

## Green tea polyphenols: biology and therapeutic implications in cancer

Sharmila Shankar, Suthakar Ganapathy, Rakesh K. Srivastava

Department of Biochemistry, University of Texas Health Science Center at Tyler, Tyler, Texas, USA 75703

### TABLE OF CONTENTS

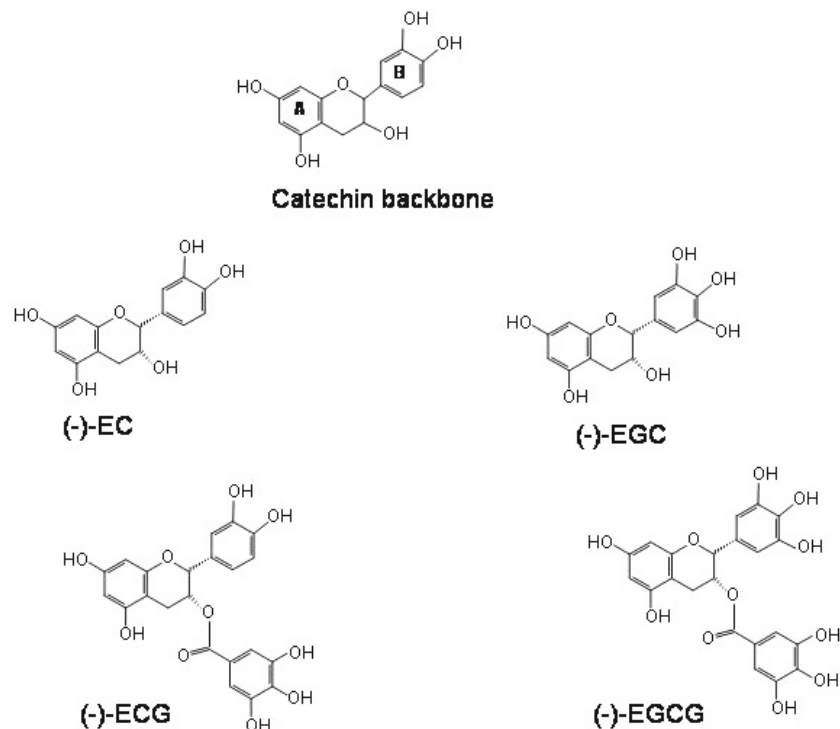
1. Abstract
2. Introduction
3. Tea Polyphenols
4. Mechanisms of Action
  - 4.1. Bcl-2 family members
  - 4.2. Matrix metalloproteinases
  - 4.3. Ras/Map Kinases
  - 4.4. PI3-kinase/AKT
  - 4.5. Cyclooxygenases
  - 4.6. Epidermal growth factor
  - 4.7. Insulin-like growth factors
  - 4.8. Transcription factors
    - 4.8.1. NF $\kappa$ B
    - 4.8.2. STAT
    - 4.8.3. AP-1
    - 4.8.4. Nrf2
5. Apoptosis and cell cycle
6. Clinical significance of EGCG
  - 6.1. Effects of EGCG on cancer
    - 6.1.1. Prostate cancer
    - 6.1.2. Lung cancer
    - 6.1.3. Skin cancer
    - 6.1.4. Pancreatic cancer
    - 6.1.5. Breast cancer
    - 6.1.6. Ovarian cancer
  - 6.2. Effects of EGCG on angiogenesis and metastasis
7. Conclusions and future directions
8. Acknowledgements
9. References

## 1. ABSTRACT

Multiple lines of evidence, mostly from population-based studies, suggest that green tea consumption is associated with reduced risk of several human malignancies such as cancer and diabetes. Epigallocatechin-3-gallate (EGCG), a major polyphenol found in green tea, is a widely studied chemopreventive agent with potential anticancer activity. Green tea polyphenols inhibit angiogenesis and metastasis, and induce growth arrest and apoptosis through regulation of multiple signaling pathways. Specifically, EGCG regulates expression of VEGF, matrix metalloproteinases, uPA, IGF-1, EGFR, cell cycle regulatory proteins and inhibits NF $\kappa$ B, PI3-K/Akt, Ras/Raf/MAPK and AP-1 signaling pathways, thereby causing strong cancer chemopreventive effects. This review discusses the molecular mechanisms of green tea polyphenols and their therapeutic implications in cancer.

## 2. INTRODUCTION

Green tea is a popular beverage consumed widely in China, Japan and India, and is a rich source of flavanoids (1-3). Flavanoids are the low molecular weight compounds divided into several different classes based on variations of the same basic structure. One such class is the flavan-3-ols, also referred to as the catechins. Catechins are especially concentrated in green tea (*Camellia sinensis*) which account for 30-40% of the dry weight of the leaves. A polyphenolic constituent, (-)-epigallocatechin-3-gallate (EGCG), is the major and most effective chemopreventive agent in green tea. Epidemiologic and rodent carcinogenesis studies have provided evidence that green tea has chemopreventive effects for a wide range of malignancies (4-11). The consumption of green tea is associated with a lower risk of several types of cancer, including stomach, esophagus, prostate and lung (2, 3, 12, 13). It has also been reported that the quantity of green tea



**Figure 1.** Structure of the major catechins found in green tea. The structures of catechin backbone, (-)-epicatechin, (EC); (-)-epicatechin-3-gallate, (ECG); (-)-epigallocatechin, (EGC); (+)-gallocatechin, (GC); and (+)-gallocatechin-3-gallate, (EGCG) are shown.

consumed plays an important role in reducing cancer risk and in delaying cancer outbreak and recurrence. It acts as an antioxidant, antiproliferative, antitumor, and anti-angiogenic agent, and thus a novel candidate for chemoprevention (2, 3, 13). Mechanistic studies have indicated that EGCG exerts various anticancer effects, including suppression of growth factor-mediated proliferation (6), inhibition of transformation (5), and repression of angiogenesis (8, 10). Tea and tea polyphenols have shown inhibitory activity during the initiation, promotion, and progression stages of carcinogenesis (9, 11). *In vitro*, tea polyphenols, especially EGCG, have been shown to cause growth inhibition and apoptosis in several human tumor cell lines, including melanoma, breast cancer, lung cancer, leukemia, and colon cancer (3, 13-16). The main objective of this review is to discuss the molecular mechanisms of green tea polyphenols and their therapeutic implications in cancer.

### 3. TEA POLYPHENOLS

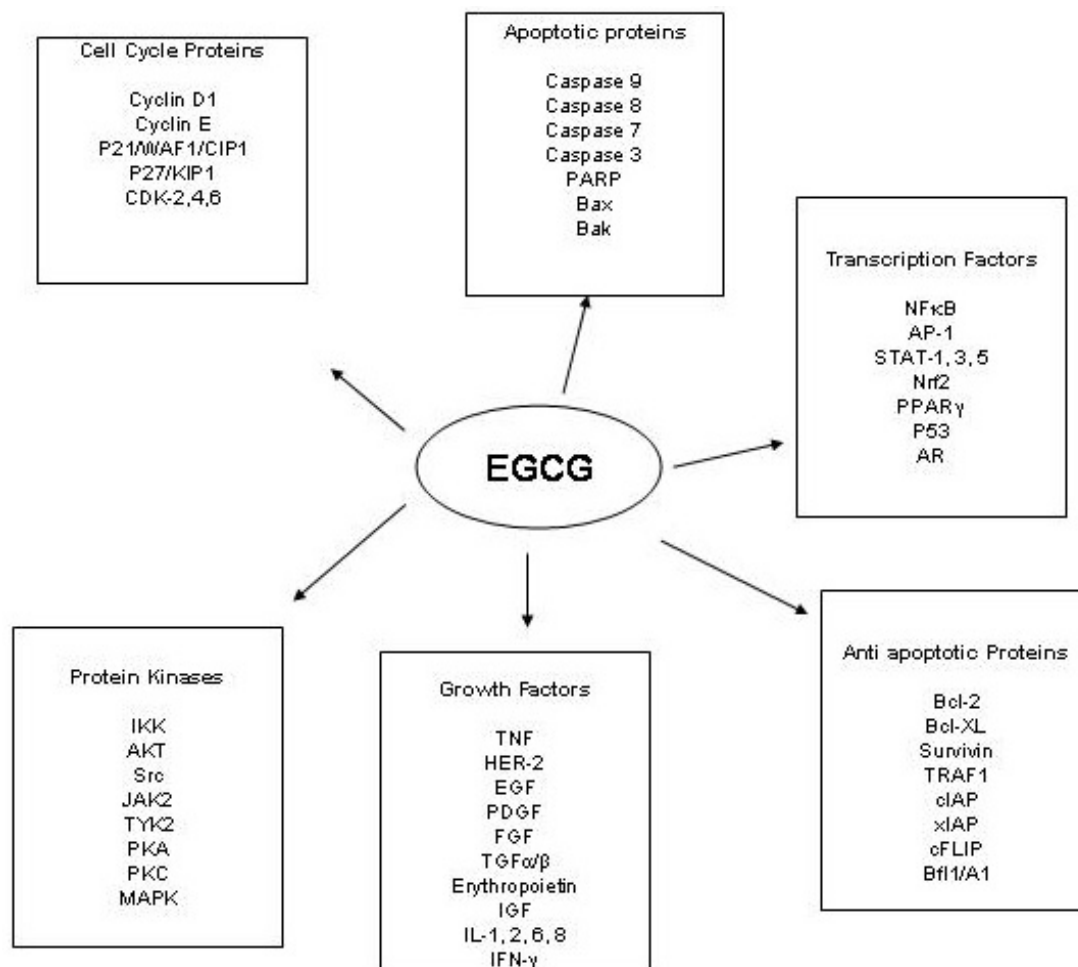
EGCG is the major constituent found in green tea. Several other polyphenolic compounds found in lower abundance in green tea include (-)-epicatechin-3-gallate, (ECG); (-)-epigallocatechin, (EGC); (-)-epicatechin, (EC); (+)-gallocatechin, (GC); (+)-gallocatechin-3-gallate, (GCG) and catechin (Figure 1). Several *in vitro* and animal models suggest that green tea polyphenols inhibit a vast array of biomedically relevant molecular targets and disease related cellular processes (17, 18).

### 4. MECHANISMS OF ACTION

Green tea polyphenols induce growth arrest and apoptosis, and inhibit angiogenesis and metastasis through regulation of multiple signaling pathways. Specifically, EGCG regulates expression of Bcl-2 family members, VEGF, MMPs, uPA, IGF-1, EGFR, cell cycle inhibitors (p21<sup>WAF1/CIP1</sup>, and p27<sup>KIP1</sup>), and inhibits survival signaling pathways such as NFκB, PI3-K/Akt, and Ras/Raf/MAPK, thereby causing strong cancer chemopreventive effects (Figure 2). The molecular mechanisms of EGCG and related polyphenols are summarized below.

#### 4.1. Bcl-2 family members

Recently, the Bcl-2 gene family has emerged as critical regulators of apoptosis in a variety of physiological and pathological processes (19, 20). Study of the mechanism of apoptosis by Bcl-2-related genes offers new possibilities for prevention and treatment of several human diseases (21, 22). Some of the proteins within this family, including Bcl-2 and Bcl-X<sub>L</sub>, inhibit apoptosis, and others, such as Bcl-X<sub>S</sub>, Bax and Bak, promote apoptosis (19, 20). Indeed, the ratio between these two subsets determines, in part, the susceptibility of cells to a death signal (23). Furthermore, these death-inducing proteins heterodimerize with members of the death-inhibitory family (19). The “BH-3-only protein” PUMA is an essential mediator of p53-dependent and -independent apoptosis *in vitro* and *in vivo* (24-26). This suggests that PUMA can bypass the p53 control mechanism of apoptosis *in vivo*, and thus enhance



**Figure 2.** Molecular targets of green tea polyphenols. Green tea polyphenols exert their effect on multiple signaling pathways. They regulate cell cycle proteins (cyclin D1, cyclin E, p21<sup>WAF1/CIP1</sup>, p27<sup>KIP1</sup> and CDK-2/4/6), protein kinases (IKK, AKT, Src, JAK2, TYK2, MAPK, PKA and PKC), growth factors (EGF, HER-2, PDGF, FGF, TGF-α/β, Erythropoietin, IGF-I, IL-1/2/6/8, and IFNγ), transcription factors (NFκB, AP-1, STAT-1/3/5, Nrf2, PPAR, p53 and AR), proapoptotic proteins (caspases, PARP, Bax and Bak) and antiapoptotic proteins (Bcl-2, Bcl-X<sub>L</sub>, survivin, cIAP1, xIAP, cFLIP, TRAF1 and Bfl1/A1).

the therapeutic potential of EGCG in cells harboring wild type and mutated p53. PUMA is likely to play a role in mediating cell death through the cytochrome c/Apaf-1-dependent pathway. We and others have shown that overexpression of Bcl-2 or Bcl-X<sub>L</sub> protects a wide variety of cell types from many death-inducing stimuli (27-33). We and others have shown that EGCG inhibits the expression of antiapoptotic proteins Bcl-2, Bcl-X<sub>L</sub> and induces expressions of proapoptotic proteins such as Bax, Bak, PUMA, Noxa and Bim, suggesting that the Bcl-2 family members play major roles in inducing cell growth and apoptosis (34-39).

#### 4.2. Matrix metalloproteinases

Proteolytic degradation of components like collagen, proteoglycan, laminin, elastin and fibronectin in the extracellular matrix is considered to be the prerequisite for tumor invasion and metastasis. Matrix

metalloproteinases (MMPs) can degrade essentially all of the protein components of the extracellular matrix (40, 41). In addition, these MMPs also substantially contribute to angiogenesis, differentiation, proliferation and apoptosis (40-42). Hence, MMPs are important regulators of tumor growth both at the primary site and in distant metastases and considered important targets for cancer therapy (43). Among the various types of MMPs, gelatinase A (MMP-2) and gelatinase B (MMP-9) seem to play an important role in tumor invasion and metastasis (44). The expression of MMPs is primarily regulated through AP-1 via mitogen activated protein kinase (MAPK) pathway (45, 46). Thus, MMPs and their regulatory pathways have been considered the promising targets for anticancer drugs and chemopreventive agents (47).

EGCG has been shown to inhibit metastasis of lung cancer cells by inhibiting MMP-9 (48). EGCG and

EGC inhibited Interleukin (IL)-1 $\beta$ -induced expression of the collagenases, MMP-1, MMP-3 and MMP-13, and the stromelysin in human tendon-derived fibroblasts, and had a smaller effect on MMP-2 mRNA expression, which was not stimulated by IL-1 $\beta$  (49). GTP and EGCG significantly inhibited the expression of VEGF, MMP-2 and MMP-9 in prostate cancer cells of TRAMP mice and in DU-145 cells, respectively (46, 50). EGCG inhibited the phorbol 12-myristate 13-acetate (PMA)-induced cell invasiveness and MMP-9 expression in human gastric cancer AGS cells (51). EGCG also abrogated the PMA-induced activation of ERK and JNK, which are upstream modulators of AP-1 (51). Overall, these results suggest that EGCG may exert at least part of its anti-invasive effect in several cancers by controlling MMP expression through the suppression of MAPK and AP-1 activation.

### 4.3. RAS/MAP kinases

The Ras proteins are small (21 kDa) GTP-binding, membrane-associated proteins (52). They are in their activated state when bound to GTP, and are inactivated by GTP hydrolysis. This intrinsic GTPase activity is enhanced by association with GTPase-activating protein (52). The Ras proteins transduce signals from ligand-activated tyrosine kinase receptors to downstream effectors (53). Activating mutations can impair GTP hydrolysis and lead to constitutively activated Ras that impacts the cellular phenotype (54). Oncogenic Ras can lead to cellular transformation (55), presumably by perturbing its signal transduction pathways. Ras regulates multiple signaling pathways (56). Three major groups of MAP kinases are found in mammalian cells: extracellular signal-regulated protein kinase (ERK) (57), p38 MAP kinase (58), and c-Jun N-terminal kinase (JNK) (59-61). MAP kinases regulate many cellular activities, which range from gene expression to mitosis, movement, metabolism, and apoptosis. These MAP kinases are activated by the dual phosphorylations of neighboring threonine and tyrosine residues in response to various extracellular stimuli (62, 63). Specifically, p38 and JNK have been implicated in stress-responsive signaling leading to the initiation of adaptive events such as gene expression, differentiation, metabolism, and apoptosis (59, 60, 64). ERKs are often activated by growth signals, such as epidermal growth factor (EGF) or platelet-derived growth factor (65).

EGCG induces JNK pathway which causes the release of cytochrome c and apoptosis in colon cancer cells (66). EGCG-induced JNK activation was blocked by the antioxidants glutathione and N-acetyl-L-cysteine, suggesting the involvement of oxidative stress in EGCG-induced apoptosis (66). EGCG-induces p57/KIP2 via the p38 MAPK signaling pathway in oral carcinoma cells (67). In p57-negative tumor cells, JNK signaling mediates EGCG-induced apoptosis, and exogenous expression of p57 suppresses EGCG-induced apoptosis via inhibition of JNK. Furthermore, restoration of p57 expression in tumor cells significantly reduced tumorigenicity in athymic mice, suggesting that p57 expression may be useful as a target for cancer therapies.

### 4.4. PI3-kinase/AKT

PTEN (phosphatase and tensin homolog deleted on chromosome 10, also called MMAC1 or TEP1) is a tumor suppressor gene identified on human chromosome 10q23 (68-70). PTEN is frequently deleted or mutated in a wide range of human cancers, including glioblastoma (71), melanoma (72), and prostate (73), breast (74), and endometrial cancers (75). Germ line PTEN mutations are present in patients with Cowden disease and Bannayan-Zonana syndrome (76, 77). Besides functioning as a tumor suppressor, PTEN is also essential for embryonic development (78-80). Phosphatidylinositol 3,4,5-trisphosphate (PIP<sub>3</sub>) is a substrate of PTEN (81-83). Many tumor-associated missense mutations cluster around the phosphatase domain, and most remaining mutations are predicted to truncate the protein due to nonsense or frameshift mutations (68, 69, 84), suggesting that the phosphatase activity of PTEN plays important roles in PTEN function. Recent evidence demonstrates the ability of PTEN to directly dephosphorylate position D3 of PIP<sub>3</sub> (81). PTEN increases sensitivity to cell death in response to several apoptotic stimuli by negatively regulating the PI3K/Akt pathway (82). In addition to its role in regulating the PI3K/Akt cell survival pathway, PTEN also inhibits growth factor-induced Shc phosphorylation and suppresses the MAP kinase signaling pathway (85), suggesting that PTEN has roles in independent signaling pathways. We have shown that PTEN overexpression in human prostate cancer cells induces apoptosis, regardless of the presence of endogenous PTEN, and Akt was identified as a key molecule in this effect (86).

Akt is one of the most frequently activated protein kinases in human cancer (87). Hyperactivation of Akt is associated with resistance to apoptosis, increased cell growth, cell proliferation, and cellular energy metabolism (87, 88). Thus, Akt contributes to tumor growth and progression by promoting cell invasiveness and angiogenesis (86, 89-94). Overexpression of Akt has been reported in a variety of human cancers, and cells expressing elevated levels of Akt are less sensitive to apoptosis stimuli (95-98). Mammalian cells express three highly homologous Akt isoforms (Akt-1-3) that are encoded by separate genes and share over 80% amino acid sequence identity. Upon activation, growth factor receptors activate the catalytic p110 subunit of PI3K via recruitment of the corresponding p85 regulatory subunit or via ras activation, which can directly activate p110. p110 then phosphorylates phosphoinositides (PI) at the D3 position of the inositol ring to generate PI (3,4,5) P<sub>3</sub> (PIP<sub>3</sub>). The rate limiting steps in Akt activation is the binding of PIP<sub>3</sub> to the PH domain of Akt and subsequent translocation of Akt to the plasma membrane. Akt is then phosphorylated by PI3K-dependent kinase-1 (PDK1) at a threonine residue in the catalytic domain (Thr308), and by another kinase PDK2 at a serine residue (Ser473) in the carboxy-terminal hydrophobic motif. Phosphorylation at both sites is required for full activation of Akt. Antagonizing PI3K activity negatively regulates Akt activity. Once activated, however, Akt exerts antiapoptotic effects through phosphorylation of substrates such as Bad (99, 100) and caspase-9 (101) that directly regulate the apoptotic

machinery, or human telomerase reverse transcriptase subunit (102), forkhead transcription family members (103, 104) and IB kinases (105) that indirectly inhibit apoptosis (106).

EGCG has been shown to inhibit cell growth and proliferation by inhibiting PI-3K/Akt pathway in breast, bladder, prostate, and cervical cancers (35, 107-109). Green tea extract and EGCG inhibited serum-induced HIF-1 $\alpha$  protein and VEGF expression by interfering with the PI3-K/Akt/mammalian target of rapamycin signaling pathways in human cervical carcinoma and hepatoma cells (110). EGCG inhibited tyrosine phosphorylation of PDGF $\beta$ -receptor and downstream activation of ERK and PI3-K/Akt pathways in pancreatic cancer cells (111). Treatment with EGCG inhibited the constitutive activation of the EGFR, Stat3, and Akt in YCU-H891 head and neck squamous cell carcinoma (HNSCC) and MDA-MB-231 breast carcinoma cell lines (112). Furthermore, EGCG inhibited PI-3K/Akt activation that, in turn, resulted in modulation of Bcl-2 family proteins, leading to enhanced apoptosis of bladder cancer T24 cells (35). These findings suggest that PI3-K/Akt pathway could be a major target for chemoprevention.

### 4.5. Cyclooxygenases

Cyclooxygenase (COX) is an enzyme that is responsible for formation of important biological mediators called prostanoids (including prostaglandins, prostacyclin and thromboxane). Pharmacological inhibition of COX can provide relief from the symptoms of inflammation and pain. Currently three COX isoenzymes are known - COX-1, COX-2 and COX-3. COX-3 is a splice variant of COX-1 which retains intron one and has a frameshift mutation (113). Different tissues express varying levels of COX-1 and COX-2. Although both enzymes act basically in the same fashion, selective inhibition can make a difference in terms of side-effects. COX-1 is considered a constitutive enzyme, being found in most mammalian cells. More recently it has been shown to be upregulated in various carcinomas and to have a central role in tumorigenesis. COX-2, on the other hand, is undetectable in most normal tissues. It is an inducible enzyme, becoming abundant in activated macrophages and other cells at sites of inflammation.

EGCG inhibits COX-2 without affecting COX-1 expression at both the mRNA and protein levels, in androgen-sensitive LNCaP and androgen-insensitive PC-3 human prostate carcinoma cells (114). EGCG inhibited the expression of COX-2 and the production of PGE-2 (115). The effect of EGCG on COX-2 expression resulted in decreased COX-2 promoter activity via inhibition of NF $\kappa$ B activation (116). EGCG also promoted rapid mRNA decay mediated through the COX-2 3'untranslated region (3'UTR). In conclusion, these data suggest that inhibition of COX-2 is a mechanism for the anti-proliferative effect of green tea, and emphasizes the role of green tea polyphenols in cancer prevention and treatment.

EGCG treatment to colon cancer cells resulted in a strong activation of AMPK (AMP-activated protein

kinase) and an inhibition of COX-2 expression (117). The decreased COX-2 expression as well as prostaglandin E2 secretion by EGCG was completely abolished by inhibiting AMPK by an AMPK inhibitor, compound C. In another study, cotreatment of lung cancer cells with EGCG plus celecoxib (a cyclooxygenase-2 inhibitor) strongly induced the expression of GADD153 (growth arrest and DNA damage-inducible 153), while neither EGCG nor celecoxib alone was effective (118). However, cotreatment did not induce expression of other apoptosis related genes, p21<sup>WAF1/CIP1</sup> and GADD45. Upregulation of GADD153 by cotreatment with EGCG plus celecoxib correlated with induction of apoptosis.

### 4.6. Epidermal growth factor

The epidermal growth factor receptor (EGFR) belongs to a family of receptor tyrosine kinases in mammals which is composed of four members: EGFR/ERB1, ERB2/ HER2/neu, ERB3, and ERB4 (119, 120). EGFR is an 1186 amino acid residue transmembrane glycoprotein (121). The binding of specific polypeptide ligands results in phosphorylation of multiple tyrosine residues in the COOH-terminal tail, triggering the cellular signaling pathway that regulates fundamental cellular processes such as proliferation, migration, differentiation and survival. Although the ERB family is regarded as the prototypical group of the receptor tyrosine kinase (RTK) family, an important defining feature of the ERB network is that two members of the family, ERB2 and ERB3, are non-autonomous. ERB2 lacks the capacity to interact with a growth-factor ligand, whereas the kinase activity of ERB3 is defective. Despite this lack of autonomy, both ERB2 and ERB3 form heterodimeric complexes with other ERB receptors that are capable of generating potent cellular signals. EGFR is over expressed in many types of tumor cells, such as breast, brain, bladder, lung, gastric, head & neck, cervix, ovary and endometrium (122).

EGCG caused a decrease in the phosphorylated forms of EGFR and HER2 proteins, and subsequently caused a decrease in the phosphorylated forms of the ERK and Akt proteins (123). Similar effects of these compounds were seen when the cells were stimulated with TGF $\alpha$ . Reporter assays indicated that EGCG inhibited the transcriptional activity of the AP-1, c-fos, NF $\kappa$ B, and cyclin D1 promoters (123). In head and neck squamous cell carcinoma, EGCG inhibited phosphorylation of the EGFR, Stat3 and ERK proteins, and also inhibited basal and TGF $\beta$ -stimulated c-fos and cyclin D1 promoter activity (124). In cervical cancer, EGCG inhibited EGF-dependent activation of EGFR, and EGFR-dependent activation of the ERK1/2 (107). EGCG also inhibited EGFR-dependent AKT activity. The EGCG-dependent reduction in ERK and AKT activity is associated with reduced phosphorylation of downstream substrates, including p90RSK, FKHR, and BAD. In another study, EGCG markedly inhibited EGF-induced cell transformation of mouse epidermal JB6 Cl 41 cells (125), and EGF-induced activation of AP-1, and PI3K (125). Overall, these studies demonstrate that targeting the EGF signaling pathway by EGCG may be an effective strategy for prevention and treatment of cancers.

### 4.7. Insulin-like growth factors

The insulin-like growth factors (IGFs) play an important role in normal growth and development (126). Evidence suggests they may also regulate the growth of several cancer cell types (126-131). This regulation is mediated by interactions between the receptors and ligands. There is now ample evidence to suggest that these interactions are also influenced by extracellular IGF binding proteins (IGFBPs). Six different IGFBPs have been cloned. Some species may act to inhibit the mitogenic effects of the IGFs. Furthermore, inhibitory binding proteins could be used as neutralizers of IGF action. The insulin-like growth factor (IGF)-I receptor (IGF-IR) is a tyrosine kinase receptor that is activated by the binding of secreted growth factors, IGF-I or IGF-II. The IGF-IR is a heterotetrameric transmembrane glycoprotein with two identical  $\alpha$ -subunits, which are responsible for ligand binding, and two identical  $\beta$ -subunits, which contain a juxtamembrane domain, an ATP binding pocket, an intracellular tyrosine kinase domain, and COOH terminus, and are joined by disulfide bridges (132). On ligand interaction with the IGF-IR $\alpha$  subunit, residues in the tyrosine kinase domain of the  $\beta$ -subunit are autophosphorylated. Additional phosphorylation sites adjacent to these tyrosine residues can serve as a docking site for the adaptor protein, insulin receptor substrate-1 (IRS-1), which mediates activity through the regulatory subunits of PI-3K. The receptor can also recruit the Src homology-2 domain containing transforming protein, leading to activation of the Ras/Raf/ERK pathway (126, 133).

EGCG is a highly potent inhibitor of IGF-IR tyrosine kinase activity and malignant cell growth (134). Furthermore, IGF-IR autophosphorylation in the presence of increasing ATP concentrations was unaltered by EGCG treatment. Thus, EGCG can block IGF-IR kinase activity and phosphorylation of its downstream targets, resulting in an inhibition of IGF-IR-mediated cell proliferation and transformation. Green tea polyphenols inhibited IGF-1 and IGFBP-3 in TRAMP mice (135). Neutralization of IGF-I with an antihuman IGF-I antibody reduced viability of the human glioblastoma cell lines (136), suggesting that EGCG has an inhibitory effect on malignant brain tumors, and IGF-I may be involved in the effects of EGCG. Overall, these studies demonstrate that targeting the IGF-I signaling pathway by EGCG may be useful for prevention and treatment of cancer.

### 4.8. Transcription factors

#### 4.8.1. NF $\kappa$ B

Nuclear factor- $\kappa$ B (NF $\kappa$ B) is a family of closely related protein dimers that bind a common sequence motif in DNA called the  $\kappa$ B site, which was originally discovered in B cells. Under resting conditions, NF $\kappa$ B dimers reside in the cytoplasm. NF $\kappa$ B is activated by free radicals, inflammatory stimuli, cytokines, carcinogens, tumor promoters, endotoxins  $\gamma$ -radiation, ultraviolet light and X-rays. Upon activation, it is translocated to the nucleus, where it induces the expression of more than 200 genes that have been shown to suppress apoptosis and induce cellular transformation, proliferation, invasion, metastasis, chemo-

resistance, radio-resistance and inflammation. Many of the target genes that are activated are critical to the establishment of the early and late stages of aggressive cancers including expression of cyclin D1, Bcl-2, Bcl-X<sub>L</sub>, MMPs and VEGF.

Phosphorylation of I $\kappa$ B by I $\kappa$ B kinase causes ubiquitination and degradation of I $\kappa$ B, thus releasing NF $\kappa$ B that then translocates to the nucleus. Phosphorylation and activation of I $\kappa$ B kinase is controlled by and NF $\kappa$ B inducing kinases and there is crosstalk between activation of the MAPK, ERK pathway and the NF $\kappa$ B-inducing kinase/I $\kappa$ B kinase/NF $\kappa$ B pathway. EGCG has been shown to inhibit NF $\kappa$ B activity in human colon, prostate cancer cells (137-143). Treatment of normal human epidermal keratinocytes with EGCG was found to inhibit UVB-mediated activation of NF $\kappa$ B (144). The cleavage of RelA/p65 subunit of NF $\kappa$ B was blocked by a pan caspase inhibitor N-benzyloxycarbonyl-Val-Ala-Asp(OMe)-fluoromethylketone (Z-VAD-FMK) during EGCG-mediated apoptosis (140). EGCG can suppress NF $\kappa$ B activation as well as other pro-survival pathways such as PI3K/AKT/mTOR and MAPKs in human bronchial epithelial cells, which may contribute to its ability to suppress inflammation, proliferation and angiogenesis induced by cigarette smoke (145). Thus, NF $\kappa$ B is considered as a target for preventing cancer, and modulation of this pathway by EGCG could contribute to its chemopreventive potential.

#### 4.8.2. STAT

Signal transducer and activator of transcription (STAT) proteins are the signaling molecules that are activated by phosphorylation through janus kinase (JAK) or cytokine receptors, G-protein-coupled receptors, or growth factor receptors, or by intracellular non-receptor tyrosine kinase recruitment (146, 147). Seven mammalian STAT proteins have been identified. STAT3 and STAT5 have been implicated in multiple myeloma, lymphomas, leukemias, and several solid tumors making these proteins logical targets for cancer therapy. These STAT proteins contribute to cell survival and growth by preventing apoptosis through increased expression of anti-apoptotic proteins, such as Bcl-2 and Bcl-X<sub>L</sub>. STAT3 has been shown to directly activate VEGF gene, which is responsible for increased angiogenesis. Elevated STAT3 activity has been detected in head and neck squamous cell carcinoma (148), leukemias (149), lymphomas (150) and multiple myeloma (151). EGCG has been shown to suppress STAT activation in tumor cells and down regulate the phosphorylation of STAT3 (124).

Consumption of green tea is able to mediate cardioprotection and enhance cardiac function during ischemia/reperfusion injury. Green tea extract (GTE) and EGCG inhibit STAT1 activation and protect the myocardium against ischaemia/reperfusion injury (152, 153). Because GTE-mediated cardioprotection is achieved, at least in part, through inhibition of STAT1 activity, a similar action can be implemented in the clinical setting to minimize STAT1 activation levels in patients with acute coronary artery disease.

### 4.8.3. AP-1

Activator Protein 1 (AP-1) transcription factor is a protein dimer composed of members of the basic region leucine zipper protein superfamily, specifically, the Jun, Fos, and activating transcription factor proteins (154). AP-1 activity has been implicated in various cellular functions including proliferation, transformation, differentiation, and apoptosis (155, 156). High AP-1 activity has also been shown to be involved in the tumor promotion and progression of various types of cancers, such as lung, breast, and skin cancer (157, 158). AP-1 regulates many genes that contain the specific DNA sequences in the promoter region collectively called the TPA response element. One class of genes that AP-1 regulates is matrix metalloproteinases, which catalyze the proteolytic cleavage of extracellular matrix components; AP-1 activity has been associated with invasive and metastatic characteristics of cancer cells (159, 160). Recently, EGCG was shown to inhibit AP1 activity through the inhibition of MAPK, specifically, the JNK (161). EGCG increases involucrin gene expression, suggesting that it enhances normal human keratinocyte differentiation (162, 163). EGCG increases hINV (AP1 factor-regulated human involucrin) promoter activity that requires the presence of an intact hINV promoter AP1 factor binding site (163). Fra-1, Fra-2, FosB, JunB, JunD, c-Jun, and c-Fos levels are increased by EGCG treatment, as is AP1 factor binding to hINV promoter AP1 site. EGCG response requires Ras, MEKK1, MEK3, and p38 kinases. These studies demonstrate that in normal human keratinocytes, EGCG markedly increases, via a MAPK signaling mechanism, AP1 factor-associated responses. The mechanisms of action of other tea polyphenols are not well understood.

### 4.8.4. Nrf2

Nuclear Factor-E2-related Factor 2 (Nrf2) is a key transcriptional factor that activates the antioxidant-reactive element (ARE) and in turn regulates the expression of antioxidant phase II detoxifying enzymes (164). It is interesting to note that the promoter region of heme oxygenase-1 gene (HO-1) contains the ARE sequence (165). The mode of transcriptional activation of Nrf2 is not fully understood. Even though, it was considered that several upstream signaling kinases, including protein kinase C (PKC), phosphoinositol 3-kinase (PI3K), and mitogen-activated protein kinases (p38, ERK1/2, and JNK) regulate Nrf2/ARE activity (164-166). However, it is still unclear which kinase acts as an upstream mediator of Nrf2. It was observed that human lung adenocarcinoma A549 cells, which belong to NSCLC, were significantly resistant to the induction of apoptosis by EGCG and these cells express high levels of constitutive HO-1 and Nrf2, as compared with other human cancer cells (167). Although Nrf2 plays a critical role in protection against pulmonary fibrosis, presumably through enhancement of cellular antioxidant capacity (168), HO-1/Nrf2 activation as a defense mechanism in carcinoma cells during lung carcinogenesis may lead to their resistance to chemopreventive and chemotherapeutic regimens.

## 5. Apoptosis and cell cycle

Apoptosis is the protective mechanism through which unwanted cells are eliminated from the system. This is essential for normal development, turnover and replacement of cells in the living system and serves as the protective mechanism against cancer. It was reported that EGCG induces cell cycle arrest and apoptosis in many cancer cells without affecting the normal cells (169, 170). EGCG induces the expression of Cdk inhibitor p21<sup>WAF1/CIP1</sup>, and p27<sup>KIP1</sup>, decreases the expression of cyclin D1 and inhibits Cdk2 and Cdk4 kinases (171-173). Thus, EGCG either exerts its growth-inhibitory effects through modulation of the activities of several key G1 regulatory proteins such as Cdk2 and Cdk4 or mediates the induction of p21 and p27. In regards to apoptosis, EGCG activates caspase-3 and caspase-9, regulates mitochondrial functions (release of cytochrome c and Smac/DIABLO, and depolarization of mitochondrial membranes), and cleaves PARP. These physiological events are critical for the mitochondrial-dependent apoptosis or cell-intrinsic pathway of apoptosis (174-177).

## 6. CLINICAL SIGNIFICANCE OF EGCG

### 6.1. Effects of EGCG on cancer

#### 6.1.1. Prostate cancer

Prostate cancer is the leading cause of cancer-related deaths among males in the U.S. Green tea and its major constituent EGCG inhibit the growth of a variety of human prostate cancer-cell lines. EGCG treatment of prostate cancer cells resulted in dose-dependent inhibition of cell growth, cell cycle arrest, and induction of apoptosis (171, 178-185). Chemoprevention involving the use of natural or synthetic agents to suppress, block or reverse the process of carcinogenesis could be an effective approach to reduce the incidence of prostate cancer. Indeed, prostate cancer represents an excellent candidate disease for chemoprevention, because it is typically diagnosed in elderly men. Consistent with this assumption, there is intense activity in defining chemopreventive agents and molecular targets for prostate cancer chemoprevention. Among the many such agents that are available, for a variety of reasons, naturally occurring nontoxic dietary substances are preferred. EGCG and related catechins were found to inhibit the growth of prostate cancer in cell culture, and xenografts and TRAMP models (12, 50, 171, 179-181, 184-187). A preclinical study has demonstrated the accumulation of tea polyphenols (PP) and theaflavins in the small and large intestine, liver, and prostate of C57BL/6 mice which received decaffeinated black tea diet (185). In a human clinical trial, tea PPs were greater in prostate samples from men consuming black (BT) and green tea (GT) than in men consuming a caffeine-matched soda control (SC) (185). Although tea PP were not detectable in serum, *ex vivo* LNCaP prostate cancer cell proliferation was less when cells were grown in media containing patient serum collected after BT and GT consumption relative to baseline serum. Thus, the tea polyphenols and theaflavins are bioavailable in the prostate where they may be active in the prevention of prostate cancer.

### 6.1.2. Lung Cancer

Green tea extracts at dose of 2% and EGCG at 1.2 mM in the drinking water inhibits lung tumorigenesis in mice treated with a potent nitrosoamine found in tobacco smoke (188). Oral administration of green tea and EGCG inhibits metastasis of Lewis lung carcinoma LL2 cells in mice (189). Since superoxide can enhance the invasiveness of tumor cells, EGCG radical scavenging activity may be related to its inhibition of cancer-cell invasion and metastasis (189). Green tea at a dose of 0.6% as the sole source of drinking water, reduces tumor multiplicity in transgenic mice treated with 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK) (190). Drinking tea for 4-8 weeks reduces NNK-induced expression of mouse lung oncogenes, such as *c-myc*, *c-raf* and *c-H-ras*, suggesting a possible mechanism of green tea action through modulation of oncogene expression (191).

### 6.1.3. Skin cancer

Skin is the largest body organ, and serves as a protective barrier against environmental insults such as UV radiation-induced damage. Much of the deleterious effect of solar UV radiation is caused by UVB (290-320 nm). UVB induces skin cells to produce reactive oxygen species (ROS), eicosanoids, proteinases, and cytokines; the inhibition of these mediators is considered to reduce skin damage. Antioxidants such as ascorbic acid and  $\alpha$ -tocopherol have been demonstrated to produce photoprotective effects in some *in vitro* and *in vivo* studies (192). It was reported that EGCG treatment inhibits UVB-induced infiltration of leukocytes (macrophage/neutrophils), a potential source of generation of ROS and prostaglandin metabolites, which play critical roles in skin tumor promotion in multistage skin carcinogenesis (193, 194). Furthermore, topical treatment with EGCG prevented UV-induced suppression of the contact hypersensitivity in wild-type (WT) mice but had no effect in IL-12 knockout mice (195). Injection of anti-IL-12 monoclonal antibody to WT mice blocked the preventive effect of EGCG on UV-induced immunosuppression. These studies suggest that EGCG can prevent UV-induced immunosuppression, and this may contribute to the chemopreventive activity of EGCG in prevention of photocarcinogenesis.

### 6.1.4. Pancreatic cancer

Cancer of the pancreas is the fourth leading cause of cancer death in the United States. This year approximately 32,000 Americans will die from cancer of the pancreas. With an overall 5-year survival rate of 3% (196), pancreatic cancer has one of the poorest prognoses among all cancers (197). Aside from its silent nature and tendency for late discovery, pancreatic cancer also shows unusual resistance to chemotherapy and radiation. Only 20% of pancreatic cancer patients are eligible for surgical resection, which currently remains the only potentially curative therapy (198). The operations are very complex, and unless performed by surgeons specially trained and experienced in this procedure, they can be associated with very high rates of operative morbidity and mortality. Unfortunately, many cancers of the pancreas are not resectable at the time of diagnosis. There are limited

treatment options available for this disease because chemo- and radio-therapies are largely ineffective, and metastatic disease frequently redevelops even after surgery. Therefore, there is an urgent need to discover novel and effective chemopreventive approaches for pancreatic cancer.

Despite these successes, there is clearly a great need to improve our understanding of the fundamental nature of cancer of the pancreas. Ductal cancer of the pancreas putatively evolves through multistage neoplastic transformation processes that are reflected in a series of histologically well-defined precursor lesions termed pancreatic intraepithelial neoplasias (PanIN) (199). On the molecular level, the interplay between different signaling pathways remains an area of active investigation. Mutations in the K-ras gene occur early, the inactivation of the p16<sup>INK4A</sup> gene at intermediate stages, and the inactivation of p53 and DPC/Smad4 at a relatively late stage (200, 201). On the tissue level, the cell type that gives rise to ductal adenocarcinoma is not well understood. Proposed cellular origins for pancreatic carcinoma include duct cells (199, 202), islet cells (203, 204), acinar cells (205-207), or rare undifferentiated precursor cells (208). Centroacinar cells have emerged as a candidate cell of origin based upon the persistent activation of the Notch pathway in these cells in adulthood (209). Although centroacinar cells constitute the terminal cells of the ductal system and contain ultrastructural features of ductal cells, the precise lineage of centroacinar cells has not yet been elucidated.

Pancreatic cancer becomes clinically apparent at late stages and it resists all forms of conventional chemotherapy and radiotherapy (196, 197). Therefore, understanding the pathogenesis of the preinvasive stage, and developing effective strategies to prevent pancreatic neoplasms are of paramount importance. It has been reported that the quantity of green tea consumed, plays an important role in reducing cancer risk and in delaying cancer outbreak and recurrence. EGCG can exert a growth-suppressive effect on human pancreatic cancer cells *in vitro* (13-15). It induces cell cycle arrest and apoptosis in pancreatic cancer cells and thus holds great promise for development as a chemopreventive agent. In addition, EGCG causes Bax oligomerization, generates reactive oxygen species (ROS), and depolarizes mitochondrial membranes to facilitate cytochrome *c* release into cytosol (14). Furthermore, EGCG activates c-Jun N-terminal kinase (JNK) in pancreatic carcinoma cells (14). Black and green tea extracts, GTP, and EGCG decreased the expression of the K-ras gene, and inhibited growth of pancreatic cancer cells (13). Thus, EGCG may be a potent biologic inhibitor of human pancreatic carcinomas, reducing their proliferative and invasive activities.

### 6.1.5. Breast cancer

EGCG was reported to be cytotoxic toward breast cancer cells. EGCG inhibited cell proliferation in MCF-7, BT474, Hs578t, MDA-MB-231, MBA-MB-468 and BT-20 cells, but had no effect on normal mammary epithelial cells (7, 38, 108, 175, 210-218). EGCG and



related polyphenols induced apoptosis in breast cancer cells through caspase activation and mitochondrial damage (7, 38, 108, 175, 210-218). EGCG inhibits the expression of cyclin D, cyclin E, CDK4, CDK1 and PCNA, which are correlated with cell cycle arrest at G1 (214). Studies in the ER negative cell line, MDA-MB-231, showed a very similar trend, with increased protein expression of p21<sup>WAF1/CIP1</sup> and p27<sup>KIP1</sup> following EGCG treatment (112). EGCG induced apoptosis in T-47D cells through caspase cascade and the cells were detained at the G1 phase. The rate of apoptosis and activity of caspase-3 induced by EGCG was time and dose dependent (215). Nude mice inoculated with human breast cancer MDA-MB-231 cells and treated with GTP and EGCG were effective in delaying the tumor incidence as well as reducing the tumor burden compared to control (214). GTP and EGCG treatment also induced tumor cell apoptosis and inhibited proliferation in xenografted nude mice. Thus, GTP and EGCG treatment inhibits proliferation and induce apoptosis of breast cancer cells *in vitro* and *in vivo*. All together, these studies strongly suggest that GTP and EGCG have anti-tumor properties.

### 6.1.6. Ovarian cancer

EGCG inhibited ovarian cancer cell growth, caused cell cycle arrest and induced apoptosis in SKOV-3 (p53 negative), OVCAR-3 (mutant type p53) and PA-1 (wild type p53) cells (219). The cell cycle was arrested at the G1 phase by EGCG in SKOV-3 and OVCAR-3 cells, whereas in PA-1 cells it exerted its effect at the G1/S phase. EGCG differentially regulated the expression of Bax, p21, Rb, cyclin D1, CDK4, Bcl-X<sub>L</sub>, showing a possible gene regulatory role of EGCG. The continual expression in p21<sup>WAF1</sup> suggests that EGCG acts in the same way with p53 proteins to facilitate apoptosis. Bax, PCNA, and Bcl-X are important in EGCG-mediated apoptosis. In contrast, CDK4 and Rb are not important in ovarian cancer cell growth inhibition. EGCG can inhibit ovarian cancer cell growth through induction of apoptosis and cell cycle arrest as well as regulation of cell cycle-related proteins. Thereby, the EGCG-mediated apoptosis can be applied to an advanced strategy in the development of a potential drug against ovarian cancer.

The endothelin (ET) A receptor (ET(A)R)/ET-1 autocrine pathway is overexpressed in ovarian carcinoma and triggers tumor growth, neoangiogenesis, and invasion. These latter tumor-promoting effects are mediated through the activation of cyclooxygenase (COX)-1- and COX-2-dependent pathways by ET-1. Pretreatment of HEY and OVCA 433 ovarian carcinoma cell lines with green tea and EGCG inhibited ET-1/ET(A)R expression, ET(A)R-mediated COX-1/2 mRNA expression, and COX-2 promoter activity (220). These effects were associated with a significant reduction in the COX-1/2-derived prostaglandin E2 (PGE2) production. These results provide a novel insight into the mechanism by which EGCG, by affecting ET(A)R-dependent COX-1/2 pathways may inhibit ovarian tumors suggesting that EGCG may be useful in preventing and treating ovarian carcinoma in which activation of ET(A)R by ET-1 plays a critical role in tumor growth and progression. The EGCG-induced inhibitory

effects were also associated with a decrease in ET(A)R-dependent activation of the ERK1/2 and p38 MAPKs and PI-3K pathway (39).

Eight ovarian cancer cell lines were tested (SKOV3, CAOV3, OVCAR3, OVCAR10, A2780, CP70, C30, and C200) and showed IC50s for EGCG at the micromolar range, including ones that are resistant to the chemotherapeutic drug cisplatin (221). The ovarian cancer cells were sensitive to H<sub>2</sub>O<sub>2</sub> at similar concentrations, and EGCG treatment led to enhanced intracellular H<sub>2</sub>O<sub>2</sub> (221). Neutralization with pyruvate, a scavenger of H<sub>2</sub>O<sub>2</sub>, suggests that the toxicity of EGCG may be mediated by oxidative stress from the free radical. Addition of Tempol, a superoxide dismutase mimetic, demonstrates that H<sub>2</sub>O<sub>2</sub> might be generated endogenously from superoxide. The toxicity of cisplatin and the development of cisplatin resistance are major obstacles in treatment of ovarian cancer. Interestingly, the addition of EGCG amplified the toxicity of cisplatin. EGCG significantly increased cisplatin potency in SKOV3, CAOV3, and C200 cells, the latter being a cell line induced to have several hundred fold resistant to cisplatin above the parental line. These findings suggest that EGCG may accentuate oxidative stress to inhibit growth of ovarian cancer cells and sensitize them to cisplatin.

### 6.2. Effects of EGCG on angiogenesis and metastasis

Angiogenesis, the formation of new capillaries from pre-existing vessels, is required for several physiological processes as well as pathological conditions. It is composed of several steps which include the degradation of vascular basement membrane matrix by protease, migration and proliferation of endothelial cells into interstitium, endothelial tube formation, recruitment and attachment of mesenchymal cells to the endothelial cells tube, and maturation of blood vessels with the formation of vascular basement membrane (222). Upon angiogenic stimuli, proteases represented by matrix metalloprotease (MMP) are activated and, in turn, degrade vascular basement membranes, thus leading to the migration of activated endothelial cells into interstitium. Several proangiogenic (e.g. VEGF, bFGF, PlGF, G-CSF, HGF, angiopoietin-1/2, angiogenin, proliferin, PDGF-BB, TGF- $\alpha$ , TGF- $\beta$ , TNF- $\alpha$ , IL-3, and IL-8) and antiangiogenic factors (e.g. IFN, PF4, angiostatin, endostatin, vasostatin, arrestin, angioarrestin, vasohibin, IL-4, IL-10, IL-12, IP-10, MIG) have been reported (222, 223).

VEGF is a mitogen for endothelial cells and is associated with tumor-induced angiogenesis. VEGF binds to VEGF receptors and is responsible for most of its mitogenic and chemotactic effects. EGCG inhibited microvessel density, endothelial cell growth, chemotaxis, invasion, VEGF receptor phosphorylation and induced apoptosis (224-226). EGCG inhibited phosphorylation of VEGF receptors and ERK1/2, and mRNA expression of the early growth response factor-1 (227). The inhibition of VEGF binding to its receptors may contribute to the antiangiogenic and cancer chemopreventive effects of EGCG. EGCG may exert at least part of its anticancer effect by inhibiting angiogenesis through blocking the

induction of VEGF. VEGF-induced IL-8 production at the mRNA and protein levels are also suppressed with EGCG (228). Furthermore, EGCG may exert the anti-angiogenic effect by inhibiting the PDGF-induced VEGF expression at multiple signaling levels (229).

Matrix metalloproteinases (MMP) that participate in extracellular matrix degradation are involved in the development of metastasis, epidermal detachment and hepatic fibrosis. MMP-2 and MMP-9, seems to play an important role in tumor invasion and metastasis. EGCG has been shown to affect MMP-2 and MMP-9 activity both directly and indirectly in endothelial cells thereby inhibit or delay cancer invasion, metastasis, and angiogenesis via modulations in MMPs (46, 180, 230-235). MMP7 was shown to degrade *ex vivo* on healthy normal skin collagen VII and fibrillin 1. MMP7 could take an active part in the epidermal detachment occurring during recessive dystrophic epidermolysis bullosa (RDEB). EGCG inhibited MMP7 and developed a good protection of collagen type VII and fibrillin 1 susceptible of being degraded by MMP7 (231). Thus, EGCG could be used beneficially in patients suffering from RDEB. EGCG may also exert anti-fibrogenic activity (236). It inhibited expression of MMP-2 mRNA and protein in rat hepatic stellate cells (HSC) (236). EGCG treatment also reduced concanavalin A (ConA)-induced activation of secreted MMP-2 and reduced MT1-MMP activity. In addition, EGCG inhibited either HSC migration or invasion. The abilities of EGCG to suppress MMP-2 activation and HSC invasiveness suggest that EGCG may be useful in the treatment and prevention of hepatic fibrosis.

During tumor neovascularization, vascular endothelial growth factor and ephrin (Eph) families emerge as critical mediators of angiogenesis. EGCG inhibited ephrin-A1-mediated endothelial cell migration, as well as tumor angiogenesis (237). Furthermore, EGCG inhibited the ephrin-A1-mediated phosphorylation of EphA2 and ERK-1/2. Taken together, these data indicate that activation of ERK-1/2 plays an essential role in ephrin-A1-mediated cell migration, and suggest a novel anti-angiogenic role of EGCG in cancer chemoprevention.

The endothelin A receptor (ET(A)R)/endothelin-1 (ET-1) axis is overexpressed in ovarian carcinoma representing a novel therapeutic target. Treatment with green tea or EGCG inhibited ET(A)R and ET-1 expression and reduced the basal and ET-1-induced cell proliferation and invasion (39). The EGCG-induced inhibitory effects were associated with a decrease of ET(A)R-dependent activation of the ERK-1/2 and p38 MAPKs and PI3-K pathways. Remarkably, EGCG treatment resulted in a lowering of basal and ET-1-induced angiogenesis and invasiveness mediators, such as vascular endothelial growth factor and tumor proteinase activation. EGCG inhibited HEY ovarian carcinoma xenografts, and this effect was associated with a reduction in ET-1, ET(A)R, and vascular endothelial growth factor expression, microvessel density, and proliferation index. These results provide a novel insight into the mechanism by which EGCG, affecting multiple ET(A)R-dependent pathways, may inhibit ovarian carcinoma growth.

Catechin, conjugated with fatty acid (acyl-catechin), strongly inhibited DNA polymerase, HL-60 cancer cell growth, and angiogenesis (238). Catechin conjugated with stearic acid [(2R,3S)-3',4',5,7-tetrahydroxyflavan-3-yl octadecanoate; catechin-C18] was the strongest inhibitor in DNA polymerase  $\alpha$  and  $\beta$  and angiogenesis. Catechin-C18 also suppressed human endothelial cell (HUVEC) tube formation on the reconstituted basement membrane, suggesting that it may affect not only DNA polymerases but also signal transduction pathways in HUVECs. Based on these studies, it appears that acyl-catechins target both DNA polymerases and angiogenesis as anticancer agents. Furthermore, acylation of catechin may be an effective chemical modification to improve the anticancer activity of catechin.

## 7. CONCLUSIONS AND FUTURE DIRECTIONS

EGCG is a major constituent in green tea which is a popular beverage (next to water) and shown to afford protection against many cancer types. Green tea and its polyphenolic antioxidants are much more potent than vitamin C and vitamin E scavenging potentially carcinogenic free radicals. Dozens of studies have also demonstrated almost one-third of the cancers are caused by dietary and lifestyle-related habits hence, manipulation of the diet are being recognized as a potential strategy against cancer. Extensive *in vitro* investigations using both hormone responsive and non-responsive cell lines have shown that EGCG induces apoptosis and alters the expression of cell cycle regulatory proteins that are critical for cell survival and apoptosis. Stereoselective total synthesis of EGCG, and its structurally related catechins in the laboratory, could provide new sources of these compounds for investigational and biomedical use. Like most chemopreventive agents, EGCG also possesses limited systemic bioavailability. Furthermore, auto-oxidation of EGCG may be another problem of its reduced biological activity in humans. Recent studies have demonstrated the ability of EGCG to bind high affinity binding proteins as possible direct targets for the action.

Green tea polyphenols inhibit metastasis through regulation of urokinase and matrix metalloproteinases. Polyphenols reduced angiogenesis, in part by decreasing vascular endothelial growth factor production and receptor phosphorylation. Interestingly, EGCG reduced dihydrofolate reductase activity, which would affect nucleic acid and protein synthesis. Furthermore, it also acted as an aryl hydrocarbon receptor antagonist by directly binding the receptor's molecular chaperone, heat shock protein 90.

In conclusion, green and black tea polyphenols act at numerous points regulating cancer cell growth, survival, angiogenesis, and metastasis, and possess several other health benefits. These agents have shown promising results by their ability to inhibit carcinogenesis in laboratory studies. If these effects can be successfully translated into human studies then these agents may prove to be valuable adjuvant therapies in the future. Data from

clinical trials of tea polyphenols are needed to define the optimal dosing, schedule, toxicities, and clinical efficacy before widespread use can be recommended.

## 8. ACKNOWLEDGEMENTS

This work was supported by grants from the National Institutes of Health and the Department of Defense. We thank all the lab members for critically reading the manuscript.

## 12. REFERENCES

- Ahn, W. S., S. W. Huh, S. M. Bae, I. P. Lee, J. M. Lee, S. E. Namkoong, C. K. Kim & J. I. Sin: A major constituent of green tea, EGCG, inhibits the growth of a human cervical cancer cell line, CaSki cells, through apoptosis, G (1) arrest, and regulation of gene expression. *DNA Cell Biol*, 22, 217-24 (2003)
- Manson, M. M., P. B. Farmer, A. Gescher & W. P. Steward: Innovative agents in cancer prevention. *Recent Results Cancer Res*, 166, 257-75 (2005)
- Lambert, J. D. & C. S. Yang: Mechanisms of cancer prevention by tea constituents. *J Nutr*, 133, 3262S-3267S (2003)
- Muto, S., T. Yokoi, Y. Gondo, M. Katsuki, Y. Shioyama, K. Fujita & T. Kamataki: Inhibition of benzo[a]pyrene-induced mutagenesis by (-)-epigallocatechin gallate in the lung of rpsL transgenic mice. *Carcinogenesis*, 20, 421-4 (1999)
- Sachinidis, A., C. Seul, S. Seewald, H. Ahn, Y. Ko & H. Vetter: Green tea compounds inhibit tyrosine phosphorylation of PDGF beta-receptor and transformation of A172 human glioblastoma. *FEBS Lett*, 471, 51-5 (2000)
- Ermakova, S., B. Y. Choi, H. S. Choi, B. S. Kang, A. M. Bode & Z. Dong: The intermediate filament protein vimentin is a new target for epigallocatechin gallate. *J Biol Chem*, 280, 16882-90 (2005)
- Ermakova, S. P., B. S. Kang, B. Y. Choi, H. S. Choi, T. F. Schuster, W. Y. Ma, A. M. Bode & Z. Dong: (-)-Epigallocatechin gallate overcomes resistance to etoposide-induced cell death by targeting the molecular chaperone glucose-regulated protein 78. *Cancer Res*, 66, 9260-9 (2006)
- Cao, Y. & R. Cao: Angiogenesis inhibited by drinking tea. *Nature*, 398, 381 (1999)
- Bode, A. M. & Z. Dong: Targeting signal transduction pathways by chemopreventive agents. *Mutat Res*, 555, 33-51 (2004)
- Jung, Y. D. & L. M. Ellis: Inhibition of tumour invasion and angiogenesis by epigallocatechin gallate (EGCG), a major component of green tea. *Int J Exp Pathol*, 82, 309-16 (2001)
- Lambert, J. D. & C. S. Yang: Cancer chemopreventive activity and bioavailability of tea and tea polyphenols. *Mutat Res*, 523-524, 201-8 (2003)
- Park, O. J. & Y. J. Surh: Chemopreventive potential of epigallocatechin gallate and genistein: evidence from epidemiological and laboratory studies. *Toxicol Lett*, 150, 43-56 (2004)
- Lyn-Cook, B. D., T. Rogers, Y. Yan, E. B. Blann, F. F. Kadlubar & G. J. Hammons: Chemopreventive effects of tea extracts and various components on human pancreatic and prostate tumor cells *in vitro*. *Nutr Cancer*, 35, 80-6 (1999)
- Qanungo, S., M. Das, S. Haldar & A. Basu: Epigallocatechin-3-gallate induces mitochondrial membrane depolarization and caspase-dependent apoptosis in pancreatic cancer cells. *Carcinogenesis*, 26, 958-67 (2005)
- Tan, M., A. Norwood, M. May, M. Tucci & H. Benghuzzi: Effects of (-)-epigallocatechin gallate and thymoquinone on proliferation of a PANC-1 cell line in culture. *Biomed Sci Instrum*, 42, 363-71 (2006)
- Takada, M., Y. Nakamura, T. Koizumi, H. Toyama, T. Kamigaki, Y. Suzuki, Y. Takeyama & Y. Kuroda: Suppression of human pancreatic carcinoma cell growth and invasion by epigallocatechin-3-gallate. *Pancreas*, 25, 45-8 (2002)
- Doss, M. X., S. P. Potta, J. Hescheler & A. Sachinidis: Trapping of growth factors by catechins: a possible therapeutic target for prevention of proliferative diseases. *J Nutr Biochem*, 16, 259-66 (2005)
- Khan, N., F. Afaq, M. Saleem, N. Ahmad & H. Mukhtar: Targeting multiple signaling pathways by green tea polyphenol (-)-epigallocatechin-3-gallate. *Cancer Res*, 66, 2500-5 (2006)
- Korsmeyer, S. J.: Regulators of cell death. *Trends Genet*, 11, 101-5 (1995)
- Cory, S. & J. M. Adams: The Bcl2 family: regulators of the cellular life-or-death switch. *Nat Rev Cancer*, 2, 647-56 (2002)
- Green, D. R.: Apoptotic pathways: the roads to ruin. *Cell*, 94, 695-8 (1998)
- Reed, J. C.: Regulation of apoptosis by bcl-2 family proteins and its role in cancer and chemoresistance. *Curr Opin Oncol*, 7, 541-6 (1995)
- Oltvai, Z. N., C. L. Millman & S. J. Korsmeyer: Bcl-2 heterodimerizes *in vivo* with a conserved homolog, Bax, that accelerates programmed cell death. *Cell*, 74, 609-19 (1993)
- Jeffers, J. R., E. Parganas, Y. Lee, C. Yang, J. Wang, J. Brennan, K. H. MacLean, J. Han, T. Chittenden, J. N. Ihle, P. J. McKinnon, J. L. Cleveland & G. P. Zambetti: Puma is an essential mediator of p53-dependent and -independent apoptotic pathways. *Cancer Cell*, 4, 321-8 (2003)
- Villunger, A., E. M. Michalak, L. Coultas, F. Mullauer, G. Bock, M. J. Ausserlechner, J. M. Adams & A. Strasser: p53- and drug-induced apoptotic responses mediated by BH3-only proteins puma and noxa. *Science*, 302, 1036-8 (2003)
- Villunger, A., C. Scott, P. Bouillet & A. Strasser: Essential role for the BH3-only protein Bim but redundant roles for Bax, Bcl-2, and Bcl-w in the control of granulocyte survival. *Blood*, 101, 2393-400 (2003)
- Osford, S. M., C. L. Dallman, P. W. Johnson, A. Ganesan & G. Packham: Current strategies to target the anti-apoptotic Bcl-2 protein in cancer cells. *Curr Med Chem*, 11, 1031-9 (2004)
- Chan, S. L. & V. C. Yu: Proteins of the bcl-2 family in apoptosis signalling: from mechanistic insights to therapeutic opportunities. *Clin Exp Pharmacol Physiol*, 31, 119-28 (2004)

29. Kirkin, V., S. Joos & M. Zornig: The role of Bcl-2 family members in tumorigenesis. *Biochim Biophys Acta*, 1644, 229-49 (2004)
30. Haldar, S., A. Basu & C. M. Croce: Bcl2 is the guardian of microtubule integrity. *Cancer Res*, 57, 229-33 (1997)
31. Srivastava, R. K., Q. S. Mi, J. M. Hardwick & D. L. Longo: Deletion of the loop region of Bcl-2 completely blocks 28. paclitaxel-induced apoptosis. *Proc Natl Acad Sci U S A*, 96, 3775-80 (1999)
32. Srivastava, R. K., C. Y. Sasaki, J. M. Hardwick & D. L. Longo: Bcl-2-mediated drug resistance: inhibition of apoptosis by blocking nuclear factor of activated T lymphocytes (NFAT)-induced Fas ligand transcription. *J Exp Med*, 190, 253-65 (1999)
33. Srivastava, R. K., S. J. Sollott, L. Khan, R. Hansford, E. G. Lakatta & D. L. Longo: Bcl-2 and Bcl-X (L) block thapsigargin-induced nitric oxide generation, c-Jun NH (2)-terminal kinase activity, and apoptosis. *Mol Cell Biol*, 19, 5659-74 (1999)
34. Siddiqui, I. A., N. Zaman, M. H. Aziz, S. R. Reagan-Shaw, S. Sarfaraz, V. M. Adhami, N. Ahmad, S. Raisuddin & H. Mukhtar: Inhibition of CWR22R{nu}1 tumor growth and PSA secretion in athymic nude mice by green and black teas. *Carcinogenesis*, 27, 833-9 (2006)
35. Qin, J., L. P. Xie, X. Y. Zheng, Y. B. Wang, Y. Bai, H. F. Shen, L. C. Li & R. Dahiya: A component of green tea, (-)-epigallocatechin-3-gallate, promotes apoptosis in T24 human bladder cancer cells via modulation of the PI3K/Akt pathway and Bcl-2 family proteins. *Biochem Biophys Res Commun*, 354, 852-7 (2007)
36. Zhao, Y., L. F. Yang, M. Ye, H. H. Gu & Y. Cao: Induction of apoptosis by epigallocatechin-3-gallate via mitochondrial signal transduction pathway. *Prev Med*, 39, 1172-9 (2004)
37. Chung, J. H., J. H. Han, E. J. Hwang, J. Y. Seo, K. H. Cho, K. H. Kim, J. I. Youn & H. C. Eun: Dual mechanisms of green tea extract (EGCG)-induced cell survival in human epidermal keratinocytes. *Faseb J*, 17, 1913-5 (2003)
38. Baliga, M. S., S. Meleth & S. K. Katiyar: Growth inhibitory and antimetastatic effect of green tea polyphenols on metastasis-specific mouse mammary carcinoma 4T1 cells *in vitro* and *in vivo* systems. *Clin Cancer Res*, 11, 1918-27 (2005)
39. Spinella, F., L. Rosano, V. Di Castro, S. Decandia, A. Albini, M. R. Nicotra, P. G. Natali & A. Bagnato: Green tea polyphenol epigallocatechin-3-gallate inhibits the endothelin axis and downstream signaling pathways in ovarian carcinoma. *Mol Cancer Ther*, 5, 1483-92 (2006)
40. Yu, M., H. Sato, M. Seiki, S. Spiegel & E. W. Thompson: Calcium influx inhibits MT1-MMP processing and blocks MMP-2 activation. *FEBS Lett*, 412, 568-72 (1997)
41. Chambers, A. F. & L. M. Matrisian: Changing views of the role of matrix metalloproteinases in metastasis. *J Natl Cancer Inst*, 89, 1260-70 (1997)
42. John, A. & G. Tuszynski: The role of matrix metalloproteinases in tumor angiogenesis and tumor metastasis. *Pathol Oncol Res*, 7, 14-23 (2001)
43. Nelson, A. R., B. Fingleton, M. L. Rothenberg & L. M. Matrisian: Matrix metalloproteinases: biologic activity and clinical implications. *J Clin Oncol*, 18, 1135-49 (2000)
44. Kleiner, D. E. & W. G. Stetler-Stevenson: Matrix metalloproteinases and metastasis. *Cancer Chemother Pharmacol*, 43 Suppl, S42-51 (1999)
45. Sternlicht, M. D. & Z. Werb: How matrix metalloproteinases regulate cell behavior. *Annu Rev Cell Dev Biol*, 17, 463-516 (2001)
46. Vayalil, P. K. & S. K. Katiyar: Treatment of epigallocatechin-3-gallate inhibits matrix metalloproteinases-2 and -9 via inhibition of activation of mitogen-activated protein kinases, c-jun and NF-kappaB in human prostate carcinoma DU-145 cells. *Prostate*, 59, 33-42 (2004)
47. Brawley, O. W., S. Barnes & H. Parnes: The future of prostate cancer prevention. *Ann N Y Acad Sci*, 952, 145-52 (2001)
48. Yang, J., D. Wei & J. Liu: Repressions of MMP-9 expression and NF-kappa B localization are involved in inhibition of lung carcinoma 95-D cell invasion by (-)-epigallocatechin-3-gallate. *Biomed Pharmacother*, 59, 98-103 (2005)
49. Corps, A. N., V. A. Curry, D. J. Buttle, B. L. Hazleman & G. P. Riley: Inhibition of interleukin-1beta-stimulated collagenase and stromelysin expression in human tendon fibroblasts by epigallocatechin gallate ester. *Matrix Biol*, 23, 163-9 (2004)
50. Adhami, V. M., N. Ahmad & H. Mukhtar: Molecular targets for green tea in prostate cancer prevention. *J Nutr*, 133, 2417S-2424S (2003)
51. Kim, H. S., M. H. Kim, M. Jeong, Y. S. Hwang, S. H. Lim, B. A. Shin, B. W. Ahn & Y. D. Jung: EGCG blocks tumor promoter-induced MMP-9 expression via suppression of MAPK and AP-1 activation in human gastric AGS cells. *Anticancer Res*, 24, 747-53 (2004)
52. Boguski, M. S. & F. McCormick: Proteins regulating Ras and its relatives. *Nature*, 366, 643-54 (1993)
53. Bokoch, G. M. & C. J. Der: Emerging concepts in the Ras superfamily of GTP-binding proteins. *Faseb J*, 7, 750-9 (1993)
54. Gibbs, J. B., I. S. Sigal, M. Poe & E. M. Scolnick: Intrinsic GTPase activity distinguishes normal and oncogenic ras p21 molecules. *Proc Natl Acad Sci U S A*, 81, 5704-8 (1984)
55. Shih, C. & R. A. Weinberg: Isolation of a transforming sequence from a human bladder carcinoma cell line. *Cell*, 29, 161-9 (1982)
56. Campbell, S. L., R. Khosravi-Far, K. L. Rossman, G. J. Clark & C. J. Der: Increasing complexity of Ras signaling. *Oncogene*, 17, 1395-413 (1998)
57. Schaeffer, H. J. & M. J. Weber: Mitogen-activated protein kinases: specific messages from ubiquitous messengers. *Mol Cell Biol*, 19, 2435-44 (1999)
58. Han, J. & R. J. Ulevitch: Emerging targets for anti-inflammatory therapy. *Nat Cell Biol*, 1, E39-40 (1999)
59. Davis, R. J.: Signal transduction by the JNK group of MAP kinases. *Cell*, 103, 239-52 (2000)
60. Chang, L. & M. Karin: Mammalian MAP kinase signalling cascades. *Nature*, 410, 37-40 (2001)
61. Robinson, M. J. & M. H. Cobb: Mitogen-activated protein kinase pathways. *Curr Opin Cell Biol*, 9, 180-6 (1997)
62. Woessmann, W., Y. H. Meng & N. F. Mivechi: An essential role for mitogen-activated protein kinases, ERKs,

in preventing heat-induced cell death. *J Cell Biochem*, 74, 648-62 (1999)

63. Kyriakis, J. M. & J. Avruch: Sounding the alarm: protein kinase cascades activated by stress and inflammation. *J Biol Chem*, 271, 24313-6 (1996)

64. Ono, K. & J. Han: The p38 signal transduction pathway: activation and function. *Cell Signal*, 12, 1-13 (2000)

65. Lubinus, M., K. E. Meier, E. A. Smith, K. C. Gause, E. C. LeRoy & M. Trojanowska: Independent effects of platelet-derived growth factor isoforms on mitogen-activated protein kinase activation and mitogenesis in human dermal fibroblasts. *J Biol Chem*, 269, 9822-5 (1994)

66. Chen, C., G. Shen, V. Hebbar, R. Hu, E. D. Owuor & A. N. Kong: Epigallocatechin-3-gallate-induced stress signals in HT-29 human colon adenocarcinoma cells. *Carcinogenesis*, 24, 1369-78 (2003)

67. Yamamoto, T., H. Digumarthi, Z. Aranbayeva, J. Wataha, J. Lewis, R. Messer, H. Qin, D. Dickinson, T. Osaki, G. S. Schuster & S. Hsu: EGCG-targeted p57/KIP2 reduces tumorigenicity of oral carcinoma cells: Role of c-Jun N-terminal kinase. *Toxicol Appl Pharmacol* (2006)

68. Li, J., C. Yen, D. Liaw, K. Podsypanina, S. Bose, S. I. Wang, J. Puc, C. Miliareis, L. Rodgers, R. McCombie, S. H. Bigner, B. C. Giovanella, M. Ittmann, B. Tycko, H. Hibshoosh, M. H. Wigler & R. Parsons: PTEN, a putative protein tyrosine phosphatase gene mutated in human brain, breast, and prostate cancer. *Science*, 275, 1943-7 (1997)

69. Steck, P. A., M. A. Pershouse, S. A. Jasser, W. K. Yung, H. Lin, A. H. Ligon, L. A. Langford, M. L. Baumgard, T. Hattier, T. Davis, C. Frye, R. Hu, B. Swedlund, D. H. Teng & S. V. Tavtigian: Identification of a candidate tumour suppressor gene, MMAC1, at chromosome 10q23.3 that is mutated in multiple advanced cancers. *Nat Genet*, 15, 356-62 (1997)

70. Li, D. M. & H. Sun: TEP1, encoded by a candidate tumor suppressor locus, is a novel protein tyrosine phosphatase regulated by transforming growth factor beta. *Cancer Res*, 57, 2124-9 (1997)

71. Wang, S. I., J. Puc, J. Li, J. N. Bruce, P. Cairns, D. Sidransky & R. Parsons: Somatic mutations of PTEN in glioblastoma multiforme. *Cancer Res*, 57, 4183-6 (1997)

72. Guldberg, P., P. Thor Straten, A. Birck, V. Ahrenkiel, A. F. Kirkin & J. Zeuthen: Disruption of the MMAC1/PTEN gene by deletion or mutation is a frequent event in malignant melanoma. *Cancer Res*, 57, 3660-3 (1997)

73. Cairns, P., K. Okami, S. Halachmi, N. Halachmi, M. Esteller, J. G. Herman, J. Jen, W. B. Isaacs, G. S. Bova & D. Sidransky: Frequent inactivation of PTEN/MMAC1 in primary prostate cancer. *Cancer Res*, 57, 4997-5000 (1997)

74. Rhei, E., L. Kang, F. Bogomolny, M. G. Federici, P. I. Borgen & J. Boyd: Mutation analysis of the putative tumor suppressor gene PTEN/MMAC1 in primary breast carcinomas. *Cancer Res*, 57, 3657-9 (1997)

75. Tashiro, H., M. S. Blazes, R. Wu, K. R. Cho, S. Bose, S. I. Wang, J. Li, R. Parsons & L. H. Ellenson: Mutations in PTEN are frequent in endometrial carcinoma but rare in other common gynecological malignancies. *Cancer Res*, 57, 3935-40 (1997)

76. Liaw, D., D. J. Marsh, J. Li, P. L. Dahia, S. I. Wang, Z. Zheng, S. Bose, K. M. Call, H. C. Tsou, M. Peacocke, C.

Eng & R. Parsons: Germline mutations of the PTEN gene in Cowden disease, an inherited breast and thyroid cancer syndrome. *Nat Genet*, 16, 64-7 (1997)

77. Marsh, D. J., P. L. Dahia, Z. Zheng, D. Liaw, R. Parsons, R. J. Gorlin & C. Eng: Germline mutations in PTEN are present in Bannayan-Zonana syndrome. *Nat Genet*, 16, 333-4 (1997)

78. Di Cristofano, A., B. Pesce, C. Cordon-Cardo & P. P. Pandolfi: Pten is essential for embryonic development and tumour suppression. *Nat Genet*, 19, 348-55 (1998)

79. Suzuki, A., J. L. de la Pompa, V. Stambolic, A. J. Elia, T. Sasaki, I. del Barco Barrantes, A. Ho, A. Wakeham, A. Itie, W. Khoo, M. Fukumoto & T. W. Mak: High cancer susceptibility and embryonic lethality associated with mutation of the PTEN tumor suppressor gene in mice. *Curr Biol*, 8, 1169-78 (1998)

80. Podsypanina, K., L. H. Ellenson, A. Nemes, J. Gu, M. Tamura, K. M. Yamada, C. Cordon-Cardo, G. Cattoretti, P. E. Fisher & R. Parsons: Mutation of Pten/Mmac1 in mice causes neoplasia in multiple organ systems. *Proc Natl Acad Sci USA*, 96, 1563-8 (1999)

81. Maehama, T. & J. E. Dixon: The tumor suppressor, PTEN/MMAC1, dephosphorylates the lipid second messenger, phosphatidylinositol 3,4,5-trisphosphate. *J Biol Chem*, 273, 13375-8 (1998)

82. Stambolic, V., A. Suzuki, J. L. de la Pompa, G. M. Brothers, C. Mirtsos, T. Sasaki, J. Ruland, J. M. Penninger, D. P. Siderovski & T. W. Mak: Negative regulation of PKB/Akt-dependent cell survival by the tumor suppressor PTEN. *Cell*, 95, 29-39 (1998)

83. Tamura, M., J. Gu, K. Matsumoto, S. Aota, R. Parsons & K. M. Yamada: Inhibition of cell migration, spreading, and focal adhesions by tumor suppressor PTEN. *Science*, 280, 1614-7 (1998)

84. Teng, D. H., R. Hu, H. Lin, T. Davis, D. Iliev, C. Frye, B. Swedlund, K. L. Hansen, V. L. Vinson, K. L. Gumpfer, L. Ellis, A. El-Naggar, M. Frazier, S. Jasser, L. A. Langford, J. Lee, G. B. Mills, M. A. Pershouse, R. E. Pollack, C. Tornos, P. Troncoso, W. K. Yung, G. Fujii, A. Berson, P. A. Steck & *et al.*: MMAC1/PTEN mutations in primary tumor specimens and tumor cell lines. *Cancer Res*, 57, 5221-5 (1997)

85. Gu, J., M. Tamura & K. M. Yamada: Tumor suppressor PTEN inhibits integrin- and growth factor-mediated mitogen-activated protein (MAP) kinase signaling pathways. *J Cell Biol*, 143, 1375-83 (1998)

86. Chen, X., H. Thakkar, F. Tyan, S. Gim, H. Robinson, C. Lee, S. K. Pandey, C. Nwokorie, N. Onwudiwe & R. K. Srivastava: Constitutively active Akt is an important regulator of TRAIL sensitivity in prostate cancer. *Oncogene*, 20, 6073-83 (2001)

87. Downward, J.: PI 3-kinase, Akt and cell survival. *Semin Cell Dev Biol*, 15, 177-82 (2004)

88. Harrington, L. S., G. M. Findlay & R. F. Lamb: Restraining PI3K: mTOR signalling goes back to the membrane. *Trends Biochem Sci*, 30, 35-42 (2005)

89. Dudek, H., S. R. Datta, T. F. Franke, M. J. Birnbaum, R. Yao, G. M. Cooper, R. A. Segal, D. R. Kaplan & M. E. Greenberg: Regulation of neuronal survival by the serine-threonine protein kinase Akt. *Science*, 275, 661-5 (1997)

90. Kaufmann, S. H. & M. O. Hengartner: Programmed cell death: alive and well in the new millennium. *Trends Cell Biol*, 11, 526-34 (2001)

91. Kennedy, S. G., A. J. Wagner, S. D. Conzen, J. Jordan, A. Bellacosa, P. N. Tsichlis & N. Hay: The PI 3-kinase/Akt signaling pathway delivers an anti-apoptotic signal. *Genes Dev*, 11, 701-13 (1997)
92. Khwaja, A.: Akt is more than just a Bad kinase. *Nature*, 401, 33-4 (1999)
93. Kulik, G. & M. J. Weber: Akt-dependent and -independent survival signaling pathways utilized by insulin-like growth factor I. *Mol Cell Biol*, 18, 6711-8 (1998)
94. Kandasamy, K. & R. K. Srivastava: Role of the phosphatidylinositol 3'-kinase/PTEN/Akt kinase pathway in tumor necrosis factor-related apoptosis-inducing ligand-induced apoptosis in non-small cell lung cancer cells. *Cancer Res*, 62, 4929-37 (2002)
95. Asano, T., Y. Yao, J. Zhu, D. Li, J. L. Abbruzzese & S. A. Reddy: The PI 3-kinase/Akt signaling pathway is activated due to aberrant Pten expression and targets transcription factors NF-kappaB and c-Myc in pancreatic cancer cells. *Oncogene*, 23, 8571-80 (2004)
96. Cicens, J., P. Urban, V. Vuaroqueaux, M. Labuhn, W. Kung, E. Wight, M. Mayhew, U. Eppenberger & S. Eppenberger-Castori: Increased level of phosphorylated akt measured by chemiluminescence-linked immunosorbent assay is a predictor of poor prognosis in primary breast cancer overexpressing ErbB-2. *Breast Cancer Res*, 7, R394-401 (2005)
97. Semba, S., T. Moriya, W. Kimura & M. Yamakawa: Phosphorylated Akt/PKB controls cell growth and apoptosis in intraductal papillary-mucinous tumor and invasive ductal adenocarcinoma of the pancreas. *Pancreas*, 26, 250-7 (2003)
98. Yao, Z., Y. Okabayashi, Y. Yutsudo, T. Kitamura, W. Ogawa & M. Kasuga: Role of Akt in growth and survival of PANC-1 pancreatic cancer cells. *Pancreas*, 24, 42-6 (2002)
99. Datta, S. R., H. Dudek, X. Tao, S. Masters, H. Fu, Y. Gotoh & M. E. Greenberg: Akt phosphorylation of BAD couples survival signals to the cell-intrinsic death machinery. *Cell*, 91, 231-41 (1997)
100. del Peso, L., M. Gonzalez-Garcia, C. Page, R. Herrera & G. Nunez: Interleukin-3-induced phosphorylation of BAD through the protein kinase Akt. *Science*, 278, 687-9 (1997)
101. Cardone, M. H., N. Roy, H. R. Stennicke, G. S. Salvesen, T. F. Franke, E. Stanbridge, S. Frisch & J. C. Reed: Regulation of cell death protease caspase-9 by phosphorylation. *Science*, 282, 1318-21 (1998)
102. Kang, S. S., T. Kwon, D. Y. Kwon & S. I. Do: Akt protein kinase enhances human telomerase activity through phosphorylation of telomerase reverse transcriptase subunit. *J Biol Chem*, 274, 13085-90 (1999)
103. Brunet, A., A. Bonni, M. J. Zigmond, M. Z. Lin, P. Juo, L. S. Hu, M. J. Anderson, K. C. Arden, J. Blenis & M. E. Greenberg: Akt promotes cell survival by phosphorylating and inhibiting a Forkhead transcription factor. *Cell*, 96, 857-68 (1999)
104. Kops, G. J., N. D. de Ruiter, A. M. De Vries-Smits, D. R. Powell, J. L. Bos & B. M. Burgering: Direct control of the Forkhead transcription factor AFX by protein kinase B. *Nature*, 398, 630-4 (1999)
105. Romashkova, J. A. & S. S. Makarov: NF-kappaB is a target of AKT in anti-apoptotic PDGF signalling. *Nature*, 401, 86-90 (1999)
106. Ozes, O. N., L. D. Mayo, J. A. Gustin, S. R. Pfeffer, L. M. Pfeffer & D. B. Donner: NF-kappaB activation by tumour necrosis factor requires the Akt serine-threonine kinase. *Nature*, 401, 82-5 (1999)
107. Sah, J. F., S. Balasubramanian, R. L. Eckert & E. A. Rorke: Epigallocatechin-3-gallate inhibits epidermal growth factor receptor signaling pathway. Evidence for direct inhibition of ERK1/2 and AKT kinases. *J Biol Chem*, 279, 12755-62 (2004)
108. Bigelow, R. L. & J. A. Cardelli: The green tea catechins, (-)-Epigallocatechin-3-gallate (EGCG) and (-)-Epicatechin-3-gallate (ECG), inhibit HGF/Met signaling in immortalized and tumorigenic breast epithelial cells. *Oncogene*, 25, 1922-30 (2006)
109. Siddiqui, I. A., V. M. Adhami, F. Afaq, N. Ahmad & H. Mukhtar: Modulation of phosphatidylinositol-3-kinase/protein kinase B- and mitogen-activated protein kinase-pathways by tea polyphenols in human prostate cancer cells. *J Cell Biochem*, 91, 232-42 (2004)
110. Zhang, Q., X. Tang, Q. Lu, Z. Zhang, J. Rao & A. D. Le: Green tea extract and (-)-epigallocatechin-3-gallate inhibit hypoxia- and serum-induced HIF-1alpha protein accumulation and VEGF expression in human cervical carcinoma and hepatoma cells. *Mol Cancer Ther*, 5, 1227-38 (2006)
111. Masamune, A., K. Kikuta, M. Satoh, N. Suzuki & T. Shimosegawa: Green tea polyphenol epigallocatechin-3-gallate blocks PDGF-induced proliferation and migration of rat pancreatic stellate cells. *World J Gastroenterol*, 11, 3368-74 (2005)
112. Masuda, M., M. Suzui, J. T. Lim, A. Deguchi, J. W. Soh & I. B. Weinstein: Epigallocatechin-3-gallate decreases VEGF production in head and neck and breast carcinoma cells by inhibiting EGFR-related pathways of signal transduction. *J Exp Ther Oncol*, 2, 350-9 (2002)
113. Cuendet, M. & J. M. Pezzuto: The role of cyclooxygenase and lipoxygenase in cancer chemoprevention. *Drug Metabol Drug Interact*, 17, 109-57 (2000)
114. Hussain, T., S. Gupta, V. M. Adhami & H. Mukhtar: Green tea constituent epigallocatechin-3-gallate selectively inhibits COX-2 without affecting COX-1 expression in human prostate carcinoma cells. *Int J Cancer*, 113, 660-9 (2005)
115. Jeong, Y. I., I. D. Jung, J. S. Lee, C. M. Lee, J. D. Lee & Y. M. Park: (-)-Epigallocatechin gallate suppresses indoleamine 2,3-dioxygenase expression in murine dendritic cells: Evidences for the COX-2 and STAT1 as potential targets. *Biochem Biophys Res Commun*, 354, 1004-9 (2007)
116. Peng, G., D. A. Dixon, S. J. Muga, T. J. Smith & M. J. Wargovich: Green tea polyphenol (-)-epigallocatechin-3-gallate inhibits cyclooxygenase-2 expression in colon carcinogenesis. *Mol Carcinog*, 45, 309-19 (2006)
117. Hwang, J. T., J. Ha, I. J. Park, S. K. Lee, H. W. Baik, Y. M. Kim & O. J. Park: Apoptotic effect of EGCG in HT-29 colon cancer cells via AMPK signal pathway. *Cancer Lett*, 247, 115-21 (2007)

118. Suganuma, M., M. Kurusu, K. Suzuki, E. Tasaki & H. Fujiki: Green tea polyphenol stimulates cancer preventive effects of celecoxib in human lung cancer cells by upregulation of GADD153 gene. *Int J Cancer*, 119, 33-40 (2006)
119. Ullrich, A., L. Coussens, J. S. Hayflick, T. J. Dull, A. Gray, A. W. Tam, J. Lee, Y. Yarden, T. A. Libermann, J. Schlessinger & *et al.*: Human epidermal growth factor receptor cDNA sequence and aberrant expression of the amplified gene in A431 epidermoid carcinoma cells. *Nature*, 309, 418-25 (1984)
120. Saloman, D. S., C. Bianco, A. D. Ebert, N. I. Khan, M. De Santis, N. Normanno, C. Wechselberger, M. Seno, K. Williams, M. Sanicola, S. Foley, W. J. Gullick & G. Persico: The EGF-CFC family: novel epidermal growth factor-related proteins in development and cancer. *Endocr Relat Cancer*, 7, 199-226 (2000)
121. Wells, A.: EGF receptor. *Int J Biochem Cell Biol*, 31, 637-43 (1999)
122. Ullrich, A. & J. Schlessinger: Signal transduction by receptors with tyrosine kinase activity. *Cell*, 61, 203-12 (1990)
123. Shimizu, M., A. Deguchi, J. T. Lim, H. Moriwaki, L. Kopelovich & I. B. Weinstein: (-)-Epigallocatechin gallate and polyphenon E inhibit growth and activation of the epidermal growth factor receptor and human epidermal growth factor receptor-2 signaling pathways in human colon cancer cells. *Clin Cancer Res*, 11, 2735-46 (2005)
124. Masuda, M., M. Suzui & I. B. Weinstein: Effects of epigallocatechin-3-gallate on growth, epidermal growth factor receptor signaling pathways, gene expression, and chemosensitivity in human head and neck squamous cell carcinoma cell lines. *Clin Cancer Res*, 7, 4220-9 (2001)
125. Ichimatsu, D., M. Nomura, S. Nakamura, S. Moritani, K. Yokogawa, S. Kobayashi, T. Nishioka & K. I. Miyamoto: Structure-activity relationship of flavonoids for inhibition of epidermal growth factor-induced transformation of JB6 Cl 41 cells. *Mol Carcinog* (2007)
126. Pollak, M. N., E. S. Schernhammer & S. E. Hankinson: Insulin-like growth factors and neoplasia. *Nat Rev Cancer*, 4, 505-18 (2004)
127. Leng, S. L., K. S. Leeding, P. R. Gibson & L. A. Bach: Insulin-like growth factor-II renders LIM 2405 human colon cancer cells resistant to butyrate-induced apoptosis: a potential mechanism for colon cancer cell survival *in vivo*. *Carcinogenesis*, 22, 1625-31 (2001)
128. Tanno, S., S. Tanno, Y. Mitsuuchi, D. A. Altomare, G. H. Xiao & J. R. Testa: AKT activation up-regulates insulin-like growth factor I receptor expression and promotes invasiveness of human pancreatic cancer cells. *Cancer Res*, 61, 589-93 (2001)
129. Wu, J. D., A. Odman, L. M. Higgins, K. Haugk, R. Vessella, D. L. Ludwig & S. R. Plymate: *In vivo* effects of the human type I insulin-like growth factor receptor antibody A12 on androgen-dependent and androgen-independent xenograft human prostate tumors. *Clin Cancer Res*, 11, 3065-74 (2005)
130. Min, Y., Y. Adachi, H. Yamamoto, A. Insumran, Y. Arimura, T. Endo, Y. Hinoda, C. T. Lee, S. Nadaf, D. P. Carbone & K. Imai: Insulin-like growth factor I receptor blockade enhances chemotherapy and radiation responses and inhibits tumour growth in human gastric cancer xenografts. *Gut*, 54, 591-600 (2005)
131. Bahr, C. & B. Groner: The insulin like growth factor-1 receptor (IGF-1R) as a drug target: novel approaches to cancer therapy. *Growth Horm IGF Res*, 14, 287-95 (2004)
132. Hernandez-Sanchez, C., V. Blakesley, T. Kalebic, L. Helman & D. LeRoith: The role of the tyrosine kinase domain of the insulin-like growth factor-I receptor in intracellular signaling, cellular proliferation, and tumorigenesis. *J Biol Chem*, 270, 29176-81 (1995)
133. Larsson, O., A. Girnita & L. Girnita: Role of insulin-like growth factor 1 receptor signalling in cancer. *Br J Cancer*, 92, 2097-101 (2005)
134. Li, M., Z. He, S. Ermakova, D. Zheng, F. Tang, Y. Y. Cho, F. Zhu, W. Y. Ma, Y. Sham, E. A. Rogozin, A. M. Bode, Y. Cao & Z. Dong: Direct inhibition of insulin-like growth factor-I receptor kinase activity by (-)-epigallocatechin-3-gallate regulates cell transformation. *Cancer Epidemiol Biomarkers Prev*, 16, 598-605 (2007)
135. Adhami, V. M., I. A. Siddiqui, N. Ahmad, S. Gupta & H. Mukhtar: Oral consumption of green tea polyphenols inhibits insulin-like growth factor-I-induced signaling in an autochthonous mouse model of prostate cancer. *Cancer Res*, 64, 8715-22 (2004)
136. Yokoyama, S., H. Hirano, N. Wakimaru, K. P. Sarker & J. Kuratsu: Inhibitory effect of epigallocatechin-gallate on brain tumor cell lines *in vitro*. *Neuro-oncol*, 3, 22-8 (2001)
137. Lin, J. K., Y. C. Liang & S. Y. Lin-Shiau: Cancer chemoprevention by tea polyphenols through mitotic signal transduction blockade. *Biochem Pharmacol*, 58, 911-5 (1999)
138. Ahn, S. C., G. Y. Kim, J. H. Kim, S. W. Baik, M. K. Han, H. J. Lee, D. O. Moon, C. M. Lee, J. H. Kang, B. H. Kim, Y. H. Oh & Y. M. Park: Epigallocatechin-3-gallate, constituent of green tea, suppresses the LPS-induced phenotypic and functional maturation of murine dendritic cells through inhibition of mitogen-activated protein kinases and NF-kappaB. *Biochem Biophys Res Commun*, 313, 148-55 (2004)
139. Yan, Z., T. Yong-Guang, L. Fei-Jun, T. Fa-Qing, T. Min & C. Ya: Interference effect of epigallocatechin-3-gallate on targets of nuclear factor kappaB signal transduction pathways activated by EB virus encoded latent membrane protein 1. *Int J Biochem Cell Biol*, 36, 1473-81 (2004)
140. Gupta, S., K. Hastak, F. Afaq, N. Ahmad & H. Mukhtar: Essential role of caspases in epigallocatechin-3-gallate-mediated inhibition of nuclear factor kappa B and induction of apoptosis. *Oncogene*, 23, 2507-22 (2004)
141. Ahmad, N., S. Gupta & H. Mukhtar: Green tea polyphenol epigallocatechin-3-gallate differentially modulates nuclear factor kappaB in cancer cells versus normal cells. *Arch Biochem Biophys*, 376, 338-46 (2000)
142. Song, X. Z., Z. G. Bi & A. E. Xu: Green tea polyphenol epigallocatechin-3-gallate inhibits the expression of nitric oxide synthase and generation of nitric oxide induced by ultraviolet B in HaCaT cells. *Chin Med J (Engl)*, 119, 282-7 (2006)
143. Wheeler, D. S., J. D. Catravas, K. Odoms, A. Denenberg, V. Malhotra & H. R. Wong: Epigallocatechin-3-gallate, a green tea-derived polyphenol, inhibits IL-1 beta-dependent proinflammatory signal transduction in cultured respiratory epithelial cells. *J Nutr*, 134, 1039-44 (2004)

144. Afaq, F., V. M. Adhami, N. Ahmad & H. Mukhtar: Inhibition of ultraviolet B-mediated activation of nuclear factor kappaB in normal human epidermal keratinocytes by green tea Constituent (-)-epigallocatechin-3-gallate. *Oncogene*, 22, 1035-44 (2003)
145. Syed, D. N., F. Afaq, M. H. Kweon, N. Hadi, N. Bhatia, V. S. Spiegelman & H. Mukhtar: Green tea polyphenol EGCG suppresses cigarette smoke condensate-induced NF-kappaB activation in normal human bronchial epithelial cells. *Oncogene*, 26, 673-82 (2007)
146. Yu, C. L., D. J. Meyer, G. S. Campbell, A. C. Lerner, C. Carter-Su, J. Schwartz & R. Jove: Enhanced DNA-binding activity of a Stat3-related protein in cells transformed by the Src oncoprotein. *Science*, 269, 81-3 (1995)
147. Bromberg, J. F., C. M. Horvath, D. Besser, W. W. Lathem & J. E. Darnell, Jr.: Stat3 activation is required for cellular transformation by v-src. *Mol Cell Biol*, 18, 2553-8 (1998)
148. Grandis, J. R., S. D. Drenning, A. Chakraborty, M. Y. Zhou, Q. Zeng, A. S. Pitt & D. J. Tweardy: Requirement of Stat3 but not Stat1 activation for epidermal growth factor receptor-mediated cell growth *In vitro*. *J Clin Invest*, 102, 1385-92 (1998)
149. Carlesso, N., D. A. Frank & J. D. Griffin: Tyrosyl phosphorylation and DNA binding activity of signal transducers and activators of transcription (STAT) proteins in hematopoietic cell lines transformed by Bcr/Abl. *J Exp Med*, 183, 811-20 (1996)
150. Weber-Nordt, R. M., C. Egen, J. Wehinger, W. Ludwig, V. Gouilleux-Gruart, R. Mertelsmann & J. Finke: Constitutive activation of STAT proteins in primary lymphoid and myeloid leukemia cells and in Epstein-Barr virus (EBV)-related lymphoma cell lines. *Blood*, 88, 809-16 (1996)
151. Bharti, A. C., N. Donato & B. B. Aggarwal: Curcumin (diferuloylmethane) inhibits constitutive and IL-6-inducible STAT3 phosphorylation in human multiple myeloma cells. *J Immunol*, 171, 3863-71 (2003)
152. Stephanou, A.: Role of STAT-1 and STAT-3 in ischaemia/reperfusion injury. *J Cell Mol Med*, 8, 519-25 (2004)
153. Townsend, P. A., T. M. Scarabelli, E. Pasini, G. Gitti, M. Menegazzi, H. Suzuki, R. A. Knight, D. S. Latchman & A. Stephanou: Epigallocatechin-3-gallate inhibits STAT-1 activation and protects cardiac myocytes from ischemia/reperfusion-induced apoptosis. *Faseb J*, 18, 1621-3 (2004)
154. Karin, M., Z. Liu & E. Zandi: AP-1 function and regulation. *Curr Opin Cell Biol*, 9, 240-6 (1997)
155. Dong, Z., M. J. Birrer, R. G. Watts, L. M. Matrisian & N. H. Colburn: Blocking of tumor promoter-induced AP-1 activity inhibits induced transformation in JB6 mouse epidermal cells. *Proc Natl Acad Sci U S A*, 91, 609-13 (1994)
156. Huang, C., W. Y. Ma, M. R. Young, N. Colburn & Z. Dong: Shortage of mitogen-activated protein kinase is responsible for resistance to AP-1 transactivation and transformation in mouse JB6 cells. *Proc Natl Acad Sci U S A*, 95, 156-61 (1998)
157. Dumont, J. A., A. J. Bitonti, C. D. Wallace, R. J. Baumann, E. A. Cashman & D. E. Cross-Doersen: Progression of MCF-7 breast cancer cells to antiestrogen-resistant phenotype is accompanied by elevated levels of AP-1 DNA-binding activity. *Cell Growth Differ*, 7, 351-9 (1996)
158. Saez, E., S. E. Rutberg, E. Mueller, H. Oppenheim, J. Smoluk, S. H. Yuspa & B. M. Spiegelman: c-fos is required for malignant progression of skin tumors. *Cell*, 82, 721-32 (1995)
159. Crawford, H. C. & L. M. Matrisian: Mechanisms controlling the transcription of matrix metalloproteinase genes in normal and neoplastic cells. *Enzyme Protein*, 49, 20-37 (1996)
160. Wright, J. H., S. McDonnell, G. Portella, G. T. Bowden, A. Balmain & L. M. Matrisian: A switch from stromal to tumor cell expression of stromelysin-1 mRNA associated with the conversion of squamous to spindle carcinomas during mouse skin tumor progression. *Mol Carcinog*, 10, 207-15 (1994)
161. Dong, Z., W. Ma, C. Huang & C. S. Yang: Inhibition of tumor promoter-induced activator protein 1 activation and cell transformation by tea polyphenols, (-)-epigallocatechin gallate, and theaflavins. *Cancer Res*, 57, 4414-9 (1997)
162. Eckert, R. L., J. F. Crish, T. Efimova & S. Balasubramanian: Opposing action of curcumin and green tea polyphenol in human keratinocytes. *Mol Nutr Food Res*, 50, 123-9 (2006)
163. Balasubramanian, S., T. Efimova & R. L. Eckert: Green tea polyphenol stimulates a Ras, MEKK1, MEK3, and p38 cascade to increase activator protein 1 factor-dependent involucrin gene expression in normal human keratinocytes. *J Biol Chem*, 277, 1828-36 (2002)
164. Lee, J. S. & Y. J. Surh: Nrf2 as a novel molecular target for chemoprevention. *Cancer Lett*, 224, 171-84 (2005)
165. Kobayashi, A., M. I. Kang, Y. Watai, K. I. Tong, T. Shibata, K. Uchida & M. Yamamoto: Oxidative and electrophilic stresses activate Nrf2 through inhibition of ubiquitination activity of Keap1. *Mol Cell Biol*, 26, 221-9 (2006)
166. Martin, D., A. I. Rojo, M. Salinas, R. Diaz, G. Gallardo, J. Alam, C. M. De Galarreta & A. Cuadrado: Regulation of heme oxygenase-1 expression through the phosphatidylinositol 3-kinase/Akt pathway and the Nrf2 transcription factor in response to the antioxidant phytochemical carnosol. *J Biol Chem*, 279, 8919-29 (2004)
167. Kweon, M. H., V. M. Adhami, J. S. Lee & H. Mukhtar: Constitutive overexpression of Nrf2-dependent heme oxygenase-1 in A549 cells contributes to resistance to apoptosis induced by epigallocatechin 3-gallate. *J Biol Chem*, 281, 33761-72 (2006)
168. Cho, H. Y., S. P. Reddy, M. Yamamoto & S. R. Kleberger: The transcription factor NRF2 protects against pulmonary fibrosis. *Faseb J*, 18, 1258-60 (2004)
169. Ahmad, N., D. K. Feyes, A. L. Nieminen, R. Agarwal & H. Mukhtar: Green tea constituent epigallocatechin-3-gallate and induction of apoptosis and cell cycle arrest in human carcinoma cells. *J Natl Cancer Inst*, 89, 1881-6 (1997)
170. Yang, C. S., P. Maliakal & X. Meng: Inhibition of carcinogenesis by tea. *Annu Rev Pharmacol Toxicol*, 42, 25-54 (2002)



171. Gupta, S., T. Hussain & H. Mukhtar: Molecular pathway for (-)-epigallocatechin-3-gallate-induced cell cycle arrest and apoptosis of human prostate carcinoma cells. *Arch Biochem Biophys*, 410, 177-85 (2003)
172. Xu, F. & Y. S. Zhen: (-)-Epigallocatechin-3-gallate enhances anti-tumor effect of cytosine arabinoside on HL-60 cells. *Acta Pharmacol Sin*, 24, 163-8 (2003)
173. Liang, Y. C., S. Y. Lin-Shiau, C. F. Chen & J. K. Lin: Inhibition of cyclin-dependent kinases 2 and 4 activities as well as induction of Cdk inhibitors p21 and p27 during growth arrest of human breast carcinoma cells by (-)-epigallocatechin-3-gallate. *J Cell Biochem*, 75, 1-12 (1999)
174. Hofmann, C. S. & G. E. Sonenshein: Green tea polyphenol epigallocatechin-3 gallate induces apoptosis of proliferating vascular smooth muscle cells via activation of p53. *Faseb J*, 17, 702-4 (2003)
175. Roy, A. M., M. S. Baliga & S. K. Katiyar: Epigallocatechin-3-gallate induces apoptosis in estrogen receptor-negative human breast carcinoma cells via modulation in protein expression of p53 and Bax and caspase-3 activation. *Mol Cancer Ther*, 4, 81-90 (2005)
176. Sen, P., P. K. Chakraborty & S. Raha: Tea polyphenol epigallocatechin 3-gallate impedes the anti-apoptotic effects of low-grade repetitive stress through inhibition of Akt and NFkappaB survival pathways. *FEBS Lett*, 580, 278-84 (2006)
177. Kuhn, D., W. H. Lam, A. Kazi, K. G. Daniel, S. Song, L. M. Chow, T. H. Chan & Q. P. Dou: Synthetic peracetate tea polyphenols as potent proteasome inhibitors and apoptosis inducers in human cancer cells. *Front Biosci*, 10, 1010-23 (2005)
178. Gupta, S., N. Ahmad, A. L. Nieminen & H. Mukhtar: Growth inhibition, cell-cycle dysregulation, and induction of apoptosis by green tea constituent (-)-epigallocatechin-3-gallate in androgen-sensitive and androgen-insensitive human prostate carcinoma cells. *Toxicol Appl Pharmacol*, 164, 82-90 (2000)
179. Tam, N. N., A. Nyska, R. R. Maronpot, G. Kissling, L. Lomnitski, A. Suttie, S. Bakshi, M. Bergman, S. Grossman & S. M. Ho: Differential attenuation of oxidative/nitrosative injuries in early prostatic neoplastic lesions in TRAMP mice by dietary antioxidants. *Prostate*, 66, 57-69 (2006)
180. Pezzato, E., L. Sartor, I. Dell'Aica, R. Dittadi, M. Gion, C. Belluco, M. Lise & S. Garbisa: Prostate carcinoma and green tea: PSA-triggered basement membrane degradation and MMP-2 activation are inhibited by (-)-epigallocatechin-3-gallate. *Int J Cancer*, 112, 787-92 (2004)
181. Caporali, A., P. Davalli, S. Astancolle, D. D'Arca, M. Brausi, S. Bettuzzi & A. Corti: The chemopreventive action of catechins in the TRAMP mouse model of prostate carcinogenesis is accompanied by clusterin over-expression. *Carcinogenesis*, 25, 2217-24 (2004)
182. Yu, H. N., J. J. Yin & S. R. Shen: Growth inhibition of prostate cancer cells by epigallocatechin gallate in the presence of Cu2+. *J Agric Food Chem*, 52, 462-6 (2004)
183. Yu, H. N., S. R. Shen & J. J. Yin: Effects of interactions of EGCG and Cd (2+) on the growth of PC-3 cells and their mechanisms. *Food Chem Toxicol*, 45, 244-9 (2007)
184. Stuart, E. C., M. J. Scandlyn & R. J. Rosengren: Role of epigallocatechin gallate (EGCG) in the treatment of breast and prostate cancer. *Life Sci*, 79, 2329-36 (2006)
185. Henning, S. M., W. Aronson, Y. Niu, F. Conde, N. H. Lee, N. P. Seeram, R. P. Lee, J. Lu, D. M. Harris, A. Moro, J. Hong, L. Pak-Shan, R. J. Barnard, H. G. Ziaee, G. Csathy, V. L. Go, H. Wang & D. Heber: Tea polyphenols and theaflavins are present in prostate tissue of humans and mice after green and black tea consumption. *J Nutr*, 136, 1839-43 (2006)
186. Wang, S. I. & H. Mukhtar: Gene expression profile in human prostate LNCaP cancer cells by (-)-epigallocatechin-3-gallate. *Cancer Lett*, 182, 43-51 (2002)
187. Ren, F., S. Zhang, S. H. Mitchell, R. Butler & C. Y. Young: Tea polyphenols down-regulate the expression of the androgen receptor in LNCaP prostate cancer cells. *Oncogene*, 19, 1924-32 (2000)
188. Chung, F. L.: The prevention of lung cancer induced by a tobacco-specific carcinogen in rodents by green and black Tea. *Proc Soc Exp Biol Med*, 220, 244-8 (1999)
189. Sazuka, M., S. Murakami, M. Isemura, K. Satoh & T. Nukiwa: Inhibitory effects of green tea infusion on *in vitro* invasion and *in vivo* metastasis of mouse lung carcinoma cells. *Cancer Lett*, 98, 27-31 (1995)
190. Zhang, Z., Q. Liu, L. E. Lantry, Y. Wang, G. J. Kelloff, M. W. Anderson, R. W. Wiseman, R. A. Lubet & M. You: A germ-line p53 mutation accelerates pulmonary tumorigenesis: p53-independent efficacy of chemopreventive agents green tea or dexamethasone/myo-inositol and chemotherapeutic agents taxol or adriamycin. *Cancer Res*, 60, 901-7 (2000)
191. Hu, G., C. Han & J. Chen: Inhibition of oncogene expression by green tea and (-)-epigallocatechin gallate in mice. *Nutr Cancer*, 24, 203-9 (1995)
192. Mukhtar, H. & C. A. Elmets: Photocarcinogenesis: mechanisms, models and human health implications. *Photochem Photobiol*, 63, 356-7 (1996)
193. Katiyar, S. K., M. S. Matsui, C. A. Elmets & H. Mukhtar: Polyphenolic antioxidant (-)-epigallocatechin-3-gallate from green tea reduces UVB-induced inflammatory responses and infiltration of leukocytes in human skin. *Photochem Photobiol*, 69, 148-53 (1999)
194. Katiyar, S., C. A. Elmets & S. K. Katiyar: Green tea and skin cancer: photoimmunology, angiogenesis and DNA repair. *J Nutr Biochem* (2006)
195. Meeran, S. M., S. K. Mantena & S. K. Katiyar: Prevention of ultraviolet radiation-induced immunosuppression by (-)-epigallocatechin-3-gallate in mice is mediated through interleukin 12-dependent DNA repair. *Clin Cancer Res*, 12, 2272-80 (2006)
196. Warshaw, A. L. & C. Fernandez-del Castillo: Pancreatic carcinoma. *N Engl J Med*, 326, 455-65 (1992)
197. Magee, C. J., P. Ghaneh & J. P. Neoptolemos: Surgical and medical therapy for pancreatic carcinoma. *Best Pract Res Clin Gastroenterol*, 16, 435-55 (2002)
198. Yeo, T. P., R. H. Hruban, S. D. Leach, R. E. Wilentz, T. A. Sohn, S. E. Kern, C. A. Iacobuzio-Donahue, A. Maitra, M. Goggins, M. I. Canto, R. A. Abrams, D. Laheru, E. M. Jaffee, M. Hidalgo & C. J. Yeo: Pancreatic cancer. *Curr Probl Cancer*, 26, 176-275 (2002)
199. Hruban, R. H., N. V. Adsay, J. Albores-Saavedra, C. Compton, E. S. Garrett, S. N. Goodman, S. E. Kern, D. S. Klimstra, G. Kloppel, D. S. Longnecker, J. Luttges & G. J. Offerhaus: Pancreatic intraepithelial neoplasia: a new nomenclature and classification system for pancreatic duct lesions. *Am J Surg Pathol*, 25, 579-86 (2001)

200. Bardeesy, N. & R. A. DePinho: Pancreatic cancer biology and genetics. *Nat Rev Cancer*, 2, 897-909 (2002)
201. Hilgers, W. & S. E. Kern: Molecular genetic basis of pancreatic adenocarcinoma. *Genes Chromosomes Cancer*, 26, 1-12 (1999)
202. Cubilla, A. L. & P. J. Fitzgerald: Morphological lesions associated with human primary invasive nonendocrine pancreas cancer. *Cancer Res*, 36, 2690-8 (1976)
203. Yoshida, T. & D. Hanahan: Murine pancreatic ductal adenocarcinoma produced by *in vitro* transduction of polyoma middle T oncogene into the islets of Langerhans. *Am J Pathol*, 145, 671-84 (1994)
204. Pour, P. M., K. K. Pandey & S. K. Batra: What is the origin of pancreatic adenocarcinoma? *Mol Cancer*, 2, 13 (2003)
205. Meszoely, I. M., A. L. Means, C. R. Scoggins & S. D. Leach: Developmental aspects of early pancreatic cancer. *Cancer J*, 7, 242-50 (2001)
206. Wagner, M., F. R. Greten, C. K. Weber, S. Koschnick, T. Mattfeldt, W. Deppert, H. Kern, G. Adler & R. M. Schmid: A murine tumor progression model for pancreatic cancer recapitulating the genetic alterations of the human disease. *Genes Dev*, 15, 286-93 (2001)
207. Bockman, D. E., J. Guo, P. Buchler, M. W. Muller, F. Bergmann & H. Friess: Origin and development of the precursor lesions in experimental pancreatic cancer in rats. *Lab Invest*, 83, 853-9 (2003)
208. Leach, S. D.: Mouse models of pancreatic cancer: the fur is finally flying! *Cancer Cell*, 5, 7-11 (2004)
209. Miyamoto, Y., A. Maitra, B. Ghosh, U. Zechner, P. Argani, C. A. Iacobuzio-Donahue, V. Sriuranpong, T. Iso, I. M. Meszoely, M. S. Wolfe, R. H. Hruban, D. W. Ball, R. M. Schmid & S. D. Leach: Notch mediates TGF alpha-induced changes in epithelial differentiation during pancreatic tumorigenesis. *Cancer Cell*, 3, 565-76 (2003)
210. Conney, A. H.: Enzyme induction and dietary chemicals as approaches to cancer chemoprevention: the Seventh DeWitt S. Goodman Lecture. *Cancer Res*, 63, 7005-31 (2003)
211. Mittal, A., M. S. Pate, R. C. Wylie, T. O. Tollefsbol & S. K. Katyar: EGCG down-regulates telomerase in human breast carcinoma MCF-7 cells, leading to suppression of cell viability and induction of apoptosis. *Int J Oncol*, 24, 703-10 (2004)
212. Chen, Z. P., J. B. Schell, C. T. Ho & K. Y. Chen: Green tea epigallocatechin gallate shows a pronounced growth inhibitory effect on cancerous cells but not on their normal counterparts. *Cancer Lett*, 129, 173-9 (1998)
213. Kavanagh, K. T., L. J. Hafer, D. W. Kim, K. K. Mann, D. H. Sherr, A. E. Rogers & G. E. Sonenshein: Green tea extracts decrease carcinogen-induced mammary tumor burden in rats and rate of breast cancer cell proliferation in culture. *J Cell Biochem*, 82, 387-98 (2001)
214. Thangapazham, R. L., A. K. Singh, A. Sharma, J. Warren, J. P. Gaddipati & R. K. Maheshwari: Green tea polyphenols and its constituent epigallocatechin gallate inhibits proliferation of human breast cancer cells *in vitro* and *in vivo*. *Cancer Lett*, 245, 232-41 (2007)
215. Zhao, X., H. Tian, X. Ma & L. Li: Epigallocatechin gallate, the main ingredient of green tea induces apoptosis in breast cancer cells. *Front Biosci*, 11, 2428-33 (2006)
216. Guo, S., J. Lu, A. Subramanian & G. E. Sonenshein: Microarray-assisted pathway analysis identifies mitogen-activated protein kinase signaling as a mediator of resistance to the green tea polyphenol epigallocatechin 3-gallate in her-2/neu-overexpressing breast cancer cells. *Cancer Res*, 66, 5322-9 (2006)
217. Kim, J., X. Zhang, K. M. Rieger-Christ, I. C. Summerhayes, D. E. Wazer, K. E. Paulson & A. S. Yee: Suppression of Wnt signaling by the green tea compound (-)-epigallocatechin 3-gallate (EGCG) in invasive breast cancer cells. Requirement of the transcriptional repressor HBP1. *J Biol Chem*, 281, 10865-75 (2006)
218. Lin, J. K. & S. Y. Lin-Shiau: Mechanisms of hypolipidemic and anti-obesity effects of tea and tea polyphenols. *Mol Nutr Food Res*, 50, 211-7 (2006)
219. Huh, S. W., S. M. Bae, Y. W. Kim, J. M. Lee, S. E. Namkoong, I. P. Lee, S. H. Kim, C. K. Kim & W. S. Ahn: Anticancer effects of (-)-epigallocatechin-3-gallate on ovarian carcinoma cell lines. *Gynecol Oncol*, 94, 760-8 (2004)
220. Spinella, F., L. Rosano, S. Decandia, V. Di Castro, A. Albini, G. Elia, P. G. Natali & A. Bagnato: Antitumor effect of green tea polyphenol epigallocatechin-3-gallate in ovarian carcinoma cells: evidence for the endothelin-1 as a potential target. *Exp Biol Med (Maywood)*, 231, 1123-7 (2006)
221. Chan, M. M., K. J. Soprano, K. Weinstein & D. Fong: Epigallocatechin-3-gallate delivers hydrogen peroxide to induce death of ovarian cancer cells and enhances their cisplatin susceptibility. *J Cell Physiol*, 207, 389-96 (2006)
222. Kalluri, R. & V. P. Sukhatme: Fibrosis and angiogenesis. *Curr Opin Nephrol Hypertens*, 9, 413-8 (2000)
223. Folkman, J.: Angiogenesis and proteins of the hemostatic system. *J Thromb Haemost*, 1, 1681-2 (2003)
224. Lee, Y. K., N. D. Bone, A. K. Strega, T. D. Shanafelt, D. F. Jelinek & N. E. Kay: VEGF receptor phosphorylation status and apoptosis is modulated by a green tea component, epigallocatechin-3-gallate (EGCG), in B-cell chronic lymphocytic leukemia. *Blood*, 104, 788-94 (2004)
225. Jung, Y. D., M. S. Kim, B. A. Shin, K. O. Chay, B. W. Ahn, W. Liu, C. D. Bucana, G. E. Gallick & L. M. Ellis: EGCG, a major component of green tea, inhibits tumour growth by inhibiting VEGF induction in human colon carcinoma cells. *Br J Cancer*, 84, 844-50 (2001)
226. Fassina, G., R. Vene, M. Morini, S. Minghelli, R. Benelli, D. M. Noonan & A. Albini: Mechanisms of inhibition of tumor angiogenesis and vascular tumor growth by epigallocatechin-3-gallate. *Clin Cancer Res*, 10, 4865-73 (2004)
227. Neuhaus, T., S. Pabst, S. Stier, A. A. Weber, K. Schror, A. Sachinidis, H. Vetter & Y. D. Ko: Inhibition of the vascular-endothelial growth factor-induced intracellular signaling and mitogenesis of human endothelial cells by epigallocatechin-3 gallate. *Eur J Pharmacol*, 483, 223-7 (2004)
228. Rodriguez, S. K., W. Guo, L. Liu, M. A. Band, E. K. Paulson & M. Meydani: Green tea catechin, epigallocatechin-3-gallate, inhibits vascular endothelial growth factor angiogenic signaling by disrupting the formation of a receptor complex. *Int J Cancer*, 118, 1635-44 (2006)

229. Park, J. S., M. H. Kim, H. J. Chang, K. M. Kim, S. M. Kim, B. A. Shin, B. W. Ahn & Y. D. Jung: Epigallocatechin-3-gallate inhibits the PDGF-induced VEGF expression in human vascular smooth muscle cells via blocking PDGF receptor and Erk-1/2. *Int J Oncol*, 29, 1247-52 (2006)
230. Annabi, B., Y. T. Lee, C. Martel, A. Pilorget, J. P. Bahary & R. Beliveau: Radiation induced-tubulogenesis in endothelial cells is antagonized by the antiangiogenic properties of green tea polyphenol (-) epigallocatechin-3-gallate. *Cancer Biol Ther*, 2, 642-9 (2003)
231. Changotade, S. I., A. Assoumou, F. Gueniche, F. Fioretti, S. Segulier, Y. de Prost, C. Bodemer, G. Godeau & K. Senni: Epigallocatechin Gallate's Protective Effect against MMP7 in Recessive Dystrophic Epidermolysis Bullosa Patients. *J Invest Dermatol*, 127, 821-8 (2007)
232. Annabi, B., J. C. Currie, A. Moghrabi & R. Beliveau: Inhibition of HuR and MMP-9 expression in macrophage-differentiated HL-60 myeloid leukemia cells by green tea polyphenol EGCG. *Leuk Res* (2006)
233. Ngameni, B., M. Touaibia, R. Patnam, A. Belkaid, P. Sonna, B. T. Ngadjui, B. Annabi & R. Roy: Inhibition of MMP-2 secretion from brain tumor cells suggests chemopreventive properties of a furanocoumarin glycoside and of chalcones isolated from the twigs of *Dorstenia turbinata*. *Phytochemistry*, 67, 2573-9 (2006)
234. Roomi, M. W., V. Ivanov, T. Kalinovsky, A. Niedzwiecki & M. Rath: Modulation of human renal cell carcinoma 786-0 MMP-2 and MMP-9 activity by inhibitors and inducers *in vitro*. *Med Oncol*, 23, 245-50 (2006)
235. Kim, S. H., H. J. Park, C. M. Lee, I. W. Choi, D. O. Moon, H. J. Roh, H. K. Lee & Y. M. Park: Epigallocatechin-3-gallate protects toluene diisocyanate-induced airway inflammation in a murine model of asthma. *FEBS Lett*, 580, 1883-90 (2006)
236. Zhen, M. C., X. H. Huang, Q. Wang, K. Sun, Y. J. Liu, W. Li, L. J. Zhang, L. Q. Cao & X. L. Chen: Green tea polyphenol epigallocatechin-3-gallate suppresses rat hepatic stellate cell invasion by inhibition of MMP-2 expression and its activation. *Acta Pharmacol Sin*, 27, 1600-7 (2006)
237. Tang, F. Y., E. P. Chiang & C. J. Shih: Green tea catechin inhibits ephrin-A1-mediated cell migration and angiogenesis of human umbilical vein endothelial cells. *J Nutr Biochem* (2006)
238. Matsubara, K., A. Saito, A. Tanaka, N. Nakajima, R. Akagi, M. Mori & Y. Mizushima: Catechin conjugated with fatty acid inhibits DNA polymerase and angiogenesis. *DNA Cell Biol*, 25, 95-103 (2006)

**Key Words:** Chemoprevention, Green Tea, EGCG, Polyphenols, Cancer, Apoptosis, Review

**Send correspondence to:** Dr Rakesh K. Srivastava, Department of Biochemistry, The University of Texas Health Science Center at Tyler, 11937 US Highway 271, Tyler, Texas 75708-3154, Tel: 903-877-7559, Fax: 903-877-5320, E-mail: rakesh.srivastava@uthct.edu

<http://www.bioscience.org/current/vol12.htm>