of the intrinsic apoptotic pathway (the release of cytochrome *c* and the activation of caspase-9) during reperfusion, but not ischemia, has been observed in chick cardiomyocytes (244), and activation of caspase-8 and caspase-3 occurs in response to hypoxia or ischemia (245). However, no information exists regarding the influence of testosterone on myocardial proapoptotic signaling cascades following acute I/R. In our study, we observed decreased activation of pro-apoptotic signaling cascade (caspase-3, caspase-11) in castrated males and TRB-treated males compared to untreated male hearts following I/R, whereas anti-apoptotic Bcl-2 was increased in the latter group. Clinically, anabolic androgenic steroid abuse has been associated with myocardial ischemia and sudden cardiac death (246). Some studies have demonstrated that anabolic androgenic steroids induce injury and apoptosis in myocardial cells (205, 234) and skeletal muscle cells in culture (247). Our observations of decreased apoptotic signaling with testosterone depletion or testosterone receptor blockade are consistent with these studies.

One of the first large-scale studies of the cardioprotective role of estrogen investigated the ability of hormone replacement therapy to prevent secondary coronary events. The Heart and Estrogen/progestin Replacement Study (248) (HERS) was a randomized trial involving nearly 3,000 postmenopausal women under eighty years of age (mean of 66.7 years) who had a history of coronary artery disease. The women were treated with either placebo or combination hormone therapy (0.625 mg conjugated estrogen and 2.5 mg medroxyprogesterone daily). Interestingly, no significant differences in primary outcome (CVD death or nonfatal myocardial infarction) or secondary outcome (stroke, peripheral arterial disease, congestive heart failure, resuscitated arrest, unstable angina, or resuscitated arrest) were noted between the hormone treated groups and those treated with placebo after a four-year follow-up, despite a favorable effect of the hormone therapy on the lipid profile. It was found,

however, that more women in the hormone-treated group suffered coronary events during the first year of treatment than those taking the placebo. Women in the hormone treated group also experienced more thrombosis events such as pulmonary embolism. During years four and five of the study, however, hormone-treated women had fewer cardiovascular events. DM Herrington, *et al* (249) also investigated the ability of HRT (combination as well as unopposed estrogen) to provide secondary protection against cardiovascular (CV) events by administering unopposed, conjugated estrogen, estrogen and medroxyprogesterone, or placebo to postmenopausal women with coronary disease. It was concluded that HRT administration did not prevent the progression of atherosclerotic plaques, despite, once again, a favorable effect on the lipid profile. The Estrogen in the Prevention of Atherosclerosis Trial (250) was organized to assess estrogen's ability to attenuate the progression of subclinical atherosclerotic plaques, and the protocol included postmenopausal women with no history of CVD receiving unopposed doses of beta-estradiol to ensure no interference from progesterone. This trial concluded that unopposed estradiol treatment can slow the progression of subclinical atherosclerosis to much the same extent as traditional lipid-lowering therapy (250).

Other studies, along with HERS, investigating estrogen's preventative effects have also written of a seemingly higher "early-harm" risk of cardiovascular events during the first year of treatment in post-menopausal women taking hormone replacement therapy (44). These studies included not only secondary prevention studies, but primary prevention studies as well. One such primary prevention study is the Women's Health Initiative (224), which administered combination hormone therapy to healthy postmenopausal women. It was found (and the trial ceased) at the five-year follow-up due to a higher occurrence of stroke, cardiac events, thromboembolic events, and invasive breast cancer among those on a hormone regimen. It was determined by this trial that HRT should not be used as a primary preventative measure for cardiovascular disease. The Nurses Health Study (251), as well as observational studies that show evidence of early-harm for women who have prior CVD and are beginning hormone regimens (252, 253), supported the findings of HERS (254). Studies that were administering estrogen to men with coronary artery disease also were stopped early for safety consideration because those receiving hormone therapy were suffering more cardiac events than the control group (255). It is expected that this high-risk period for HRT use (the first year of therapy) is so pronounced because it is during this time that the events occur in those with pre-existing risk factors, such as those with a tendency to develop thromboemboli (252, 254). Several studies have documented that the net effect of estrogen on the coagulation system is procoagulant (249, 256). Another possible explanation for this finding could be that progesterone interferes with some of the acute beneficial effects of estrogen on vasoreactivity (257-259). Cannon, *et al* (260), found that estrogen administration tends to increase levels of matrix metalloproteinase-9, which is critical to T-cell migration and tends to promote lesion rupture. This could

imply that although the long-term increase in MMP-9 may be able to prevent the aggregation of matrix proteins, this acute effect of estrogen may put women with vulnerable plaques at risk during early use (260).

All that can be derived from these studies at this point is that although estrogen therapy may possibly be able to play a role in the prevention of the formation of the plaques associated with atherosclerosis or to slow progression during very early stages of the disease, HRT cannot repair damage that is already done and is not as effective at slowing progression at later stages of the disease. Therefore, the timing of the initiation of hormone treatment relative to age and menopause may be a crucial point to consider (261). There are a couple of possible explanations for this observation (261). First, early-stage atherosclerotic lesions are composed of less dense fibrous tissue than the later-stage lesions (262), and aging can decrease the expression of the estrogen  $\alpha$ -receptor, making the estrogen less able to exert its effects (263). Overall, more clinical studies are needed to adequately assess both the primary and secondary protective capabilities of estrogen. Special attention should be given to the nature and health status of the study participants, which form of HRT (combination or estrogen) and dosage that is given, and to the mode of administration (oral or transdermal), as these are often linked to differences in the beneficial or detrimental effects of estrogen therapy (254).

It is well-established that estrogen has favorable effects on some of the major risk factors for the development of atherosclerosis and therefore CVD. Although there is some concern as to estrogen's tendency to significantly increase plasma triglyceride levels (264), estrogen therapy has also been shown to reduce levels of low-density lipoprotein (LDL) and increase levels of high-density lipoprotein (HDL) in postmenopausal women. This restores the lipid profile back to one that resembles the profile of premenopausal women, who are at lower risk for CVD (265). Estrogen administration has also been shown to decrease levels of collagen production in rabbit aortas, which has implications in the formation of advanced atherosclerotic plaques (266). Additionally, Godsland and coworkers (267) demonstrated the estrogen may have a favorable effect on insulin resistance. Estrogen has also been targeted as having a role in the change in fat distribution that many women experience after menopause. Menopause usually results in an abdominal concentration of body fat (as opposed to peripheral), which is a risk factor for cardiovascular disease. HRT can prevent increases in abdominal fat and distribute it elsewhere, therefore having a favorable effect on cardiovascular risk (268). It is, however, estimated that the above-mentioned effects account for only about 30 to 50 percent of estrogen's cardioprotective effects (269). There has been intense investigation into the other possibilities for estrogen's effects. Postmenopausal women experience changes in the levels of many factors involved with hemostasis that put them at higher risk for thrombosis. Therefore much investigation has been directed at determining estrogen's effects on coagulation factors (232). Endothelial function and how estrogen may alter

vasoreactivity have also been investigated (232), as have the hormone's effects on inflammatory markers such as the cytokines, C-reactive protein, and the adhesion molecules (270).

Several studies are also investigating the effects of tamoxifen, an estrogen receptor modulator used to treat breast cancer (271), and raloxifene, an estrogen receptor modulator used in the treatment of osteoporosis (272). Tamoxifen, although it acts as an estrogen antagonist in the breast (273), often mimics that actions of estrogen in tissues other than the breast, and has also been shown to have favorable effects on lipid profiles (274). Raloxifene, although it also demonstrates an ability to lower low-density lipoprotein levels (275), does not demonstrate estrogen-like effects in the uterus or breast (272, 276).

Much research attention has been given to the effects of estrogen and hormone replacement therapy on blood coagulation and the formation of emboli, as these are major risk factors for cardiovascular disease and myocardial infarction (277), and it has been demonstrated that levels of several coagulation factors such as factors VII and VIII as well as fibrinogen increase after menopause (278-282). Much of the attention given to hormone therapy's impact on coagulation factors originated from the aftermath of the HERS study, during which the women in the hormone-treated group suffered more thromboembolic events (248, 283). Additionally, tamoxifen treatments have been linked to higher rates of thromboembolic events in women (271). Studies have produced conflicting conclusions concerning what effect estrogen has on coagulation and thrombosis, as hormone therapy effects the various fibrinolytic components differently (284). Because pro-inflammatory effects of estrogen have been found with D-dimer, metalloproteinase 9, and Factor VII, whereas anti-inflammatory effects have been found with fibrinogen, endothelial adhesion molecules, and plasminogen activator inhibitor-1, it is difficult to determine which effect will prevail (260, 285, 286). It is therefore challenging to assess whether estrogen's effects on the coagulation pathway have the power to explain some of estrogen's proposed cardioprotective effects. Much of the concern that estrogens and hormone replacement therapy may heighten risk for thromboembolism stems from the available data on the use of oral contraceptives and their tendency to increase such risk (287). Indeed, several studies have concluded that the use of oral contraceptives with high doses of estrogen is associated with a higher rate of thromboemboli formation (288). However, estrogens in hormone replacement therapy (HRT) have different effects on the separate components of the coagulation system, so many studies have assessed the effects of HRT on factors such as the level of fibrinolysis, the activation of coagulation, levels of plasminogen activator, and levels of plasminogen-activator inhibitor type 1 (287).

It has been shown that estrogen may exert anti-coagulant effects by interfering with platelet aggregation (287). After a blood vessel wall is injured, collagen fibers allow for platelets to adhere to the injury site, and the platelets simultaneously produce several compounds

involved with coagulation including a prostaglandin, thromboxane A<sub>2</sub>, which allows platelets to adhere to one another and acts as a potent vessel constrictor. Thromboxane A<sub>2</sub> is broken down to thromboxane B<sub>2</sub>, which serves as an indicator of platelet activity (287). Although it has been shown that pre-menopausal women produce significantly less thromboxane B<sub>2</sub> than age-matched men, women show a linear increase in the level of the prostaglandin during the post menopausal years whereas such an increase has not been demonstrated in men (287). This could partially explain the increased incidence of atherosclerosis and associated cardiovascular disease in post-menopausal women in comparison to pre-menopausal women. It was demonstrated that lipopolysaccharide-induced thromboxane B<sub>2</sub> levels in post-menopausal women were nearly restored to the levels seen in pre-menopausal women (levels reduced by 35%) after one year of treatment with HRT for both orally and transdermally administered estrogen, suggesting that HRT could possibly decrease platelet reactivity (289). The same study cited significantly reduced levels of tissue factor (TF) expression and decreased tumor necrosis factor release in response to HRT. Reduced levels of tissue factor lessen the ability of factor VIIa to bind to its receptor, a key step in the initiation of coagulation (289).

Nozaki, *et al* (277) also conducted a study on the effects of HRT on fibrinolytic components by administering unopposed conjugated equine estrogen (CEE) to women who had undergone hysterectomy and oophorectomy, combination hormone therapy (CEE and MPA) to a group of postmenopausal women, and a placebo to a control group. Levels of various fibrinolytic components (Factor VII, protein C, fibrinogen level, antithrombin III, plasminogen activator inhibitor-1 (PAI-1), and tissue type plasminogen) were measured at 1, 3, and 6 months of treatment—during the period of time when women are thought to be at the greatest risk for a thromboembolic event after starting hormone therapy. Whereas antithrombin III and tissue-type plasminogen levels remained stable throughout treatment, protein C activity increased roughly 10 percent in each treatment group, fibrinogen levels dropped nearly seven percent, and the levels of PAI-1 decreased by nearly sixteen percent. In both the cases of decreased levels, the unopposed CEE treatment group saw a slightly greater result than the combination therapy group, whereas the combination group saw the greater increase in protein C activity. The activity of factor VII increased by ten percent in the CEE group only. They conclude that with the exception of the raise in the factor VII levels among the CEE treated patients, HRT has no adverse effects on coagulation and fibrinolysis (277).

Further studies involving plasminogen activator inhibitor-1 have shown that postmenopausal women tend to have a higher level of PAI-1, a potent inhibitor of fibrinolysis, than premenopausal women, possibly putting them at higher risk for thromboemboli or cardiovascular events (290). Koh, *et al* (286), studied the effects of combination and unopposed estradiol therapy (orally and transdermally administered) on the levels of PAI-1 and on

the levels of D-dimer (a by-product of fibrinolysis). D-dimer levels increased proportionately with the decreasing PAI-1 levels. Orally-administered estradiol and estradiol combined with progestin led to a reduction in PAI-1 levels by approximately 50% as opposed to the transdermally-administered therapy, which showed no significant effect. This agrees with the findings of Kroon (291) and Gilbert (292) showing that passage of the hormone through the liver has an impact on estrogen's effect on PAI-1. In fact, it has been suggested that the liver, along with the endothelium, is a main source of PAI-1 (293). Van Kesteren *et al* (294) found that the administration of ethinylestradiol (100 µg/day) in combination with an anti-androgen compound to male→female transsexuals decreased both tissue plasminogen and PAI-1 levels. Sobel, *et al* (295) demonstrated that 17beta-estradiol affected plasminogen activator (tPA), which is necessary for fibrinolysis, in a biphasic dose-dependent manner using bovine aortic endothelial cells. Increased secretion of tPA from the endothelial cells was seen at lower, physiological concentrations of the hormone, whereas decreased levels of tPA were seen at higher concentrations of the hormones. The most pronounced stimulatory effect was seen at 10<sup>-12</sup> mol/L estradiol. A similar, reciprocal pattern was seen with estrogen's effect on the presence of PAI-1 antigen. Inhibition was seen at 10<sup>-12</sup> mol/L estrogen, whereas stimulation was seen at 10<sup>-7</sup> mol/L. Similar results were seen with progesterone. Because fibrinolysis is a mechanism balanced by the interaction of tPA and PAI-1, and because studies have shown that physiological levels of estradiol tend to decrease endothelial PAI-1 levels and increase tPA levels (290), these results support that restoring estrogen to normal physiological levels after menopause may be beneficial in the potentiation of fibrinolysis.

Contrary to what was laid out above about estrogen's potential effects on fibrinolysis, many studies have demonstrated that estrogen activates the coagulation system (278). Aside from the data that state oral contraceptives can increase the risk for thromboembolic events (279, 288) and the knowledge that HERS participants treated with hormones showed a higher incidence of thrombosis events, Caine *et al* (296) administered 0.625 mg or 1.25 mg CEE to healthy postmenopausal women and noted a dose-dependent increase in thrombin generation and fibrinopeptide A. Levels of antithrombin and protein S, inhibitors of thrombin activation and formation, respectively, were also shown to decrease. Scarabin *et al* (297) found that a regimen of 2mg oral estrogen with cyclic progesterone increased levels of prothrombin fragments 1 and 2 and decreased antithrombin activity in healthy postmenopausal women, whereas no result was seen with the transdermal regimen. Current evidence indicates that while estrogen therapy seems to have a favorable effect on fibrinolysis, it has a simultaneous unfavorable effect on the activation of coagulation. This led Koh *et al* (278) to wonder if the pro-fibrinolytic effects of estrogen were a secondary response to the coagulation activation induced by estrogen. 0.625mg of CEE was administered to nine postmenopausal women. After one month, eight women had lower PAI-1 activity

and seven had marginally greater t-PA activity. The ratio of t-PA to PAI-1 activity, as a measure of fibrinolytic potential (298), increased six to 650 percent in all nine women, whereas no significant change for the group was seen in coagulation activation markers prothrombin fragments 1 and 2 levels or in levels of thrombin-antithrombin. This study, although small, suggests that coagulation activation by estrogen is not necessary for the increased potentiation of fibrinolysis (278).

Once the monocytes have migrated into the vessel wall and converted to tissue macrophages, they, along with the platelets, smooth muscle cells, endothelial cells, and lymphocytes, are capable of producing a wide variety of molecules such as cytokines and growth factors that aid in the progression of atherosclerotic lesions as they act in a paracrine, juxtacrine, and autocrine manner and disrupt endothelial function (299). These growth factors and the cytokines play central roles in the development of atherosclerosis, as they are involved in nearly every aspect of the inflammatory process; they are often the stimulators of cell proliferation, migration, adhesion, and regulators of vasomotor tone (300). The interactions that estrogen has with these molecules therefore may provide insight into estrogen's cardioprotective effects. Although there is no definitive statement as to what effect estrogen has on the cytokines such as tumor necrosis factor (TNF) and interleukin-1 (IL-1), estrogen receptors have been found on human monocytes, suggesting that estrogen may modulate cytokine release (301).

A link between estrogen levels and IL-1 levels was discovered by Cannon and Dinarello (302) who found that IL-1 activity was increased in women immediately post-ovulation. However, it was then suggested that conjugated estrogens had no effect on IL-1 release in bone marrow aspirates (303). Patients who had undergone a complete hysterectomy showed higher levels of IL-1 than other postmenopausal women who were on a hormone replacement regimen (304). This decrease in IL-1 expression is apparently mediated by estrogen at the transcriptional level, as lower levels of IL-1 mRNA were found in human monocytes and peritoneal macrophages (305). IL-6 protein and its transcripts have been identified in atherosclerotic lesions as well, and it has been shown that IL-6 can induce monocyte chemotactic protein-1 in mononuclear cells (306). It was demonstrated that administration of 17 $\beta$ -estradiol to ex-vivo aortic tissue of male apolipoprotein-E knockout mice reduced IL-6 secretion by 50 percent (307). Although possible, it is unlikely that the lower levels of testosterone were responsible for the decrease in IL-6 secretion. This suggests that the suppression of IL-6 secretion from the macrophages may serve to protect by inhibiting autocrine stimulation of chemokines involved in cell recruitment (307).

Upon investigating the effects of estradiol on the trans migratory behavior and cytokine profile of autoreactive T-cells, YC Zang and colleagues (308) found that estradiol can inhibit T cell transmigration at an estradiol level mimicking pregnancy. Estradiol also

upregulated the T cell secretion of IL-10 and decreased secretion of TNF in a non antigen-dependent manner. It was also shown that estradiol can inhibit nuclear transcription factor kappa B (NF-kappaB) (308), which is typically activated by injury and the associated cytokines (300). Because NF-kappa B regulates a number of genes involved in the immune response, it may be through NF-kappa B that estrogen exerts some of its effects on the markers of inflammation. For example, Selzman, *et al* (309) demonstrated that the mitogenic effect of TNF on human vascular smooth muscle cells is dependent on NF kappaB activation, as the administration of an NF kappaB inhibitor led to a decrease in vascular cell proliferation and a decrease in the levels of IL-6 as well. Zang, *et al* (308) additionally noted a decreased level of MMP expression in T-cells treated with estradiol; however, several other studies have found HRT to be associated with higher MMP-9 levels. Hormone replacement therapy was found to be associated with higher levels of MMP-9, which digests the matrix and makes lesions more prone to rupture (310), in a study by Cannon, *et al* (311). Zanger, *et al* (260) also reported that women on hormone replacement therapy tended to have higher levels of serum MMP-9 than those women taking a placebo.

Aune and coworkers (289) found that after twelve months of HRT, the levels of TNF produced by lipopolysaccharide-stimulated macrophages had decreased significantly in both patients receiving estrogen orally and those receiving treatment transdermally. Pacifici, *et al* (304) determined that the spontaneous macrophage secretion of TNF and IL-1 was decreased in women taking hormone therapy in comparison to postmenopausal women not on a regimen. Other studies, such as that conducted by Stock, *et al* (281), found that estrogen had little or no effect on spontaneous macrophage IL-1 secretion. Zhang and coworkers (312) additionally found that pre-incubating murine bone marrow-derived macrophages in 17 $\beta$ -estradiol significantly inhibited lipopolysaccharide-induced TNF release. Kamada *et al* (313) studied the levels of fifteen cytokines in postmenopausal women on HRT and those who were not on a regimen. Although higher levels of TNF and lower levels of transforming growth factor beta1 (TGF) were found in the late postmenopausal women, no significant differences were found in cytokine levels between the group on HRT and the group not receiving hormones. The women on the hormone therapy did experience a significant increase of colony-stimulating factor, which is known to decrease serum cholesterol (313).

Insulin-like growth factor, basic fibroblast growth factor, PDGF, and epidermal growth factor are the main growth factors involved in the proliferation of smooth muscle cells, endothelial cells, and matrix proteins (299), and therefore have been pinpointed at possibly playing an important role in the development of atherosclerotic lesions. It has been shown that one of the earliest nuclear events associated with the action of growth factors is the induction of the proto-oncogene *c-fos*. Breast cancer research has shown that estrogen may upregulate this oncogene and cause cell proliferation synergistically with IGF (314). Although most studies investigating the linkage

between estrogen and growth factor signaling pathways have been conducted using tissue other than vascular tissue (299), it has been shown that postmenopausal women on HRT show lower levels of serum IGF-1 when compared to those not on a hormone regimen (315). In a study on the effects of estrogen on transplant atherosclerosis, Saito *et al* (316) determined that estrogen administration to rats undergoing cardiovascular transplantation inhibited the typical increase in insulin-like growth factor-1 that occurs during the onset of posttransplantation atherosclerosis. The estrogen regimen (20 micrograms/kg daily for 2 days prior to surgery and until euthanasia) also decreased expression of the major histocompatibility complex class II antigen (316). Recent findings have shown that TNF can downregulate insulin-like growth factor-1 and upregulate insulin-like growth factor binding protein-3 in vascular smooth muscle cells (224). Selzman, *et al* (299) found that estrogen replacement could decrease the aortic accumulation of TGF in sheep, indicating that estrogen may modulate the growth factor's impact on VSMC proliferation. The *in vitro* incubation of human aortic smooth muscle cells with estrogen showed increased levels of TGF secretion and increased potentiation of TGF's inhibition of VSMC proliferation.

Shanker, *et al* (317), determined that the addition of estrogen to cells from the monocyte/macrophage THP-1 line treated with 12-O-tetradecanoylphorbol 13 acetate (TPA) increased the level of PDGF-A transcript by 61 and 190 percent after 48 and 96 hours, respectively. Results for PDGF-B were not conclusive. The addition of estrogen to cells stimulated with TPA as well as with lipopolysaccharide caused no significant change in the amount of the PDGF-A transcript. It has been shown that the differentiation of THP-1 line cells as induced by TPA is similar to the transformation of monocytes to macrophages (138, 318), which is crucial during initiation and progression of atherosclerosis. Due to the fact that the actual role of PDGF in atherogenesis is still not clear, it is difficult to assess whether this effect of estrogen is cardioprotective or not. It has been suggested that PDGF-A may play an important role in early-stage atherosclerotic lesions, and that PDGF-A can inhibit the migratory activity of cultured smooth muscle cells (319). Estrogen's upregulation of PDGF-A (317) in the context of this study may be a mechanism by which it decreases the level of cell migration early in the lesion formation process. This is difficult to confirm, as most studies investigating the proliferative effects of PDGF involve late-stage lesions (317). It has been previously shown that estrogen can inhibit the migration of vascular smooth muscle cells *in vitro* (320). Kikuchi (321) *et al*, however, concluded that the addition of either estradiol or estrone sulfate to cultured VSMC decreased transcript levels for PDGF-A, IL-1, and IL-6. These results were similar to those of Okubo, *et al* (321) Dai-Do, *et al* (272), who found that PDGF-BB stimulated DNA synthesis in VSMCs cultured from both postmenopausal females and age-matched males, and that the administration of estradiol inhibited the ability of both bFGF and PDGF to upregulate DNA synthesis in both males and females. Saito, *et al* (316) additionally found that the administration of 20 µg/kg per day to male rats

undergoing aortic allograft transplantation diminished the post-transplant increases in PDGF, IGF-1, and bFGF that were seen with the placebo group.

Another study conducted by Somjen *et al* (322) showed that 17β estradiol administration had differential effects on DNA synthesis in vascular smooth muscle cells and in endothelial cells. Low doses (0.3 nmol/L) of estradiol stimulated DNA synthesis whereas higher doses (30 nmol/L) inhibited DNA synthesis as measured by [<sup>3</sup>H] thymidine incorporation in VSMCs. In contrast, estradiol increased DNA synthesis in a dose-dependent manner in endothelial cells (E304 cells). High doses of estradiol inhibited PDGF and IGF-1 induced DNA synthesis in the VSMCs, whereas estradiol increased the growth factor-induced DNA synthesis in endothelial cells. It is suggested that estrogen's inhibition of VSMC proliferation and stimulation of DNA synthesis in the endothelium may characterize its role in vascular remodeling (322). When cells were incubated with tamoxifen and raloxifene, the modulators had the same effect on DNA synthesis as estrogen in both cell types. When cells were incubated with a combination of 0.3 nmol/L estradiol and either tamoxifen or raloxifene, the estradiol-induced increase in DNA synthesis was inhibited in endothelial cells but not in VSMCs (322).

Another marker of inflammation, C-reactive protein (CRP), has been shown to be an independent risk factor for CVD in both sexes (270, 323). High serum levels of this marker was the strongest overall predictor for cardiovascular disease in women overall (270). Vicenzino Paseri and coworkers (324) investigated the effect of C-reactive protein on several other factors in the inflammatory response, including vascular cell adhesion molecule (VCAM-1), intercellular adhesion molecule (ICAM-1), and E-selectin. Incubation of human coronary artery endothelium in human serum with CRP led to an increase in the expression of ICAM-1 and VCAM-1 from 5 µg/ml to 50 µg/ml and also increased the expression of E-selectin. CRP also induces tissue factor (325). It is unclear at this point whether CRP itself plays a direct role in atherosclerosis, or whether it simply serves as a marker of inflammation. (324) A few studies have concluded that HRT administered to postmenopausal women actually raises the level of CRP. Ridker and colleagues (326) conducted a study of 493 healthy postmenopausal women and found that those on HRT generally had higher levels of CRP than those not taking supplemental hormones. During the PEPI trial, (285) levels of CRP rose over the three-year period in women taking each of the four different hormone regimens (progesterone included). The serum concentration of CRP correlates to that of TNF-α (270) and IL-6, which is the main inducer of hepatic CRP production (270, 327). It was found by Walsh, *et al* (328) the increases in CRP levels in response to HRT administration were indeed higher for those women with high IL-6 levels, and that raloxifene, an estrogen receptor modulator, is not associated with higher CRP levels in postmenopausal women. However, this increased level of CRP was also seen in women after their IL-6 levels had decreased, suggesting that estrogen may exert its effects on CRP production by acting on the liver directly (328).



Estrogen has also been shown to affect various adhesion molecules and chemotactic proteins that are involved in the pathogenesis of atherosclerosis. Caulin-Glaser, *et al* (329) incubated human umbilical vein endothelial cells in estradiol and found that it strongly inhibited IL-1 mediated membrane E-selectin and vascular cell adhesion molecule-1 induction. Estradiol treatment also inhibited intercellular adhesion molecule-1 hyperinduction. This could possibly be a mode by which estrogen exerts its cardioprotective effects, as lower levels of these molecules lessen the ability of leukocytes and other inflammatory molecules to bind to endothelial cell injury sites. Caulin-Glaser (330) also found during a human study that men with CAD, as well as postmenopausal women with CAD not on a hormone regimen, had higher levels of circulating cellular adhesion molecules in comparison to the women with CAD taking estrogen. Those postmenopausal women with CAD taking estrogen, however, had increased levels of VCAM-1 than those not taking estrogen, as did the premenopausal women with CAD. In a study involving healthy individuals, Van Baal *et al* (331) found that the administration of unopposed estrogen or the combination of estrogen/trimegestone for twelve weeks to postmenopausal women without CAD led to decreases in ICAM-1, VCAM-1, and thrombomodulin. A decrease in endothelin levels was seen only in the group given combination therapy. There is also evidence that the effect that HRT or estrogen treatment may have on the expression of various adhesion molecules may be a function of its mode of administration. Seljeflot *et al* (332) reported that after twelve months of HRT, lower levels of E-selectin, ICAM-1, and VCAM-1 were seen in postmenopausal women with CAD who were receiving the treatment transdermally rather than orally. Oger (333) found that the lowering effect of HRT on ICAM-1 was dependent on treatment duration for those on a transdermal regimen, whereas this was not seen among those taking HRT orally.

Additionally, it was discovered in a study using human aortic endothelial cells and THP-1 monocytes that 48 hours of incubation with 17 $\beta$  estradiol decreased monocyte adhesion by 28 percent, as well as decreased cytokine (TNF, IL-1 or both)-induced adhesion by 30-35 percent after a four hour addition of cytokines. Of importance, however, was that the estradiol could not inhibit endothelial cell-monocyte adhesion if the cells were incubated with the cytokines for 24 hours (334). Conversely, Zhang, *et al* (221) found that both estradiol and testosterone increase the expression of E-selectin and VCAM-1 in a receptor-mediated manner in human umbilical vein endothelial cells incubated simultaneously with TNF and the steroid hormone. The conclusion that estradiol upregulates expression of adhesion molecules as induced by TNF is supported by the work of Cid *et al* (54). It was suggested, however, that estradiol could decrease expression of monocyte-chemotactic protein-1 (MCP-1) in human artery endothelial cells. Estradiol decreased MCP-1 mRNA expression by 30 percent, although this effect was seen only in the arterial cells and not in umbilical vein cells (335). This suggests that estrogen may function to attenuate macrophage recruitment.

It has long been suspected that at least some of estrogen's proposed ability to protect from the progression of atherosclerosis and cardiovascular disease comes from its effect on endothelial function, as the endothelium and its metabolically active cells are crucial in the development of atherosclerosis (336). The endothelium serves as a physical barrier of protection for the underlying components of the blood vessel, as well as a "docking point" for monocytes, leukocytes, and neutrophils (300). It also functions to prevent clotting and promote smooth blood flow through the vessel (300). The endothelium is also largely responsible for regulating vascular tone, as it is a source of vessel constrictors such as thromboxane, free radicals, endothelins, and cytokines as well as of vessel relaxants such as prostacyclin, and nitric oxide. (300) It was also recently documented that estradiol may lessen the extent of TNF-induced apoptosis among endothelial cells (337). As mentioned previously, Aune *et al* (289) found that after one year of hormone replacement therapy, there was a significant decrease in the production of the lipopolysaccharide-induced prostaglandin thromboxane B<sub>2</sub>, a potent vasoconstrictor.

Similarly, it has been suggested that estrogen can regulate the secretion of the prostaglandin prostacyclin, which acts as a vasodilator and inhibitor of platelet aggregation (299), although the results have been contradictory. Higher levels of 6-keto-prostaglandin were noted in the uterine arteries of premenopausal, but not postmenopausal women (338). Findings supporting this data have been obtained by tissue culture (339). O'Sullivan and colleagues (340) placed ovariectomized female monkeys on an atherogenic diet and treated a select group with Premarin and found that there was an 165 percent increase in arachidonate-induced prostacyclin production by vessels treated with estrogen, although there was no difference in the basal prostacyclin release. Additionally, prostacyclin production was inversely related to plaque size (340). Ospina *et al* (341) found that estrogen treatment increases cerebrovascular prostacyclin production in the rat by increasing levels of cyclooxygenase-1 as well as prostacyclin synthase. It was also found that estrogen can stimulate prostacyclin synthesis in ovine pulmonary artery endothelium (342). Conversely, Redmond and coworkers (343) found that estradiol had an inhibitory effect on flow- and acute hypoxia-induced prostacyclin release from endothelial cells. This was supported by a more recent study by Christodoulakos *et al* (335) which concluded that 17 $\beta$  estradiol and raloxifene had a lowering (unfavorable) effect on prostacyclin levels.

van Kesteren *et al* (294) conducted a study using a subset of male  $\rightarrow$  female and female  $\rightarrow$  male transsexuals in order to assess the effects of transgender hormone therapy on coagulation factors as well as on markers of endothelial functioning such as big endothelin-1 (ET-1). Baseline levels were similar for the vasoconstrictor ET-1 between groups. The administration of estrogens to the M  $\rightarrow$  F transsexuals led to significant decreases in the level of big ET-1, whereas the F  $\rightarrow$  M transsexuals receiving testosterone treatment saw an increase in ET-1. Other human studies have also shown that estradiol administration can attenuate ET-1 induced vessel constriction (335, 344,

345). Lee and colleagues (345) concluded that intracoronary treatment with estrogen decreased levels of ET-1 in post-coronary angioplasty patients. Saitta, *et al* (344) found that estrogen therapy with norethisterone acetate as well as raloxifene therapy lowered ET-1 levels in healthy postmenopausal women after a sixth-month trial. It was noted by Jhund, *et al* (346), however, that estrogen treatment administered to women with coronary artery disease lost its ability to modulate ET-1-induced vasoconstriction after three months. In addition to its action as a vasoconstrictor, ET-1 has also been shown to promote endothelial cell adhesion molecule expression (347).

It has also been suggested that estrogen may have an effect on the production of nitric oxide, a vasodilator that is also capable of preventing platelet aggregation, leukocyte chemotaxis, and T-cell proliferation (348) within the endothelium. Wagner and colleagues (349) investigated the hypothesis that estrogen exhibited an anti-atherosclerotic effect by shifting the balance between nitric oxide (NO) and the free radical superoxide in the vessel wall. They showed the 17beta estradiol caused a concentration-dependent decrease in the expression of NADPH oxidase, an enzyme required for the generation of superoxide. 17beta estradiol also increased the production of nitric oxide in the vessel wall by a factor of two over the same concentration range. Incubation of the human vein umbilical endothelial cells with the estrogen also decreased the ability of the cells to produce superoxide upon phorbol ester stimulation. It appears estradiol may act as an antioxidant which can improve the NO/superoxide ratio and therefore normalize the response to injury. Saito *et al* (348) concluded that while estrogen therapy can suppress cytokine-inducible nitric oxide synthase (iNOS) (possibly via estrogen's effects on NF kappa-B) in macrophages and smooth muscle cells, it can also upregulate endothelial nitric oxide synthase (eNOS). In this case, estrogen is indirectly blocking the cytokines' ability to activate the inflammatory pathway. The study by Christodoulakos, *et al* (335) found that both the administration of estrogen plus norethisterone acetate and estrogen and norethisterone plus raloxifene decreased ET-1 levels and increased the nitric oxide/ET-1 ratio to produce a possibly cardioprotective effect. The study by Saitta (344) also concluded that estrogen treatments and raloxifene treatments can increase NO production.

Another recent study focused in on the questions as to whether estrogen's cardioprotective effect is a function of its effect on nitric oxide production (350). After treating human aortic endothelial cells with 17beta estradiol and measuring eNOS expression after 24 hour and 7 day regimens using intro-arterial infusion of interbranchial N<sup>G</sup>-monomethyl-L-arginine, it was determined that there was no change in eNOS expression after acute or chronic estradiol treatment. A study by the same group (350) in which the vasoconstrictor response to a substrate inhibitor of nitric oxide synthase was measured in 10 healthy postmenopausal women receiving 80µg transdermal estrogen daily for four weeks also showed no difference in vasoconstrictor responses as mediated by nitric oxide.

It has also been shown that estrogen administration may effect the vasodilator response to acetylcholine (350). There is evidence that oestrogen modulates the vasodilator response to acetylcholine in rabbit aortas, (351) and that continuous estrogen administration can reduce the acetylcholine-induced vasoconstriction in atherosclerotic ovariectomized monkeys (352).

Collins, *et al* (353) demonstrated that estrogen could decrease or abolish the amount of vasoconstriction in response to acetylcholine (Ach) in post-menopausal women with coronary artery disease, and that this was a phenomena seen in females, but not in males. This attenuation of Ach-induced vasoconstriction in women by estrogen was also supported by the work of Reis, *et al* (354).

Estrogen has also been shown to have some protective capabilities against ischemia-reperfusion injury in various organs. Shi *et al* (207) used ovariectomized rats to assess estrogen's ability to protect from cortical ischemia-reperfusion injury by inducing one hour of middle cerebral artery occlusion and using sequential diffusion weighted MRI to obtain resultant lesion size. It was shown that while both the groups of ovariectomized rats (one receiving 100µg/kg estrogen 2 hours prior to the procedure, the other one not receiving estrogen) developed similar lesions in response to ischemia, the cortical-area lesions of the rats receiving estrogen decreased in size by 50-60 percent during reperfusion, whereas such a decrease was not seen in the rats not on an estrogen regimen. This and other studies (355-357) suggest that estrogen may act as a neuroprotective agent and selectively protect cortical tissue from ischemic damage during occlusion and therefore may be useful in the treatment of stroke. Additionally, Nonaka *et al* (358) discovered that the administration of estradiol reduced leukocyte accumulation by 35.7 percent 24 hours post-retinal ischemia reperfusion. There was also evidence that the administration of the estradiol lessened the extent of retinal damage 168 hours post-reperfusion.

Various animal studies have also studied the effects of estrogen on ischemia reperfusion injury in cardiac muscle. Squadrito, *et al* (359) exposed rats treated with 17beta estradiol and untreated rats to one hour of left coronary artery occlusion followed by one hour of reperfusion, using various markers of injury such as levels of necrosis, creatinine phosphokinase, TNF, and ICAM-1 to indicate the level of damage. Administration of estradiol to rats 5 minutes after the induction of injury decreased levels of necrosis, decreased levels of macrophage and serum TNF as well as creatinine phosphokinase activity, and prevented an increase in ICAM-1 staining. Zhai, *et al* (360) found that hearts from female, ovariectomized rats given estrogen supplements and from sham-operated female rats had higher coronary flow rates and left ventricular pressures as well as higher nitrate production post-ischemia/reperfusion than ovariectomized rats not undergoing hormone treatment. However, nitrate production and coronary flow rate were not consistently different throughout reperfusion. The same investigators

also found the dietary phytoestrogens may lessen the extent of ischemia-reperfusion injury in female rats (361). McNulty, *et al* (362) found, however, no difference in infarct size or cardiac function between estrogen treated rats and untreated rats after ischemia-reperfusion injury irrespective of gender. It has also been shown through experiments with canine hearts that estrogen treatment may limit the size of infarction as mediated by activation of mitochondrial ATP-sensitive potassium channels in myocardial tissue in both male and female dogs (97).

Yet another recent path of study has focused on estrogen's ability to modulate the cardiac heat shock protein 70, whose expression can be stimulated by exercise (363). Hsp 70, which encodes intracellular proteins that can prevent protein abnormalities and serve in cytoprotection (363), has been found to have the ability to protect from cardiac injury and even improve ATP synthesis (364, 365). A close temporal relationship has also been documented between the accumulation of Hsp 70 and recovery from ischemia-reperfusion injury (366). Paroo, *et al* (367) found that male rats exercised on treadmills experienced a two-fold increase in the expression of Hsp 70, whereas this increase was not seen in exercised female rats. Additionally, ovariectomized female rats also had a significant increase in Hsp 70 levels, and the administration of supplemental estrogen to these rats reversed that effect and was accompanied by lower levels of exercise-induced Hsp 70. It was also concluded that although cardiac cells contain both estrogen receptors alpha and beta, that this is not the mechanism by which estrogen mediates the Hsp 70 expression as treatment with tamoxifen (and estrogen antagonist) did not change the response in comparison to controls. Additional findings that 17alpha estradiol (which does not activate receptors) also showed the ability to inhibit exercise induced expression of Hsp70 further supports that estrogen does not mediate Hsp 70 via its receptors. (367) The authors speculate that estrogen may mediate the response through a membrane-stabilizing mechanism, and that although females do show less of a stress response to exercise, this can be disadvantageous as it is an indication that certain protective forces against ischemia-reperfusion injury are not initiated (366). This could partially explain why premenopausal women who do suffer cardiac events have a higher mortality rate than men who suffer similar events (368).

Due to the apparent rise in cardiac events in postmenopausal women as compared to premenopausal women, the vast majority of research involving the steroid sex hormones and influences on cardiac functioning have focused on estrogen and its possible cardioprotective role. However, several of the studies investigating the effects of estrogens on factors involved in the development of atherosclerosis and cardiovascular disease also conducted trials using testosterone or its derivatives, as testosterone's effects are not exclusively immunosuppressive (43). For example, Somjen *et al* (322) concluded that dihydroxytestosterone (DHT) had very similar effects to those of estrogen on DNA synthesis in endothelial cells and vascular smooth muscle cells. DHT dose-dependently increased DNA synthesis in endothelial cells as measured

by [<sup>3</sup>H] thymidine incorporation. In VSMCs, testosterone enhanced DNA synthesis at low concentrations (3 nmol/L) and inhibited synthesis at higher concentrations (300 nmol/L).

The androgen antagonist flutamide inhibited DHT's actions on both endothelial and VSM cells. Van Kesteren *et al* (294) studied female → male transsexuals taking testosterone hormone therapy and found significant increases in endothelin-1 levels. Levels of plasminogen activator and PAI-1 did not change. Additionally, Sobel and colleagues (295) found that incubation of bovine aortic endothelial cells with testosterone tended to increase tissue plasminogen activator at low concentrations of hormone ( $10^{-12}$  M) and decrease its secretion at higher concentrations ( $10^{-7}$  M). Testosterone at a concentration of  $10^{-7}$  mol/L also increased the secretion of PAI-1. Webb, *et al* (369) found that intracoronary infusion of testosterone improves coronary blood flow possibly by inducing vessel dilation in men with coronary artery disease, although dilation was shown to be reduced in women taking high doses of androgens (370). Studies have also found that higher levels of testosterone in men are associated with higher levels of high density lipoprotein, contrary to popular belief.

Zhang, *et al* (221) reported that testosterone increases the TNF- $\alpha$  induced expression of VCAM-1 and E-selectin in endothelial cells by a receptor-mediated mechanism. This report agrees with the findings of McCrohon, *et al* (371), who found that dihydroxytestosterone increases the expression of endothelial VCAM-1. A conflicting report by Mukherjee, *et al* (372) proposed that testosterone inhibited TNF induced VCAM-1 expression in human umbilical vein endothelial cells by conversion to estradiol via the enzyme aromatase. They found that dihydroxytestosterone, an unaromatizable testosterone, had no effect on VCAM-1 mRNA expression, whereas testosterone administration (30nM-1 $\mu$ M) decreased VCAM-1 expression in a dose-dependent manner. Because the addition of anastrozole (an aromatase inhibitor) lessened testosterone's inhibitory effect on VCAM-1 expression, it was concluded that testosterone exerts its effect on the expression of VCAM-1 by converting to estradiol via aromatase present in endothelial cells (372). Dihydroxytestosterone has also been linked to increases in platelet accumulation (373).

## 7. SUMMARY AND PERSPECTIVE

The known mechanisms of myocardial injury are becoming increasingly complex. Not long ago, mechanisms of necrosis and ion dyshomeostasis were appreciated as the primary culprits in myocardial injury. Our progressive understanding of the mechanisms of apoptosis, stunning, hibernation, and preconditioning has served to bring us closer to enhanced myocardial preservation but, at the same time, has made understanding injury more complex and difficult. Although estrogen and testosterone have long been respected for their potent effects, our recent understanding of acute inflammation following ischemia has lit the spotlight on another theater in which these players are active. Testosterone appears to

be a potent promoter of proinflammatory and proapoptotic signaling processes in the heart and estrogen appears to have the counter effect. The results of our studies, and those of many of our colleagues around the country, most notably Dr. Chaudry, have revealed that there is much work to do in further understanding the role that sex hormones may have in altering the course of inflammation during ischemia, sepsis, and trauma.

### 8. ACKNOWLEDGEMENT

This work was supported in part by NIH R01GM070628, the Clarian Values Fund, the Showalter Trust, and the Cryptic Masons Medical Research Foundation. I thank Drs. M. Wang, B. Tsai, J. Pitcher, A. Kher, and K. Meldrum for their commitment, dedication, and hard work.

### 9. REFERENCES

1. Wang M, L Baker, BM Tsai, KK Meldrum, and DR Meldrum: Sex Differences in the Myocardial Inflammatory Response to Ischemia/Reperfusion Injury. *Am J Physiol Endocrinol Metab* 288, E321-E326 (2005)
2. Wang M, BM Tsai, A Kher, LB Baker, GM Wairiuko, and DR Meldrum: The Role of Endogenous Testosterone in Myocardial Proinflammatory and Proapoptotic Signaling after Acute Ischemia-Reperfusion. *Am J Physiol Heart Circ Physiol* 288, H221-H226 (2005)
3. Pitcher JM, BM Tsai, M Wang, RD Nagy, and DR Meldrum: Is the preconditioning threshold higher in females? *J Surg Res* In press (2004)
4. Mendelsohn ME and RH Karas: The protective effects of estrogen on the cardiovascular system. *N Engl J Med* 340, 1801-1811 (1999)
5. Meldrum DR: Tumor necrosis factor in the heart. *Am J Physiol* 274, R577-595 (1998)
6. Meldrum DR: Vascular endothelial growth factor, polycystic ovary syndrome, and ovarian hyperstimulation syndrome. *Fertil Steril* 78, 1170-1171 (2002)
7. Meldrum DR, A Ayala, and IH Chaudry: Energetics of defective macrophage antigen presentation following hemorrhage. *Surgery* 112, 150-158 (1992)
8. Meldrum DR, A Ayala, and IH Chaudry: Mechanism of diltiazem's immunomodulatory effects following hemorrhage and resuscitation. *Am J Physiol* 265, C412-C421 (1993)
9. Meldrum DR, A Ayala, and IH Chaudry: Energetics of lymphocyte "burn-out" in late sepsis: Adjuvant treatment with ATP-MgCl<sub>2</sub> restores energetics and decreases lethality. *J Surg Res* 57, 55-61 (1994)
10. Meldrum DR, A Ayala, MM Perrin, W Ertel, and IH Chaudry: Diltiazem restores IL-2, IL-3, IL-6, and IFN-gamma synthesis and decreases host susceptibility to sepsis following hemorrhage. *J Surg Res* 51, 158-164 (1991)
11. Meldrum DR, A Ayala, P Wang, W Ertel, and IH Chaudry: Association between decreased splenic ATP levels and immunodepression: amelioration with ATP-MgCl<sub>2</sub>. *Am J Physiol* 261, R351-357 (1991)
12. Meldrum DR, BS Cain, JC Cleveland, Jr., X Meng, A Ayala, A Banerjee, and AH Harken: Adenosine decreases post-ischaemic cardiac TNF-alpha production: anti-inflammatory implications for preconditioning and transplantation. *Immunology* 92, 472-477 (1997)
13. Meldrum DR, JC Cleveland, X Meng, BC Sheridan, A Banerjee, and AH Harken: Hemorrhage induces acute cardioadaptation to ischemia reperfusion by an  $\alpha_1$ -adrenoceptor mediated, protein synthesis independent mechanism. *Am J Physiol* 272, R718-R725 (1997)
14. Meldrum DR, JC Cleveland, MB Mitchell, RT Rowland, A Banerjee, and AH Harken: Constructive priming of myocardium against ischemia-reperfusion injury. *Shock* 6, 238-242 (1996)
15. Meldrum DR, JC Cleveland, MB Mitchell, BC Sheridan, F Robertson, AH Harken, and A Banerjee: Protein kinase C mediates Ca<sup>2+</sup> induced cardioadaptation to ischemia-reperfusion injury. *Am J Physiol* 271, R1718-R1726 (1996)
16. Meldrum DR, JC Cleveland, EE Moore, DA Partrick, A Banerjee, and AH Harken: Adaptive and maladaptive mechanisms of cellular priming. *Ann Surg* 226, 587-598. (1997)
17. Meldrum DR, JC Cleveland, RT Rowland, A Banerjee, and AH Harken: Calcium induced inotropy is in part mediated by protein kinase C. *J Surg Res* 63, 400-405 (1996)
18. Meldrum DR, JC Cleveland, RT Rowland, A Banerjee, AH Harken, and X Meng: Early and delayed preconditioning: Differential mechanisms and additive protection. *Am J Physiol* 273, H725-H733 (1997)
19. Meldrum DR, JC Cleveland, BC Sheridan, X Meng, F Robertson, BS Cain, AH Harken, and A Banerjee: Protein kinase C isoform diversity in preconditioning. *J Surg Res* 69, 183-187 (1997)
20. Meldrum DR, JC Cleveland, BC Sheridan, RT Rowland, A Banerjee, and AH Harken: Differential effects of adenosine preconditioning on the post-ischemic rat myocardium. *J Surg Res* 65, 156-164 (1996)
21. Meldrum DR, JC Cleveland, BC Sheridan, RT Rowland, A Banerjee, and AH Harken: Cardiac surgical implications of calcium dyshomeostasis in the heart. *Ann Thorac Surg* 61, 1273-1280 (1996)
22. Meldrum DR, JC Cleveland, BC Sheridan, RT Rowland, A Banerjee, and AH Harken: Cardiac

preconditioning with calcium: Clinically accessible myocardial protection. *J Thorac Cardiovasc Surg* 112, 778-786 (1996)

23. Meldrum DR, JC Cleveland, BC Sheridan, RT Rowland, CH Selzman, A Banerjee, and AH Harken: Alpha-adrenergic activation of myocardial NFkB during hemorrhage. *J Surg Res* 69, 268-276 (1997)

24. Meldrum DR, CA Dinarello, JC Cleveland, Jr., BS Cain, BD Shames, X Meng, and AH Harken: Hydrogen peroxide induces tumor necrosis factor alpha-mediated cardiac injury by a P38 mitogen-activated protein kinase-dependent mechanism. *Surgery* 124, 291-296; discussion 297 (1998)

25. Meldrum DR, CA Dinarello, JC Cleveland, BS Cain, BD Shames, X Meng, and AH Harken: Ischemic preconditioning decreases post-ischemic myocardial TNF: Potential ultimate effector mechanism of preconditioning. *Circulation* 98, 214-219 (1998)

26. Meldrum DR, CA Dinarello, JC Cleveland, BS Cain, BD Shames, X Meng, and AH Harken: Feature Article. Human myocardial TNF production following acute global ischemia and reperfusion injury in vivo. *J Mol Cell Cardiol* 30, 1683-1689 (1998)

27. Meldrum DR, CA Dinarello, JC Cleveland, L Shapiro, BC Sheridan, and AH Harken: p38 MAP kinase mediates myocardial ischemia reperfusion injury. *Surg Forum* In press (1997)

28. Meldrum DR, CA Dinarello, BD Shames, JC Cleveland, Jr., BS Cain, A Banerjee, X Meng, and AH Harken: Ischemic preconditioning decreases postischemic myocardial tumor necrosis factor-alpha production. Potential ultimate effector mechanism of preconditioning. *Circulation* 98, II214-218; discussion II218-219 (1998)

29. Meldrum DR and KK Donnahoo: Role of TNF in mediating renal insufficiency following cardiac surgery: evidence of a postbypass cardiorenal syndrome. *J Surg Res* 85, 185-199 (1999)

30. Meldrum DR, RC McIntyre, BC Sheridan, JC Cleveland, Jr., DA Fullerton, and AH Harken: L-arginine decreases alveolar macrophage proinflammatory monokine production during acute lung injury by a nitric oxide synthase-dependent mechanism. *J Trauma* 43, 888-893 (1997)

31. Meldrum DR, X Meng, CA Dinarello, A Ayala, BS Cain, BD Shames, L Ao, A Banerjee, and AH Harken: Human myocardial tissue TNFalpha expression following acute global ischemia in vivo. *J Mol Cell Cardiol* 30, 1683-1689 (1998)

32. Meldrum DR, X Meng, BD Shames, B Pomerantz, KK Donnahoo, A Banerjee, and AH Harken: Liposomal delivery of heat-shock protein 72 into the heart prevents

endotoxin-induced myocardial contractile dysfunction. *Surgery* 126, 135-141 (1999)

33. Meldrum DR, X Meng, BC Sheridan, RC McIntyre, Jr., AH Harken, and A Banerjee: Tissue-specific protein kinase C isoforms differentially mediate macrophage TNFalpha and IL-1beta production. *Shock* 9, 256-260 (1998)

34. Meldrum DR, MB Mitchell, A Banerjee, and AH Harken: Cardiac preconditioning: Induction of endogenous tolerance to ischemia-reperfusion injury. *Arch Surg* 128, 1208-1211 (1993)

35. Meldrum DR, DA Partrick, JC Cleveland, Jr., R Shenkar, KK Meldrum, A Raiesdana, A Ayala, JW Brown, and AH Harken: On-pump coronary artery bypass surgery activates human myocardial NF-kappaB and increases TNF-alpha in the heart. *J Surg Res* 112, 175-179 (2003)

36. Meldrum DR, R Shenkar, BC Sheridan, BS Cain, E Abraham, and AH Harken: Hemorrhage activates myocardial NFkappaB and increases TNF-alpha in the heart. *J Mol Cell Cardiol* 29, 2849-2854 (1997)

37. Meldrum DR, BC Sheridan, JC Cleveland, DA Fullerton, A Banerjee, and AH Harken: Neutrophils are required for endotoxin induced myocardial cross-tolerance to ischemia-reperfusion injury. *Arch Surg* 131, 1203-1208 (1996)

38. Meldrum KK, DR Meldrum, KL Hile, EB Yerkes, A Ayala, MP Cain, RC Rink, AJ Casale, and MA Kaefer: p38 MAPK mediates renal tubular cell TNF-alpha production and TNF-alpha-dependent apoptosis during simulated ischemia. *Am J Physiol Cell Physiol* 281, C563-570 (2001)

39. Meldrum KK, DR Meldrum, X Meng, L Ao, and AH Harken: TNF-alpha-dependent bilateral renal injury is induced by unilateral renal ischemia-reperfusion. *Am J Physiol Heart Circ Physiol* 282, H540-546 (2002)

40. Meldrum KK, DR Meldrum, SF Sezen, JK Crone, and AL Burnett: Heat shock prevents simulated ischemia-induced apoptosis in renal tubular cells via a PKC-dependent mechanism. *Am J Physiol Regul Integr Comp Physiol* 281, R359-364 (2001)

41. Meldrum DR: Tech.Sight. Sequencing genomes and beyond. *Science* 292, 515-517 (2001)

42. Meldrum KK, AL Burnett, X Meng, R Misseri, MB Shaw, JP Gearhart, and DR Meldrum: Liposomal delivery of heat shock protein 72 into renal tubular cells blocks nuclear factor-kappaB activation, tumor necrosis factor-alpha production, and subsequent ischemia-induced apoptosis. *Circ Res* 92, 293-299 (2003)

43. Ansar Ahmed S, WJ Penhale, and N Talal: Sex hormones, immune responses, and autoimmune diseases. Mechanisms of sex hormone action. *Am J Pathol* 121, 531-551 (1985)



44. Rossouw JE: Hormones, genetic factors, and gender differences in cardiovascular disease. *Cardiovasc Res* 53, 550-557 (2002)
45. Kapadia S, J Lee, G Torre-Amione, HH Birdsall, TS Ma, and DL Mann: Tumor necrosis factor-alpha gene and protein expression in adult feline myocardium after endotoxin administration. *J Clin Invest* 96, 1042-1052 (1995)
46. Kumar A, V Thota, L Dee, J Olson, E Uretz, and JE Parrillo: Tumor necrosis factor alpha and interleukin 1beta are responsible for in vitro myocardial cell depression induced by human septic shock serum. *J Exp Med* 183, 949-958 (1996)
47. Maass DL, J White, and JW Horton: IL-1beta and IL-6 act synergistically with TNF-alpha to alter cardiac contractile function after burn trauma. *Shock* 18, 360-366 (2002)
48. Cain BS, DR Meldrum, CA Dinarello, X Meng, KS Joo, A Banerjee, and AH Harken: Tumor necrosis factor-alpha and interleukin-1beta synergistically depress human myocardial function. *Crit Care Med* 27, 1309-1318 (1999)
49. Tracey KJ, Y Fong, DG Hesse, KR Manogue, AT Lee, GC Kuo, SF Lowry, and A Cerami: Anti-cachectin/TNF monoclonal antibodies prevent septic shock during lethal bacteraemia. *Nature* 330, 662-664 (1987)
50. Sakamoto A, J Matsumura, S Mii, Y Gotoh, and R Ogawa: A prostaglandin E2 receptor subtype EP4 agonist attenuates cardiovascular depression in endotoxin shock by inhibiting inflammatory cytokines and nitric oxide production. *Shock* 22, 76-81 (2004)
51. Arumugam TV, IA Shiels, TM Woodruff, DN Granger, and SM Taylor: The role of the complement system in ischemia-reperfusion injury. *Shock* 21, 401-409 (2004)
52. Angele MK, MG Schwacha, A Ayala, and IH Chaudry: Effect of gender and sex hormones on immune responses following shock. *Shock* 14, 81-90 (2000)
53. Baker L, KK Meldrum, M Wang, R Sankula, R Vanam, A Raiesdana, B Tsai, K Hile, JW Brown, and DR Meldrum: The role of estrogen in cardiovascular disease. *J Surg Res* 115, 325-344 (2003)
54. Becker RC, M Terrin, R Ross, GL Knatterud, P Desvigne-Nickens, JM Gore, and E Braunwald: Comparison of clinical outcomes for women and men after acute myocardial infarction. The Thrombolysis in Myocardial Infarction Investigators. *Ann Intern Med* 120, 638-645 (1994)
55. Kostis JB, AC Wilson, K O'Dowd, P Gregory, S Chelton, NM Cosgrove, A Chirala, and T Cui: Sex differences in the management and long-term outcome of acute myocardial infarction. A statewide study. MIDAS Study Group. Myocardial Infarction Data Acquisition System. *Circulation* 90, 1715-1730 (1994)
56. Demirovic J, H Blackburn, PG McGovern, R Luepker, JM Sprafka, and D Gilbertson: Sex differences in early mortality after acute myocardial infarction (the Minnesota Heart Survey) *Am J Cardiol* 75, 1096-1101 (1995)
57. Vaccarino V, HM Krumholz, LF Berkman, and RI Horwitz: Sex differences in mortality after myocardial infarction. Is there evidence for an increased risk for women? *Circulation* 91, 1861-1871 (1995)
58. Sonke GS, R Beaglehole, AW Stewart, R Jackson, and FM Stewart: Sex differences in case fatality before and after admission to hospital after acute cardiac events: analysis of community based coronary heart disease register. *BMJ* 313, 853-855 (1996)
59. Tunstall-Pedoe H, C Morrison, M Woodward, B Fitzpatrick, and G Watt: Sex differences in myocardial infarction and coronary deaths in the Scottish MONICA population of Glasgow 1985 to 1991. Presentation, diagnosis, treatment, and 28-day case fatality of 3991 events in men and 1551 events in women. *Circulation* 93, 1981-1992 (1996)
60. Chambless L, U Keil, A Dobson, M Mahonen, K Kuulasmaa, AM Rajakangas, H Lowel, and H Tunstall-Pedoe: Population versus clinical view of case fatality from acute coronary heart disease: results from the WHO MONICA Project 1985-1990. Multinational MONItoring of Trends and Determinants in CARdiovascular Disease. *Circulation* 96, 3849-3859 (1997)
61. Napolitano LM, ME Greco, A Rodriguez, JA Kufera, RS West, and TM Scalea: Gender differences in adverse outcomes after blunt trauma. *J Trauma* 50, 274-280 (2001)
62. Oberholzer A, M Keel, R Zellweger, U Steckholzer, O Trentz, and W Ertel: Incidence of septic complications and multiple organ failure in severely injured patients is sex specific. *J Trauma* 48, 932-937 (2000)
63. Gannon CJ, LM Napolitano, M Pasquale, JK Tracy, and RJ McCarter: A statewide population-based study of gender differences in trauma: validation of a prior single-institution study. *J Am Coll Surg* 195, 11-18 (2002)
64. George RL, G McGwin, Jr., J Metzger, IH Chaudry, and LW Rue, 3<sup>rd</sup>: The association between gender and mortality among trauma patients as modified by age. *J Trauma* 54, 464-471 (2003)
65. Wohltmann CD, GA Franklin, PW Boaz, FA Luchette, PA Kearney, JD Richardson, and DA Spain: A multicenter evaluation of whether gender dimorphism affects survival after trauma. *Am J Surg* 181, 297-300 (2001)
66. Mostafa G, T Huynh, RF Sing, WS Miles, HJ Norton, and MH Thomason: Gender-related outcomes in trauma. *J Trauma* 53, 430-434; discussion 434-435 (2002)
67. Offner PJ, EE Moore, and WL Biffl: Male gender is a risk factor for major infections after surgery. *Arch Surg* 134, 935-938; discussion 938-940 (1999)

68. Gannon CJ, M Pasquale, JK Tracy, RJ McCarter, and LM Napolitano: Male gender is associated with increased risk for postinjury pneumonia. *Shock* 21, 410-414 (2004)
69. Eachempati SR, L Hydo, and PS Barie: Gender-based differences in outcome in patients with sepsis. *Arch Surg* 134, 1342-1347 (1999)
70. Schroder J, V Kahlke, M Book, and F Stuber: Gender differences in sepsis: genetically determined? *Shock* 14, 307-310; discussion 310-303 (2000)
71. Schroder J, V Kahlke, KH Staubach, P Zabel, and F Stuber: Gender differences in human sepsis. *Arch Surg* 133, 1200-1205 (1998)
72. Wichmann MW, D Inthorn, HJ Andress, and FW Schildberg: Incidence and mortality of severe sepsis in surgical intensive care patients: the influence of patient gender on disease process and outcome. *Intensive Care Med* 26, 167-172 (2000)
73. Muller MJ, SP Pegg, and MR Rule: Determinants of death following burn injury. *Br J Surg* 88, 583-587 (2001)
74. O'Keefe GE, JL Hunt, and GF Purdue: An evaluation of risk factors for mortality after burn trauma and the identification of gender-dependent differences in outcomes. *J Am Coll Surg* 192, 153-160 (2001)
75. Cumming J, GF Purdue, JL Hunt, and GE O'Keefe: Objective estimates of the incidence and consequences of multiple organ dysfunction and sepsis after burn trauma. *J Trauma* 50, 510-515 (2001)
76. Delyani JA, T Murohara, TO Nossuli, and AM Lefer: Protection from myocardial reperfusion injury by acute administration of 17 beta-estradiol. *J Mol Cell Cardiol* 28, 1001-1008 (1996)
77. Booth EA, M Marchesi, EJ Kilbourne, and BR Lucchesi: 17Beta-estradiol as a receptor-mediated cardioprotective agent. *J Pharmacol Exp Ther* 307, 395-401 (2003)
78. Node K, M Kitakaze, H Kosaka, T Minamino, H Funaya, and M Hori: Amelioration of ischemia- and reperfusion-induced myocardial injury by 17beta-estradiol: role of nitric oxide and calcium-activated potassium channels. *Circulation* 96, 1953-1963 (1997)
79. Hale SL, Y Birnbaum, and RA Kloner: beta-Estradiol, but not alpha-estradiol, reduced myocardial necrosis in rabbits after ischemia and reperfusion. *Am Heart J* 132, 258-262 (1996)
80. Kolodgie FD, A Farb, SH Litovsky, J Narula, LA Jeffers, SJ Lee, and R Virmani: Myocardial protection of contractile function after global ischemia by physiologic estrogen replacement in the ovariectomized rat. *J Mol Cell Cardiol* 29, 2403-2414 (1997)
81. Kim YD, B Chen, J Beauregard, P Kouretas, G Thomas, MY Farhat, AK Myers, and DE Lees: 17 beta-Estradiol prevents dysfunction of canine coronary endothelium and myocardium and reperfusion arrhythmias after brief ischemia/reperfusion. *Circulation* 94, 2901-2908 (1996)
82. McHugh NA, SM Cook, JL Schairer, MM Bidgoli, and GF Merrill: Ischemia- and reperfusion-induced ventricular arrhythmias in dogs: effects of estrogen. *Am J Physiol* 268, H2569-2573 (1995)
83. Beer S, M Reincke, M Kral, SZ Lie, S Steinhauer, HH Schmidt, B Alolio, and S Neubauer: Susceptibility to cardiac ischemia/reperfusion injury is modulated by chronic estrogen status. *J Cardiovasc Pharmacol* 40, 420-428 (2002)
84. Zhai P, TE Eurell, R Cotthaus, EH Jeffery, JM Bahr, and DR Gross: Effect of estrogen on global myocardial ischemia-reperfusion injury in female rats. *Am J Physiol Heart Circ Physiol* 279, H2766-2775 (2000)
85. Horton JW, DJ White, and DL Maass: Gender-related differences in myocardial inflammatory and contractile responses to major burn trauma. *Am J Physiol Heart Circ Physiol* 286, H202-213 (2004)
86. Wichmann MW, R Zellweger, CM DeMaso, A Ayala, and IH Chaudry: Enhanced immune responses in females, as opposed to decreased responses in males following haemorrhagic shock and resuscitation. *Cytokine* 8, 853-863 (1996)
87. Angele MK, MW Knoferl, MG Schwacha, A Ayala, WG Cioffi, KI Bland, and IH Chaudry: Sex steroids regulate pro- and anti-inflammatory cytokine release by macrophages after trauma-hemorrhage. *Am J Physiol* 277, C35-42 (1999)
88. Zellweger R, MW Wichmann, A Ayala, S Stein, CM DeMaso, and IH Chaudry: Females in proestrus state maintain splenic immune functions and tolerate sepsis better than males. *Crit Care Med* 25, 106-110 (1997)
89. Angele MK, A Ayala, WG Cioffi, KI Bland, and IH Chaudry: Testosterone: the culprit for producing splenocyte immune depression after trauma hemorrhage. *Am J Physiol* 274, C1530-1536 (1998)
90. Angele MK, A Ayala, BA Monfils, WG Cioffi, KI Bland, and IH Chaudry: Testosterone and/or low estradiol: normally required but harmful immunologically for males after trauma-hemorrhage. *J Trauma* 44, 78-85 (1998)
91. Messingham KA, M Shirazi, LA Duffner, MA Emanuele, and EJ Kovacs: Testosterone receptor blockade restores cellular immunity in male mice after burn injury. *J Endocrinol* 169, 299-308 (2001)
92. Ba ZF, P Wang, DJ Koo, M Zhou, WG Cioffi, KI Bland, and IH Chaudry: Testosterone receptor blockade

after trauma and hemorrhage attenuates depressed adrenal function. *Am J Physiol Regul Integr Comp Physiol* 279, R1841-1848 (2000)

93. Remmers DE, P Wang, WG Cioffi, KI Bland, and IH Chaudry: Testosterone receptor blockade after trauma-hemorrhage improves cardiac and hepatic functions in males. *Am J Physiol* 273, H2919-2925 (1997)

94. Angele MK, MW Wichmann, A Ayala, WG Cioffi, and IH Chaudry: Testosterone receptor blockade after hemorrhage in males. Restoration of the depressed immune functions and improved survival following subsequent sepsis. *Arch Surg* 132, 1207-1214 (1997)

95. Knoferl MW, MK Angele, MG Schwacha, TS Anantha Samy, KI Bland, and IH Chaudry: Immunoprotection in proestrus females following trauma-hemorrhage: the pivotal role of estrogen receptors. *Cell Immunol* 222, 27-34 (2003)

96. Knoferl MW, MK Angele, MD Diodato, MG Schwacha, A Ayala, WG Cioffi, KI Bland, and IH Chaudry: Female sex hormones regulate macrophage function after trauma-hemorrhage and prevent increased death rate from subsequent sepsis. *Ann Surg* 235, 105-112 (2002)

97. Jarrar D, P Wang, WG Cioffi, KI Bland, and IH Chaudry: The female reproductive cycle is an important variable in the response to trauma-hemorrhage. *Am J Physiol Heart Circ Physiol* 279, H1015-1021 (2000)

98. Lloyd GW, NR Patel, E McGing, AF Cooper, D Brennand-Roper, and G Jackson: Does angina vary with the menstrual cycle in women with premenopausal coronary artery disease? *Heart* 84, 189-192 (2000)

99. Kawano H, T Motoyama, M Ohgushi, K Kugiyama, H Ogawa, and H Yasue: Menstrual cyclic variation of myocardial ischemia in premenopausal women with variant angina. *Ann Intern Med* 135, 977-981 (2001)

100. Clark PI, SP Glasser, GH Lyman, J Krug-Fite, and A Root: Relation of results of exercise stress tests in young women to phases of the menstrual cycle. *Am J Cardiol* 61, 197-199 (1988)

101. Durant R, K Klouche, S Delbosc, M Morena, L Amigues, JJ Beraud, B Canaud, and JP Cristol: Superoxide anion overproduction in sepsis: effects of vitamin E and simvastatin. *Shock* 22, 34-39 (2004)

102. Huot J, F Houle, F Marceau, and J Landry: Oxidative stress-induced actin reorganization mediated by the p38 mitogen-activated protein kinase/heat shock protein 27 pathway in vascular endothelial cells. *Circ Res* 80, 383-392 (1997)

103. Guyton KZ, Y Liu, M Gorospe, Q Xu, and NJ Holbrook: Activation of mitogen-activated protein kinase by H<sub>2</sub>O<sub>2</sub>. Role in cell survival following oxidant injury. *J Biol Chem* 271, 4138-4142 (1996)

104. Mihm S, D Galter, and W Droge: Modulation of transcription factor NF kappa B activity by intracellular glutathione levels and by variations of the extracellular cysteine supply. *Faseb J* 9, 246-252 (1995)

105. Liu SL, S Degli Esposti, T Yao, AM Diehl, and MA Zern: Vitamin E therapy of acute CCl<sub>4</sub>-induced hepatic injury in mice is associated with inhibition of nuclear factor kappa B binding. *Hepatology* 22, 1474-1481 (1995)

106. Ballard-Croft C, DJ White, DL Maass, DP Hybki, and JW Horton: Role of p38 mitogen-activated protein kinase in cardiac myocyte secretion of the inflammatory cytokine TNF-alpha. *Am J Physiol Heart Circ Physiol* 280, H1970-1981 (2001)

107. Cain BS, DR Meldrum, X Meng, CA Dinarello, BD Shames, A Banerjee, and AH Harken: p38 MAPK inhibition decreases TNF-alpha production and enhances postischemic human myocardial function. *J Surg Res* 83, 7-12 (1999)

108. Wang M, R Sankula, BM Tsai, KK Meldrum, M Turrentine, KL March, JW Brown, CA Dinarello, and DR Meldrum: P38 MAPK mediates myocardial proinflammatory cytokine production and endotoxin-induced contractile suppression. *Shock* 21, 170-174 (2004)

109. Kita T, H Yamaguchi, H Sato, K Kasai, T Tanaka, and N Tanaka: Role of p38 mitogen-activated protein kinase pathway on renal failure in the infant rat after burn injury. *Shock* 21, 535-542 (2004)

110. Lub-De Hooge MN, S De Jong, C Vermot-Desroches, JE Tulleken, EG De Vries, and JG Zijlstra: Endotoxin increases plasma soluble tumor necrosis factor-related apoptosis-inducing ligand level mediated by the p38 mitogen-activated protein kinase signaling pathway. *Shock* 22, 186-188 (2004)

111. Lee JC, JT Laydon, PC McDonnell, TF Gallagher, S Kumar, D Green, D McNulty, MJ Blumenthal, JR Heys, SW Landvatter, and et al: A protein kinase involved in the regulation of inflammatory cytokine biosynthesis. *Nature* 372, 739-746 (1994)

112. Schafer PH, SA Wadsworth, L Wang, and JJ Siekierka: p38 alpha mitogen-activated protein kinase is activated by CD28-mediated signaling and is required for IL-4 production by human CD4+CD45RO+ T cells and Th2 effector cells. *J Immunol* 162, 7110-7119 (1999)

113. Beyaert R, A Cuenda, W Vanden Berghe, S Plaisance, JC Lee, G Haegeman, P Cohen, and W Fiers: The p38/RK mitogen-activated protein kinase pathway regulates interleukin-6 synthesis response to tumor necrosis factor. *Embo J* 15, 1914-1923 (1996)

114. Marie C, S Roman-Roman, and G Rawadi: Involvement of mitogen-activated protein kinase pathways in interleukin-8 production by human monocytes and polymorphonuclear cells stimulated with lipopolysaccharide or Mycoplasma fermentans membrane lipoproteins. *Infect Immun* 67, 688-693 (1999)

115. Zechner D, R Craig, DS Hanford, PM McDonough, RA Sabbadini, and CC Glembotski: MKK6 activates myocardial cell NF-kappaB and inhibits apoptosis in a p38 mitogen-activated protein kinase-dependent manner. *J Biol Chem* 273, 8232-8239 (1998)
116. Brown MA and WK Jones: NF-kappaB action in sepsis: the innate immune system and the heart. *Front Biosci* 9, 1201-1217 (2004)
117. Carter AB, KL Knudtson, MM Monick, and GW Hunninghake: The p38 mitogen-activated protein kinase is required for NF-kappaB-dependent gene expression. The role of TATA-binding protein (TBP) *J Biol Chem* 274, 30858-30863 (1999)
118. Nick JA, NJ Avdi, SK Young, LA Lehman, PP McDonald, SC Frasch, MA Billstrom, PM Henson, GL Johnson, and GS Worthen: Selective activation and functional significance of p38alpha mitogen-activated protein kinase in lipopolysaccharide-stimulated neutrophils. *J Clin Invest* 103, 851-858 (1999)
119. Maass DL, DP Hybki, J White, and JW Horton: The time course of cardiac NF-kappaB activation and TNF-alpha secretion by cardiac myocytes after burn injury: contribution to burn-related cardiac contractile dysfunction. *Shock* 17, 293-299 (2002)
120. Lum RT, SS Kerwar, SM Meyer, MG Nelson, SR Schow, D Shiffman, MM Wick, and A Joly: A new structural class of proteasome inhibitors that prevent NF-kappa B activation. *Biochem Pharmacol* 55, 1391-1397 (1998)
121. Wesche H, WJ Henzel, W Shillinglaw, S Li, and Z Cao: MyD88: an adapter that recruits IRAK to the IL-1 receptor complex. *Immunity* 7, 837-847 (1997)
122. Burns K, F Martinon, C Esslinger, H Pahl, P Schneider, JL Bodmer, F Di Marco, L French, and J Tschopp: MyD88, an adapter protein involved in interleukin-1 signaling. *J Biol Chem* 273, 12203-12209 (1998)
123. Wang C, L Deng, M Hong, GR Akkaraju, J Inoue, and ZJ Chen: TAK1 is a ubiquitin-dependent kinase of MKK and IKK. *Nature* 412, 346-351 (2001)
124. Levy RJ and CS Deutschman: Evaluating myocardial depression in sepsis. *Shock* 22, 1-10 (2004)
125. Krown KA, K Yasui, MJ Brooker, AE Dubin, C Nguyen, GL Harris, PM McDonough, CC Glembotski, PT Palade, and RA Sabbadini: TNF alpha receptor expression in rat cardiac myocytes: TNF alpha inhibition of L-type Ca2+ current and Ca2+ transients. *FEBS Lett* 376, 24-30 (1995)
126. Yokoyama T, L Vaca, RD Rossen, W Durante, P Hazarika, and DL Mann: Cellular basis for the negative inotropic effects of tumor necrosis factor-alpha in the adult mammalian heart. *J Clin Invest* 92, 2303-2312 (1993)
127. Oral H, GW Dorn, 2nd, and DL Mann: Sphingosine mediates the immediate negative inotropic effects of tumor necrosis factor-alpha in the adult mammalian cardiac myocyte. *J Biol Chem* 272, 4836-4842 (1997)
128. Goldhaber JJ, KH Kim, PD Natterson, T Lawrence, P Yang, and JN Weiss: Effects of TNF-alpha on [Ca2+]i and contractility in isolated adult rabbit ventricular myocytes. *Am J Physiol* 271, H1449-1455 (1996)
129. Finkel MS, CV Oddis, TD Jacob, SC Watkins, BG Hattler, and RL Simmons: Negative inotropic effects of cytokines on the heart mediated by nitric oxide. *Science* 257, 387-389 (1992)
130. Kurrelmeyer KM, LH Michael, G Baumgarten, GE Taffet, JJ Peschon, N Sivasubramanian, ML Entman, and DL Mann: Endogenous tumor necrosis factor protects the adult cardiac myocyte against ischemic-induced apoptosis in a murine model of acute myocardial infarction. *Proc Natl Acad Sci U S A* 97, 5456-5461 (2000)
131. Reinhart K, C Wiegand-Lohnert, F Grimminger, M Kaul, S Withington, D Treacher, J Eckart, S Willatts, C Bouza, D Krausch, F Stockenhuber, J Eiselstein, L Daum, and J Kempeni: Assessment of the safety and efficacy of the monoclonal anti-tumor necrosis factor antibody-fragment, MAK 195F, in patients with sepsis and septic shock: a multicenter, randomized, placebo-controlled, dose-ranging study. *Crit Care Med* 24, 733-742 (1996)
132. Wertheimer SJ, CL Myers, RW Wallace, and TP Parks: Intercellular adhesion molecule-1 gene expression in human endothelial cells. Differential regulation by tumor necrosis factor-alpha and phorbol myristate acetate. *J Biol Chem* 267, 12030-12035 (1992)
133. Youker K, CW Smith, DC Anderson, D Miller, LH Michael, RD Rossen, and ML Entman: Neutrophil adherence to isolated adult cardiac myocytes. Induction by cardiac lymph collected during ischemia and reperfusion. *J Clin Invest* 89, 602-609 (1992)
134. Kukiella GL, CW Smith, AM Manning, KA Youker, LH Michael, and ML Entman: Induction of interleukin-6 synthesis in the myocardium. Potential role in postreperfusion inflammatory injury. *Circulation* 92, 1866-1875 (1995)
135. Entman ML, K Youker, T Shoji, G Kukiella, SB Shappell, AA Taylor, and CW Smith: Neutrophil induced oxidative injury of cardiac myocytes. A compartmented system requiring CD11b/CD18-ICAM-1 adherence. *J Clin Invest* 90, 1335-1345 (1992)
136. Romson JL, BG Hook, SL Kunkel, GD Abrams, MA Schork, and BR Lucchesi: Reduction of the extent of ischemic myocardial injury by neutrophil depletion in the dog. *Circulation* 67, 1016-1023 (1983)

137. Jolly SR, WJ Kane, BG Hook, GD Abrams, SL Kunkel, and BR Lucchesi: Reduction of myocardial infarct size by neutrophil depletion: effect of duration of occlusion. *Am Heart J* 112, 682-690 (1986)
138. Litt MR, RW Jeremy, HF Weisman, JA Winkelstein, and LC Becker: Neutrophil depletion limited to reperfusion reduces myocardial infarct size after 90 minutes of ischemia. Evidence for neutrophil-mediated reperfusion injury. *Circulation* 80, 1816-1827 (1989)
139. Granger DN: Role of xanthine oxidase and granulocytes in ischemia-reperfusion injury. *Am J Physiol* 255, H1269-1275 (1988)
140. Patel KD, GA Zimmerman, SM Prescott, RP McEver, and TM McIntyre: Oxygen radicals induce human endothelial cells to express GMP-140 and bind neutrophils. *J Cell Biol* 112, 749-759 (1991)
141. Shingu M and M Nobunaga: Chemotactic activity generated in human serum from the fifth component of complement by hydrogen peroxide. *Am J Pathol* 117, 201-206 (1984)
142. Akgur FM, MF Brown, GB Zibari, JC McDonald, CJ Epstein, CR Ross, and DN Granger: Role of superoxide in hemorrhagic shock-induced P-selectin expression. *Am J Physiol Heart Circ Physiol* 279, H791-797 (2000)
143. Lakshminarayanan V, DW Beno, RH Costa, and KA Roebuck: Differential regulation of interleukin-8 and intercellular adhesion molecule-1 by H<sub>2</sub>O<sub>2</sub> and tumor necrosis factor- $\alpha$  in endothelial and epithelial cells. *J Biol Chem* 272, 32910-32918 (1997)
144. Sellak H, E Franzini, J Hakim, and C Pasquier: Reactive oxygen species rapidly increase endothelial ICAM-1 ability to bind neutrophils without detectable upregulation. *Blood* 83, 2669-2677 (1994)
145. Jolly SR, WJ Kane, MB Bailie, GD Abrams, and BR Lucchesi: Canine myocardial reperfusion injury. Its reduction by the combined administration of superoxide dismutase and catalase. *Circ Res* 54, 277-285 (1984)
146. Wang P, H Chen, H Qin, S Sankarapandi, MW Becher, PC Wong, and JL Zweier: Overexpression of human copper, zinc-superoxide dismutase (SOD1) prevents postischemic injury. *Proc Natl Acad Sci U S A* 95, 4556-4560 (1998)
147. Chen Z, B Siu, YS Ho, R Vincent, CC Chua, RC Hamdy, and BH Chua: Overexpression of MnSOD protects against myocardial ischemia/reperfusion injury in transgenic mice. *J Mol Cell Cardiol* 30, 2281-2289 (1998)
148. Flaherty JT, B Pitt, JW Gruber, RR Heuser, DA Rothbaum, LR Burwell, BS George, DJ Kereiakes, D Deitchman, N Gustafson, and et al: Recombinant human superoxide dismutase (h-SOD) fails to improve recovery of ventricular function in patients undergoing coronary angioplasty for acute myocardial infarction. *Circulation* 89, 1982-1991 (1994)
149. Murohara Y, Y Yui, R Hattori, and C Kawai: Effects of superoxide dismutase on reperfusion arrhythmias and left ventricular function in patients undergoing thrombolysis for anterior wall acute myocardial infarction. *Am J Cardiol* 67, 765-767 (1991)
150. Kim NG, H Lee, E Son, OY Kwon, JY Park, JH Park, GJ Cho, WS Choi, and K Suk: Hypoxic induction of caspase-11/caspase-1/interleukin-1 $\beta$  in brain microglia. *Brain Res Mol Brain Res* 114, 107-114 (2003)
151. Hur J, SY Kim, H Kim, S Cha, MS Lee, and K Suk: Induction of caspase-11 by inflammatory stimuli in rat astrocytes: lipopolysaccharide induction through p38 mitogen-activated protein kinase pathway. *FEBS Lett* 507, 157-162 (2001)
152. Deshpande R, H Khalili, RG Pergolizzi, SD Michael, and MD Chang: Estradiol down-regulates LPS-induced cytokine production and NF $\kappa$ B activation in murine macrophages. *Am J Reprod Immunol* 38, 46-54 (1997)
153. Mizushima Y, P Wang, D Jarrar, WG Cioffi, KI Bland, and IH Chaudry: Estradiol administration after trauma-hemorrhage improves cardiovascular and hepatocellular functions in male animals. *Ann Surg* 232, 673-679 (2000)
154. Wang M, B Tsai, JW Brown, and DR Meldrum: Gender differences in myocardial apoptotic signaling and recovery following acute injury. *Shock* 21(Suppl 2), 23 (2004)
155. Angele MK, S Nitsch, MW Knoferl, A Ayala, P Angele, FW Schildberg, KW Jauch, and IH Chaudry: Sex-specific p38 MAP kinase activation following trauma-hemorrhage: involvement of testosterone and estradiol. *Am J Physiol Endocrinol Metab* 285, E189-196 (2003)
156. McHugh NA, GF Merrill, and SR Powell: Estrogen diminishes postischemic hydroxyl radical production. *Am J Physiol* 274, H1950-1954 (1998)
157. Kim YD, MY Farhat, AK Myers, P Kouretas, KW DeGroot, A Pacquing, PW Ramwell, JP Suyderhoud, and DE Lees: 17-Beta estradiol regulation of myocardial glutathione and its role in protection against myocardial stunning in dogs. *J Cardiovasc Pharmacol* 32, 457-465 (1998)
158. Barp J, AS Araujo, TR Fernandes, KV Rigatto, S Llesuy, A Bello-Klein, and P Singal: Myocardial antioxidant and oxidative stress changes due to sex hormones. *Braz J Med Biol Res* 35, 1075-1081 (2002)
159. Squadrito F, D Altavilla, G Squadrito, GM Campo, M Arlotta, V Arcoraci, L Minutoli, M Serrano, A Saitta, and AP Caputi: 17Beta-oestradiol reduces cardiac leukocyte



accumulation in myocardial ischaemia reperfusion injury in rat. *Eur J Pharmacol* 335, 185-192 (1997)

160. Sumeray MS, DD Rees, and DM Yellon: Infarct size and nitric oxide synthase in murine myocardium. *J Mol Cell Cardiol* 32, 35-42 (2000)

161. Xi L, NC Jarrett, ML Hess, and RC Kukreja: Essential role of inducible nitric oxide synthase in monophosphoryl lipid A-induced late cardioprotection: evidence from pharmacological inhibition and gene knockout mice. *Circulation* 99, 2157-2163 (1999)

162. Weiner CP, I Lizasoain, SA Baylis, RG Knowles, IG Charles, and S Moncada: Induction of calcium-dependent nitric oxide synthases by sex hormones. *Proc Natl Acad Sci U S A* 91, 5212-5216 (1994)

163. Nuedling S, S Kahlert, K Loebbert, PA Doevendans, R Meyer, H Vetter, and C Grohe: 17 Beta-estradiol stimulates expression of endothelial and inducible NO synthase in rat myocardium in-vitro and in-vivo. *Cardiovasc Res* 43, 666-674 (1999)

164. Kleinert H, T Wallerath, C Euchenhofer, I Ihrig-Biedert, H Li, and U Forstermann: Estrogens increase transcription of the human endothelial NO synthase gene: analysis of the transcription factors involved. *Hypertension* 31, 582-588 (1998)

165. Grohe C, S Kahlert, K Lobbart, and H Vetter: Expression of oestrogen receptor alpha and beta in rat heart: role of local oestrogen synthesis. *J Endocrinol* 156, R1-7 (1998)

166. Ma XL, AS Weyrich, DJ Lefer, and AM Lefer: Diminished basal nitric oxide release after myocardial ischemia and reperfusion promotes neutrophil adherence to coronary endothelium. *Circ Res* 72, 403-412 (1993)

167. Lefer AM and DJ Lefer: The role of nitric oxide and cell adhesion molecules on the microcirculation in ischaemia-reperfusion. *Cardiovasc Res* 32, 743-751 (1996)

168. Hoshida S, N Yamashita, J Igarashi, M Nishida, M Hori, T Kamada, T Kuzuya, and M Tada: Nitric oxide synthase protects the heart against ischemia-reperfusion injury in rabbits. *J Pharmacol Exp Ther* 274, 413-418 (1995)

169. Steenbergen C, E Murphy, JA Watts, and RE London: Correlation between cytosolic free calcium, contracture, ATP, and irreversible ischemic injury in perfused rat heart. *Circ Res* 66, 135-146 (1990)

170. Marban E, Y Koretsune, M Corretti, VP Chacko, and H Kusuoka: Calcium and its role in myocardial cell injury during ischemia and reperfusion. *Circulation* 80, IV17-22 (1989)

171. Quignard JF, JM Frapier, MC Harricane, B Albat, J Nargeot, and S Richard: Voltage-gated calcium channel

currents in human coronary myocytes. Regulation by cyclic GMP and nitric oxide. *J Clin Invest* 99, 185-193 (1997)

172. Mery PF, C Pavoine, L Belhassen, F Pecker, and R Fischmeister: Nitric oxide regulates cardiac Ca<sup>2+</sup> current. Involvement of cGMP-inhibited and cGMP-stimulated phosphodiesterases through guanylyl cyclase activation. *J Biol Chem* 268, 26286-26295 (1993)

173. Zahradnikova A and O Krizanov: Nitric oxide and its effects on the calcium transport systems in the myocardium. *Gen Physiol Biophys* 16, 197-214 (1997)

174. Zahradnikova A, I Minarovic, RC Venema, and LG Meszaros: Inactivation of the cardiac ryanodine receptor calcium release channel by nitric oxide. *Cell Calcium* 22, 447-454 (1997)

175. Ziolo MT, H Katoh, and DM Bers: Expression of inducible nitric oxide synthase depresses beta-adrenergic-stimulated calcium release from the sarcoplasmic reticulum in intact ventricular myocytes. *Circulation* 104, 2961-2966 (2001)

176. Shinbo A and T Iijima: Potentiation by nitric oxide of the ATP-sensitive K<sup>+</sup> current induced by K<sup>+</sup> channel openers in guinea-pig ventricular cells. *Br J Pharmacol* 120, 1568-1574 (1997)

177. Sasaki N, T Sato, A Ohler, B O'Rourke, and E Marban: Activation of mitochondrial ATP-dependent potassium channels by nitric oxide. *Circulation* 101, 439-445 (2000)

178. Kim HW, NA Steenaart, DG Ferguson, and EG Kranias: Functional reconstitution of the cardiac sarcoplasmic reticulum Ca<sup>2+</sup>(+)-ATPase with phospholamban in phospholipid vesicles. *J Biol Chem* 265, 1702-1709 (1990)

179. Kranias EG: Regulation of Ca<sup>2+</sup> transport by cyclic 3',5'-AMP-dependent and calcium-calmodulin-dependent phosphorylation of cardiac sarcoplasmic reticulum. *Biochim Biophys Acta* 844, 193-199 (1985)

180. Lindemann JP, LR Jones, DR Hathaway, BG Henry, and AM Watanabe: beta-Adrenergic stimulation of phospholamban phosphorylation and Ca<sup>2+</sup>-ATPase activity in guinea pig ventricles. *J Biol Chem* 258, 464-471 (1983)

181. Wu G, SL Yang, C Hsu, RC Yang, HK Hsu, N Liu, J Yang, LW Dong, and MS Liu: Transcriptional regulation of cardiac sarcoplasmic reticulum calcium-ATPase gene during the progression of sepsis. *Shock* 22, 46-50 (2004)

182. Golden KL, QI Fan, B Chen, J Ren, J O'Connor, and JD Marsh: Adrenergic stimulation regulates Na<sup>(+)</sup>/Ca<sup>(2+)</sup>Exchanger expression in rat cardiac myocytes. *J Mol Cell Cardiol* 32, 611-620 (2000)

183. Thawornkaiwong A, S Preawnim, and J Wattanapermpool: Upregulation of beta 1-adrenergic

- receptors in ovariectomized rat hearts. *Life Sci* 72, 1813-1824 (2003)
184. Kam KW, JS Qi, M Chen, and TM Wong: Estrogen reduces cardiac injury and expression of beta1-adrenoceptor upon ischemic insult in the rat heart. *J Pharmacol Exp Ther* 309, 8-15 (2004)
185. Cross HR, EG Kranias, E Murphy, and C Steenbergen: Ablation of PLB exacerbates ischemic injury to a lesser extent in female than male mice: protective role of NO. *Am J Physiol Heart Circ Physiol* 284, H683-690 (2003)
186. Cross HR, E Murphy, and C Steenbergen: Ca(2+) loading and adrenergic stimulation reveal male/female differences in susceptibility to ischemia-reperfusion injury. *Am J Physiol Heart Circ Physiol* 283, H481-489 (2002)
187. Golden KL, JD Marsh, and Y Jiang: Castration reduces mRNA levels for calcium regulatory proteins in rat heart. *Endocrine* 19, 339-344 (2002)
188. Tani M and JR Neely: Role of intracellular Na+ in Ca2+ overload and depressed recovery of ventricular function of reperfused ischemic rat hearts. Possible involvement of H+-Na+ and Na+-Ca2+ exchange. *Circ Res* 65, 1045-1056 (1989)
189. Murphy E, M Perlman, RE London, and C Steenbergen: Amiloride delays the ischemia-induced rise in cytosolic free calcium. *Circ Res* 68, 1250-1258 (1991)
190. Murphy E, HR Cross, and C Steenbergen: Is Na/Ca exchange during ischemia and reperfusion beneficial or detrimental? *Ann N Y Acad Sci* 976, 421-430 (2002)
191. Cross HR, L Lu, C Steenbergen, KD Philipson, and E Murphy: Overexpression of the cardiac Na+/Ca2+ exchanger increases susceptibility to ischemia/reperfusion injury in male, but not female, transgenic mice. *Circ Res* 83, 1215-1223 (1998)
192. Sugishita K, Z Su, F Li, KD Philipson, and WH Barry: Gender influences [Ca(2+)](i) during metabolic inhibition in myocytes overexpressing the Na(+)-Ca(2+) exchanger. *Circulation* 104, 2101-2106 (2001)
193. Cleveland JC, Jr., DR Meldrum, RT Rowland, A Banerjee, and AH Harken: Adenosine preconditioning of human myocardium is dependent upon the ATP-sensitive K+ channel. *J Mol Cell Cardiol* 29, 175-182 (1997)
194. Downey JM and MV Cohen: Signal transduction in ischemic preconditioning. *Adv Exp Med Biol* 430, 39-55 (1997)
195. Light PE, AA Sabir, BG Allen, MP Walsh, and RJ French: Protein kinase C-induced changes in the stoichiometry of ATP binding activate cardiac ATP-sensitive K+ channels. A possible mechanistic link to ischemic preconditioning. *Circ Res* 79, 399-406 (1996)
196. Miyamae M, SA Camacho, MW Weiner, and VM Figueredo: Attenuation of postischemic reperfusion injury is related to prevention of [Ca2+]m overload in rat hearts. *Am J Physiol* 271, H2145-2153 (1996)
197. Delcamp TJ, C Dales, L Ralenkotter, PS Cole, and RW Hadley: Intramitochondrial [Ca2+] and membrane potential in ventricular myocytes exposed to anoxia-reoxygenation. *Am J Physiol* 275, H484-494 (1998)
198. Jovanovic N, S Jovanovic, A Jovanovic, and A Terzic: Gene delivery of Kir6.2/SUR2A in conjunction with pinacidil handles intracellular Ca2+ homeostasis under metabolic stress. *Faseb J* 13, 923-929 (1999)
199. Holmuhamedov EL, L Wang, and A Terzic: ATP-sensitive K+ channel openers prevent Ca2+ overload in rat cardiac mitochondria. *J Physiol* 519 Pt 2, 347-360 (1999)
200. Lee TM, SF Su, CC Tsai, YT Lee, and CH Tsai: Cardioprotective effects of 17 beta-estradiol produced by activation of mitochondrial ATP-sensitive K(+) channels in canine hearts. *J Mol Cell Cardiol* 32, 1147-1158 (2000)
201. Tsai CH, SF Su, TF Chou, and TM Lee: Differential effects of sarcolemmal and mitochondrial K(ATP) channels activated by 17 beta-estradiol on reperfusion arrhythmias and infarct sizes in canine hearts. *J Pharmacol Exp Ther* 301, 234-240 (2002)
202. Lee TM, TF Chou, and CH Tsai: Differential role of K(ATP) channels activated by conjugated estrogens in the regulation of myocardial and coronary protective effects. *Circulation* 107, 49-54 (2003)
203. Ling S, A Dai, MR Williams, K Myles, RJ Dilley, PA Komesaroff, and K Sudhir: Testosterone (T) enhances apoptosis-related damage in human vascular endothelial cells. *Endocrinology* 143, 1119-1125 (2002)
204. Verzola D, MT Gandolfo, F Salvatore, B Villaggio, F Gianiorio, P Traverso, G Deferrari, and G Garibotto: Testosterone promotes apoptotic damage in human renal tubular cells. *Kidney Int* 65, 1252-1261 (2004)
205. Zaugg M, NZ Jamali, E Lucchinetti, W Xu, M Alam, SA Shafiq, and MA Siddiqui: Anabolic-androgenic steroids induce apoptotic cell death in adult rat ventricular myocytes. *J Cell Physiol* 187, 90-95 (2001)
206. Pelzer T, M Schumann, M Neumann, T deJager, M Stimpel, E Serfling, and L Neyses: 17beta-estradiol prevents programmed cell death in cardiac myocytes. *Biochem Biophys Res Commun* 268, 192-200 (2000)
207. Camper-Kirby D, S Welch, A Walker, I Shiraishi, KD Setchell, E Schaefer, J Kajstura, P Anversa, and MA Sussman: Myocardial Akt activation and gender: increased nuclear activity in females versus males. *Circ Res* 88, 1020-1027 (2001)
208. Fujio Y, T Nguyen, D Wencker, RN Kitsis, and K Walsh: Akt promotes survival of cardiomyocytes in vitro

and protects against ischemia-reperfusion injury in mouse heart. *Circulation* 101, 660-667 (2000)

209. Matsui T, L Li, F del Monte, Y Fukui, TF Franke, RJ Hajjar, and A Rosenzweig: Adenoviral gene transfer of activated phosphatidylinositol 3'-kinase and Akt inhibits apoptosis of hypoxic cardiomyocytes in vitro. *Circulation* 100, 2373-2379 (1999)

210. Dudek H, SR Datta, TF Franke, MJ Birnbaum, R Yao, GM Cooper, RA Segal, DR Kaplan, and ME Greenberg: Regulation of neuronal survival by the serine-threonine protein kinase Akt. *Science* 275, 661-665 (1997)

211. Datta SR, H Dudek, X Tao, S Masters, H Fu, Y Gotoh, and ME Greenberg: Akt phosphorylation of BAD couples survival signals to the cell-intrinsic death machinery. *Cell* 91, 231-241 (1997)

212. Cardone MH, N Roy, HR Stennicke, GS Salvesen, TF Franke, E Stanbridge, S Frisch, and JC Reed: Regulation of cell death protease caspase-9 by phosphorylation. *Science* 282, 1318-1321 (1998)

213. Brunet A, A Bonni, MJ Zigmond, MZ Lin, P Juo, LS Hu, MJ Anderson, KC Arden, J Blenis, and ME Greenberg: Akt promotes cell survival by phosphorylating and inhibiting a Forkhead transcription factor. *Cell* 96, 857-868 (1999)

214. Suzuki K, B Murtuza, RT Smolenski, IA Sammut, N Suzuki, Y Kaneda, and MH Yacoub: Overexpression of interleukin-1 receptor antagonist provides cardioprotection against ischemia-reperfusion injury associated with reduction in apoptosis. *Circulation* 104, I308-I303 (2001)

215. Gwechenberger M, LH Mendoza, KA Youker, NG Frangogiannis, CW Smith, LH Michael, and ML Entman: Cardiac myocytes produce interleukin-6 in culture and in viable border zone of reperfused infarctions. *Circulation* 99, 546-551 (1999)

216. Daftarian PM, A Kumar, M Kryworuchko, and F Diaz-Mitoma: IL-10 production is enhanced in human T cells by IL-12 and IL-6 and in monocytes by tumor necrosis factor- $\alpha$ . *J Immunol* 157, 12-20 (1996)

217. Fiorentino DF, A Zlotnik, TR Mosmann, M Howard, and A O'Garra: IL-10 inhibits cytokine production by activated macrophages. *J Immunol* 147, 3815-3822 (1991)

218. Mosmann TR: Properties and functions of interleukin-10. *Adv Immunol* 56, 1-26 (1994)

219. de Waal Malefyt R, J Abrams, B Bennett, CG Figdor, and JE de Vries: Interleukin 10(IL-10) inhibits cytokine synthesis by human monocytes: an autoregulatory role of IL-10 produced by monocytes. *J Exp Med* 174, 1209-1220 (1991)

220. Salituro FG, UA Germann, KP Wilson, GW Bemis, T Fox, and MS Su: Inhibitors of p38 MAP kinase: therapeutic

intervention in cytokine-mediated diseases. *Curr Med Chem* 6, 807-823 (1999)

221. Liao P, SQ Wang, S Wang, M Zheng, SJ Zhang, H Cheng, Y Wang, and RP Xiao: p38 Mitogen-activated protein kinase mediates a negative inotropic effect in cardiac myocytes. *Circ Res* 90, 190-196 (2002)

222. Dougherty CJ, LA Kubasiak, H Prentice, P Andreka, NH Bishopric, and KA Webster: Activation of c-Jun N-terminal kinase promotes survival of cardiac myocytes after oxidative stress. *Biochem J* 362, 561-571 (2002)

223. Pantos C, V Malliopolou, I Paizis, P Moraitis, I Mourouzis, S Tzeis, E Karamanoli, DD Cokkinos, H Carageorgiou, D Varonos, and DV Cokkinos: Thyroid hormone and cardioprotection: study of p38 MAPK and JNKs during ischaemia and at reperfusion in isolated rat heart. *Mol Cell Biochem* 242, 173-180 (2003)

224. Anwar A, AA Zahid, KJ Scheidegger, M Brink, and P Delafontaine: Tumor necrosis factor- $\alpha$  regulates insulin-like growth factor-1 and insulin-like growth factor binding protein-3 expression in vascular smooth muscle. *Circulation* 105, 1220-1225 (2002)

225. Tsai BM, M Wang, M Clauss, P Sun, and DR Meldrum: Endothelial monocyte-activating polypeptide II causes NOS-dependent pulmonary artery vasodilation: a novel effect for a proinflammatory cytokine. *Am J Physiol Regul Integr Comp Physiol* 287, R767-R771 (2004)

226. Peng T, X Lu, M Lei, and Q Feng: Endothelial nitric-oxide synthase enhances lipopolysaccharide-stimulated tumor necrosis factor- $\alpha$  expression via cAMP-mediated p38 MAPK pathway in cardiomyocytes. *J Biol Chem* 278, 8099-8105 (2003)

227. Song X, G Li, J Vaage, and G Valen: Effects of sex, gonadectomy, and oestrogen substitution on ischaemic preconditioning and ischaemia-reperfusion injury in mice. *Acta Physiol Scand* 177, 459-466 (2003)

228. Cava sin MA, SS Sankey, AL Yu, S Menon, and XP Yang: Estrogen and testosterone have opposing effects on chronic cardiac remodeling and function in mice with myocardial infarction. *Am J Physiol Heart Circ Physiol* 284, H1560-H1569 (2003)

229. Krieg M, K Smith, and W Bartsch: Demonstration of a specific androgen receptor in rat heart muscle: relationship between binding, metabolism, and tissue levels of androgens. *Endocrinology* 103, 1686-1694 (1978)

230. Marsh JD, MH Lehmann, RH Ritchie, JK Gwathmey, GE Green, and RJ Schiebinger: Androgen receptors mediate hypertrophy in cardiac myocytes. *Circulation* 98, 256-261 (1998)

231. Wehling M: Specific, nongenomic actions of steroid hormones. *Annu Rev Physiol* 59, 365-393 (1997)

232. Chou TM, K Sudhir, SJ Hutchison, E Ko, TM Amidon, P Collins, and K Chatterjee: Testosterone induces dilation of canine coronary conductance and resistance arteries in vivo. *Circulation* 94, 2614-2619 (1996)
233. Yue P, K Chatterjee, C Beale, PA Poole-Wilson, and P Collins: Testosterone relaxes rabbit coronary arteries and aorta. *Circulation* 91, 1154-1160 (1995)
234. Welder AA, JW Robertson, RD Fugate, and RB Melchert: Anabolic-androgenic steroid-induced toxicity in primary neonatal rat myocardial cell cultures. *Toxicol Appl Pharmacol* 133, 328-342 (1995)
235. Remmers DE, WG Cioffi, KI Bland, P Wang, MK Angele, and IH Chaudry: Testosterone: the crucial hormone responsible for depressing myocardial function in males after trauma-hemorrhage. *Ann Surg* 227, 790-799 (1998)
236. Barkett M, D Xue, HR Horvitz, and TD Gilmore: Phosphorylation of IkappaB-alpha inhibits its cleavage by caspase CPP32 in vitro. *J Biol Chem* 272, 29419-29422 (1997)
237. Ghayur T, S Banerjee, M Hugunin, D Butler, L Herzog, A Carter, L Quintal, L Sekut, R Talanian, M Paskind, W Wong, R Kamen, D Tracey, and H Allen: Caspase-1 processes IFN-gamma-inducing factor and regulates LPS-induced IFN-gamma production. *Nature* 386, 619-623 (1997)
238. Ramage P, D Cheneval, M Chvei, P Graff, R Hemmig, R Heng, HP Kocher, A Mackenzie, K Memmert, L Revesz, and et al: Expression, refolding, and autocatalytic proteolytic processing of the interleukin-1 beta-converting enzyme precursor. *J Biol Chem* 270, 9378-9383 (1995)
239. Dinarello CA and BJ Pomerantz: Proinflammatory cytokines in heart disease. *Blood Purif* 19, 314-321 (2001)
240. Miyawaki H, Y Wang, and M Ashraf: Oxidant stress with hydrogen peroxide attenuates calcium paradox injury: role of protein kinase C and ATP-sensitive potassium channel. *Cardiovasc Res* 37, 691-699 (1998)
241. Kang SJ, S Wang, H Hara, EP Peterson, S Namura, S Amin-Hanjani, Z Huang, A Srinivasan, KJ Tomaselli, NA Thornberry, MA Moskowitz, and J Yuan: Dual role of caspase-11 in mediating activation of caspase-1 and caspase-3 under pathological conditions. *J Cell Biol* 149, 613-622 (2000)
242. Guerra S, A Leri, X Wang, N Finato, C Di Loreto, CA Beltrami, J Kajstura, and P Anversa: Myocyte death in the failing human heart is gender dependent. *Circ Res* 85, 856-866 (1999)
243. Konopleva M, S Zhao, Z Xie, H Segall, A Younes, DF Claxton, Z Estrov, SM Kornblau, and M Andreeff: Apoptosis. Molecules and mechanisms. *Adv Exp Med Biol* 457, 217-236 (1999)
244. Vanden Hoek TL, Y Qin, K Wojcik, CQ Li, ZH Shao, T Anderson, LB Becker, and KJ Hamann: Reperfusion, not simulated ischemia, initiates intrinsic apoptosis injury in chick cardiomyocytes. *Am J Physiol Heart Circ Physiol* 284, H141-150 (2003)
245. O'Mahoney ME, S Logue, E Szegezdi, C Stenson-Cox, U Fitzgerald, and A Samali: Hypoxia and ischemia induce nuclear condensation and caspase activation in cardiomyocytes. *Ann N Y Acad Sci* 1010, 728-732 (2003)
246. Dickerman RD, F Schaller, I Prather, and WJ McConathy: Sudden cardiac death in a 20-year-old bodybuilder using anabolic steroids. *Cardiology* 86, 172-173 (1995)
247. Abu-Shakra S, MS Alhalabi, FC Nachtman, RA Schemidt, and WS Brusilow: Anabolic steroids induce injury and apoptosis of differentiated skeletal muscle. *J Neurosci Res* 47, 186-197 (1997)
248. Grady D, W Applegate, T Bush, C Furberg, B Riggs, and SB Hulley: Heart and Estrogen/progestin Replacement Study (HERS): design, methods, and baseline characteristics. *Control Clin Trials* 19, 314-335 (1998)
249. Grady D, NK Wenger, D Herrington, S Khan, C Furberg, D Hunninghake, E Vittinghoff, and S Hulley: Postmenopausal hormone therapy increases risk for venous thromboembolic disease. The Heart and Estrogen/progestin Replacement Study. *Ann Intern Med* 132, 689-696 (2000)
250. Hodis HN, WJ Mack, RA Lobo, D Shoupe, A Sevanian, PR Mahrer, RH Selzer, CR Liu Cr, CH Liu Ch, and SP Azen: Estrogen in the prevention of atherosclerosis. A randomized, double-blind, placebo-controlled trial. *Ann Intern Med* 135, 939-953 (2001)
251. Grodstein F, JE Manson, and MJ Stampfer: Postmenopausal hormone use and secondary prevention of coronary events in the nurses' health study. a prospective, observational study. *Ann Intern Med* 135, 1-8 (2001)
252. Heckbert SR, RC Kaplan, NS Weiss, BM Psaty, D Lin, CD Furberg, JR Starr, GD Anderson, and AZ LaCroix: Risk of recurrent coronary events in relation to use and recent initiation of postmenopausal hormone therapy. *Arch Intern Med* 161, 1709-1713 (2001)
253. Alexander KP, LK Newby, AS Hellkamp, RA Harrington, ED Peterson, S Kopecky, A Langer, P O'Gara, CM O'Connor, RN Daly, RM Califf, S Khan, and V Fuster: Initiation of hormone replacement therapy after acute myocardial infarction is associated with more cardiac events during follow-up. *J Am Coll Cardiol* 38, 1-7 (2001)
254. Collins P: Clinical cardiovascular studies of hormone replacement therapy. *Am J Cardiol* 90, 30F-34F (2002)
255. Canner PL and M Halperin: Implications of findings in the coronary drug project for secondary prevention trials

in coronary heart disease. The coronary; drug project research group. *Circulation* 63, 1342-1350 (1981)

256. Castellsague J, S Perez Gutthann, and LA Garcia Rodriguez: Recent epidemiological studies of the association between hormone replacement therapy and venous thromboembolism. A review. *Drug Saf* 18, 117-123 (1998)

257. Kawano H, T Motoyama, N Hirai, T Yoshimura, K Kugiyama, H Ogawa, H Okamura, and H Yasue: Effect of medroxyprogesterone acetate plus estradiol on endothelium-dependent vasodilation in postmenopausal women. *Am J Cardiol* 87, 238-240, A239 (2001)

258. Miyagawa K, J Rosch, F Stanczyk, and K Hermsmeyer: Medroxyprogesterone interferes with ovarian steroid protection against coronary vasospasm. *Nat Med* 3, 324-327 (1997)

259. Williams JK, EK Honore, SA Washburn, and TB Clarkson: Effects of hormone replacement therapy on reactivity of atherosclerotic coronary arteries in cynomolgus monkeys. *J Am Coll Cardiol* 24, 1757-1761 (1994)

260. Zanger D, BK Yang, J Ardans, MA Waclawiw, G Csako, LM Wahl, and RO Cannon, 3<sup>rd</sup>: Divergent effects of hormone therapy on serum markers of inflammation in postmenopausal women with coronary artery disease on appropriate medical management. *J Am Coll Cardiol* 36, 1797-1802 (2000)

261. Hodis HN and WJ Mack: Atherosclerosis imaging methods: assessing cardiovascular disease and evaluating the role of estrogen in the prevention of atherosclerosis. *Am J Cardiol* 89, 19E-27E; discussion 27E (2002)

262. Dollar AL, AH Kragel, DJ Fernicola, MA Waclawiw, and WC Roberts: Composition of atherosclerotic plaques in coronary arteries in women less than 40 years of age with fatal coronary artery disease and implications for plaque reversibility. *Am J Cardiol* 67, 1223-1227 (1991)

263. Losordo DW, M Kearney, EA Kim, J Jekanowski, and JM Isner: Variable expression of the estrogen receptor in normal and atherosclerotic coronary arteries of premenopausal women. *Circulation* 89, 1501-1510 (1994)

264. Walsh BW, I Schiff, B Rosner, L Greenberg, V Ravnkar, and FM Sacks: Effects of postmenopausal estrogen replacement on the concentrations and metabolism of plasma lipoproteins. *N Engl J Med* 325, 1196-1204 (1991)

265. Nabulsi AA, AR Folsom, A White, W Patsch, G Heiss, KK Wu, and M Szklo: Association of hormone-replacement therapy with various cardiovascular risk factors in postmenopausal women. The Atherosclerosis Risk in Communities Study Investigators. *N Engl J Med* 328, 1069-1075 (1993)

266. Fischer GM and ML Swain: Effects of estradiol and progesterone on the increased synthesis of collagen in atherosclerotic rabbit aortas. *Atherosclerosis* 54, 177-185 (1985)

267. Godsland IF, K Gangar, C Walton, MP Cust, MI Whitehead, V Wynn, and JC Stevenson: Insulin resistance, secretion, and elimination in postmenopausal women receiving oral or transdermal hormone replacement therapy. *Metabolism* 42, 846-853 (1993)

268. Haarbo J, U Marslew, A Gotfredsen, and C Christiansen: Postmenopausal hormone replacement therapy prevents central distribution of body fat after menopause. *Metabolism* 40, 1323-1326 (1991)

269. Stampfer MJ and GA Colditz: Estrogen replacement therapy and coronary heart disease: a quantitative assessment of the epidemiologic evidence. *Prev Med* 20, 47-63 (1991)

270. Ridker PM, CH Hennekens, JE Buring, and N Rifai: C-reactive protein and other markers of inflammation in the prediction of cardiovascular disease in women. *N Engl J Med* 342, 836-843 (2000)

271. Fisher B, J Costantino, C Redmond, R Poisson, D Bowman, J Couture, NV Dimitrov, N Wolmark, DL Wickerham, ER Fisher, and et al: A randomized clinical trial evaluating tamoxifen in the treatment of patients with node-negative breast cancer who have estrogen-receptor-positive tumors. *N Engl J Med* 320, 479-484 (1989)

272. Dai-Do D, E Espinosa, G Liu, TJ Rabelink, F Julmy, Z Yang, F Mahler, and TF Luscher: 17 beta-estradiol inhibits proliferation and migration of human vascular smooth muscle cells: similar effects in cells from postmenopausal females and in males. *Cardiovasc Res* 32, 980-985 (1996)

273. Osborne CK: Tamoxifen in the treatment of breast cancer. *N Engl J Med* 339, 1609-1618 (1998)

274. Guetta V, RM Lush, WD Figg, MA Waclawiw, and RO Cannon, 3<sup>rd</sup>: Effects of the antiestrogen tamoxifen on low-density lipoprotein concentrations and oxidation in postmenopausal women. *Am J Cardiol* 76, 1072-1073 (1995)

275. Walsh BW, LH Kuller, RA Wild, S Paul, M Farmer, JB Lawrence, AS Shah, and PW Anderson: Effects of raloxifene on serum lipids and coagulation factors in healthy postmenopausal women. *Jama* 279, 1445-1451 (1998)

276. Anzano MA, CW Peer, JM Smith, LT Mullen, MW Shrader, DL Logsdon, CL Driver, CC Brown, AB Roberts, and MB Sporn: Chemoprevention of mammary carcinogenesis in the rat: combined use of raloxifene and 9-cis-retinoic acid. *J Natl Cancer Inst* 88, 123-125 (1996)

277. Nozaki M, R Ogata, K Koera, K Hashimoto, and H Nakano: Changes in coagulation factors and fibrinolytic components of postmenopausal women receiving continuous hormone replacement therapy. *Climacteric* 2, 124-130 (1999)
278. Koh KK, MK Horne, 3rd, and RO Cannon, 3<sup>rd</sup>: Effects of hormone replacement therapy on coagulation, fibrinolysis, and thrombosis risk in postmenopausal women. *Thromb Haemost* 82, 626-633 (1999)
279. Meade TW, AP Haines, JD Imeson, Y Stirling, and SG Thompson: Menopausal status and haemostatic variables. *Lancet* 1, 22-24 (1983)
280. Scarabin PY, C Bonithon-Kopp, L Bara, A Malmejac, L Guize, and M Samama: Factor VII activation and menopausal status. *Thromb Res* 57, 227-234 (1990)
281. Folsom AR, KK Wu, CE Davis, MG Conlan, PD Sorlie, and M Szklo: Population correlates of plasma fibrinogen and factor VII, putative cardiovascular risk factors. *Atherosclerosis* 91, 191-205 (1991)
282. Barrett-Connor E, S Slone, G Greendale, D Kritzer-Silverstein, M Espeland, SR Johnson, M Waclawiw, and SE Fineberg: The Postmenopausal Estrogen/Progestin Interventions Study: primary outcomes in adherent women. *Maturitas* 27, 261-274 (1997)
283. McPherson R: Is hormone replacement therapy cardioprotective? Decision-making after the heart and estrogen/progestin replacement study. *Can J Cardiol* 16 Suppl A, 14A-19A (2000)
284. Cushman M: Effects of hormone replacement therapy and estrogen receptor modulators on markers of inflammation and coagulation. *Am J Cardiol* 90, 7F-10F (2002)
285. Cushman M, C Legault, E Barrett-Connor, ML Stefanick, C Kessler, HL Judd, PA Sakkinen, and RP Tracy: Effect of postmenopausal hormones on inflammation-sensitive proteins: the Postmenopausal Estrogen/Progestin Interventions (PEPI) Study. *Circulation* 100, 717-722 (1999)
286. Koh KK, R Mincemoyer, MN Bui, G Csako, F Pucino, V Guetta, M Waclawiw, and RO Cannon, 3<sup>rd</sup>: Effects of hormone-replacement therapy on fibrinolysis in postmenopausal women. *N Engl J Med* 336, 683-690 (1997)
287. Oian P and B Osterud: Monocyte-platelet function and protection against cardiovascular disease. *Maturitas* 23 Suppl, S57-60 (1996)
288. Mammen EF: Oral contraceptives and blood coagulation: a critical review. *Am J Obstet Gynecol* 142, 781-790 (1982)
289. Aune B, P Oian, I Omsjo, and B Osterud: Hormone replacement therapy reduces the reactivity of monocytes and platelets in whole blood--a beneficial effect on atherogenesis and thrombus formation? *Am J Obstet Gynecol* 173, 1816-1820 (1995)
290. Gebara OC, MA Mittleman, P Sutherland, I Lipinska, T Matheney, P Xu, FK Welty, PW Wilson, D Levy, JE Muller, and et al: Association between increased estrogen status and increased fibrinolytic potential in the Framingham Offspring Study. *Circulation* 91, 1952-1958 (1995)
291. Kroon UB, G Silfverstolpe, and L Tengborn: The effects of transdermal estradiol and oral conjugated estrogens on haemostasis variables. *Thromb Haemost* 71, 420-423 (1994)
292. Gilbert J, A Estelles, A Cano, F Espana, R Barrachina, S Grancha, J Aznar, and M Tortajada: The effect of estrogen replacement therapy with or without progestogen on the fibrinolytic system and coagulation inhibitors in postmenopausal status. *Am J Obstet Gynecol* 173, 1849-1854 (1995)
293. Kooistra T: The use of cultured human endothelial cells and hepatocytes as an in vitro model system to study modulation of endogenous fibrinolysis. *Fibrinolysis* 4, 33-39 (1990)
294. van Kesteren PJ, T Kooistra, M Lansink, GJ van Kamp, H Asscheman, LJ Gooren, JJ Emeis, UM Vischer, and CD Stehouwer: The effects of sex steroids on plasma levels of marker proteins of endothelial cell functioning. *Thromb Haemost* 79, 1029-1033 (1998)
295. Sobel MI, CA Winkel, LB Macy, P Liao, and TD Bjornsson: The regulation of plasminogen activators and plasminogen activator inhibitor type 1 in endothelial cells by sex hormones. *Am J Obstet Gynecol* 173, 801-808 (1995)
296. Caine YG, KA Bauer, S Barzegar, H ten Cate, FM Sacks, BW Walsh, I Schiff, and RD Rosenberg: Coagulation activation following estrogen administration to postmenopausal women. *Thromb Haemost* 68, 392-395 (1992)
297. Scarabin PY, M Alhenc-Gelas, G Plu-Bureau, P Taisne, R Agher, and M Aiach: Effects of oral and transdermal estrogen/progesterone regimens on blood coagulation and fibrinolysis in postmenopausal women. A randomized controlled trial. *Arterioscler Thromb Vasc Biol* 17, 3071-3078 (1997)
298. Chandler WL, SL Trimble, SC Loo, and D Mornin: Effect of PAI-1 levels on the molar concentrations of active tissue plasminogen activator (t-PA) and t-PA/PAI-1 complex in plasma. *Blood* 76, 930-937 (1990)
299. Selzman CH, JS Gaynor, AS Turner, TA Whitehill, LD Horwitz, and AH Harken: Estrogen replacement inhibits intimal hyperplasia and the accumulation and effects of transforming growth factor beta1. *J Surg Res* 80, 380-385 (1998)

300. Selzman CH, SA Miller, and AH Harken: Therapeutic implications of inflammation in atherosclerotic cardiovascular disease. *Ann Thorac Surg* 71, 2066-2074 (2001)
301. Weusten JJ, MA Blankenstein, FH Gmelig-Meyling, HJ Schuurman, L Kater, and JH Thijssen: Presence of oestrogen receptors in human blood mononuclear cells and thymocytes. *Acta Endocrinol (Copenh)* 112, 409-414 (1986)
302. Cannon JG and CA Dinarello: Increased plasma interleukin-1 activity in women after ovulation. *Science* 227, 1247-1249 (1985)
303. Kassem M, S Khosla, TC Spelsberg, and BL Riggs: Cytokine production in the bone marrow microenvironment: failure to demonstrate estrogen regulation in early postmenopausal women. *J Clin Endocrinol Metab* 81, 513-518 (1996)
304. Pacifici R, C Brown, E Puscheck, E Friedrich, E Slatopolsky, D Maggio, R McCracken, and LV Avioli: Effect of surgical menopause and estrogen replacement on cytokine release from human blood mononuclear cells. *Proc Natl Acad Sci U S A* 88, 5134-5138 (1991)
305. Polan ML, J Loukides, P Nelson, S Carding, M Diamond, A Walsh, and K Bottomly: Progesterone and estradiol modulate interleukin-1 beta messenger ribonucleic acid levels in cultured human peripheral monocytes. *J Clin Endocrinol Metab* 69, 1200-1206 (1989)
306. Biswas P, F Delfanti, S Bernasconi, M Mengozzi, M Cota, N Polentarutti, A Mantovani, A Lazzarin, S Sozzani, and G Poli: Interleukin-6 induces monocyte chemotactic protein-1 in peripheral blood mononuclear cells and in the U937 cell line. *Blood* 91, 258-265 (1998)
307. Sukovich DA, K Kauser, FD Shirley, V DelVecchio, M Halks-Miller, and GM Rubanyi: Expression of interleukin-6 in atherosclerotic lesions of male ApoE-knockout mice: inhibition by 17beta-estradiol. *Arterioscler Thromb Vasc Biol* 18, 1498-1505 (1998)
308. Zang YC, JB Halder, J Hong, VM Rivera, and JZ Zhang: Regulatory effects of estradiol on T cell migration and cytokine profile: inhibition of transcription factor NF-kappa B. *J Neuroimmunol* 124, 106-114 (2002)
309. Selzman CH, BD Shames, LL Reznikov, SA Miller, X Meng, HA Barton, A Werman, AH Harken, CA Dinarello, and A Banerjee: Liposomal delivery of purified inhibitory-kappaBalpha inhibits tumor necrosis factor-alpha-induced human vascular smooth muscle proliferation. *Circ Res* 84, 867-875 (1999)
310. Libby P: Molecular bases of the acute coronary syndromes. *Circulation* 91, 2844-2850 (1995)
311. Cannon RO, 3rd, BK Yang, and J Ardans: Increased serum matrix levels of metalloproteinase-9 expression in postmenopausal women on estrogen therapy. *J Am Coll Cardiol* 35, 303 (1995)
312. Zhang X, L Wang, H Zhang, D Guo, Z Qiao, and J Qiao: Estrogen inhibits lipopolysaccharide-induced tumor necrosis factor-alpha release from murine macrophages. *Methods Find Exp Clin Pharmacol* 23, 169-173 (2001)
313. Kamada M, M Irahara, M Maegawa, Y Ohmoto, T Takeji, T Yasui, and T Aono: Postmenopausal changes in serum cytokine levels and hormone replacement therapy. *Am J Obstet Gynecol* 184, 309-314 (2001)
314. Fisher B, JP Costantino, DL Wickerham, CK Redmond, M Kavanah, WM Cronin, V Vogel, A Robidoux, N Dimitrov, J Atkins, M Daly, S Wieand, E Tan-Chiu, L Ford, and N Wolmark: Tamoxifen for prevention of breast cancer: report of the National Surgical Adjuvant Breast and Bowel Project P-1 Study. *J Natl Cancer Inst* 90, 1371-1388 (1998)
315. Friend KE, ML Hartman, SS Pezzoli, JL Clasey, and MO Thorner: Both oral and transdermal estrogen increase growth hormone release in postmenopausal women--a clinical research center study. *J Clin Endocrinol Metab* 81, 2250-2256 (1996)
316. Saito S, N Motomura, H Lou, PW Ramwell, and ML Foegh: Specific effects of estrogen on growth factor and major histocompatibility complex class II antigen expression in rat aortic allograft. *J Thorac Cardiovasc Surg* 114, 803-809; discussion 809-810 (1997)
317. Shanker G, M Sorci-Thomas, and MR Adams: Estrogen modulates the inducible expression of platelet-derived growth factor mRNA by monocyte/macrophages. *Life Sci* 56, 499-507 (1995)
318. Auwerx J: The human leukemia cell line, THP-1: a multifaceted model for the study of monocyte-macrophage differentiation. *Experientia* 47, 22-31 (1991)
319. Koyama N, N Morisaki, Y Saito, and S Yoshida: Regulatory effects of platelet-derived growth factor-AA homodimer on migration of vascular smooth muscle cells. *J Biol Chem* 267, 22806-22812 (1992)
320. Kolodgie FD, A Jacob, PS Wilson, GC Carlson, A Farb, A Verma, and R Virmani: Estradiol attenuates directed migration of vascular smooth muscle cells in vitro. *Am J Pathol* 148, 969-976 (1996)
321. Kikuchi N, M Urabe, K Iwasa, T Okubo, H Tsuchiya, T Hosoda, H Tatsumi, and H Honjo: Atheroprotective effect of estradiol and estrone sulfate on human vascular smooth muscle cells. *J Steroid Biochem Mol Biol* 72, 71-78 (2000)
322. Somjen D, F Kohen, A Jaffe, Y Amir-Zaltsman, E Knoll, and N Stern: Effects of gonadal steroids and their antagonists on DNA synthesis in human vascular cells. *Hypertension* 32, 39-45 (1998)



323. Ridker PM, MJ Stampfer, and N Rifai: Novel risk factors for systemic atherosclerosis: a comparison of C-reactive protein, fibrinogen, homocysteine, lipoprotein(a), and standard cholesterol screening as predictors of peripheral arterial disease. *Jama* 285, 2481-2485 (2001)
324. Pasceri V, JT Willerson, and ET Yeh: Direct proinflammatory effect of C-reactive protein on human endothelial cells. *Circulation* 102, 2165-2168 (2000)
325. Cermak J, NS Key, RR Bach, J Balla, HS Jacob, and GM Vercellotti: C-reactive protein induces human peripheral blood monocytes to synthesize tissue factor. *Blood* 82, 513-520 (1993)
326. Ridker PM, CH Hennekens, N Rifai, JE Buring, and JE Manson: Hormone replacement therapy and increased plasma concentration of C-reactive protein. *Circulation* 100, 713-716 (1999)
327. Baumann H and J Gauldie: The acute phase response. *Immunol Today* 15, 74-80 (1994)
328. Walsh BW, DA Cox, A Sashegyi, RA Dean, RP Tracy, and PW Anderson: Role of tumor necrosis factor- $\alpha$  and interleukin-6 in the effects of hormone replacement therapy and raloxifene on C-reactive protein in postmenopausal women. *Am J Cardiol* 88, 825-828 (2001)
329. Caulin-Glaser T, CA Watson, R Pardi, and JR Bender: Effects of 17 $\beta$ -estradiol on cytokine-induced endothelial cell adhesion molecule expression. *J Clin Invest* 98, 36-42 (1996)
330. Caulin-Glaser T, WJ Farrell, SE Pfau, B Zaret, K Bunger, JF Setaro, JJ Brennan, JR Bender, MW Cleman, HS Cabin, and MS Remetz: Modulation of circulating cellular adhesion molecules in postmenopausal women with coronary artery disease. *J Am Coll Cardiol* 31, 1555-1560 (1998)
331. Van Baal WM, JJ Emeis, P Kenemans, H Kessel, ER Peters-Muller, CG Schalkwijk, MJ van der Mooren, and CD Stehouwer: Short-term hormone replacement therapy: reduced plasma levels of soluble adhesion molecules. *Eur J Clin Invest* 29, 913-921 (1999)
332. Seljeflot I, H Arnesen, AE Hofstad, and I Os: Reduced expression of endothelial cell markers after long-term transdermal hormone replacement therapy in women with coronary artery disease. *Thromb Haemost* 83, 944-948 (2000)
333. Oger E, M Alhenc-Gelas, G Plu-Bureau, L Mennen, M Cambillau, L Guize, Y Pujol, and P Scarabin: Association of circulating cellular adhesion molecules with menopausal status and hormone replacement therapy. Time-dependent change in transdermal, but not oral estrogen users. *Thromb Res* 101, 35-43 (2001)
334. Mikkola TS and RW St Clair: Estradiol reduces basal and cytokine induced monocyte adhesion to endothelial cells. *Maturitas* 41, 313-319 (2002)
335. Christodoulakos G, C Panoulis, E Kouskouni, C Chondros, S Dendrinis, and G Creatsas: Effects of estrogen-progestin and raloxifene therapy on nitric oxide, prostacyclin and endothelin-1 synthesis. *Gynecol Endocrinol* 16, 9-17 (2002)
336. Cain BS, DR Meldrum, CH Selzman, JC Cleveland, Jr., X Meng, BC Sheridan, A Banerjee, and AH Harken: Surgical implications of vascular endothelial physiology. *Surgery* 122, 516-526 (1997)
337. Spyridopoulos I, AB Sullivan, M Kearney, JM Isner, and DW Losordo: Estrogen-receptor-mediated inhibition of human endothelial cell apoptosis. Estradiol as a survival factor. *Circulation* 95, 1505-1514 (1997)
338. Steinleitner A, FZ Stanczyk, JH Levin, G d'Ablaing, 3rd, MA Vijod, VL Shahbazian, and RA Lobo: Decreased in vitro production of 6-keto-prostaglandin F1  $\alpha$  by uterine arteries from postmenopausal women. *Am J Obstet Gynecol* 161, 1677-1681 (1989)
339. Mikkola T, P Turunen, K Avela, A Orpana, L Viinikka, and O Ylikorkala: 17  $\beta$ -estradiol stimulates prostacyclin, but not endothelin-1, production in human vascular endothelial cells. *J Clin Endocrinol Metab* 80, 1832-1836 (1995)
340. O'Sullivan MG, JA Goodrich, and MR Adams: Increased prostacyclin synthesis by atherosclerotic arteries from estrogen-treated monkeys. *Life Sci* 69, 395-401 (2001)
341. Ospina JA, DN Krause, and SP Duckles: 17 $\beta$ -estradiol increases rat cerebrovascular prostacyclin synthesis by elevating cyclooxygenase-1 and prostacyclin synthase. *Stroke* 33, 600-605 (2002)
342. Sherman TS, KL Chambliss, LL Gibson, MC Pace, ME Mendelsohn, SL Pfister, and PW Shaul: Estrogen acutely activates prostacyclin synthesis in ovine fetal pulmonary artery endothelium. *Am J Respir Cell Mol Biol* 26, 610-616 (2002)
343. Redmond EM, MN Cherian, and RC Wetzel: 17  $\beta$ -Estradiol inhibits flow- and acute hypoxia-induced prostacyclin release from perfused endocardial endothelial cells. *Circulation* 90, 2519-2524 (1994)
344. Saitta A, D Altavilla, D Cucinotta, N Morabito, N Frisina, F Corrado, R D'Anna, A Lasco, G Squadrito, A Gaudio, F Cancellieri, V Arcoraci, and F Squadrito: Randomized, double-blind, placebo-controlled study on effects of raloxifene and hormone replacement therapy on plasma no concentrations, endothelin-1 levels, and endothelium-dependent vasodilation in postmenopausal women. *Arterioscler Thromb Vasc Biol* 21, 1512-1519 (2001)

345. Che W, N Lerner-Marmarosh, Q Huang, M Osawa, S Ohta, M Yoshizumi, M Glassman, JD Lee, C Yan, BC Berk, and J Abe: Insulin-like growth factor-1 enhances inflammatory responses in endothelial cells: role of Gab1 and MEKK3 in TNF-alpha-induced c-Jun and NF-kappaB activation and adhesion molecule expression. *Circ Res* 90, 1222-1230 (2002)
346. Jhund PS, N Dawson, AP Davie, N Sattar, J Norrie, KP O'Kane, and JJ McMurray: Attenuation of endothelin-1 induced vasoconstriction by 17beta estradiol is not sustained during long-term therapy in postmenopausal women with coronary heart disease. *J Am Coll Cardiol* 37, 1367-1373 (2001)
347. Ishizuka T, M Takamizawa-Matsumoto, K Suzuki, and A Kurita: Endothelin-1 enhances vascular cell adhesion molecule-1 expression in tumor necrosis factor alpha-stimulated vascular endothelial cells. *Eur J Pharmacol* 369, 237-245 (1999)
348. Saito S, RS Aras, H Lou, PW Ramwell, and ML Foegh: Effects of estrogen on nitric oxide synthase expression in rat aorta allograft and smooth muscle cells. *J Heart Lung Transplant* 18, 937-945 (1999)
349. Wagner AH, MR Schroeter, and M Hecker: 17beta-estradiol inhibition of NADPH oxidase expression in human endothelial cells. *Faseb J* 15, 2121-2130 (2001)
350. Duncan AC, JR Petrie, MJ Brosnan, AM Devlin, RA Bass, DS Charnock-Jones, JM Connell, AF Dominiczak, and MA Lumsden: Is estradiol cardioprotection a nitric oxide-mediated effect? *Hum Reprod* 17, 1918-1924 (2002)
351. Gisclard V, VM Miller, and PM Vanhoutte: Effect of 17 beta-estradiol on endothelium-dependent responses in the rabbit. *J Pharmacol Exp Ther* 244, 19-22 (1988)
352. Williams JK, MR Adams, DM Herrington, and TB Clarkson: Short-term administration of estrogen and vascular responses of atherosclerotic coronary arteries. *J Am Coll Cardiol* 20, 452-457 (1992)
353. Collins P, GM Rosano, PM Sarrel, L Ulrich, S Adamopoulos, CM Beale, JG McNeill, and PA Poole-Wilson: 17 beta-Estradiol attenuates acetylcholine-induced coronary arterial constriction in women but not men with coronary heart disease. *Circulation* 92, 24-30 (1995)
354. Reis SE, ST Gloth, RS Blumenthal, JR Resar, HA Zacur, G Gerstenblith, and JA Brinker: Ethinyl estradiol acutely attenuates abnormal coronary vasomotor responses to acetylcholine in postmenopausal women. *Circulation* 89, 52-60 (1994)
355. Yang SH, J Shi, AL Day, and JW Simpkins: Estradiol exerts neuroprotective effects when administered after ischemic insult. *Stroke* 31, 745-749; discussion 749-750 (2000)
356. Toung TJ, RJ Traystman, and PD Hurn: Estrogen-mediated neuroprotection after experimental stroke in male rats. *Stroke* 29, 1666-1670 (1998)
357. Simpkins JW, G Rajakumar, YQ Zhang, CE Simpkins, D Greenwald, CJ Yu, N Bodor, and AL Day: Estrogens may reduce mortality and ischemic damage caused by middle cerebral artery occlusion in the female rat. *J Neurosurg* 87, 724-730 (1997)
358. Nonaka A, J Kiryu, A Tsujikawa, K Yamashiro, K Miyamoto, H Nishiwaki, M Mandai, Y Honda, and Y Ogura: Administration of 17beta-estradiol attenuates retinal ischemia-reperfusion injury in rats. *Invest Ophthalmol Vis Sci* 41, 2689-2696 (2000)
359. Squadrito F, D Altavilla, G Squadrito, GM Campo, M Arlotta, V Arcoraci, L Minutoli, A Saitta, and AP Caputi: The involvement of tumour necrosis factor-alpha in the protective effects of 17 beta oestradiol in splanchnic ischaemia-reperfusion injury. *Br J Pharmacol* 121, 1782-1788 (1997)
360. Zhai P, TE Eurell, PS Cooke, DB Lubahn, and DR Gross: Myocardial ischemia-reperfusion injury in estrogen receptor-alpha knockout and wild-type mice. *Am J Physiol Heart Circ Physiol* 278, H1640-1647 (2000)
361. Zhai P, TE Eurell, RP Cotthaus, EH Jeffery, JM Bahr, and DR Gross: Effects of dietary phytoestrogen on global myocardial ischemia-reperfusion injury in isolated female rat hearts. *Am J Physiol Heart Circ Physiol* 281, H1223-1232 (2001)
362. McNulty PH, D Jagasia, JM Whiting, and T Caulin-Glaser: Effect of 6-wk estrogen withdrawal or replacement on myocardial ischemic tolerance in rats. *Am J Physiol Heart Circ Physiol* 278, H1030-1034 (2000)
363. Benjamin IJ and E Christians: Exercise, estrogen, and ischemic cardioprotection by heat shock protein 70. *Circ Res* 90, 833-835 (2002)
364. Benjamin IJ and DR McMillan: Stress (heat shock) proteins: molecular chaperones in cardiovascular biology and disease. *Circ Res* 83, 117-132 (1998)
365. Dillmann WH: Small heat shock proteins and protection against injury. *Ann N Y Acad Sci* 874, 66-68 (1999)
366. Karmazyn M, K Mailer, and RW Currie: Acquisition and decay of heat-shock-enhanced postischemic ventricular recovery. *Am J Physiol* 259, H424-431 (1990)
367. Paroo Z, JV Haist, M Karmazyn, and EG Noble: Exercise improves postischemic cardiac function in males but not females: consequences of a novel sex-specific heat shock protein 70 response. *Circ Res* 90, 911-917 (2002)

## Sex hormones in cardiac injury and protection

368. Vaccarino V, TR Holford, and HM Krumholz: Pulse pressure and risk for myocardial infarction and heart failure in the elderly. *J Am Coll Cardiol* 36, 130-138 (2000)

369. Hayward CS, CM Webb, and P Collins: Hormone replacement therapy and heart-rate variability. *Lancet* 354, 256 (1999)

370. McCredie RJ, JA McCrohon, L Turner, KA Griffiths, DJ Handelsman, and DS Celermajer: Vascular reactivity is impaired in genetic females taking high-dose androgens. *J Am Coll Cardiol* 32, 1331-1335 (1998)

371. McCrohon JA, W Jessup, DJ Handelsman, and DS Celermajer: Androgen exposure increases human monocyte adhesion to vascular endothelium and endothelial cell expression of vascular cell adhesion molecule-1. *Circulation* 99, 2317-2322 (1999)

372. Mukherjee TK, H Dinh, G Chaudhuri, and L Nathan: Testosterone attenuates expression of vascular cell adhesion molecule-1 by conversion to estradiol by aromatase in endothelial cells: implications in atherosclerosis. *Proc Natl Acad Sci U S A* 99, 4055-4060 (2002)

373. Ayashi A, R Mathur, and P Halushka: Testosterone increases human platelet thromboxane A2 receptor density and aggregation responses. *Circulation* 91, 2742-2747 (1995)

**Key Words:** Estrogen, testosterone, heart, injury, cytokines, TNF, IL-1, IL-6, Ischemia, Myocardium, Sex Hormones, Gender, Review

**Send correspondence to:** Dr. Daniel R. Meldrum, Departments of Surgery and Cellular and Integrative Physiology, Indiana University Medical Center, 545 Barnhill Drive, Emerson Hall Room 215, Indianapolis, Indiana 46202, Tel:317-313-5217, Fax:317-274-2940, E-mail: dmeldrum@iupui.edu

<http://www.bioscience.org/current/vol10.htm>