

Heart failure and cachexia: insights offered from molecular biology

Viviane MA Conraads, Vicky Y Hoymans, Christiaan J Vrints

Department of Cardiology, Centre for Cell Therapy and Regenerative Medicine, University Hospital Antwerp, Belgium

TABLE OF CONTENTS

1. Abstract
2. Chronic heart failure: beyond the limits of the heart
3. Cardiac cachexia: is it relevant?
4. Morphologic, metabolic and functional changes in skeletal muscle
5. How do CHF patients lose skeletal muscle mass?
 - 5.1. Substrate utilization
 - 5.2. Processes involved in the loss of skeletal muscle protein
6. Disruption of the neurohormonal balance: mediators of skeletal muscle wasting
 - 6.1. "Classical" neuroendocrine pathways: catecholamines and the RAAS
 - 6.2. Anabolic/catabolic imbalance
 - 6.3. Inflammation and oxidative stress
 - 6.4. Leptin
 - 6.5. Ghrelin
7. Heart failure related myopathy; where does exercise training come in?
8. Conclusions
9. References

1. ABSTRACT

Chronic heart failure (CHF) is an enormous medical and communal burden. The syndrome is common, carries a grim prognosis and severely impacts quality of life. Those patients who develop cardiac cachexia combat both important disability and a poor outlook. Muscle wasting is a critical component of cachexia. The pathophysiological determinants are numerous and some of them are common to other chronic severe illnesses. There is increasing awareness, however, that heart failure related myopathy is a distinct entity, characterized by specific functional, structural and morphologic changes and the involvement of several neurohormonal pathways, catabolic processes, a pro-inflammatory environment and increased oxidative stress. Although clear-cut evidence based solutions for the problem are not readily available, the modulating effects of regular exercise in CHF patients suggest that physical training should at least be incorporated in the essentially multi-disciplinary approach.

2. CHRONIC HEART FAILURE: BEYOND THE LIMITS OF THE HEART

Pathophysiological insights into the syndrome of chronic heart failure (CHF) have witnessed a paradigm shift. Until two decades ago, central haemodynamic disturbances seemed to perfectly fit the cardinal symptoms of CHF, with *fatigue* being the clinical translation of "forward failure" and *dyspnoea* reflecting the consequences of "backward failure". However, several observations have undermined the haemodynamic model of CHF. Left ventricular dysfunction, for instance, does not necessarily concur with compromised exercise capacity in patients with CHF (1). In addition, restoration of haemodynamics to near normal with inotropic drugs or after cardiac transplantation does not acutely improve physical capacity (2). Two important developments have dramatically changed heart failure research. First, there is the recognition that neurohormonal adaptation, providing initial haemostasis, eventually gives rise to a vicious circle, which encompasses

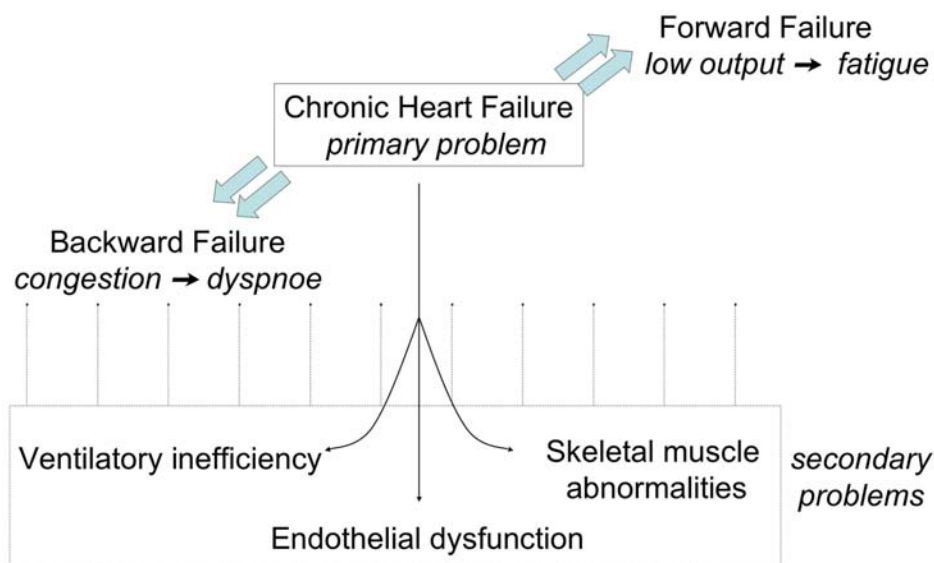


Figure 1. Chronic heart failure: a systemic disease. Primary cardiac dysfunction will lead to both symptoms of backward (dyspnoea) and forward failure (fatigue). With time, however, secondary maladaptations occur. Peripheral endothelial dysfunction, ventilatory inefficiency and skeletal muscle abnormalities have major impact on the symptoms and progression of the disease.

further deterioration of cardiac function and ventricular remodelling. Overstimulation of the sympathetic nervous system and the renin-angiotensin-aldosterone pathway (RAAS), a low-grade inflammatory state and a profound anabolic-catabolic dysbalance are central elements of the disturbed neurohormonal network. Second, the concept of CHF as a mere cardiologic entity has been modified. The syndrome is now considered to be a multi-faceted systemic disease. CHF comprises an inciting cardiac event, which thereafter triggers the onset of several peripheral maladaptive processes (3). Three principal players in this field account for a significant part of heart failure symptoms. First, both at rest and during exercise, peripheral vasomotor tone is increased and vasodilatory responses are reduced. Mediated by the above-described neurohormonal disturbances, oxidative stress and deficient NO-mediated endothelial function play an essential role in this process. Secondly, excessive ventilation, particularly during exercise, causes a shift in the slope of the linear relationship between minute ventilation and CO₂ production. This ventilatory inefficiency is thought to origin from ventilation/perfusion defects and more importantly, from an overactive muscle ergoreflex system. Lastly, and this is the main focus of this review, muscular alterations covering the entire range of macroscopic (i.e. loss of muscle bulk) (4) to histological (5) and ultrastructural changes (6, 7), as well as functional abnormalities related to fibre shift (from slow oxidative type I to glycolytic type II fibres) (5-8) have been implicated. Figure 1 illustrates the intricate coupling of these different elements of the CHF-phenotype.

3. CARDIAC CACHEXIA: IS IT RELEVANT?

Within certain limits, a unifying pathogenesis, shared by several severe chronic diseases, ultimately leads

to a general wasting syndrome. Although starvation can be relevant in patients with CHF, cancer, chronic obstructive pulmonary disease and AIDS, the fact that tissue loss in these patients is not limited to the fat compartment, suggests that other processes are at play. The occurrence of cachexia, irrespective of the underlying disease, infers poor prognosis and physical limitation. Anker and colleagues were the first to suggest a simple, objective criterion for the definition of cardiac cachexia. In the absence of other cachectic disease states, clinical cardiac cachexia can be diagnosed when non-intentional weight loss of > 7.5% of the previous normal and “dry” weight occurs over a period of at least 6 months. In a group of 171 patients with CHF, these investigators demonstrated that the cachectic state is an independent risk factor for poor outcome with a mortality rate after 18 months of 50% compared to 17% in non-cachectic patients (9). Later on, studying 1929 patients enrolled in the SOLVD trial, the poor outlook for weight-losing CHF patients was confirmed. At 8 months follow-up, all pre-defined cut points (i.e. 5, 7.5, 10 and 15%) for weight loss were significantly associated with impaired survival after adjustment for demographic characteristics, functional class, cardiac function and treatment allocation. Weight loss of 6% or more at any time was the strongest predictor of impaired survival (10) and its prevalence attained 12-15% of CHF patients in NYHA functional class II-IV. When judging a patient’s condition, it is important to stress the fact that the *process* of weight loss cannot be interchanged with low body weight per se. Other confounding factors that complicate objective assessment result from fluctuating body weight related to congestion and diuretic treatment. Besides being a negative prognostic indicator, skeletal muscle wasting obviously affects global exercise performance and muscle strength, which are reflected in fatigue and low quality of life. Preservation of muscle mass is an independent determinant of peak oxygen

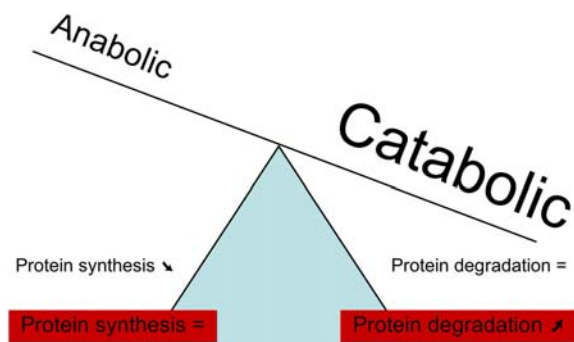


Figure 2. Cardiac cachexia: anabolic/catabolic imbalance. Despite the fact that both protein synthesis and degradation could be implied in muscular wasting, most of the evidence points into the direction of increased protein breakdown as a result of increased catabolism.

consumption (VO_2peak) in CHF patients (11). Harrington *et al.* (12) compared symptomatic with asymptomatic CHF patients (VO_2peak 14.6 ± 1.3 versus 27.1 ± 1.6 ml/kg/min), who were otherwise well matched for demographic characteristics and left ventricular function. Besides differences in peak leg blood flow, there was a significant reduction in cross sectional area (CSA) of the quadriceps muscle, which concurred with reduced muscle strength and increased fatigue in the symptomatic group. In addition, overactivation of the muscle ergoreflex, being a driving force for hyperventilation during exercise in CHF, appears strongly related to reduced muscle mass (13). Although the functional implications of skeletal muscle wasting catch the eye, tissue loss is not limited to the muscular compartment. Patients with cardiac cachexia suffer from a general loss of fat, lean, bone tissue and bone density (14).

4. MORPHOLOGIC, METABOLIC AND FUNCTIONAL CHANGES IN SKELETAL MUSCLE

Reduced quadriceps and total leg muscle CSA partly accounts for impaired maximal isometric strength and early fatigability in CHF patients. After correction for CSA, however, lower strength per unit muscle when compared to controls, implicates that muscle “quality” is also affected (12). Several histological and biochemical changes have been repeatedly reported in an attempt to explain the functional deficit of peripheral skeletal muscle in the setting of CHF. Increased collagen content (fibrosis) differentiated the skeletal musculature from cachectic versus non-cachectic CHF patients in a study by Filippatos *et al.* (15). The correlation between endothelial cells/fibre and VO_2peak in class II/III CHF patients suggests that reduced microvascular density of skeletal muscle might precede other muscle alterations (16). Ultrastructural abnormalities, such as lower mitochondrial volume (6), a decrease in the surface density of cytochrome c oxidase-positive mitochondria, of mitochondrial cristae and mitochondrial inner border membrane corroborate with the long known typical shift towards type IIx fibres (7). With the use of ^{31}P nuclear magnetic resonance spectroscopy, a rapid depletion of phosphocreatine and a lower rate of

resynthesis during exercise have been shown (17). Hambrecht and colleagues (18) demonstrated that reduced availability of mitochondrial creatine kinase (mi-CK), responsible for transfer of high-energy phosphates from the mitochondrion to myosin filaments, might account in part for compromised energetics. The simplified view of a muscle myopathy in CHF characterized by a shift from an oxidative to a more glycolytic phenotype is in contrast with the complex and incompletely understood alterations in skeletal muscle mitochondrial function (19). In a very elegant experiment, Mettauer *et al.* (20) compared oxidative capacity of skeletal muscle of CHF patients with sedentary and active controls. Despite lower VO_2peak , muscle oxidative capacity and regulation were comparable in the CHF and sedentary group, suggesting that disease-specific metabolic muscular changes in CHF patients take place upstream of mitochondria.

5. HOW DO CHF PATIENTS LOSE SKELETAL MUSCLE MASS?

5.1. Substrate utilization

Figure 2 illustrates a simplified view of what cardiac cachexia is all about; a profound imbalance between anabolic and catabolic factors, eventually culminating in loss of muscular mass. In theory, 2 possible scenarios exist. However, most of the gathered evidence to date, points towards the direction of protein degradation overpowering protein synthesis. The profound shift in substrate utilisation in CHF patients entails increased circulating levels and oxidation of free fatty acids (FFA), decreased skeletal muscle uptake and use of glucose and, importantly, increased protein turnover and breakdown (21). The interplay between the hyperadrenergic status, promoting lipolysis and thereby increasing FFA concentration, and the presence of insulin resistance (22), favours this less efficient metabolic divergence. Myofibrillar proteins are particularly important in the preservation of muscle mass and function. Myosin heavy chain (MHC) comprises about a quarter of cellular protein mass and is a key structural and functional component of myofibrils. Toth and colleagues (23) demonstrated that, in contrast to actin, MHC content in vastus lateralis biopsies from CHF patients was decreased in relation to disease severity. Since no changes in fractional MHC synthesis occurred, this finding suggests increased protein breakdown as a plausible mechanism. Differences in myofibrillar gene expression are suggested to explain the specific skeletal muscle myofibrillar protein phenotype (i.e.; shift from Type I to type Type IIx fibres) (24).

The relevance of nutrition adequacy, energy availability and basal metabolic rate is still debated. Poehlman *et al.* (25), comparing CHF patients (mainly NYHA class III) and matched controls, showed a 18% higher resting metabolic rate, which was significantly correlated with reduced fat-free mass. Data obtained from non-obese and clinically stable CHF patients indicate that inadequate calorie and protein intake, together with reduced energy availability might trigger the development of muscle wasting (26). Higher resting energy expenditure has been attributed to sympathetic overactivity, whereas

stimulated thermogenesis might result from increased leptin levels. Whole-body protein metabolism in CHF patients appeared to be affected only if resting hypermetabolism coexisted (27). Of note, in this study, a pro-inflammatory status favoured protein breakdown. Overall, several components of the neuro-hormonal network, involved in the development of cachexia, affect food intake, energy expenditure and substrate utilization (see further).

5.2. Processes involved in the loss of skeletal muscle protein

There is a very tight regulation of intracellular protein degradation, for which multiple proteolytic systems exist. Of particular interest in the setting of cardiac cachexia, is the ubiquitin-proteasome system (UPS) (28). In this system, proteins are first tagged for degradation by ubiquitin. The process of ubiquitination is repeated, until a chain of several ubiquitin molecules are linked to each other and to the protein substrate. This is an energy consuming process involving several enzymes and carrier proteins. Ubiquitin is activated by the E1 enzyme, resulting in the formation of ubiquitin-E1 thiol ester, which is then recognized by E2 enzymes, to which ubiquitin is transferred. E2 serves as an ubiquitin-carrier protein and delivers ubiquitin to the target protein with the aid of E3 (ubiquitin ligase), which is responsible for the target selection and specificity. After polyubiquitination, the modified protein is rapidly degraded by a very large cytosolic proteolytic complex, the 26S proteasome, again requiring ATP to function. It appears that multiple types of skeletal muscle atrophy involve a common program of UPS-related changes in gene expression. There appears to be an impressive upregulation of the E3 ligases atrogin 1/MAFbx and MuRF-1 in fasted mice, in rats with cancer cachexia, streptozotocin-induced diabetes, uraemia and after disuse (29, 30). Recently, the upregulation of these E3 ligases has been demonstrated in atrophic muscle (31), as well as in the myocardium in animal heart failure models (32) and in skeletal muscle biopsies of CHF patients.

Apoptosis or programmed cell death has also been put forward as a mechanism contributing to the reduction of muscle mass. Although the potential consequences of the loss of a single nucleus in a multi-nucleated muscle fibre seem limited at first glance, the nuclear domain theory supports the notion that each nucleus controls a specific cytoplasmatic territory. Data have been published on the presence of apoptotic nuclei in skeletal muscles of CHF patients. In a small selective patient group scheduled for surgical revascularisation, skeletal muscle contained a higher number of TUNEL positive nuclei compared to controls (33). Tissue concentrations of caspase 3 and ubiquitin were increased, whereas the anti-apoptotic protein bcl-2 was decreased. There was an inverse relationship between TUNEL positive nuclei and both fibre cross sectional area and $\text{VO}_{2\text{peak}}$. Adams *et al.* confirmed these findings (34). A larger group of CHF patients was divided according to the presence of apoptosis or not. Besides a significantly lower exercise capacity, the apoptosis-positive subgroup was characterized by increased iNOS and a lower bcl-2 expression. However, the lack of specificity of the TUNEL technique, which is

often used to detect DNA fragmentation, might lead to overestimation of the relevance of apoptosis in this particular context. In our experience, skeletal muscle apoptosis in mild to moderate CHF patients could not be confirmed (35). Despite the presence of TUNEL positive nuclei in biopsies taken from the vastus lateralis muscle of CHF patients, confirmatory immunohistochemical analyses, using antibodies against cleaved caspase 3 and cleaved poly ADP-ribose polymerase (PARP) were negative. In addition, several TUNEL-positive nuclei also stained positive with SC35 splicing antibodies, indicating active gene transcription and thus precluding apoptosis.

Skeletal muscle satellite cells reside under the basement membrane, surrounding myofibres. Considered to guard muscular integrity, they awake from a quiescent state in case of muscle damage and re-enter the cell cycle. By fusing together or to existing myofibres, respectively, replacement or repair of injured tissue takes place. Although speculative, the limited proliferative capacity of residing satellite cells (≈ 50 -60 doublings) might be overwhelmed by repeated skeletal muscle injury (i.e.; inflammation, oxidative stress, catecholamines). In addition, several neurohormonal maladaptations that characterise CHF might disorganise the satellite cell population. Recent studies have demonstrated the presence of another population of progenitor cells, identified as c-kit positive stem cells (36). Compared to healthy individuals, their number in skeletal muscle biopsies of CHF patients is significantly lower. If these cells actually contribute to skeletal muscle regeneration, the poor recruitment of circulating stem cells in CHF might be another limiting factor for skeletal muscle repair.

6. DISRUPTION OF THE NEUROHORMONAL BALANCE: MEDIATORS OF SKELETAL MUSCLE WASTING

There is no doubt that numerous factors act upon the skeletal muscle in CHF, but the neurohormonal environment seems to be critical in this regard (Figure 3).

6.1. “Classical” neuroendocrine pathways: catecholamines and the RAAS

There is a significant upregulation of so-called stresshormones in cachectic versus non-cachectic CHF patients (37). It has long been known that catecholamines catalyze the vicious circle that characterizes the CHF syndrome. Besides the devastating consequences of their toxic myocardial and vascular effects, the peripheral musculature is also submitted to mainly beta-2 receptor mediated adrenergic effects. Both necrotic and apoptotic catecholamine-induced cell death, especially of slow-twitch fibres, have been described (38). Preliminary evidence suggests that weight gain, especially in cachectic CHF patients, can be obtained with the use of beta-blocker therapy (39, 40). Of note, there is important crosstalk between the peripheral skeletal muscle and the sympathetic nervous system. Indeed, increased ergoreflex sensitivity, which is related to reduced muscle mass in CHF patients, directly stimulates sympathetic drive, resulting in disruption of the autonomic nervous system (12).

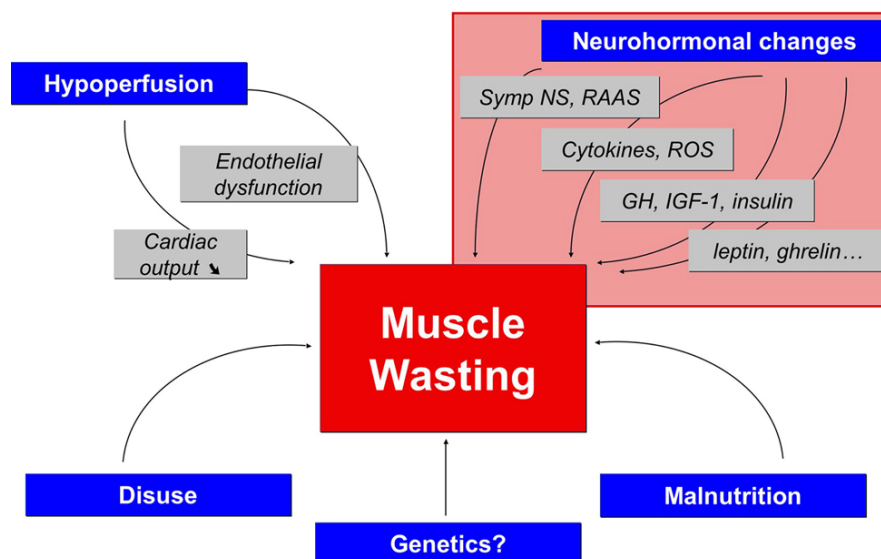


Figure 3. Determinants of muscle wasting in chronic heart failure. Several factors are responsible for the muscle changes observed in the setting of chronic heart failure. Despite the initial compensatory function, however, disturbance of the neurohormonal network is particularly relevant in the pathogenesis of muscular wasting. Symp NS: sympathetic nervous system, RAAS: renin-angiotensin-aldosterone system, ROS: reactive oxygen species, GH: growth hormone, IGF-1: insulin-like growth factor -1.

In animal experiments, angiotensin II induces pressor-independent weight loss, probably due to a significant anorexigenic effect. However, the observed concomitant increased protein breakdown and the reduction in circulating and local skeletal insulin-like growth factor 1 (IGF-1) levels also implicate metabolic consequences (41, 42). The muscular upregulation of *Murf-1* and *MAFbx* mRNA implies an important role for the UPS. In transgenic mice with skeletal muscle specific IGF-1 expression, these changes were blocked (43). Skeletal muscles express angiotensin (AT)-1 receptors and the administration of angiotensin II in the rat results in myocyte apoptotic cell death (44). Dalla Libera *et al.* showed that in the monocrotaline induced CHF model, AT1 receptor blockade protected apoptosis-dependent skeletal muscle atrophy and CHF-related changes in the MHC pattern (45). Aldosterone injections induced skeletal muscle apoptosis in the soleus muscle of the rat, an effect that could be prevented by the prior administration of spironolactone (46). Similar to catecholamines, and probably mediated through the sympathetic nervous system, angiotensin II appears to induce lipolysis in subcutaneous and visceral fat (47).

6.2. Anabolic/catabolic imbalance

Anabolic deficiency is related to abnormalities in 3 anabolic endocrine systems: the gonadal, adrenal and somatotrophic axes. The cortisol/dehydroepiandrosterone (DHEAS) ratio in cachectic CHF patients is significantly increased and clearly related to lean tissue mass (14). In men with CHF, reduced testosterone, DHEAS and IGF-1 levels were independent markers of poor prognosis (48). Acquired growth-hormone (GH) resistance is a feature of severe catabolism. Its presence in other critical illnesses has stimulated interest in a possible role in the pathophysiology

of cardiac cachexia. GH exerts catabolic actions via activation of somatomedins, of which IGF-1 is considered a main component. The classical pattern of GH resistance entails an increased GH level without a proportional rise in IGF-1. Niebauer *et al.* (49) divided CHF patients according to their IGF-1 level and showed a significant skeletal muscle functional and mass deficit in the legs of those with low levels. The ratio of IGF-1/GH appears significantly reduced in cachectic CHF patients (50). Interestingly, the most important predictor of this ratio was the level of GH-binding protein, which is identical to the GH receptor ectodomain and reflects the cellular GH receptor status. Anker and co-workers meticulously investigated the GH-IGF-1 axis and proposed that the increase in IGF-1 levels upon GH treatment in CHF patients depends on circulating concentrations of GH-binding protein. These observed changes in circulating levels could be just the tip of the iceberg, since Hambrecht and colleagues (51) demonstrated that even in non-cachectic CHF patients, local skeletal muscle IGF-1 production is decreased, whereas the IGF-1 receptor is upregulated. In this study, local IGF-1 expression was related to muscle CSA. Transgenic overexpression of locally acting IGF-1 inhibited UPS-mediated muscle atrophy in a CHF mice model (52). In addition, preliminary evidence supports the concept that local IGF-1 may act as a regenerative agent, increasing stem cell recruitment to sites of muscle injury and resulting in accelerated myogenic differentiation (53).

Chronic heart failure is to be considered an insulin-resistant state (22), which has an important effect on survival (54). Besides a shift in substrate utilisation, insulin resistance is inversely related to exercise capacity. This effect might be partly mediated by functional skeletal

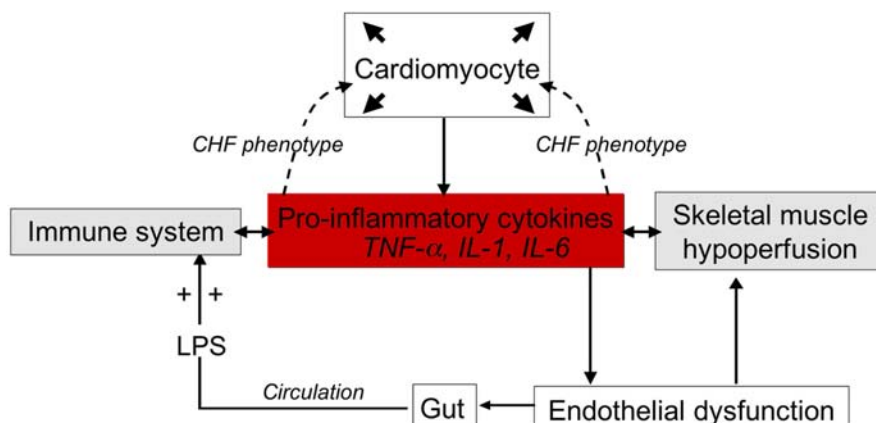


Figure 4. Pro-inflammatory adaptations in the setting of chronic heart failure. It is very likely that pro-inflammatory cytokines are produced in variable organs and tissues in the setting of chronic heart failure. Moreover, these cytokines not only contribute to the circulating pool, they also have para- and endocrine function, mostly with detrimental consequences. Whereas circulating pro-inflammatory cytokines are capable of inducing the heart failure phenotype, the local production by cardiomyocytes is probably the result of haemodynamic overload (arrows). Pro-inflammatory cytokines reduce endothelial vasodilatory capacity, which in turn leads to skeletal muscle and gut underperfusion. Skeletal muscles release pro-inflammatory cytokines secondary to hypoxia. The translocation of lipopolysaccharide (LPS) or endotoxin into the circulation could be an important trigger for immune activation.

muscle impairment, but muscle wasting as a result from activation of the UPS through suppression of PI3K/Akt signalling has also recently been described (55).

6.3. Inflammation and oxidative stress

The seminal work of Levine and colleagues, published in 1990, opened a whole new area of heart failure related research (56). These authors documented elevated levels of the pro-inflammatory cytokine tumour necrosis factor (TNF)- α in cachectic CHF patients. Since then, it has become clear that pro-inflammatory cytokines and their receptors are produced in multiple organs and tissues in the setting of CHF (Figure 4) (57-59). The independent prognostic role of pro-inflammatory cytokines and their receptors has been demonstrated in large cohorts of CHF patients (60). The presumed catabolic role of TNF- α is reflected in its former quotation, 'cachectin'. Besides its anorexigenic effect, skeletal muscle atrophy and weakness are elicited by this cytokine through a variety of mechanisms. First, TNF- α and interleukin (IL)-1 impair myogenesis through the activation of the transcription factor nuclear factor- κ B, which may lead to the inability of satellite cells to differentiate into functional fibres after damage *in vivo* (61). Secondly, the pro-apoptotic effect of TNF- α at the level of the skeletal muscle could be relevant (62). Finally, TNF-related muscle atrophy is mediated by p38 MAPK signalled expression of MAFbx, resulting in activation of the UPS (63). Reactive oxygen species (ROS) are likely to function as a second messenger after TNF-receptor binding. Cytokines and circulating soluble TNF receptor levels correlate with exercise performance of CHF patients (64). The local mRNA expression of IL-1, IL-6, TNF- α and iNOS illustrates the para- and autocrine function of these cytokines (65). In a rat model, Janssens *et al.* (66) demonstrated that even short-term exposure to IL-6 significantly reduces the cross-section of both type I and type II muscle fibres. The intricate link between local

inflammation and oxidative stress is reflected by the concomitant increased expression of pro-inflammatory cytokines and the downregulation of radical scavenger enzymes, resulting in increased rates of apoptosis when compared to sedentary controls (67).

6.4. Leptin

Recently, several new candidates for lean and fat body mass regulation in CHF have been studied. Leptin is the adipocyte product of the ob gene, with a direct inhibitory effect on neuropeptide Y, which is located mainly in the hypothalamus (68). Leptin induces saturation, it reduces lipid synthesis and increases energy expenditure and thermogenesis. The adipocyte origin of leptin explains its relationship and regulation by body weight and the amount of fat tissue. This negative feedback mechanism appears to be disturbed, since fat-corrected leptin levels in CHF patients are higher than normal. However, no difference between cachectic and non-cachectic CHF patients has been reported (69). The close relationship of leptin levels with the previously mentioned GH-BP suggests that leptin might play a role in GH resistance.

6.5. Ghrelin

Ghrelin is a GH releasing peptide derived from the stomach that stimulates food intake resulting in a positive energy balance. It also has anti-inflammatory, anti-apoptotic and vasodilatory effects and it reduces the activation of the central nervous system. Nagaya and colleagues (70) nicely demonstrated that cachectic CHF patients show significantly increased levels of circulating ghrelin, which tended to increase with the severity of the disease and were related to plasma levels of GH and TNF- α . These researchers proposed that ghrelin levels are elevated in CHF as a sort of compensatory mechanism. Intravenous infusion of ghrelin in 10 CHF patients apparently augmented food intake without an impressive

rise in body weight, but with a significant increase in lean body mass (71). In addition, left ventricular function and exercise capacity significantly improved.

7. HEART FAILURE RELATED MYOPATHY; WHERE DOES EXERCISE TRAINING COME IN?

Scepticism surrounding the mere existence of a specific myopathy related to CHF has fuelled the comparison with other chronic illnesses that involve skeletal muscle wasting, with age-related sarcopenia and with sedentary normal subjects.

There appears to be a very specific process of age-related loss of muscle mass, termed “age-related motor-unit remodelling”, which encompasses the transition towards a slower phenotype (more type I fibres) and more pronounced atrophy of type II fibres. This shift appears to be related to denervation and subsequent reinnervation of type II fibres, although part of the denervated fibres is lost (72). Without going into detail, it is obvious from the well-known phenotypic changes observed in CHF patients, which involve exactly the opposite processes (i.e. shift from type I to type II fibres), that CHF related myopathy is distinct. Elderly CHF patients, however, might exhibit a “mixed” pattern, resulting in an even greater vulnerability to the disabling consequences of skeletal muscle adaptations (73).

Although the “chicken and egg” discussion will not be easily resolved, the previously described neurohormonal and molecular abnormalities that go hand in hand with the development of muscle wasting strongly suggest that this clinical entity cannot be simply attributed to muscle disuse. The implementation of exercise training in the CHF population, however, offers an effective non-pharmacological opportunity to improve exercise capacity and quality of life and also provides survival benefit (74). The fact that muscle hypertrophy, increased muscle strength, reduced muscle fatigability and enhanced aerobic capacity co-occur with the reversal of several pathways that are considered characteristic of cachexia, underscores the existence of a specific CHF related myopathy.

In a series of randomized exercise-based studies, Hambrecht *et al.* (7, 75) showed that endurance training in CHF patients induces a re-shift in fibre phenotype, together with a partial restoration of metabolic abnormalities at the level of the skeletal muscle. Improved aerobic metabolism is illustrated by the observed increase in volume density of mitochondria and of cytochrome c oxidase-positive mitochondria in biopsies taken from the vastus lateralis muscle (75). There is now ample evidence that exercise training has anti-inflammatory effects, both at the level of the skeletal muscle itself (65), and in the peripheral circulation (76, 77). Interestingly, besides the significantly down-regulated expression of TNF- α , IL-1 β and IL-6 in the skeletal muscle of CHF patients after a 6-month program of endurance training, Gielen *et al.* (65) also found a 50% lower expression of iNOS. Later studies confirmed the anti-oxidative capacity of regular physical exercise by demonstrating the augmented activity of radical scavenger

enzymes (i.e. catalase and glutathione peroxidase activity), together with decreased nitrotyrosine production (67). Exercise training reduces the level of circulating neurohormones (64, 78) and muscle sympathetic nerve activity (MSNA) (79). In line with the well-known stimulatory effect of muscular stretch on IGF-1 expression in animal studies, exercise training led to a strong increase in local skeletal muscle IGF-1 expression, despite the lack of changes in circulating levels of both GH and IGF-1 (80).

An important issue that has been insufficiently addressed until now, is the choice of training modalities for this specific population. Although it is tempting to speculate that resistance training will preferentially tackle and reverse muscle atrophy, a direct comparison of endurance and resistance training is urgently needed. There is evidence to support the notion that resistance training in these patients is safe and effective (81). In addition, many of the observed effects of endurance training have been duplicated with the use of resistance training in CHF patients. This is true for changes in the autonomic nervous system (82) and restoration of peripheral endothelial function (82, 83). Comprehensive training programs, combining dynamic resistive exercise with regular endurance training are gradually gaining popularity in severely debilitated CHF patients. Neuromodulation (64) as well as anti-inflammatory properties (76) have been ascribed to combined resistance-endurance exercise training.

8. CONCLUSIONS

It has taken the medical community a long time to fully recognize the detrimental consequences of muscular wasting and this is particularly true for cardiologists. The term cachexia already appears in ancient literature. Although Hippocrates associated “melting of the thighs” with poor prognosis, basic scientists have only recently begun to explore this fascinating field. Cachexia characterizes the end stage of numerous chronic illnesses. Therefore, important breakthroughs in deciphering this complex disease state are expected to surface from various domains in medicine. Pathophysiological insight is a requisite for the development of therapeutic strategies designed to reverse or halt muscular wasting. Although post hoc analyses of large randomized drug trials provide substantial support for the idea that optimal evidence based CHF treatment is beneficial, a specific anabolic approach for cardiac patients with cachexia is still lacking. The complexity of this disorder will necessitate a multi-faceted and multi-disciplinary approach. Although detailed information is beyond the scope of this review, efforts to evaluate the effects of exercise training, nutritional optimization, neurohormonal modulation, anti-inflammatory approaches, GH and ghrelin infusions are being actively pursued.

9. REFERENCES

1. Franciosa J. A., B. J. Baker & L. Seth: Pulmonary versus systemic hemodynamics in determining exercise capacity

- of patients with chronic left ventricular failure. *Am Heart J* 110, 807-813 (1985)
2. Kobashigawa J. A., D.A. Leaf, N. Lee, M. P. Gleeson, H. Liu, M. A. Hamilton, J. D. Moriguchi, N. Kawata, K. Einhorn, E. Herlihy & H. Laks: A controlled trial of exercise rehabilitation after heart transplantation. *N Engl J Med* 340, 272-277 (1999)
3. Conraads V., J. Bosmans & C. Vrints: Chronic heart failure: an example of a systemic chronic inflammatory disease resulting in cachexia. *Int J Cardiol* 85, 33-49 (2002)
4. Harrington D., S. D. Anker, T. P. Chua, K. M. Webb-Peploe, P. P. Ponikowski, P. A. Poole-Wilson & A. J. Coats: Skeletal muscle function and its relation to exercise tolerance in chronic heart failure. *J Am Coll Cardiol* 30, 1758-1764 (1997)
5. Massie B. M., A. Simonini, P. Sahgal, L. Wells & G. A. Dudley: Relation of systemic and local muscle exercise capacity to skeletal muscle characteristics in men with congestive heart failure. *J Am Coll Cardiol* 27, 140-145 (1996)
6. Drexler H., U. Riede, T. Münzel, H. König, E. Funke & H. Just: Alterations of skeletal muscle in chronic heart failure. *Circulation* 85, 1751-1759 (1992)
7. Hambrecht R., E. Fiehn, J. T. Yu, J. Niebauer, C. Weigl, L. Hilbrich, V. Adams & U. Riede, G. Schuler: Effects of endurance training on mitochondrial ultrastructure and fiber type distribution in skeletal muscle of patients with stable chronic heart failure. *J Am Coll Cardiol* 29, 1067-1073 (1997)
8. Sullivan M. J., H. J. Green & F. R. Cobb: Skeletal muscle biochemistry and histology in ambulatory patients with long-term heart failure. *Circulation* 81, 518-527 (1990)
9. Anker S. D., P. Ponikowski, S. Varney, T. P. Chua, A. L. Clark, K. M. Weber-Peploe, D. Harrington, W. J. Kox, P. A. Poole-Wilson & A. J. Coats: Wasting as independent risk factor for mortality in chronic heart failure. *Lancet* 349, 1050-1053 (1997)
10. Anker S. D., A. Negassa, A. J. Coats, R. Afzal, P. A. Poole-Wilson, J. N. Cohn & S. Yusuf: Prognostic importance of weight loss in chronic heart failure and the effect of treatment with angiotensin-converting-enzyme inhibitors: an observational study. *Lancet* 361, 1077-1083 (2003)
11. Ciccoira M., L. Zanolla, L. Franceschini, A. Rossi, G. Golia, M. Zamboni, P. Tosoni & P. Zardini: Skeletal muscle mass independently predicts peak oxygen consumption and ventilatory response during exercise in noncachectic patients with chronic heart failure. *J Am Coll Cardiol* 37, 2080-2085 (2001)
12. Harrington D., S. D. Anker & A. J. Coats: Preservation of exercise capacity and lack of peripheral changes in asymptomatic patients with severely impaired left ventricular function. *Eur Heart J* 22, 392-399 (2001)
13. Piepoli M., A. Kaczmarek, D. P. Francis, L. C. Davies, M. Rauchhaus, E. A. Jankowska, S. D. Anker, A. Capucci, W. Banasiak & P. Ponikowski: Reduced peripheral skeletal muscle mass and abnormal reflex physiology in chronic heart failure. *Circulation* 114, 126-134 (2006)
14. Anker S. D., P. P. Ponikowski, A. L. Clark, F. Leyva, M. Rauchhaus, M. Kemp, M. M. Teixeira, P. G. Hellewel, J. Hooper, P. A. Poole-Wilson & A. J. Coats: Cytokines and neurohormones relating to body composition alterations in the wasting syndrome of chronic heart failure. *Eur Heart J* 20, 683-693 (1999)
15. Filippatos G. S., C. Kanatselos, D. D. Manolatos, B. Vougas, A. Sideris, D. Kardara, S. D. Anker, F. Kardaras & B. Uhal: Studies on apoptosis and fibrosis in skeletal musculature; a comparison of heart failure patients with and without cardiac cachexia. *Int J Cardiol* 90, 107-113 (2003).
16. Duscha B.D., W. E. Kraus, S. J. Keteyian, M. J. Sullivan, H. J. Green, F. H. Schachat, A. M. Pippen, C. A. Brawner, J. M. Blank & B. H. Annex: Capillary density of skeletal muscle: a contributing mechanism for exercise intolerance in class II-III chronic heart failure independent of other peripheral alterations. *J Am Coll Cardiol* 33, 1956-1963 (1999).
17. Mancini D.M., N. Ferraro, M. Tuchler, B. Chance & J. R. Wilson: Detection of abnormal calf muscle metabolism in patients with heart failure using phosphorus-31 nuclear magnetic resonance. *Am J Cardiol* 26, 1234-1240 (1988)
18. Hambrecht R., V. Adams, S. Gielen, A. Linke, S. Mobius-Winkler, J. Yu, J. Niebauer, H. Jiang, E. Fiehn & G. Schuler: Exercise intolerance in patients with chronic heart failure and increased expression of inducible nitric oxide synthase in the skeletal muscle. *J Am Coll Cardiol* 33, 174-179 (1999).
19. Ventura-Clapier R., A. Garnier & V. Veksler: Energy metabolism in heart failure. *J Physiol* 555, 1-13 (2004)
20. Mettauer B., J. Zoll, H. Sanchez, E. Lampert, F. Ribera, V. Veksler, X. Bigard, P. Mateo, E. Epailly, J. Lonsdorfer & R. Ventura-Clapier : Oxidative capacity of skeletal muscle in heart failure patients versus sedentary or active control subjects. *J Am Coll Cardiol* 38, 947-954 (2001)
21. Norrelund H., H. Wiggers, M. Halbirk, J. Frystyk, A. Flyvbjerg, H. E. Botker, O. Schmitz, J. O. Jorgensen, J. S. Christiansen & N. Moller: Abnormalities of whole body protein turnover, muscle metabolism and levels of metabolic hormones in patients with chronic heart failure. *J Int Med* 260, 11-21(2006)
22. Swan J. W., S. D. Anker, C. Walton, I. F. Godsland, A. L. Clark, F. Leyva, J. C. Stevenson & A. J. Coats: Insulin resistance in chronic heart failure: relation to severity and etiology of heart failure. *J Am Coll Cardiol* 30, 527-532 (1997)
23. Toth M. J., D. E. Matthews, P. A. Ades, M. D. Tischler, P. Van Buren, M. Previs & M. M. LeWinter: Skeletal muscle myofibrillar protein metabolism in heart failure: relationship to immune activation and functional capacity. *Am J Physiol Endocrinol Metab* 288, 685-692 (2005)
24. Toth M. J., P. A. Ades, M. M. LeWinter, R. P. Tracy & A. Tchernof: Skeletal muscle myofibrillar mRNA expression in heart failure; relationship to local and circulating hormones. *J Appl Physiol* 100, 35-41 (2006)
25. Poehlman E. T., J. Scheffers, S. S. Gottlieb, M. L. Fisher & P. Vaitekevicius: Increased resting metabolic rate in patients with congestive heart failure. *Ann Int Med* 121, 860-862 (1994)
26. Aquilani R., C. Opasich, M. Verri, F. Boschi, O. Febo, E. Pains & O. Pastoris: Is nutritional intake adequate in chronic heart failure patients? *J Am Coll Cardiol* 42, 1218-1223 (2003)

27. Toth M. J. & D. E. Matthews: Whole-body protein metabolism in chronic heart failure; relationship to anabolic and catabolic hormones. *J Parenter Enteral Nutr* 30, 104-201 (2006)
28. Nandi D., P. Tahiliani, A. Kumar & D. Chandu: The ubiquitin-proteasome system. *J Biosci* 31, 137-155 (2006)
29. Lecker S. H., R. T. Jagoe, A. Gilbert, M. Gomes, V. Baracos, J. Bailey, S. R. Price, W. E. Mitch & A. L. Goldberg: Multiple types of skeletal muscle atrophy involve a common program of changes in gene expression. *FASEB* 18, 39-51 (2004)
30. Sackey J. M., J. P. K. Hyatt, A. Raffaello, R. T. Jagoe, R. R. Roy, V. R. Edgerton, S. H. Lecker & A. L. Goldberg: Rapid disuse and denervation atrophy involve transcriptional changes similar to those of muscle wasting during systemic diseases. *FASEB* 21, 140-155 (2007)
31. Schulze P. C., J. Fang, K. A. Kassik, J. Gannon, M. Cupesi, C. MacGillivray, R. T. Lee & N. Rosenthal: Transgenic overexpression of locally acting insulin-like growth factor-1 inhibits ubiquitin-mediated muscle atrophy in chronic left-ventricular dysfunction. *Circ Res* 97, 418-426 (2005)
32. Adams V., A. Linke, U. Wisloff U, C. Doring, S. Erbs, N. Krankel, C. C. Witt, S. Labeit, U. Muller-Werdan, G. Schuler & R. Hambrecht: Myocardial expression of Murf-1 and MAFbx after induction of chronic heart failure: effect on myocardial contractility. *Cardiovasc Res* 73, 120-129 (2007)
33. Vescovo G., M. Volterrani, R. Zennaro, M. Sandri, C. Ceconi, R. Lorusso, R. Ferrari, G. B. Ambrosio & M. Dalla Libera : Apoptosis in the skeletal muscle of patients with heart failure: investigation of clinical and biochemical changes. *Heart* 84, 431-437 (2000)
34. Adams V., H. Jiang, J. Yu J, S. Mobius-Winkler, E. Fiehn, A. Linke, C. Weigl, G. Schuler & R. Hambrecht: Apoptosis in skeletal myocytes of patients with chronic heart failure is associated with exercise intolerance. *J Am Coll Cardiol* 33, 959-965 (1999)
35. Conraads V. M. A., V. Y. Hoymans, T. Vermeulen, C. Vrints & W. Martinet: Apoptosis versus active gene transcription in the skeletal muscle of patients with mild to moderate chronic heart failure. Relationship with exercise capacity. (Abstract) *Eur J Cardiovasc Prev Rehabil* 13, S9 (2006)
36. Rosenthal N., A. Musaro. Gene therapy for cardiac cachexia? *Int J Cardiol* 85, 185-191 (2002)
37. Anker S.D., T. P. Chua, P. Ponikowski P, D. Harrington, J. W. Swan, W. J. Kox, P. A. Poole-Wilson & A. J. Coats: Hormonal changes and catabolic/anabolic imbalance in chronic heart failure and their importance for cardiac cachexia. *Circulation* 96, 526-534 (1997)
38. Goldspink D.F., J. G. Burniston, G. M. Ellison, W. A. Clark, L.-B. Tan L. Catecholamine-induced apoptosis and necrosis in cardiac and skeletal myocytes of the rat *in vivo*: the same or separate death pathways? *Exp Physiol* 89, 407-416 (2004)
39. Lainsack M., I. Keber & S. D. Anker: Body composition changes in patients with systolic heart failure treated with beta blockers: a pilot study. *Int J Cardiol* 106, 319-322 (2006)
40. Hryniewicz K., A. S. Androne, A. Hudaihed & S. D. Katz: Partial reversal of cachexia by beta-adrenergic receptor blocker therapy in patients with chronic heart failure. *J Card Fail* 9, 464-468 (2003)
41. Brink M., J. Wellen & P. Delafontaine: Angiotensin II causes weight loss and decreases circulating insulin-like growth factor I in rats through a pressor-independent mechanism. *J Clin Invest* 97, 2509-2516 (1996)
42. Brink M., S. R. Price, J. Chrast, J. L. Bailey, A. Anwar, W. E. Mitch & P. Delafontaine: Angiotensin II induces skeletal muscle wasting through enhanced protein degradation and down-regulates autocrine insulin-like growth factor I. *Endocrinology* 142, 1489-1496 (2001)
43. Song Y.-H., Y. Li, J. Du, W. E. Mitch, N. Rosenthal & P. Delafontaine: Muscle-specific expression of IGF-1 blocks angiotensin II-induced skeletal muscle wasting. *J Clin Invest* 115, 451-458 (2005)
44. Burniston J., A. Sain, L.-B. Tan & D. F. Goldspink DF: Angiotensin II induces apoptosis *in vivo* in skeletal as well as cardiac, muscle of the rat. *Exp Physiol* 90, 755-761 (2005)
45. Dalla Libera L., B. Ravara B, A. Angelini, K. Rossini, M. Sandri, G. Thiene, G. Battista Ambrosio & G. Vescovo: Beneficial effects on skeletal muscle of the angiotensin II type 1 receptor blocker irbesartan in experimental heart failure. *Circulation* 103, 2195-2200 (2001)
46. Burniston J. G., A. Saini, L.-B. Tan & D. F. Goldspink: Aldosterone induces myocyte apoptosis in the heart and skeletal muscles of rats *in vivo*. *J Mol Cell Cardiol* 39, 395-399 (2005)
47. Cabassi A., P. Coghi, P. Govoni, E. Barouhiel, E. Speroni, S. Cavazzine, A. M. Cantoni, R. Scandroglio & E. Fiaccadori : Sympathetic modulation by carvedilol and losartan reduces angiotensin II-mediated lipolysis in subcutaneous and visceral fat. *J Clin Endocrinol Metab* 90, 2888-2897 (2005)
48. Jankowska E. A., B. Biel, J. Majda, A. Szklarska, M. Lopuszanska, M. Medras, S. D. Anker, W. Banasiak, P. A. Poole-Wilson & P. Ponikowski: Anabolic deficiency in men with chronic heart failure. Prevalence and detrimental impact on survival. *Circulation* 114, 1829-1837 (2006)
49. Niebauer J., C. D. Pflaum, A. L. Clark AL, C. J. Strasburger, J. Hooper, P. A. Poole-Wilson, A. J. Coats & S. D. Anker: Deficient insulin-like growth factor I in chronic heart failure predicts altered body composition, anabolic deficiency, cytokine and neurohormonal activation. *J Am Coll Cardiol* 32, 393-397 (1998)
50. Anker S. D., M. Volterrani, C. D. Pflaum, C. J. Strasburger, K. J. Osterziel, W. Doehner, M. B. Ranke, P. A. Poole-Wilson, A. Glustain, R. Dietz & A. J. Coats: Acquired growth hormone resistance in patients with chronic heart failure: implications for therapy with growth hormone. *J Am Coll Cardiol* 38, 443-452 (2001)
51. Hambrecht R., P. C. Schulze, S. Gielen S, A. Linke, S. Mobius-Winkler, J. Yu, J. J. Kratzsch, G. Baldauf, M. W. Busse, A. Schubert, V. Adams & G. Schuler: Reduction of insulin-like growth factor-I expression in the skeletal muscle of noncachectic patients with chronic heart failure. *J Am Coll Cardiol* 39, 1175-1181(2002)
52. Schulze P. C., J. Fang, K. A. Kassik, J. Gannon, M. Cupesi, C. MacGillivray, R. T. Lee & N. Rosenthal. Transgenic overexpression of locally acting insulin-like growth factor-1 inhibits ubiquitin-mediated muscle atrophy in chronic left-ventricular dysfunction. *Circ Res* 97, 418-426 (2005)

53. Musaro A., C. Giacinti, G. Borsellino, G. Dobrowolny, L. Pelosi, L. Cairns, S. Ottolenghi, G. Cossu, G. Bernardi, L. Battistini, M. Molinaro & N. Rosenthal. Stem cell-mediated muscle regeneration is enhanced by local isoform of insulin-like growth factor I: *Proc Natl Acad Sci U.S.A.* 101, 1206-1210 (2004)
54. Doehner W., M. Rauchhaus, P. Ponikowski, I. F. Godsland, S. von Haehling, D. O. Okonko, F. Leyva, A. J. Proudler, A. J. Coats & S. D. Anker : Impaired insulin sensitivity as an independent risk factor for mortality in patients with stable chronic heart failure: *J Am Coll Cardiol* 46, 1019-1026 (2005)
55. Wang X., Z. Hu, J. Hu, J. Du & W. E. Mitch: Insulin resistance accelerates muscle protein degradation: activation of the ubiquitin-proteasome pathway by defects in muscle cell signaling. *Endocrinology* 147, 4160-4168 (2006)
56. Levine B., J. Kalman, L. Mayer, H. M. Fillit & M. Packer: Elevated circulating levels of tumor necrosis factor in severe chronic heart failure. *N Engl J Med* 323, 236-241 (1990)
57. Conraads V: Pro-inflammatory cytokines and their receptors in chronic heart failure: do they really matter? *Acta Cardiol* 61, 161-168 (2006)
58. Conraads V. M., J. M. Bosmans, A. J. Schuerwegh, I. Goovaerts, L. S. De Clerck, C. H. Bridts & C. J. Vrints: Intracellular Monocyte Cytokine Production and CD14 Expression Are Up-Regulated in Severe vs Mild Chronic Heart Failure: *J Heart Lung Transplant* 24, 854-859 (2005)
59. Conraads V. M., J. P. Jorens, L. S. De Clerck, H. K. Van Saene, M. M. Ieven, J. M. Bosmans, A. Schuerwegh, C. H. Bridts, F. Wuyts, W. J. Stevens, S. D. Anker, M. Rauchhaus & C. J. Vrints: Selective intestinal decontamination in advanced chronic heart failure: a pilot trial: *Eur J Heart Fail* 6, 483-491 (2004).
60. Deswal A., N. J. Petersen, A. M. Feldman, J. B. Young, B. G. White & D. L. Mann: Cytokines and cytokine receptors in advanced heart failure. An analysis of the cytokine database from vesnarinone trial (VEST). *Circulation* 103, 2055-2059 (2001)
61. Langen R. C. J., A. M. W. J. Schols, M. C. J. M. Kelders, E. F. M. Wouters & Y. M. W. Janssen-Heininger: Inflammatory cytokines inhibit myogenic differentiation through activation of nuclear factor- κ B. *FASEB J* 15, 1169-1180 (2001)
62. Dalla Libera L., R. Sabbadini, C. Renken, B. Ravara, M. Sandri, R. Betto, A. Angelini & G. Vescovo : Apoptosis in the skeletal muscle of rats with heart failure is associated with increased levels of tumor necrosis factor alpha and sphingosine: *J Mol Cell Cardiol* 33, 1871-1878 (2001)
63. Li Y. P., Y. Chen, J. John, J. Moylan, B. Jin, D. L. Mann & M. B. Reid : TNF- α acts via p38 MAPK to stimulate expression of the ubiquitin ligase atrogin 1/MAFbx in skeletal muscle. *FASEB J* 19, 362-370 (2005)
64. Conraads V.M., P. Beckers, J. Vaes, M. Martin, V. Van Hoof, C. De Maeyer, N. Possemiers, F. L. Wuyts & C. J. Vrints: Combined endurance/resistance training reduces NT-proBNP levels in patients with chronic heart failure: *Eur Heart J* 25, 1797-1805 (2004)
65. Gielen S., V. Adams, S. Mobius-Winkler, A. Linke, S. Erbs, J. Yu, W. Kempf, A. Schubert, G. Schuler & R. Hambrecht: Anti-inflammatory effects of exercise training in the skeletal muscle of patients with chronic heart failure. *J Am Coll Cardiol* 42, 861-868 (2003)
66. Janssen S. P., G. Gayan-Ramirez, A. Van den Bergh, P. Herijgers, K. Maes, E. Verbeken & M. Decramer: Interleukin-6 causes myocardial failure and skeletal muscle atrophy in rats. *Circulation* 111, 996-1005 (2005).
67. Linke A., V. Adams, P. C. Schulze, S. Erbs, S. Gielen, E. Fiehn, S. Mobius-Winkler, A. Schubert, G. Schuler & R. Hambrecht: Antioxidative effects of exercise training in patients with chronic heart failure. Increase in radical scavenger enzyme activity in skeletal muscle. *Circulation* 111, 1763-1770 (2005)
68. Korner J. & R. Leibel: To eat or not to eat – How the gut talks to the brain. *N Eng J Med* 349, 926-928 (2003)
69. Doehner W., C. D. Pflaum, M. Rauchhaus, I. F. Godsland, K. Egerer, M. Cicoria, V. G. Florea, R. Sharma, A. P. Bolger, A. J. Coats, S. D. Anker & C. J. Strasburger: Leptin, insulin sensitivity and growth hormone binding protein in chronic heart failure with and without cardiac cachexia. *Eur J Endocrin* 145, 727-735 (2001)
70. Nagaya N., M. Uematsu, M. Kojima, Y. Date, M. Nakazato, H. Okumura, H. Hosoda, W. Shimizu, M. Yamagishi, H. Oya, H. Koh, C. Yutani & K. Kangawa: Elevated circulating levels of ghrelin in the cachexia associated with chronic heart failure: relationships between ghrelin and anabolic/catabolic factors. *Circulation* 104, 2034-2038 (2001)
71. Nagaya N., J. Moriya, Y. Yasumura, M. Uematsu, F. Onon, W. Shimizu, K. Ueno, M. Kitakaze, K. Miyatake & K. Kangawa: Effects of ghrelin administration on left ventricular function, exercise capacity, and muscle wasting in patients with chronic heart failure. *Circulation* 10, 3674-3679 (2004)
72. Vandervoort A. A: Aging of the human neuromuscular system. *Muscle Nerve* 25, 1-25 (2002)
73. Gielen S., V. Adams, J. Niebauer, G. Schuler & R. Hambrecht: Aging and heart failure-similar syndromes of exercise intolerance? Implications for exercise-based interventions; *Heart Fail Monit* 4, 130-136 (2005)
74. Piepoli M. F., C. Davos, D. P. Francis & A. J. Coats, ExTraMATCH Collaborative. Exercise training meta-analysis of trials in patients with chronic heart failure (ExTraMATCH). *BMJ* 328, 189 (2004)
75. Hambrecht R., J. Niebauer, E. Fiehn, B. Kalberer, B. Offner, K. Hauer, U. Riede, G. Schlierf, W. Kubler & G. Schuler: Physical training in patients with stable chronic heart failure: effects on cardiorespiratory fitness and ultrastructural abnormalities of leg muscles. *J Am Coll Cardiol* 25, 1239-1249 (1995)
76. Conraads V., P. Beckers, J. Bosmans, L. S. De Clerck, W. J. Stevens, C. J. Vrints, D. L. Brutsaert: Combined endurance-resistance training reduces plasma TNF-alpha receptor levels in patients with chronic heart failure and coronary artery disease. *Eur Heart J* 23, 1854-1860 (2002)
77. Adamopoulos S., J. Parissis, D. Karatzas, C. Kroupis, M. Georgiadis, G. Karavolias, J. Paraskevaidis, K. Koniavitou, A. J. Coats & D. T. Kremastinos: Physical training modulates proinflammatory cytokines and the soluble Fas/Soluble Fas Ligand System in patients with chronic heart failure. *J Am Coll Cardiol* 39, 653-663 (2002)
78. Braith R. W., M. A. Welsch, M. Feigenbaum, H. A. Kluess & C. J. Pepine: Neuroendocrine activation in heart

failure is modified by endurance exercise training. *J Am Coll Cardiol* 34, 1170-1175 (1999)

79. Roveda F., H. R. Middlekauff, M. U. P. B. Rondon, S. F. Reis, M. Souza, L. Nastari, A. C. Barretto, E. M. Krieger & C. E. Negrao: The effects of exercise training on sympathetic neural activation in advanced heart failure. A randomized controlled trial. *J Am Coll Cardiol* 42, 854-860 (2003)

80. Hambrecht R., P. C. Schulze, S. Gielen, A. Linke, S. Mobius-Winkler, S. Erbs, J. Kratzsch, A. Schubert, V. Adams & G. Schuler: Effects of exercise training on insulin-like growth factor-I expression in the skeletal muscle of non-cachectic patients with chronic heart failure. *Eur J Cardiovasc Prev Rehabil* 12, 401-406 (2005)

81. Benton M. J.: Safety and efficacy of resistance training in patients with chronic heart failure: research-based evidence. *Prog Cardiovasc Nurs* 20, 17-23 (2005)

82. Selig S. E., M. F. Carey, D. G. Menzies, J. Patterson, R. H. Geerling, A. D. Williams, V. Bamroongsuk, D. Toia, H. Krum & D. L. Hare: Moderate-intensity resistance exercise training in patients with chronic heart failure improves strength, endurance, heart rate variability, and forearm blood flow. *J Cardiac Fail* 10, 21-30 (2004)

83. Maiorana A., G. O'Driscoll, L. Dembo, C. Cheetham, C. Goodman, R. Taylor & D. Green: Effect of Aerobic and Resistance Exercise Training on Vascular Function in Heart Failure. *Am J Physiol Heart Circ Physiol* 279, H1999-2005 (2000)

Key Words: chronic heart failure, cachexia, skeletal muscle, wasting, atrophy

Send correspondence to: Viviane Conraads, MD, PhD, Department of Cardiology, University Hospital Antwerp, Wilrijkstraat 10, 2650 Belgium, Tel: 00323821-4672, Fax: 00323821-3974, E-mail: Viviane.Conraads@uza.be

<http://www.bioscience.org/current/vol13.htm>