

## ANDROGEN METABOLIC GENES IN PROSTATE CANCER PREDISPOSITION AND PROGRESSION

Nick M. Makridakis<sup>1</sup>, Grant Buchanan<sup>2</sup>, Wayne Tilley<sup>2</sup> and Juergen K. V. Reichardt<sup>1,3</sup>

<sup>1</sup> Department of Biochemistry and Molecular Biology, Institute for Genetic Medicine and <sup>3</sup> Department of Preventive Medicine, USC Keck School of Medicine, 2250 Alcazar Street, Los Angeles, CA 90089-9075, USA, <sup>2</sup> Dame Roma Mitchell Cancer Research Laboratories, University of Adelaide & Hanson Institute, PO Box 14, Rundle Mall, Adelaide, SA 5000, Australia, <sup>3</sup> University of Sydney, Medical Foundation Building (K25), 92 - 94 Parramatta Rd, Camperdown, Sydney, NSW 2042, Australia

### TABLE OF CONTENTS

1. Abstract
2. Prostate cancer
3. Androgens and Prostate Cancer
4. Steroid 5 $\alpha$ -Reductase
5. Androgen Receptor
6. 3 $\beta$ -Hydroxysteroid Dehydrogenase
7. 17 $\beta$ -Hydroxysteroid Dehydrogenase
8. 17 $\alpha$ -Hydroxylase/17, 20 Lyase
9. Conclusions
10. Acknowledgements
11. References

### 1. ABSTRACT

Significant evidence implicates androgens in prostate cancer etiology. We review recent data with regard to the association between several allelic variants of specific androgen-metabolic genes and the predisposition to prostate cancer. We also review the emerging evidence regarding the role of genetic variants of these genes as well as the androgen receptor in prostate cancer progression. Based on the prostate cancer paradigm, we propose that a multidisciplinary attack on the problem—involving biochemistry, genetics, pharmacogenetics, endocrinology and molecular epidemiology—may be important for the understanding and successful treatment of complex (in terms of etiology) human diseases.

### 2. PROSTATE CANCER

It is estimated that 232,090 U.S. men will be newly diagnosed with prostate cancer in 2005 and 30,350 will die of this disease (1). Prostate cancer is the most common malignancy among men in the United States (1). Prostate cancer also represents a substantial public health problem in other industrialized nations, for example those of the European Union (2) This disease is rare in younger men before the age of 40, but the rate of increase thereafter is greater than for any other cancer(3). There is a large variation in prostate cancer rates between racial/ethnic groups. For example, in Los Angeles, African-Americans, who also have the highest prostate cancer rate in the world, have a 70% higher rate than Caucasian Americans, who

## Androgens and Prostate Cancer

have a substantially higher rate than Hispanic Americans (Latinos), while Chinese- and Japanese-Americans (i.e. Asian-Americans) have roughly one-half the rate of Caucasians (4). It is noteworthy that Chinese and Japanese men have among the lowest prostate cancer rates in the world, they are about 1/8th to 1/20th the rates of their U.S. counterparts(4). These data strongly suggest a substantial genetic component to prostate cancer risk, although environmental factors are also known to play an important role. Finally, there is significant evidence of more aggressive disease and a less favorable outcome (e.g., lower survival) for African-American prostate cancer patients than their Caucasian-American counterparts (5), which may be at least partially due to differences in socioeconomic status.

There is a significant familial and, therefore, most likely genetic component to prostate cancer risk. In younger men, the familial form is most consistent with an autosomal dominant way of inheritance (6). The location and action of putative familial loci for prostate cancer, however, remain controversial (reviewed in reference 7). Here we examine recent findings of both constitutional SNPs (single nucleotide polymorphisms) and somatic mutations in androgen-metabolic genes and the conclusions that can be drawn for prostate cancer predisposition and/or progression.

### 3. ANDROGENS AND PROSTATE CANCER

Androgens play a critical role in both normal and abnormal prostate development (Figure 1). Studies of androgens and prostate cancer go back over 60 years for which Charles Huggins won the Nobel Prize for his discoveries concerning the hormonal treatment of prostate cancer in 1966 (8, 9). In men, testosterone is synthesized in large amounts, primarily by the Leydig cells of the testes (Figure 1) (10). Testosterone is transported into the prostate where it is irreversibly metabolized intracellularly to dihydrotestosterone (DHT; Figure 1) (10). DHT (or, much less efficiently, testosterone) is bound by the intracellular androgen receptor (AR; Figure 1) (10). This complex then translocates to the cell nucleus where it activates the transcription of various genes with androgen-responsive elements (AREs) in their promoters (Figure 1) (10). DHT can also be inactivated in the prostate by further reduction to 3 $\alpha$ - or 3 $\beta$ -androstenediol by the 3 $\alpha$ - and 3 $\beta$ -hydroxysteroid dehydrogenase enzymes (10).

Numerous epidemiologic studies have examined the association between serum androgen levels and prostate cancer risk (reviewed in reference 11). In this review we will instead focus on the association between androgen metabolic genotypes and prostate cancer.

### 4. STEROID 5 $\alpha$ -REDUCTASE

Steroid 5 $\alpha$ -reductase (or testosterone 5 $\alpha$ -reductase; 10, 12) catalyzes the irreversible conversion of testosterone to DHT, the most potent androgen. Two isozymes exist: the type I steroid 5 $\alpha$ -reductase which is encoded by the SRD5A1 gene and the type II isozyme,

which in turn is encoded by the SRD5A2 gene (Table 1)(10, 12). The type I isozyme is primarily expressed in liver and in skin cells, whereas type II plays a major role in prostate development and disease predisposition. The SRD5A2 gene is located on chromosome 2 (band 2p23; Table 1) and spans over 40 kb of genomic DNA, with 5 exons and four introns (10). Many mutations in the SRD5A2 gene that cause a rare form of male pseudohermaphroditism have been identified and characterized by functional studies (for examples, see references 10 and 12).

Several investigations have examined the androgen metabolic gene SRD5A2 as a candidate gene for prostate cancer. In an extensive screen for SRD5A2 SNPs (single nucleotide polymorphisms;13) a nonsynonymous SNP was found, the A49T variant (Figure 2), which results in the replacement of the normal alanine in codon 49 with threonine (Table 1). The A49T missense substitution has been reported to significantly increase the risk of prostate cancer in different populations (14-16) and increases the apparent maximal steroid 5 $\alpha$ -reductase activity ( $V_{max}$ ) 5-fold *in vitro* (14). Therefore, the kinetic data provide a likely explanation for the increased risk associated with the A49T mutation. Other epidemiologic studies, however, have reported a non-significant association between the A49T variant and prostate cancer predisposition (17-19). Another SRD5A2 SNP, the V89L missense substitution (replacing the valine at codon 89 with the polymorphic leucine; Table 1; Figure 2) has also been associated with an increased prostate cancer risk (20). Other studies however, have found no association between the V89L SNP and prostate cancer (21). These ambiguities may perhaps be explained by the distinct racial/ethnic make-up of the samples and the modest biochemical effect of the V89L SNP discussed below.

Functional *in vitro* studies of SRD5A2 variants have provided critical mechanistic insights and often parallel *in vivo* findings (6,22,23). As noted above, the A49T substitution results in a significant increase in  $V_{max}$ , suggesting that the substantially increased risk associated with this mutation may be the result of higher intraprostatic DHT levels caused by the A49T variant. The V89L variant in contrast results in a very modest decrease in  $V_{max}$  (23). Functional data may be important in choosing candidate SNPs suitable for epidemiologic studies (24) since an almost 200-fold variation has been demonstrated in the  $V_{max}$  of various SRD5A2 variants (23). Therefore, variants with substantially higher (or lower) activity may be more suitable markers for molecular-epidemiologic association investigations than neutral polymorphisms, because mechanistically important variants may be directly responsible for the observed associations (24).

The SRD5A2 locus also encodes significant (~60-fold) pharmacogenetic variation for the competitive steroid 5 $\alpha$ -reductase inhibitor finasteride (22, 23, 25). Interestingly, finasteride has a reduced affinity for the A49T mutant enzyme *in vitro* (22, 23, 25). Similar variation was identified for many other SRD5A2 enzyme variants and for two other 5 $\alpha$ -reductase inhibitors (23, 25).



## Androgens and Prostate Cancer

**Table 1.** Characteristics of androgen-metabolic genes analyzed in prostate cancer

Gene	Gene Product	Chromosomal Location	Constitutional Mutations	Somatic Mutations
AR	Androgen receptor	Xq11-12	30,31,117	32,33,117
SRD5A2	Steroid 5 $\alpha$ -reductase	2p23	17, 24, 25	27
HSD3B2	3B-hydroxysteroid dehydrogenase	1p13	31	None reported
CYP17	Cytochrome p450c17	10q24.3	32, 33	None reported
HSD17B3	17B-hydroxysteroid dehydrogenase	9q22	42	None reported

The numbers in this table indicate individual references. The presence of constitutional and/or somatic mutations refers to DNA from either patients' lymphocytes or their affected tumor tissue. Only polymorphic variants (31) have been reported in the HSD3B2 gene.

This significant pharmacogenetic variation should be considered when prescribing steroid 5 $\alpha$ -reductase inhibitors for prostatic diseases. Recently, the PCPT (prostate cancer prevention trial) reported that treatment of men 55 years of age or older with finasteride resulted in a significant decrease in prostate cancer incidence measured over a seven-year period (26), suggesting that finasteride prevents or delays the appearance of prostate cancer. However, finasteride treatment also resulted in a significant increase in the incidence of prostate tumors of high grade (7 or higher) Gleason score (26). This unexpected finding may at least partially be explained by the presence of specific SRD5A2 variants (like A49T; Figure 2) that are not efficiently inhibited by finasteride, in a subset of the study population. For individuals carrying the A49T allele, the competitive steroid 5 $\alpha$ -reductase inhibitor dutasteride might have been a better choice (23, 25). Thus, pharmacogenetic considerations need to play an important role in future trials and treatment regimens.

In addition to affecting predisposition, the SRD5A2 gene may also play an important role in prostate cancer progression. First, common somatic genetic alterations in a (TA)<sub>n</sub> dinucleotide repeat polymorphism located in the 3'-untranslated region (UTR; Figure 2) of the SRD5A2 gene were reported in prostate tumors (27). Subsequently, the same prostate tumors were reported to contain both somatic missense mutations and other *de novo* somatic mutations (perhaps best called somatic polymorphisms) that do not alter enzyme activity, in the coding region of the SRD5A2 gene (28). Most of the recurrent somatic SRD5A2 mutations identified increased 5 $\alpha$ -reductase activity (28), suggesting that higher intraprostatic DHT steady state levels resulting from these mutations may play a role in prostate cancer progression (Figure 2). Thus, the SRD5A2 gene may be significantly involved in both predisposition and progression of prostate cancer. This dual involvement is very interesting and may open new avenues of research that may, in turn, result in more rational and/or personalized treatment of patients and even chemoprevention. Finally, the involvement of this androgen-metabolic gene in prostate cancer progression may lead to new strategies for treating androgen-independent tumors by considering both SRD5A2 and AR gene mutations (23, 25, 27, 29-33).

### 5. ANDROGEN RECEPTOR

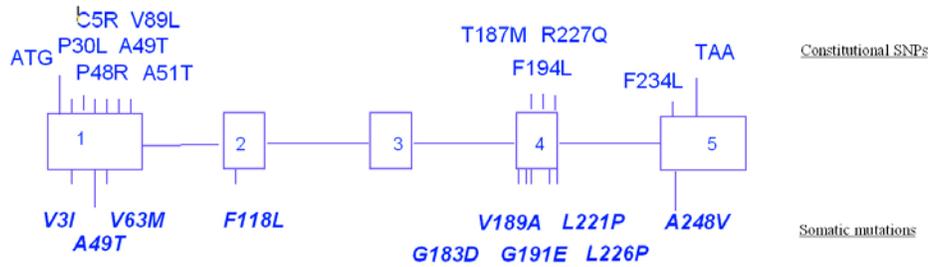
The androgen receptor (AR), which is the pivotal component of the androgen metabolic cascade / androgen signaling axis, is a member of the superfamily of nuclear transcription factors that regulate a diverse range of cellular

functions by providing a direct link between signaling molecules and gene transcription (34-36). The AR gene contains a polymorphic trinucleotide (CAG)<sub>n</sub> microsatellite (which is often simply referred to as CAG) normally distributed in the population between 6-39 repeats with an average of 21 (37, 38). The (CAG)<sub>n</sub> microsatellite encodes a polyglutamine (poly-Q) tract in the receptor, the length of which is inverse related to AR transactivation activity (39-41). The observation that the allelic frequency distribution of AR-(CAG)<sub>n</sub> repeat length paralleled the different incidence of prostate cancer amongst different U.S. racial-ethnic populations (42) led to several pilot and then case-controlled studies demonstrating an approximate 2-fold increased prostate cancer risk, decreased age of onset and/or increased risk of advanced disease for reduced AR-(CAG)<sub>n</sub> repeat length (38, 43-47). However, this association has not been consistently observed, possibly due to small sample sizes, population differences and/or failure to appropriately match cases and controls (48- 57). Importantly, recent biochemical evidence suggests that the effect of polyQ tract length on AR function may be more complex than a simple linear relationship with AR activity (39). Reassessment of cancer risk and (CAG)<sub>n</sub> repeat length using this new model is therefore clearly warranted.

AR status and a functional androgen signaling axis are particularly important for patients who are either diagnosed with, or subsequently develop, metastatic prostate cancer, as they determine the success of androgen ablation (i.e. orchidectomy, treatment with LHRH agonists/antagonists and/or AR antagonists), the only treatment option in the advanced disease setting (58, 59). Despite an initial response to these therapies in 80-90% of patients with metastatic disease, treatment is essentially palliative and disease progression eventually ensues (59, 60). Paradoxically, the failure of androgen ablation therapy in a clinical setting is accompanied by rising serum levels of the androgen regulated protein, PSA (prostate specific antigen). This observation provided the impetus to further examine the androgen signaling pathway in prostate cancer, with the result that it is now well accepted that the cellular mediator of androgen action, the AR (Table 1; Figs. 1 and 3), remains a major determinant of progression and survival in both early stage prostate cancer and in a castrate environment following androgen ablation (33, 61).

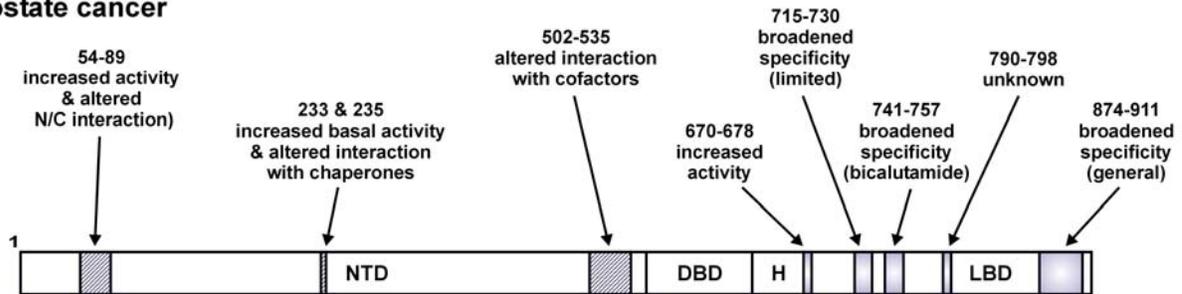
Many studies have addressed the role of AR in prostate cancer using xenograft mouse models of the human disease. Analogous to the clinical setting, prostate cancer xenograft lines typically grow only in an androgen-dependent state and are sensitive to androgen ablation, but frequently regrow in a low androgen environment

## Androgens and Prostate Cancer



**Figure 2.** Steroid 5 $\alpha$ -reductase type II missense SNPs (in constitutional DNA) and somatic mutations shown on the SRD5A2 exons. The translation start and stop codons are marked above the respective exons. SNPs in constitutional (or “germline”) and somatic mutations are indicated separately.

### Prostate cancer



### Androgen insensitivity



**Figure 3.** Collocation of androgen receptor mutations in prostate cancer and the inherited form of androgen insensitivity. The regions of the AR ligand binding domain (LBD) in which 80% of the missense AR mutations identified in prostate cancer and AIS collocate are indicated (grey boxes<sup>32,33</sup>). Regions in the AR DNA binding (DBD) and amino-terminal transactivation (NTD) domains where missense mutations have been shown to exert altered receptor function in prostate cancer<sup>33,39,71,78,79,118,119</sup> and AIS<sup>117</sup> are also designated (hatched boxes). Broadly defined effects of naturally occurring mutations detected in prostate cancer and AIS within regions of collocation are indicated.

following castration and continue to secrete PSA. In support of the hypothesis that AR continues to play an important role in growth of castrate-resistant prostate cancer, several studies have demonstrated an increase in the level of AR and/or AR regulated genes in xenograft tumors following castration in comparison to untreated tumors (62-65). In particular, comprehensive gene expression profiling determined that the AR was the only gene upregulated in the progression from androgen sensitive to castration resistant growth in all seven human prostate cancer xenograft models analyzed (63). In clinical disease, recent studies have demonstrated an increase in both AR mRNA and protein levels during progression from localized disease to metastatic castrate-resistant prostate cancer (33). Moreover, increased AR levels in clinically localized disease have been associated with relapse following radical prostatectomy, suggesting that the AR is a determinant of metastatic spread (66, 67).

Several mechanisms have been proposed to contribute to increased AR levels in castrate resistant tumors, including increased expression of the AR and AR gene amplification, the latter being demonstrated in up to 29% of castrate resistant tumors along with a concomitant increase in androgen regulated genes (68-70).

The AR has a modular structure, consisting primarily of a large amino-terminal transactivation domain (NTD), a centrally located DNA-binding domain (DBD), and a carboxy-terminal ligand binding domain (LBD) that confers high affinity and specificity for androgen binding (Figure 3). AR gene mutations have been reported in prostate cancer at a frequency of 5-50%, with a higher frequency reported in more advanced disease and following androgen ablation therapy (33). In contrast to the inherited syndrome of androgen insensitivity, where AR gene mutations result in a loss of receptor function, those

## Androgens and Prostate Cancer

detected in prostate cancer occur in distinctly different regions of the receptor, and generally confer promiscuity or increased sensitivity for receptor activation by other steroid hormones and/or by the specific antiandrogen used in clinical management of the disease (Figure 3) (32, 33, 71). These observations led to the concept of "therapy-mediated selection pressure", which is exemplified by the detection of AR gene mutations which confer enhanced receptor activity in response to flutamide but not to other antiandrogens in tumors from patients treated with hydroxyflutamide plus orchidectomy or LHRH agonists/antagonists, as part of a combined androgen blockade strategy (72, 73). Clinically, patients who progress during treatment with one antiandrogen often respond to another (74, 75). Studies of the autochthonous TRAMP model of prostate cancer (76, 77) demonstrate that therapy mediated selection of AR mutations is not limited to antiandrogen treatment. Whereas non-castrate mice develop mutations in the AR LBD, mice castrated at 12 weeks of age developed mutations in the NTD (78). Enforced expression of the AR-NTD variant, E231G, in the mouse prostate confers rapid development of prostatic intra-epithelial neoplasia that progresses to invasive and metastatic disease in 100% of mice (79). This finding suggests that the AR can also act in an appropriate environment as a proto-oncogene to promote tumor formation.

Extended culture of LNCaP human prostate cancer cells in androgen depleted media resulted in sensitization of the AR to 10-fold lower concentrations of androgens and the ability to respond to bicalutamide, without changes in AR structure or level (80). These effects may be explained by changes in the level or function of proteins that orchestrate specific aspects of AR signaling. For example, the transcriptional activity of the AR is mediated, in part, by the recruitment of coregulator proteins that enhance (coactivators) or repress (corepressors) receptor function (Figure 1). An increase in coactivator levels has been shown in prostate cancer compared to benign prostate tissues, and during the development of castrate resistant disease (81- 85). These coactivator proteins can selectively enhance the activity of the AR upon binding of alternative ligands, such as estradiol and hydroxyflutamide, sensitize the AR to lower concentrations of native and non-native ligands, or promote different modes of AR activation by altering conformational changes and homodimerisation following ligand binding (78, 82, 86-88). Conversely, decreased expression of corepressors such as NCoR and SMRT, which mediate, in part, the antagonist action of bicalutamide, flutamide and mifepristone, would facilitate increased agonist activity of these agents (89, 90). However, while more than 120 AR interacting proteins have been described, it is not currently known how the majority of these influence receptor function, or which are expressed in the prostate and have altered expression during disease etiology (Figure 1).

The evolving evidence that the AR is one of the key critical mediators of continued prostate cancer growth following hormone ablation makes a compelling argument for the development of new strategies that directly target

the receptor (33). Such new therapies are necessary for significant advances in the management of patients with advanced prostate cancer who will ultimately fail androgen ablation. Several approaches to achieve this goal have been documented. Reducing the level of AR and suppression of human prostate cancer cell growth *in vitro* and *in vivo* has been achieved with a variety of methods, incl. double-stranded RNA interference, antisense oligonucleotides, hammerhead ribozymes and analogues of the ansamycin antibiotics such as 17-allylamino-17-demethoxygeldanamycin (91- 98). Inhibition of AR function has been achieved with dominant negative AR inhibitors, microinjection of AR antibodies, "decoy" double stranded DNA fragments containing specific AR response elements and histone deacetylase inhibitors including suberoylanilide hydroxamic acid and phenylbutyrate (94, 99-102). Providing issues such as the mode of delivery and the potential for disruption of multiple signaling pathways can be circumvented, these newer approaches that directly target the AR provide considerable promise as therapeutic options for castrate-resistant prostate cancer.

### 6. 3 $\beta$ -HYDROXYSTEROID DEHYDROGENASE

DHT, the most active intraprostatic androgen, is inactivated through two reductive reactions catalyzed by 3 $\alpha$ - and 3 $\beta$ -hydroxysteroid dehydrogenase (Figure 1)(10). Therefore, 3 $\beta$ -hydroxysteroid dehydrogenase is critical for the regulation of intraprostatic DHT steady state levels by affecting its degradation rate(10). Human 3 $\beta$ -hydroxysteroid dehydrogenase can catalyze a number of steroid substrates, including DHT (103). Enzyme activity is encoded by two homologous and closely linked loci: the HSD3B1 and HSD3B2 genes, which are both located in chromosome band 1p13 (Table 1) (103, 104). The type II 3 $\beta$ -hydroxysteroid dehydrogenase enzyme encoded by the HSD3B2 gene is expressed in androgenic tissues (103) and may therefore regulate DHT levels by initiating the inactivation of this potent androgen in the prostate.

A complex (TG)<sub>n</sub> (TA)<sub>n</sub> (CA)<sub>n</sub> repeat (105) in the HSD3B2 gene has been reported to be very polymorphic, consisting of at least 25 different alleles (106). Racial/ethnic variation of this complex repeat in the HSD3B2 locus parallels prostate cancer risk (106), suggesting that this locus may also play a role in prostate cancer predisposition and/or progression (Table 1). No missense HSD3B2 mutations associated with either predisposition or progression of prostate cancer have yet been reported to date. However, SNPs that do not change amino acids have been associated with prostate cancer predisposition in both HSD3B2 and HSD3B1 genes, and these SNPs seem to have an additive effect on risk (107).

### 7. 17 $\beta$ -HYDROXYSTEROID DEHYDROGENASE

Another androgen metabolic locus that may play a role in prostate cancer predisposition is the HSD17B3 gene which encodes 17 $\beta$ -hydroxysteroid dehydrogenase (HSD) type III, also called testicular 17-ketoreductase (Figure 1) (108). This enzyme utilizes NADP(H) as a

## Androgens and Prostate Cancer

cofactor and favors the reduction of androstenedione to testosterone, in the testis (109). Thus, mutations in this gene may increase the production of T in the testes, which can directly or indirectly (through DHT) activate the AR, potentially increasing prostate cancer risk.

The HSD17B3 gene is located in chromosomal band 9q22 (Table 1) and contains 11 exons (109). At least 18 mutations and one polymorphism that cause male pseudohermaphroditism have been functionally characterized in this gene (see e.g. reference 109). Another SNP, the G289S (glycine at codon 289 replaced by serine) missense substitution, has been reported to significantly increase risk for sporadic prostate cancer (110). Thus this androgen metabolic enzyme may also be involved in prostate cancer predisposition.

### 8. 17 $\alpha$ -HYDROXYLASE/17,20 LYASE

The enzyme cytochrome p450c17 (or steroid 17  $\alpha$ -hydroxylase/17,20 lyase) catalyzes two sequential reactions in the biosynthesis of testosterone in both the gonads and the adrenals; this enzyme is the product of the CYP17 gene (Fig. 1; Table 1; 10). A polymorphic T to C transition (A1/A2 allele) has been identified in the 5' untranslated region (UTR) of the CYP17 gene (111, 112). This polymorphism has been reported at increased frequency in Caucasian prostate cancer patients, compared with controls (20, 111, 112). Another study, however, reported no significant difference in the frequency of this SNP between Caucasian prostate cancer patients and controls except for patients with a family history of the disease (108). In addition, this SNP does not appear to play any role in early-onset prostate cancer (113).

## 9. CONCLUSIONS

In this review we presented the evidence and rationale supporting the involvement of a series of androgen-metabolic candidate genes in prostate cancer predisposition and progression. Selection of these genes was guided by endocrinologic, epidemiologic, biochemical and molecular criteria. We discuss that while some of the allelic variants examined were consistently shown to be associated with increased prostate cancer risk, many other variants show significant variability in risk. The reasons for this variability are unknown, but may include small sample size, distinct racial/ ethnic background of the samples, or a minor effect on the risk exerted by these variants. Deciphering the role of such variants may be difficult without functional studies that may support their role in disease etiology and/ or progression.

The molecular-epidemiologic approach presented here for the androgen-metabolic genes has general applicability. First, SNPs (13) are identified and then their individual contributions to the phenotype are measured, in concert with other allelic variants in the same or other genes. Second, molecular-epidemiologic investigations are supported by appropriate biochemical and pharmacologic/pharmacogenetic studies (*in vitro* and/or *in vivo*) that determine the functional significance of each

allelic variant (15, 114). These multidisciplinary investigations are likely to involve epidemiologic, pharmacologic, molecular and biochemical methods. This convergence of various disciplines will probably result in an integrated molecular view of complex disease phenotypes, which can then be complemented by the analysis of environmental contributions.

In summary, the multidisciplinary analysis of androgen-metabolic genes in prostate cancer has proven useful for extending our knowledge of both the predisposition and the progression of this disease. Further investigations are likely to yield additional advances in understanding this significant public health problem. Extensions of this approach to other complex human diseases, such as benign prostatic hyperplasia (BPH), are also likely to lead to important insights into the etiology of these diseases (115).

The approach presented here focused exclusively on the sporadic form of prostate cancer. There are, however, two forms of the disease: a common, sporadic form and a much more uncommon familial form, a dichotomy typical of many complex human diseases (116). It is noteworthy in this context that none of the genes discussed in this review map to any of the regions that have been associated by linkage with the familial disease (Table 1 and data not shown). This finding suggests that the more common, sporadic form of prostate cancer and the infrequent familial form may have distinct etiologies. This hypothesis may also have significant implications for the investigation of other complex human phenotypes.

SNPs have come into widespread use already (13) Unfortunately, little attention is currently being paid to the examination of the functional significance of SNPs (24), perhaps due to the variability and hence difficulty of the assays required to functionally characterize each SNP (which makes it difficult to automate the functional analysis). The molecular epidemiologic data reviewed here have implications for association and linkage studies with SNP markers. Specifically, the epidemiologic and kinetic data on the rare A49T missense mutation in the SRD5A2 gene highlight the enormous importance of functional studies for SNP investigations. Therefore, extensive functional analyses should be pursued for all missense SNPs throughout the genome (and perhaps all SNPs), however time consuming this may be. In fact, we propose that SNPs should be classified as functionally neutral or functionally significant based on appropriate experimental data. The former may be best used in linkage studies, while the latter may be best used as "candidate alleles" (or "candidate SNPs") in candidate genes. In other words, functionally neutral SNPs are useful in identifying appropriate candidate genes for each phenotype, while functionally significant SNPs are useful for epidemiologic (i.e., association) and pharmacologic studies, because they are more likely to actually cause the phenotype (24). Therefore, the multidisciplinary analysis of prostate cancer may have resulted in the discovery of significant new strategies for human molecular genetic and epidemiologic research into multifactorial (complex) phenotypes.

10. ACKNOWLEDGMENTS

This work was supported by grants to NMM (ACS-IRG-58-007-45), WT (National Health and Medical Research Council of Australia ID#299048), GB (United States Department of Defense W81XWH-04-1-0017) and JKVR (NCI R01 68581 and NCI P01 108964 [project1]).

11. REFERENCES

1. Jemal, A., Murray, T., Ward, E., Samuels, A., Tiwari, R.C., Ghafoor, A., Feuer, E.J. and Thun, M.J. Cancer statistics, 2005. *CA Cancer J. Clin.* 55, 10, (2005)

2. Black, R.J., Bray, F., Ferlay, J. and Parkin, D.M. Cancer incidence and mortality in the European Union: cancer registry data and estimates of national incidence for 1990. *Eur. J. Cancer* 33, 1075, (1997)

3. SEER: Cancer incidence and mortality in the United States, 1973-1981. Bethesda, MD: U.S. Department of Health and Human Services, 1984. (2005)

4. Bernstein, L. and Ross, R. K. Cancer in Los Angeles County, A Portrait of Incidence and Mortality. Los Angeles, CA: University of Southern California and the California Tumor Registry, Department of Health Services, State of California, 1991. (2005)

5. Brawley, O.W., Knopf, K. and Merrill, R. The epidemiology of prostate cancer part I: descriptive epidemiology. *Semin. Urol. Oncol.* 16, 187, (1998)

6. Carter, B.S., Bova, G.S., Beaty, T.H., Steinberg, G.D., Childs, B., Isaacs, W.B. and Walsh, P.C. Hereditary prostate cancer: epidemiologic and clinical features. *J. Urol.* 150, 797, (1993)

7. Simard, J., Dumont, M., Soucy, P. and Labrie, F. Perspective: prostate cancer susceptibility genes. *Endocrinology* 143, 2029, (2002)

8. Huggins, C., Hodges, C.V. Studies on prostatic cancer: I. The effect of castration, of estrogen and of androgen injection on serum phosphatases in metastatic carcinoma of the prostate. *Cancer Res* 1, 293, (1941)

9. Huggins, C., Stevens, R.E. and Hodges, C.V. Studies on prostatic cancer. II. The effect of castration on advanced carcinoma of the prostate gland. *Arch. Surg.* 43, 209, (1941)

10. Cheng, E., Lee, C., and Grayhack, J.T. In Lepor, H., Lawson, R.K. (eds), Prostate Diseases. W.B. Saunders, Philadelphia, PA, pp. 57-71. (1993)

11. Bosland, M.C. The role of steroid hormones in prostate carcinogenesis. *J. Natl. Cancer Inst. Monogr* 39, (2000)

12. Russell, D.W., Wilson, J.D. Steroid 5 alpha-reductase: two genes/two enzymes. *Annu. Rev. Biochem.* 63, 25, (1994)

13. Syvanen, A.C., Landegren, U., Isaksson, A., Gyllenstein, U. and Brookes, A. First International SNP Meeting at Skokloster, Sweden, August 1998. Enthusiasm mixed with scepticism about single-nucleotide polymorphism markers for dissecting complex disorders. *Eur. J. Hum. Genet.* 7, 98, (1999)

14. Makridakis, N.M., Ross, R.K., Pike, M.C., Crocitto, L.E., Kolonel, L.N., Pearce, C.L., Henderson, B.E. and Reichardt, J.K. Association of mis-sense substitution in SRD5A2 gene with prostate cancer in African-American and Hispanic men in Los Angeles, USA. *Lancet* 354, 975, (1999)

15. Jaffe, J.M., Malkowicz, S.B., Walker, A.H., MacBride, S., Peschel, R., Tomaszewski, J., Van Arsdalen, K., Wein, A.J. and Rebbeck, T.R. Association of SRD5A2 genotype and pathological characteristics of prostate tumors. *Cancer Res.* 60, 1626, (2000)

16. Margiotti, K., Sangiuolo, F., De Luca, A., Froio, F., Pearce, C.L., Ricci-Barbini, V., Micali, F., Bonafe, M., Franceschi, C., Dallapiccola, B., Novelli G, Reichardt JK.. Evidence for an association between the SRD5A2 (type II steroid 5 alpha-reductase) locus and prostate cancer in Italian patients. *Dis. Markers* 16, 147, (2000)

17. Soderstrom, T., Wadelius, M., Andersson, S.O., Johansson, J.E., Johansson, S., Granath, F. and Rane, A. 5alpha-reductase 2 polymorphisms as risk factors in prostate cancer. *Pharmacogenetics* 12, 307, (2002)

18. Hsing, A.W., Chen, C., Chokkalingam, A.P., Gao, Y.T., Dightman, D.A., Nguyen, H.T., Deng, J., Cheng, J., Sesterhenn, I.A., Mostofi, F.K., Stanczyk FZ, Reichardt JK. Polymorphic markers in the SRD5A2 gene and prostate cancer risk: a population-based case-control study. *Cancer Epidemiol. Biomarkers Prev.* 10, 1077, (2001)

19. Mononen, N., Ikonen, T., Syrjakoski, K., Matikainen, M., Schleutker, J., Tammela, T.L., Koivisto, P.A. and Kallioniemi, O.P. A missense substitution A49T in the steroid 5-alpha-reductase gene (SRD5A2) is not associated with prostate cancer in Finland. *Br. J. Cancer* 84, 1344, (2001)

20. Cicek, M.S., Conti, D.V., Curran, A., Neville, P.J., Paris, P.L., Casey, G. and Witte, J.S. Association of prostate cancer risk and aggressiveness to androgen pathway genes: SRD5A2, CYP17, and the AR. *Prostate* 59, 69, (2004)

21. Pearce, C.L., Makridakis, N.M., Ross, R.K., Pike, M.C., Kolonel, L.N., Henderson, B.E. and Reichardt, J.K. Steroid 5-alpha reductase type II V89L substitution is not associated with risk of prostate cancer in a multiethnic population study. *Cancer Epidemiol. Biomarkers Prev.* 11, 417, (2002)

22. Makridakis, N., Ross, R.K., Pike, M.C., Chang, L., Stanczyk, F.Z., Kolonel, L.N., Shi, C.Y., Yu, M.C., Henderson, B.E. and Reichardt, J.K. A prevalent missense substitution that modulates activity of prostatic steroid 5alpha-reductase. *Cancer Res.* 57, 1020, (1997)

23. Makridakis, N.M., di Salle, E. and Reichardt, J.K. Biochemical and pharmacogenetic dissection of human steroid 5 alpha-reductase type II. *Pharmacogenetics* 10, 407, (2000)

24. Mehrian-Shai, R., Reichardt, J.K. A renaissance of "biochemical genetics"? SNPs, haplotypes, function, and complex diseases. *Mol. Genet. Metab* 83, 47, (2004)

25. Makridakis, N.M., Reichardt, J.K. Pharmacogenetic analysis of human steroid 5a-reductase type II: comparison of finasteride and dutasteride. *J. Mol. Endocrinol.* (in press), (2005)

26. Thompson, I.M., Goodman, P.J., Tangen, C.M., Lucia, M.S., Miller, G.J., Ford, L.G., Lieber, M.M., Cespedes, R.D., Atkins, J.N., Lippman, S.M., Carlin SM, Ryan A, Szczepanek CM, Crowley JJ, Coltman CA Jr. The influence of finasteride on the development of prostate cancer. *N. Engl. J. Med.* 349, 215, (2003)

27. Akalu, A., Dlmajian, D.A., Highshaw, R.A., Nichols, P.W. and Reichardt, J.K. Somatic mutations at the

- SRD5A2 locus encoding prostatic steroid 5 $\alpha$ -reductase during prostate cancer progression. *J. Urol.* 161, 1355, (1999)
28. Makridakis, N., Akalu, A. and Reichardt, J.K. Identification and characterization of somatic steroid 5 $\alpha$ -reductase (SRD5A2) mutations in human prostate cancer tissue. *Oncogene* 23, 7399, (2004)
29. Barrack, E.R. Androgen receptor mutations in prostate cancer. *Mt-Sinai-J-Med.* 63, 403, (1996)
30. Visakorpi, T., Hyytinen, E., Koivisto, P., Tanner, M., Keinänen, R., Palmberg, C., Palotie, A., Tammela, T., Isola, J. and Kallioniemi, O.P. In vivo amplification of the androgen receptor gene and progression of human prostate cancer. *Nat. Genet.* 9, 401, (1995)
31. Marcelli, M., Ittmann, M., Mariani, S., Sutherland, R., Nigam, R., Murthy, L., Zhao, Y., DiConcini, D., Puxeddu, E., Esen, A., Eastham J, Weigel NL, Lamb DJ. Androgen receptor mutations in prostate cancer. *Cancer Res* 60, 944, (2000)
32. Buchanan, G., Greenberg, N.M., Scher, H.I., Harris, J.M., Marshall, V.R. and Tilley, W.D. Collocation of androgen receptor gene mutations in prostate cancer. *Clin. Cancer Res.* 7, 1273, (2001)
33. Scher, H.I., Buchanan, G., Gerald, W., Butler, L.M. and Tilley, W.D. Targeting the androgen receptor: improving outcomes for castration-resistant prostate cancer. *Endocr. Relat Cancer* 11, 459, (2004)
34. Laudet, V. Evolution of the nuclear receptor superfamily: early diversification from an ancestral orphan receptor. *J. Mol. Endocrinol.* 19, 207, (1997)
35. Baker, M.E. Evolution of 17 $\beta$ -hydroxysteroid dehydrogenases and their role in androgen, estrogen and retinoid action. *Mol. Cell Endocrinol.* 171, 211, (2001)
36. Whitfield, G.K., Jurutka, P.W., Haussler, C.A. and Haussler, M.R. Steroid hormone receptors: evolution, ligands, and molecular basis of biologic function. *J. Cell Biochem. Suppl* 32-33, 110, (1999)
37. Edwards, A., Hammond, H.A., Jin, L., Caskey, C.T. and Chakraborty, R. Genetic variation at five trimeric and tetrameric tandem repeat loci in four human population groups. *Genomics* 12, 241, (1992)
38. Giovannucci, E., Stampfer, M.J., Krithivas, K., Brown, M., Dahl, D., Brufsky, A., Talcott, J., Hennekens, C.H. and Kantoff, P.W. The CAG repeat within the androgen receptor gene and its relationship to prostate cancer. *Proc. Natl. Acad. Sci. U. S. A.* 94, 3320, (1997)
39. Buchanan, G., Yang, M., Cheong, A., Harris, J.M., Irvine, R.A., Lambert, P.F., Moore, N.L., Raynor, M., Neufing, P.J., Coetzee, G.A., Tilley WD. Structural and functional consequences of glutamine tract variation in the androgen receptor. *Hum. Mol. Genet.* 13, 1677, (2004)
40. Kazemi-Esfarjani, P., Trifiro, M.A. and Pinsky, L. Evidence for a repressive function of the long polyglutamine tract in the human androgen receptor: possible pathogenetic relevance for the (CAG) $n$ -expanded neuronopathies. *Hum. Mol. Genet.* 4, 523, (1995)
41. Beilin, J., Ball, E.M., Favaloro, J.M. and Zajac, J.D. Effect of the androgen receptor CAG repeat polymorphism on transcriptional activity: specificity in prostate and non-prostate cell lines. *J. Mol. Endocrinol.* 25, 85, (2000)
42. Coetzee, G.A., Ross, R.K. Re: Prostate cancer and the androgen receptor. *J. Natl. Cancer Inst.* 86, 872, (1994)
43. Irvine, R.A., Yu, M.C., Ross, R.K. and Coetzee, G.A. The CAG and GGC microsatellites of the androgen receptor gene are in linkage disequilibrium in men with prostate cancer. *Cancer Res* 55, 1937, (1995)
44. Ingles, S.A., Ross, R.K., Yu, M.C., Irvine, R.A., La Pera, G., Haile, R.W. and Coetzee, G.A. Association of prostate cancer risk with genetic polymorphisms in vitamin D receptor and androgen receptor. *J. Natl. Cancer Inst.* 89, 166, (1997)
45. Hakimi, J.M., Schoenberg, M.P., Rondinelli, R.H., Piantadosi, S. and Barrack, E.R. Androgen receptor variants with short glutamine or glycine repeats may identify unique subpopulations of men with prostate cancer. *Clin. Cancer Res.* 3, 1599, (1997)
46. Hardy, D.O., Scher, H.I., Bogenreider, T., Sabbatini, P., Zhang, Z.F., Nanus, D.M. and Catterall, J.F. Androgen receptor CAG repeat lengths in prostate cancer: correlation with age of onset. *J. Clin. Endocrinol. Metab.* 81, 4400, (1996)
47. Stanford, J.L., Just, J.J., Gibbs, M., Wicklund, K.G., Neal, C.L., Blumenstein, B.A. and Ostrander, E.A. Polymorphic repeats in the androgen receptor gene: molecular markers of prostate cancer risk. *Cancer Res* 57, 1194, (1997)
48. Ekman, P., Gronberg, H., Matsuyama, H., Kivineva, M., Bergerheim, U.S. and Li, C. Links between genetic and environmental factors and prostate cancer risk. *Prostate* 39, 262, (1999)
49. Edwards, S.M., Badzioch, M.D., Minter, R., Hamoudi, R., Collins, N., Ardern-Jones, A., Dowe, A., Osborne, S., Kelly, J., Shearer, R., Easton DF, Saunders GF, Dearnaley DP, Eeles RA. Androgen receptor polymorphisms: association with prostate cancer risk, relapse and overall survival. *Int. J. Cancer* 84, 458, (1999)
50. Correa-Cerro, L., Wohr, G., Haussler, J., Berthon, P., Drelon, E., Mangin, P., Fournier, G., Cussenot, O., Kraus, P., Just, W., Paiss T, Cantu JM, Vogel W. (CAG) $n$ CAA and GGN repeats in the human androgen receptor gene are not associated with prostate cancer in a French-German population. *Eur. J. Hum. Genet.* 7, 357, (1999)
51. Bratt, O., Borg, A., Kristoffersson, U., Lundgren, R., Zhang, Q.X. and Olsson, H. CAG repeat length in the androgen receptor gene is related to age at diagnosis of prostate cancer and response to endocrine therapy, but not to prostate cancer risk. *Br. J. Cancer* 81, 672, (1999)
52. Lange, E.M., Chen, H., Brierley, K., Livermore, H., Wojno, K.J., Langefeld, C.D., Lange, K. and Cooney, K.A. The polymorphic exon 1 androgen receptor CAG repeat in men with a potential inherited predisposition to prostate cancer. *Cancer Epidemiol. Biomarkers Prev.* 9, 439, (2000)
53. Nam, R.K., Elhaji, Y., Krahn, M.D., Hakimi, J., Ho, M., Chu, W., Sweet, J., Trachtenberg, J., Jewett, M.A. and Narod, S.A. Significance of the cag repeat polymorphism of the androgen receptor gene in prostate cancer progression. *J. Urol.* 164, 567, (2000)
54. Latil, A.G., Azzouzi, R., Cancel, G.S., Guillaume, E.C., Cochran-Priollet, B., Berthon, P.L. and Cussenot, O. Prostate carcinoma risk and allelic variants of genes involved in androgen biosynthesis and metabolism pathways. *Cancer* 92, 1130, (2001)
55. Modugno, F., Weissfeld, J.L., Trump, D.L., Zmuda, J.M., Shea, P., Cauley, J.A. and Ferrell, R.E. Allelic

- variants of aromatase and the androgen and estrogen receptors: toward a multigenic model of prostate cancer risk. *Clin. Cancer Res.* 7, 3092, (2001)
56. Miller, E.A., Stanford, J.L., Hsu, L., Noonan, E. and Ostrander, E.A. Polymorphic repeats in the androgen receptor gene in high-risk sibships. *Prostate* 48, 200, (2001)
57. Panz, V.R., Joffe, B.I., Spitz, I., Lindenberg, T., Farkas, A. and Haffeeje, M. Tandem CAG repeats of the androgen receptor gene and prostate cancer risk in black and white men. *Endocrine*. 15, 213, (2001)
58. Santen, R.J. Clinical review 37: Endocrine treatment of prostate cancer. *J. Clin. Endocrinol. Metab.* 75, 685, (1992)
59. Thenot, S., Charpin, M., Bonnet, S. and Cavaillès, V. Estrogen receptor cofactors expression in breast and endometrial human cancer cells. *Mol. Cell Endocrinol.* 156, 85, (1999)
60. Kozlowski, J.M., Ellis, W.J. and Grayhack, J.T. Advanced prostatic carcinoma. Early versus late endocrine therapy. *Urol. Clin. North Am.* 18, 15, (1991)
61. Feldman, B.J., Feldman, D. The development of androgen-independent prostate cancer. *Nat Rev* 1, 34, (2001)
62. Gregory, C.W., Hamil, K.G., Kim, D., Hall, S.H., Pretlow, T.G., Mohler, J.L. and French, F.S. Androgen receptor expression in androgen-independent prostate cancer is associated with increased expression of androgen-regulated genes. *Cancer Res* 58, 5718, (1998)
63. Chen, C.D., Welsbie, D.S., Tran, C., Baek, S.H., Chen, R., Vessella, R., Rosenfeld, M.G. and Sawyers, C.L. Molecular determinants of resistance to antiandrogen therapy. *Nat. Med.* 10, 33, (2004)
64. Sirotiak, F.M., She, Y., Khokhar, N.Z., Hayes, P., Gerald, W. and Scher, H.I. Microarray analysis of prostate cancer progression to reduced androgen dependence: Studies in unique models contrasts early and late molecular events. *Mol. Carcinog.* 41, 150, (2004)
65. Presnell, S.C., Werdin, E.S., Maygarden, S., Mohler, J.L. and Smith, G.J. Establishment of short-term primary human prostate xenografts for the study of prostate biology and cancer. *Am. J. Pathol.* 159, 855, (2001)
66. Henshall, S.M., Quinn, D.I., Lee, C.S., Head, D.R., Golovsky, D., Brenner, P.C., Delprado, W., Stricker, P.D., Grygiel, J.J. and Sutherland, R.L. Altered expression of androgen receptor in the malignant epithelium and adjacent stroma is associated with early relapse in prostate cancer. *Cancer Res* 61, 423, (2001)
67. Ricciardelli, C., Choong, C.S., Buchanan, G., Vivekanandan, S., Neufing, P., Stahl, J., Marshall, V.R., Horsfall, D.J. and Tilley, W.D. Androgen receptor levels in prostate cancer epithelial and peritumoral stromal cells identify non-organ confined disease. *Prostate* 63, 19, (2005)
68. Koivisto, P., Kononen, J., Palmberg, C., Tammela, T., Hyytinen, E., Isola, J., Trapman, J., Cleutjens, K., Noordzij, A., Visakorpi, T., Kallioniemi O.P. Androgen receptor gene amplification: a possible molecular mechanism for androgen deprivation therapy failure in prostate cancer. *Cancer Res* 57, 314, (1997)
69. Koivisto, P.A., Helin, H.J. Androgen receptor gene amplification increases tissue PSA protein expression in hormone-refractory prostate carcinoma. *J. Pathol.* 189, 219, (1999)
70. Holzbeierlein, J., Lal, P., LaTulippe, E., Smith, A., Satagopan, J., Zhang, L., Ryan, C., Smith, S., Scher, H., Scardino, P., Reuter V, Gerald WL. Gene expression analysis of human prostate carcinoma during hormonal therapy identifies androgen-responsive genes and mechanisms of therapy resistance. *Am. J. Pathol.* 164, 217, (2004)
71. Buchanan, G., Yang, M., Nahm, S.J., Han, G., Moore, N., Bentel, J.M., Matusik, R.J., Horsfall, D.J., Marshall, V.R., Greenberg, N.M., Tilley WD. Mutations at the boundary of the hinge and ligand binding domain of the androgen receptor confer increased transactivation function. *Mol. Endocrinol.* 15, 46, 2001
72. Fenton, M.A., Shuster, T.D., Fertig, A.M., Taplin, M.E., Kolvenbag, G., Bubley, G.J. and Balk, S.P. Functional Characterization of Mutant Androgen Receptors from Androgen-independent Prostate Cancer. *Clin. Cancer Res.* 3, 1383, (1997)
73. Taplin, M.E., Bubley, G.J., Ko, Y.J., Small, E.J., Upton, M., Rajeshkumar, B. and Balk, S.P. Selection for androgen receptor mutations in prostate cancers treated with androgen antagonist. *Cancer Res* 59, 2511, (1999)
74. Kassouf, W., Tanguay, S. and Aprikian, A.G. Nilutamide as second line hormone therapy for prostate cancer after androgen ablation fails. *J. Urol.* 169, 1742, (2003)
75. Joyce, R., Fenton, M.A., Rode, P., Constantine, M., Gaynes, L., Kolvenbag, G., DeWolf, w., Balk, S., Taplin, M.E. and Bubley, G.J. High dose bicalutamide for androgen independent prostate cancer: effect of prior hormonal therapy. *J. Urol.* 159, 149, (1998)
76. Greenberg, N.M., DeMayo, F., Finegold, M.J., Medina, D., Tilley, W.D., Aspinnall, J.O., Cunha, G.R., Donjacour, A.A., Matusik, R.J. and Rosen, J.M. Prostate cancer in a transgenic mouse. *Proc. Natl. Acad. Sci. U. S. A.* 92, 3439, (1995)
77. Gingrich, J.R., Barrios, R.J., Kattan, M.W., Nahm, H.S., Finegold, M.J. and Greenberg, N.M. Androgen-independent prostate cancer progression in the TRAMP model. *Cancer Res* 57, 4687, (1997)
78. Han, G., Foster, B.A., Mistry, S., Buchanan, G., Harris, J.M., Tilley, W.D. and Greenberg, N.M. Hormone status selects for spontaneous somatic androgen receptor variants that demonstrate specific ligand and cofactor dependent activities in autochthonous prostate cancer. *J. Biol. Chem.* 276, 11204, (2001)
79. Han, G., Buchanan, G., Ittmann, M., Harris, J.M., Yu, X., DeMayo, F.J., Tilley, W. and Greenberg, N.M. Mutation of the androgen receptor causes oncogenic transformation of the prostate. *Proc. Natl. Acad. Sci. U. S. A.* 102, 1151, (2005)
80. Culig, Z., Hoffmann, J., Erdel, M., Eder, I.E., Hobisch, A., Hittmair, A., Bartsch, G., Utermann, G., Schneider, M.R., Parczyk, K., Klocker H. Switch from antagonist to agonist of the androgen receptor bicalutamide is associated with prostate tumour progression in a new model system. *Br. J. Cancer* 81, 242, (1999)
81. Debes, J.D., Tindall, D.J. The role of androgens and the androgen receptor in prostate cancer. *Cancer Lett.* 187, 1, (2002)
82. Gregory, C.W., He, B., Johnson, R.T., Ford, O.H., Mohler, J.L., French, F.S. and Wilson, E.M. A mechanism

- for androgen receptor-mediated prostate cancer recurrence after androgen deprivation therapy. *Cancer Res* 61, 4315, (2001)
83. Fujimoto, N., Yeh, S., Kang, H.Y., Inui, S., Chang, H.C., Mizokami, A. and Chang, C. Cloning and characterization of androgen receptor coactivator, ARA55, in human prostate. *J. Biol. Chem.* 274, 8316, (1999)
84. Ngan, E.S., Hashimoto, Y., Ma, Z.Q., Tsai, M.J. and Tsai, S.Y. Overexpression of Cdc25B, an androgen receptor coactivator, in prostate cancer. *Oncogene* 22, 734, (2003)
85. Debes, J.D., Sebo, T.J., Lohse, C.M., Murphy, L.M., Haugen, d.A. and Tindall, D.J. p300 in prostate cancer proliferation and progression. *Cancer Res* 63, 7638, (2003)
86. Yeh, S., Kang, H.Y., Miyamoto, H., Nishimura, K., Chang, H.C., Ting, H.J., Rahman, M., Lin, H.K., Fujimoto, N., Hu, Y.C., Mizokami, A., Huang, K.E., Chang C. Differential induction of androgen receptor transactivation by different androgen receptor coactivators in human prostate cancer DU145 cells. *Endocrine*. 11, 195, (1999)
87. Yeh, S., Miyamoto, H., Shima, H. and Chang, C. From estrogen to androgen receptor: A new pathway for sex hormones in prostate. *Proc. Natl. Acad. Sci. U. S. A.* 95, 5527, (1998)
88. He, B., Bowen, N.T., Minges, J.T. and Wilson, E.M. Androgen-induced NH<sub>2</sub>- and COOH-terminal Interaction Inhibits p160 coactivator recruitment by activation function 2. *J. Biol. Chem.* 276, 42293, (2001)
89. Hodgson, M.C., Astapova, I., Cheng, S., Lee, L.J., Verhoeven, M.C., Choi, E., Balk, S.P. and Hollenberg, A.N. The androgen receptor recruits nuclear receptor CoRepressor (N-CoR) in the presence of mifepristone via its N and C termini revealing a novel molecular mechanism for androgen receptor antagonists. *J. Biol. Chem.* 280, 6511, (2005)
90. Berrevoets, C.A., Umar, A., Trapman, J. and Brinkmann, A.O. Differential modulation of androgen receptor transcriptional activity by the nuclear receptor co-repressor (N-CoR) *Biochem. J.* 379, 731, (2004)
91. Caplen, N.J., Taylor, J.P., Statham, V.S., Tanaka, F., Fire, A. and Morgan, R.A. Rescue of polyglutamine-mediated cytotoxicity by double-stranded RNA-mediated RNA interference. *Hum. Mol. Genet.* 11, 175, (2002)
92. Eder, I.E., Culig, Z., Ramoner, R., Thurnher, M., Putz, T., Nessler-Menardi, C., Tiefenthaler, M., Bartsch, G. and Klocker, H. Inhibition of LncAP prostate cancer cells by means of androgen receptor antisense oligonucleotides. *Cancer Gene Ther.* 7, 997, (2000)
93. Eder, I.E., Hoffmann, J., Rogatsch, H., Schafer, G., Zopf, D., Bartsch, G. and Klocker, H. Inhibition of LNCaP prostate tumor growth in vivo by an antisense oligonucleotide directed against the human androgen receptor. *Cancer Gene Ther.* 9, 117, (2002)
94. Zegarra-Moro, O.L., Schmidt, L.J., Huang, H. and Tindall, D.J. Disruption of androgen receptor function inhibits proliferation of androgen-refractory prostate cancer cells. *Cancer Res* 62, 1008, (2002)
95. Solit, D.B., Zheng, F.F., Drobnyak, M., Munster, P.N., Higgins, B., Verbel, D., Heller, G., Tong, W., Cordon-Cardo, C., Agus, D.B., Scher, H.I., Rosen, N. 17-Allylamino-17-demethoxygeldanamycin induces the degradation of androgen receptor and HER-2/neu and inhibits the growth of prostate cancer xenografts. *Clin. Cancer Res.* 8, 986, (2002)
96. Grenert, J.P., Sullivan, W.P., Fadden, P., Haystead, T.A., Clark, J., Mimnaugh, E., Krutzsch, H., Ochel, H.J., Schulte, T.W., Sausville, E., Neckers, L.M., Toft, D.O. The amino-terminal domain of heat shock protein 90 (hsp90) that binds geldanamycin is an ATP/ADP switch domain that regulates hsp90 conformation. *J. Biol. Chem.* 272, 23843, (1997)
97. Prodromou, C., Roe, S.M., O'Brien, R., Ladbury, J.E., Piper, P.W. and Pearl, L.H. Identification and structural characterization of the ATP/ADP-binding site in the Hsp90 molecular chaperone. *Cell* 90, 65, (1997)
98. Stebbins, C.E., Russo, A.A., Schneider, C., Rosen, N., Hartl, F.U. and Pavletich, N.P. Crystal structure of an Hsp90-geldanamycin complex: targeting of a protein chaperone by an antitumor agent. *Cell* 89, 239, (1997)
99. Palvimo, J.J., Kallio, P.J., Ikonen, T., Mehto, M. and Janne, O.A. Dominant negative regulation of transactivation by the rat androgen receptor: roles of the N-terminal domain and heterodimer formation. *Mol. Endocrinol.* 7, 1399, (1993)
100. Bramlett, K.S., Dits, N.F., Sui, X., Jorge, M.C., Zhu, X. and Jenster, G. Repression of androgen-regulated gene expression by dominant negative androgen receptors. *Mol. Cell Endocrinol.* 183, 19, (2001)
101. Marks, P., Rifkind, R.A., Richon, V.M., Breslow, R., Miller, T. and Kelly, W.K. Histone deacetylases and cancer: causes and therapies. *Nat. Rev. Cancer* 1, 194, (2001)
102. Butler, L.M., Agus, D.B., Scher, H.I., Higgins, B., Rose, A., Cordon-Cardo, C., Thaler, H.T., Rifkind, R.A., Marks, P.A. and Richon, V.M. Suberoylanilide hydroxamic acid, an inhibitor of histone deacetylase, suppresses the growth of prostate cancer cells in vitro and in vivo. *Cancer Res* 60, 5165, (2000)
103. Labrie, F., Simard, J., Luu-The, V., Pelletier, G., Belanger, A., Lachance, Y., Zhao, H.F., Labrie, C., Breton, N., De Launoit, Y. et al. Structure and tissue-specific expression of 3 beta-hydroxysteroid dehydrogenase/5-ene-4-ene isomerase genes in human and rat classical and peripheral steroidogenic tissues. *J. Steroid Biochem. Mol. Biol.* 41, 421, (1992)
104. Berube, D., Luu, T., V, Lachance, Y., Gagne, R. and Labrie, F. Assignment of the human 3 beta-hydroxysteroid dehydrogenase gene (HSD3B) to the p13 band of chromosome 1. *Cytogenet. Cell Genet.* 52, 199, (1989)
105. Verreault, H., Dufort, I., Simard, J., Labrie, F. and Luu-The, V. Dinucleotide repeat polymorphisms in the HSD3B2 gene. *Hum. Mol. Genet.* 3, 384, (1994)
106. Devgan, S.A., Henderson, B.E., Yu, M.C., Shi, C.Y., Pike, M.C., Ross, R.K. and Reichardt, J.K. Genetic variation of 3 beta-hydroxysteroid dehydrogenase type II in three racial/ethnic groups: implications for prostate cancer risk. *Prostate* 33, 9, (1997)
107. Chang, B.L., Zheng, S.L., Hawkins, G.A., Isaacs, S.D., Wiley, K.E., Turner, A., Carpten, J.D., Bleecker, E.R., Walsh, P.C., Trent, J.M., Meyers, D.A., Isaacs, W.B., Xu, J. Joint effect of HSD3B1 and HSD3B2 genes is associated with hereditary and sporadic prostate cancer susceptibility. *Cancer Res.* 62, 1784, (2002)
108. Stanford, J.L., Noonan, E.A., Iwasaki, L., Kolb, S., Chadwick, R.B., Feng, Z. and Ostrander, E.A. A

polymorphism in the CYP17 gene and risk of prostate cancer. *Cancer Epidemiol. Biomarkers Prev.* 11, 243, (2002)

109. Geissler, W.M., Davis, D.L., Wu, L., Bradshaw, K.D., Patel, S., Mendonca, B.B., Elliston, K.O., Wilson, J.D., Russell, D.W. and Andersson, S. Male pseudohermaphroditism caused by mutations of testicular 17 beta-hydroxysteroid dehydrogenase 3. *Nat. Genet.* 7, 34, (1994)

110. Margiotti, K., Kim, E., Pearce, C.L., Spera, E., Novelli, G. and Reichardt, J.K. Association of the G289S single nucleotide polymorphism in the HSD17B3 gene with prostate cancer in Italian men. *Prostate* 53, 65, (2002)

111. Lunn, R.M., Bell, D.A., Mohler, J.L. and Taylor, J.A. Prostate cancer risk and polymorphism in 17 hydroxylase (CYP17) and steroid reductase (SRD5A2) *Carcinogenesis* 20, 1727, (1999)

112. Wadelius, M., Andersson, A.O., Johansson, J.E., Wadelius, C. and Rane, E. Prostate cancer associated with CYP17 genotype. *Pharmacogenetics* 9, 635, (1999)

113. Antognelli, C., Mearini, L., Talesa, V.N., Giannantoni, A. and Mearini, E. Association of CYP17, GSTP1, and PON1 polymorphisms with the risk of prostate cancer. *Prostate* 63, 240, (2005)

114. Makridakis, N.M., Ross, R.K., Pike, M.C., Crocitto, L.E., Kolonel, L.N., Pearce, C.L., Henderson, B.E. and Reichardt, J.K. Association of mis-sense substitution in SRD5A2 gene with prostate cancer in African-American and Hispanic men in Los Angeles, USA. *Lancet* 354, 975, (1999)

115. Montie, J.E., Pienta, K.J. Review of the role of androgenic hormones in the epidemiology of benign prostatic hyperplasia and prostate cancer. *Urology* 43, 892, (1994)

116. Ekman, P. Genetic and environmental factors in prostate cancer genesis: identifying high-risk cohorts. *Eur. Urol.* 35, 362, (1999)

117. Gottlieb, B., Beitel, L.K., Wu, J.H. and Trifiro, M. The androgen receptor gene mutations database (ARDB): 2004 update. *Hum. Mutat.* 23, 527, (2004)

118. Hara, T., Miyazaki, J., Araki, H., Yamaoka, M., Kanzaki, N., Kusaka, M. and Miyamoto, M. Novel mutations of androgen receptor: a possible mechanism of bicalutamide withdrawal syndrome. *Cancer Res* 63, 149, (2003)

119. Veldscholte, J., Ris-Stalpers, C., Kuiper, G.G., Jenster, G., Berrevoets, C., Claassen, E., van Rooij, H.C., Trapman, J., Brinkmann, A.O. and Mulder, E. A mutation in the ligand binding domain of the androgen receptor of human LNCaP cells affects steroid binding characteristics and response to anti-androgens. *Biochem. Biophys. Res. Commun.* 173, 534, (1990)

**Key Words:** Molecular Epidemiology, Androgen Metabolism, Multifactorial, Complex, Phenotype, Pharmacogenetics, Single Nucleotide Polymorphism, SNP, Review

**Send correspondence to:** Dr Juergen K. V. Reichardt, University of Sydney, Medical Foundation Building (K25), 92 - 94 Parramatta Rd, Camperdown, Sydney, NSW 2042,

Australia, Tel: 323-442-1529, Fax: 323-442-2764, E-mail: reichard@usc.edu

<http://www.bioscience.org/current/vol10.htm>