

Peroxisome proliferator-activated receptors and renal diseases

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1. ABSTRACT

Peroxisome proliferator-activated receptors (PPARs) are members of the nuclear hormone receptor superfamily of ligand-dependent transcription factors. Three isoforms of PPAR, i.e., PPAR- α , - δ , and - γ , have been identified and are differentially expressed in various tissues, including the kidney. The target genes of PPARs are involved in diverse biological processes, including adipogenesis, lipid metabolism, insulin sensitivity, inflammatory response, reproduction, and cell growth and differentiation. PPARs have been reported to protect against renal injury through indirect systemic effects and/or direct renal effects in diabetic nephropathy, glomerulonephritis, renal cell carcinoma, acute renal failure and chronic renal disease. In this review, we summarize the role of the three identified PPAR isoforms, PPAR α , - δ , and - γ , in renal physiology and discuss the renoprotective effects of PPAR ligands in various kidney diseases.

2. INTRODUCTION

The kidney plays a key role in regulating sodium and water homeostasis and blood pressure. Loss of renal function therefore causes many systemic disorders, including cardiovascular diseases and hypertension. With the prevalence of type 2 diabetes, diabetic renal complications, or diabetic nephropathy -- one of the major complications of diabetes -- has become a worldwide serious public-health concern, although glomerulonephritis, renal cell carcinoma and acute renal failure remain common renal diseases. If left untreated, these diseases progress to chronic kidney disease and, ultimately, end-stage renal disease (ESRD) (1). With ESRD, kidneys fail to function, which results in sodium and water retention and accumulation of metabolic wastes and many toxic substances. Renal system data from 2004 in the United States revealed kidney disease as a major health problem; approximately 20 million patients had the disease.

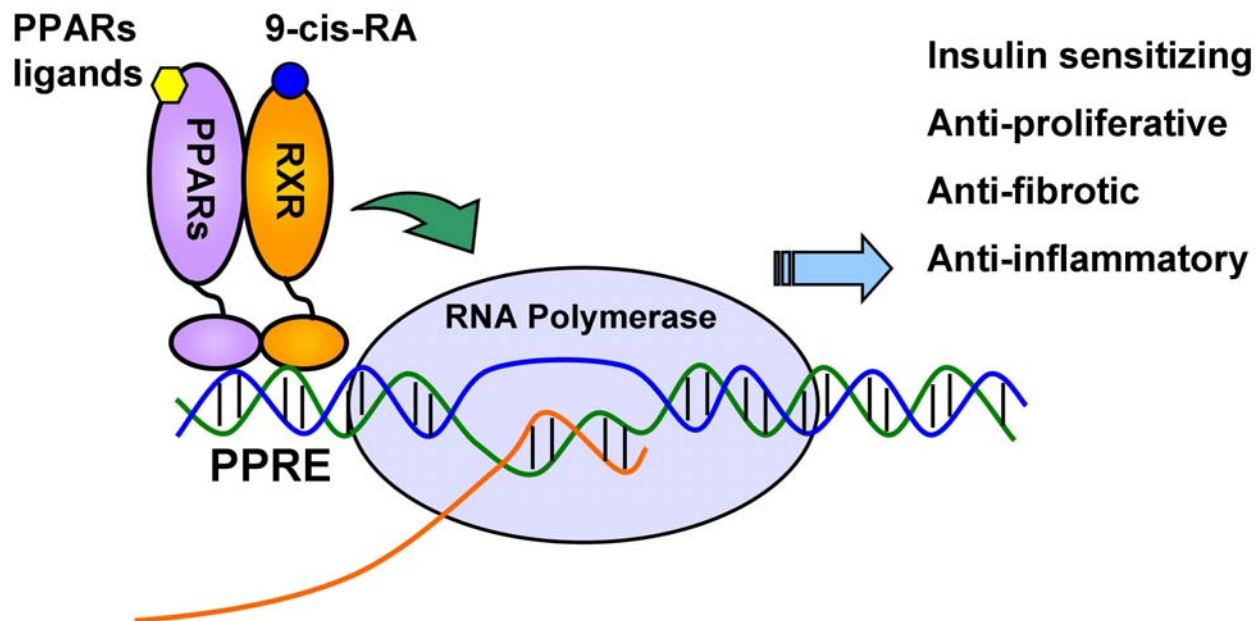


Figure 1. Schematic representation of the mode of action of PPARs. The PPAR isoforms form heterodimers with retinoid X receptor α (RXR α) in the presence of their ligands. The resulting heterodimer binds to PPAR response elements (PPRE) in the promoter regions of PPAR-driven genes, which are involved in many biological processes closely related to renoprotective effects, including insulin sensitizing, and anti-proliferative, anti-fibrotic, and anti-inflammatory actions.

Therefore, effective treatment of renal diseases is urgently needed.

As a subfamily of metabolic nuclear receptors, peroxisome proliferator-activated receptors (PPARs) participate in various biological processes, including lipid metabolism, adipogenesis, immune response, insulin sensitivity, reproduction and cell growth and differentiation (2). With the remarkable clinical effects of PPAR synthetic ligands, the role of PPARs in renal disease has received a lot of attention. In this review, we summarize the role of the three identified PPAR isoforms, PPAR α , - δ , and - γ , (3) in renal physiology and discuss the renoprotective effects of PPAR ligands in various kidney diseases.

3. PPARS: LIGANDS AND BIOLOGICAL ROLES

PPARs are members of the nuclear hormone receptor superfamily of ligand-dependent transcription factors. The three isoforms of PPAR, products of distinct genes, constitute the NR1C group in the nomenclature of nuclear receptors (4). In the presence of their specific ligands, PPARs usually heterodimerize with another nuclear receptor, retinoid X receptor α , forming a transcriptional complex that binds to a specific DNA sequence, peroxisome proliferator-response element, within the promoter regions of PPAR target genes. These genes are involved in diverse biological processes (Figure 1).

PPAR α , the first member of the PPAR subfamily identified, is highly abundant in tissues with high fatty acid oxidation activity, including the liver, kidney, intestine mucosa, heart and brown adipose (5, 6). Endogenous

ligands such as polyunsaturated fatty acids and synthetic ligands, including lipid-lowering fibrates (e.g., fenofibrate, clofibrate), can effectively activate PPAR α and regulate the transcription of an array of genes involved in lipid metabolism and inflammatory response (7, 8) (Table 1).

PPAR δ seems to be ubiquitously expressed at low levels in almost all tissues examined (6). The endogenous arachidonic-acid cyclooxygenase metabolite prostacyclin and synthetic compounds including L-165041 and GW2433 have been shown to selectively activate PPAR δ . A large body of evidence suggests that PPAR δ is involved in fatty acid and lipid metabolism and may be a pivotal factor in metabolic control (9). Recently, PPAR δ has also been reported to be important in maintaining renal cell survival in hyperosmotic medulla (10).

PPAR γ is expressed predominantly in adipose tissue, with low levels in stomach, intestine, urinary bladder, kidney, spleen, adrenal, liver, lung, brain, heart and vasculature (5, 6). PPAR γ controls adipocyte proliferation and differentiation and therefore plays an important role in regulation of lipid storage and insulin sensitivity (11). PPAR γ can be bound and activated by various small lipophilic compounds, including naturally occurring 15-deoxy- $\Delta^{12,14}$ -prostaglandin J2 (15d-PGJ2) and EETs and synthetic antidiabetic thiazolidinediones (TZDs) (e.g., rosiglitazone, pioglitazone), which are beneficial for improving insulin sensitivity. The main biological functions, ligands and distribution of expression of PPARs are summarized in Table 1.

Table 1. Biological roles, ligands and tissue distribution of PPAR isoforms.

Name	Biological functions	Ligands	Expression	Reference
PPAR α ¹	β -oxidation, fatty acid transport, lipoprotein synthesis, inflammatory response	8-S-hydroxyeicosatetraenoic acid, leukotriene B ₄ , synthetic fibrates	Abundant in liver, kidney, heart, brown adipose	8
PPAR δ	Lipid metabolism	PGI ₂ ² , synthetic compounds L-165041, GW2433	Ubiquitously expressed in almost all tissues	9
PPAR γ	Adipocyte proliferation and differentiation, lipid storage, insulin sensitivity	15d-PGJ ₂ ³ , synthetic TZDs ⁴	Predominantly expressed in adipose tissue, also mildly expressed in other tissues	11

Abbreviations: PPAR α ¹, Peroxisome proliferator-activated receptor α ; PGI₂², prostaglandin I₂; 15d-PGJ₂³, 15-deoxy- Δ 12,14-prostaglandin J₂; TZDs⁴, thiazolidinediones.

4. PPAR ligands: Clinical implications and side effects

Currently, the synthetic PPAR α ligand fenofibrate (Lipanthyl/Tricor), and PPAR γ ligands rosiglitazone (Avandia) and pioglitazone (Actos) have been used in clinical practice as lipid-lowering therapy and oral antidiabetic drugs, respectively. However, the use of these ligands may result in many side effects, including increased serum creatinine level (12), fluid retention and increased cardiovascular risk (13, 14). Recent research interest has focused on developing novel compounds possessing both PPAR α - and PPAR γ -activating properties to benefit both plasma glucose control and lipid levels. To date, many PPAR α/γ dual agonists have been developed and have shown promising therapeutic effects. However, the first generation of PPAR α/γ dual agonists, tesaglitazar (Galida) and muraglitazar (Pargluva), were withdrawn from phase III clinical trials because of their poor safety profile. The next generation of fine-tuned dual PPAR α/γ agonists prefers agonists with full PPAR α and partial PPAR γ activity.

5. PPARs: INTRARENAL LOCALIZATION

All three PPAR isoforms are functionally expressed in the kidney. PPAR α is highly expressed in the epithelial cells of proximal tubules and medullary thick ascending limbs, with much lower levels in glomerular mesangial cells (15-17), whereas PPAR γ is expressed primarily in the epithelium of distal medullary collecting ducts and to a lesser extent in the glomerular mesangial cells, endothelial cells and podocytes, proximal tubular cells, endothelial cells of renal microvasculature, and interstitial fibroblast cells (15, 18). In the kidney, PPAR δ seems to be diffusely expressed in the renal cortex and medulla, with relatively higher levels in medullary interstitial and stromal cells (15). The differential intrarenal localization of all three PPAR isoforms suggests that they play distinct roles in maintaining normal renal functions. In the following sections, we discuss the role of the three PPAR isoforms in renal pathophysiological settings and the therapeutic potential of PPAR ligands in various renal diseases, especially diabetic nephropathy.

6. PPARs: therapeutic role in renal diseases

6.1. PPAR α and renal disease

As mentioned above, in the kidney, PPAR α is selectively expressed in the proximal tubule cells, where its activation is essential for renal fatty acid metabolism, energy homeostasis, and anti-inflammatory regulation (3, 18). Large numbers of studies have indicated that PPAR α agonists significantly attenuate renal injury in various

kidney diseases such as diabetic nephropathy, acute renal failure, glomerulonephritis, and chronic renal failure (19-22). Thus, PPAR α could serve as an important renoprotective factor contributing to the prevention or delay of renal disease progression.

6.1.1. PPAR α and diabetic nephropathy

Increasing evidence suggests that PPAR α activators are effective in improving insulin resistance in type 2 diabetic patients with the insulin resistance syndrome (23). Indeed, Park *et al.* showed that fenofibrate treatment reduced fasting blood glucose, ameliorated insulin resistance, reduced hypertrophy of pancreatic islets, and reduced urinary albumin excretion in diabetic animals (19). To date, multiple mechanisms have been proposed for the hypoglycemic and insulin-sensitizing effect of PPAR α agonists. Fibrates have been reported to reduce the triglyceride content in skeletal muscle (24, 25), which is associated with improved insulin sensitivity (26). PPAR α agonists have also been found to increase hepatic fatty acid catabolism, thus resulting in decreased systemic and tissue free fatty acid content (27). Koh *et al.* showed that fenofibrate treatment prevented the development of diabetes in Otsuka Long Evans Tokushima Fatty (OLETF) rats by reducing adiposity, improving peripheral insulin sensitivity, and exerting beneficial effects on pancreatic β -cells (24). Recently, Mishra *et al.* demonstrated that PPAR α is a diabetes-induced transcription factor that helps control the renal response to lipids (28). Moreover, the renal-protective effects of fenofibrate might be achieved through the reduction of glomerular hypertrophy and mesangial matrix accumulation (19, 29). Taken together, these data suggest that PPAR α may represent a potential therapeutic target for treating insulin resistance and type 2 diabetes and preventing diabetic renal complications.

Interestingly, several recent studies have found a paradoxical phenomenon, that PPAR α deletion has a protective effect similar to that of its ligands in mice with insulin resistance induced by high-fat diet. Insulin resistance was improved in both young and old PPAR α -null mice (30-32). As expected, the old PPAR α -null mice showed milder albuminuria than wild-type mice (32). To date, the underlying mechanisms remain unknown and require further investigation.

6.1.2. PPAR α and acute renal failure

Recently, Portilla *et al.* revealed that PPAR α plays an important protective role in acute renal tubular injury induced by ischemia/reperfusion and cisplatin. Cisplatin is one of the most common antitumor agents used

in chemotherapy for malignant disease, and its major side effect is nephrotoxicity. Synthetic PPAR α ligands attenuate cisplatin-induced acute renal injury by preventing the inhibition of fatty acid oxidation (33), reducing apoptosis and necrosis of the proximal tubules through decreasing endonuclease G activity (34), and limiting inflammatory processes by blocking NF- κ B activity (21, 35). Most recently, Kamijo *et al.* demonstrated that injection of fatty acid-binding albumin into PPAR α -null mice resulted in more severe tubular lesions than in wild-type mice, which provides further evidence that PPAR α is a renoprotective factor (36). Similarly, the PPAR α agonist was shown to protect against ischemic renal injury via preservation of renal acyl CoA oxidase and cytochrome P450 4A1 gene expression through a PPAR α -dependent pathway during ischemia/reperfusion injury (37). The renal protective action of the PPAR α agonist in ischemia and nephrotoxin-induced renal tubular injury appears to be PPAR α dependent, since PPAR α gene-deficient mice subjected to renal ischemia/reperfusion or treated with nephrotoxins exhibited enhanced cortical necrosis and impaired renal function (37, 38).

6.1.3. PPAR α and glomerulonephritis

In recent years, the immunoregulatory activity of ligands for PPARs has attracted intensive attention (39, 40). Anti-glomerular basement membrane (GBM) glomerulonephritis characterized by crescent formation and necrotizing inflammation of glomerular capillaries is the most severe form of glomerulonephritis. By using a rat anti-GBM glomerulonephritis model, Saga *et al.* found that bezafibrate, a PPAR α agonist, can markedly suppress anti-GBM crescentic glomerulonephritis (41). In accordance with this result, Kamijo *et al.* recently reported that PPAR α can protect against glomerulonephritis induced by long-term exposure to the plasticizer di-(2-ethylhexyl)phthalate (42). All these findings suggest that PPAR α might be a novel therapeutic target for the treatment of glomerulonephritis.

6.2. PPAR δ and renal disease

PPAR δ plays a key role in biological processes such as fertility, lipid metabolism, bone formation, mast cell immunity, skin and brain development, wound healing, and tumorigenesis. Although PPAR δ mRNA is detected in almost all tissues and cells examined, it is relatively abundant in the kidney, with ubiquitous expression in all nephron segments (15).

Because of the high expression level in the kidney, PPAR δ participates in renal physiological regulation and pathophysiological processes. Letavernier *et al.* provided evidence that PPAR δ may protect the kidney against ischemia/reperfusion-induced acute renal failure by activating the antiapoptotic Akt signaling pathway and increasing the spread of tubular epithelial cells. In this study, PPAR δ ^{+/+} and PPAR δ ^{-/-} mutant mice showed more severe kidney dysfunction and injury than wild-type mice. Wild-type mice pre-treated with the PPAR δ agonist were completely protected against renal dysfunction (43).

Moreover, although nonsteroidal anti-inflammatory drugs (NSAIDs) are among the most

common pain relief medicines in the world, they are well recognized as a major class of therapeutic agent that causes renal papillary necrosis (44). Inhibition of PPAR δ activity may contribute to this side effect (10). Overexpression of PPAR δ can prevent medullary interstitial cell death due to reduced ability to tolerate hypertonic stress by COX2 inhibition, which suggests that PPAR δ might be an important survival factor for medullary interstitial cells in the hypertonic condition in the renal medulla.

PPAR δ may also be involved in the pathogenesis of diabetic nephropathy. Overexpression or activation of PPAR δ in skeletal muscle can significantly improve mouse endurance exercise ability, and resist to obesity with improved metabolic profiles, even in the absence of exercise (45). Regarding the direct renal effects, Escher *et al.* have recently found the PPAR δ mRNA level in the kidney remarkably down-regulated after an overnight fast and quickly restored to the normal level upon refeeding (46). In addition to nutritional regulation, the expression of PPAR δ in the kidney was also down-regulated in type 1 diabetic Akita and OVE26 mice, which resulted in decreased fatty acid oxidation and increased renal triglyceride accumulation (47). These findings provide strong evidence that PPAR δ activation may be beneficial for amelioration of diabetic nephropathy.

6.3. PPAR γ and renal disease

Although TZDs are a group of insulin sensitizers and are widely used in clinical therapy for type 2 diabetes, their renoprotective actions are just now being carefully evaluated. Increasing evidence has revealed the protective effects of PPAR γ activation on diabetic nephropathy, renal cell carcinoma, renal failure and glomerulonephritis, which strongly suggests that PPAR γ may be a potential therapeutic target for the treatment of these renal diseases (48-51). The mechanisms mediating the renoprotective effect of PPAR γ ligands may involve both systemic metabolic control and direct action on the kidney.

6.3.1. PPAR γ and diabetic nephropathy

As one of the major complications of diabetes, diabetic nephropathy is characterized by renal hypertrophy and extracellular matrix accumulation, which without effective intervention eventually progresses to fibrosis with loss of renal function. PPAR γ agonist TZDs may hold great promise for treating both insulin resistance and diabetic renal complications. The therapeutic effects of TZDs on prevention or even reversal of the progression of diabetic nephropathy are achieved possibly through both indirect systemic and direct renal effects (Figure 2).

PPAR γ exerts its insulin-sensitizing effects in adipose tissue, skeletal muscle, liver, and pancreatic β -cells. Loss-of-function mutation of PPAR γ results in severe insulin resistance, partial lipodystrophy, diabetes, hypertension and dyslipidemia in humans, in part because of excessive lipid accumulation in skeletal muscle and liver (52). PPAR γ -agonist TZDs increase the sensitivity of the liver to insulin-stimulated suppression of gluconeogenesis and enhance glucose utilization in the skeletal muscle (53). In the adipose tissue, PPAR γ activation increases glucose uptake and results in profound changes in adipokine

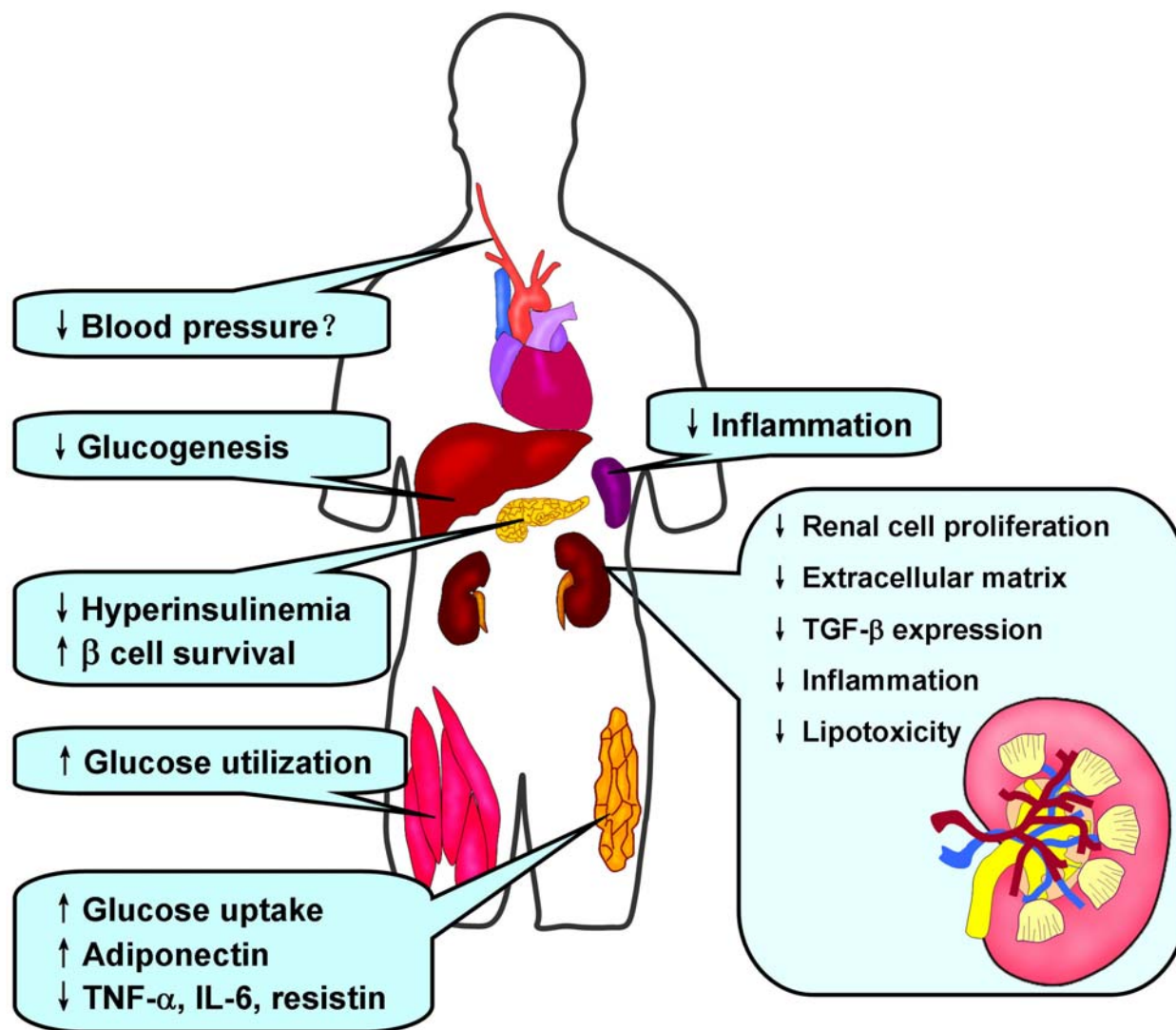


Figure 2. Summary of the therapeutic actions of PPAR γ in diabetic nephropathy. Both systemic action and direct renal effect are involved in the renoprotective effect of PPAR γ agonists. TNF- α , tumor necrosis factor α ; TGF- β , transforming growth factor β ; IL-6, interleukin 6.

expression and secretion, including suppression of insulin-desensitizing tumor necrosis factor α (TNF- α), interleukin-6 (IL-6) and resistin, and induction of insulin-sensitizing adiponectin and visfatin (54-58). In a recent study, activation of PPAR γ protected pancreatic β -cells from cytokine-induced cytotoxicity (59). In addition, PPAR γ can act as an anti-inflammatory factor to reduce the production of cytokines (TNF- α , IL-1, and IL-6) (60), probably by inhibiting the activity of pro-inflammatory transcription factors such as nuclear factor κ B (NF- κ B), activator protein 1 (AP-1) and signal transducer and activator of transcription (STAT) (61). The anti-inflammatory effect of PPAR γ is highly beneficial, since low-grade inflammation is associated with the pathogenesis of insulin resistance (62). Thus, systemic effects such as improving insulin resistance and attenuating inflammation may represent two major mechanisms mediating the beneficial effect of PPAR γ on glycemic control in type 2 diabetes, thereby

preventing the development or slowing the progression of diabetic nephropathy. PPAR γ may also benefit the kidney by lowering blood pressure. Although results remain inconclusive (63, 64), PPAR γ activation is believed to be effective in lowering blood pressure via attenuating the activity of the renin-angiotensin system and mitogen-activated protein kinase (MAPK), phosphatidylinositol 3-kinase, and ROS-generating enzymes (65-67). Interestingly, several lines of evidence point to an insulin-sensitizing action of PPAR γ antagonism (68-70). Although the underlying mechanism is unknown, these observations suggest that the PPAR γ antagonist may also have potential therapeutic implication in insulin resistance and type 2 diabetes.

Abundant evidence from studies of patients, animal models, and cell models has shown that local activation of PPAR γ in the kidney is involved in its

renoprotection. TZD therapy was repeatedly reported to be effective in reducing microalbuminuria in patients with type 2 diabetes (71-73). Compared to other oral hypoglycemic agents (including insulin, metformin, glyburide, and glibenclamide), all TZD PPAR γ agonists (troglitazone, rosiglitazone, and pioglitazone) produce similar glycemic control but appear to provide superior renal protection in patients with type 2 diabetes (74-76). For example, Miyazaki *et al.* recently reported that 12 weeks of rosiglitazone treatment significantly decreased albuminuria, which might be due at least in part to direct activation of renal PPAR γ (48). Consistent with these clinical observations, PPAR γ -agonist TZDs have been shown to improve diabetic nephropathy in animal models of both type 1 and type 2 diabetes (75, 77-80). Indeed, troglitazone treatment significantly decreased albuminuria, reduced glomerular hyperfiltration, ameliorated mesangial expansion, and inhibited renal matrix protein and TGF- β expression in the kidney of streptozotocin-induced type 1 and Zucker type 2 diabetic rats (79, 81, 82). As well, Baylis *et al.* (80) demonstrated that rosiglitazone treatment reduced albuminuria, improved glomerular filtration rate, and normalized glomerulosclerosis and tubulointerstitial fibrosis in obese type 2 diabetic rats. Importantly, a recent report showed that telmisartan is a weak PPAR γ agonist and its treatment slows the progression of diabetic nephropathy (83). This observation suggests that the renoprotective effect of angiotensin II type 1 receptor blockers may be attributed to PPAR γ activation in part.

Studies of cultured renal cells provide strong support for the possibility that direct renal action may also be involved in mediating the beneficial renal effect of PPAR γ agonists. PPAR γ agonists can inhibit the proinflammatory phenotype induced by advanced glycosylation end products (AGE) in cultured renal proximal tubular epithelial cells through STAT-1-mediated pathways involving IL-8 and intercellular adhesion molecule 1 (ICAM-1) (84). PPAR γ activators are also reported to significantly suppress the expression of transforming growth factor β (TGF- β), type IV collagen and ICAM-1 and infiltration of macrophages in the kidneys of diabetic rats, as well as inhibit NF- κ B and ICAM-1 in cultured glomerular endothelial cells and mesangial cells (82, 85, 86). In addition, PPAR γ ligand treatment inhibits cell growth and promotes cell differentiation in cultured mesangial cells (18, 87). Activation of PPAR γ markedly blocked AGE-induced MAPK activity (88) and high glucose-stimulated vascular endothelial-cell growth factor expression (89), which is consistent with the inhibitory effect of PPAR γ on cell proliferation of mesangial cells. Furthermore, TZDs ameliorate diabetic nephropathy via cell cycle-dependent mechanisms by inhibiting activity of p44/42 MAPK and bcl-2-dependent p27 (90). In the proximal tubular HK2 cells, activation of PPAR γ induces the G1-phase cell-cycle arrest and suppresses high glucose-induced AP-1 activity and monocyte chemoattractant protein 1 expression (91). Collectively, these studies suggest that PPAR γ has anti-inflammatory and antiproliferative effects in various renal cells, thereby attenuating diabetic renal complications.

In addition, increasing evidence supports the idea that antifibrotic effect of TZDs may also represent an important mechanism by which PPAR γ agonists improve diabetic nephropathy (Table 2). TZDs can ameliorate renal fibrosis by regulating many fibrosis relevant genes. Treatment of human cortical fibroblasts with pioglitazone exhibited an antiproliferative and hypertrophic effect with reduced type IV collagen and fibronectin secretion, suppressed matrix metalloproteinase-9 (MMP-9) activity, and decreased tissue inhibitor of metalloproteinase-1 (TIMP-1) and TIMP-2 production (92). Similar result was observed in human proximal tubular cells. PPAR γ agonists exerted antifibrotic actions by attenuating the increase in AP-1, TGF- β 1, and the extracellular matrix protein fibronectin (93). Antifibrotic hepatocyte growth factor (HGF) was found to be a direct target gene of PPAR γ in mesangial cells and renal interstitial fibroblasts (94). In addition, PPAR γ agonists activated c-met receptor tyrosine phosphorylation, induced Smad transcriptional co-repressor TG-interacting factor (TGIF) expression, and blocked TGF- β /Smad-mediated gene transcription in mesangial cells. Ablation of c-met receptor through the LoxP-Cre system in mesangial cells abolished the antifibrotic effect of 15d-PGJ2 (94). These antifibrotic effects of PPAR γ agonists in multiple cultured renal cells were consistent with the *in vivo* findings that PPAR γ activation improved diabetic nephropathy not only in type 1 (95), but also in type 2 diabetes (96).

Taken together, PPAR γ agonists can improve albuminuria and slow the progression of glomerulosclerosis in patients with type 2 diabetes and in animal models. Because of these desirable renoprotective effects, PPAR γ is a promising target for treating glomerular fibrotic diseases, especially diabetic nephropathy.

It is worth mentioning that combined treatment with PPAR α and PPAR γ agonists may have better therapeutic potential than each alone in the treatment of type 2 diabetes and diabetic nephropathy. Increasing evidence from both clinical trials and animal experiments show that PPAR α/γ dual agonists have striking effects on improvement of insulin resistance, hyperglycemia, dyslipidemia, blood pressure and β -cell function in type 2 diabetes (92, 97-106). In addition, the renoprotective effect of the dual agonists has been recently evaluated and shown marked reduction in albuminuria and renal glomerular fibrosis in both type 1 and type 2 diabetic mice (95, 96).

6.3.2. PPAR γ and renal cell carcinoma

Research into the impact of PPAR γ on renal cell carcinoma was initiated in 2001. Inoue *et al.* found that PPAR γ has strong immunoreactive expression in renal cancer tissues and the PPAR γ agonists inhibit the growth of renal cancer cell lines (107). The underlying mechanism might be that TZDs inhibit cell proliferation and induce apoptosis by down-regulating the expression of cyclin D1, Cdk4, vascular endothelial growth factor and basic fibroblast growth factor while up-regulating the expression of p21 and p27 (49, 108, 109). However, the *in vivo* efficacy of PPAR γ agonist in animal models has not been tested, and such studies would address the important

Table 2. Genes involved in antifibrotic action of TZDs in renal cells

Cell type	Genes relevant to fibrosis		Reference
Cortical fibroblast	↓type IV collagen ↓fibronectin	↓MMP-9 ¹ ↓TIMP-1 ² ↓TIMP-2	92
Proximal tubular cell	↓AP-1 ³ ↓TGF-β1 ⁴	↓fibronectin	93
Mesangial cell, Interstitial fibroblast	↑HGF ⁵ ↑TGIF ⁶ ↓TGF-β1 ↓PAI-1 ⁷	↓α-SMA ⁸ ↓fibronectin ↓Smad	94

Abbreviations: MMP-9¹, matrix metalloproteinase-9; TIMP-1², tissue inhibitor of metalloproteinase-1; AP-1³, activator protein 1; TGF-β1⁴, transforming growth factor β1; HGF⁵, hepatocyte growth factor; TGIF⁶, TG-interacting factor; PAI-1⁷, plasminogen activator inhibitor-1; α-SMA⁸, α-smooth muscle actin.

question of whether PPARγ could be a therapeutic target for the treatment of renal tumors.

It should be noticed that although agonists of PPARα and PPARγ are generally believed to be antitumor agents, little is known about the potential safety issues that could be involved in the use of PPARδ agonist. Unlike PPARα and PPARγ, activation of PPARδ has been reported to be associated with accelerate intestinal adenoma growth (110), suggesting PPARδ may be carcinogenic.

6.3.3. PPARγ and other kidney diseases

PPARγ activation also has a protective effect on nephritis. In a nephrotoxic serum-induced nephritic rat model, PPARγ agonists markedly alleviated crescentic glomerulonephritis by inhibiting the infiltration of ED-1-positive monocyte/macrophages and CD8-positive cells into glomeruli (111). A similar protection was observed in a nondiabetic glomerulosclerotic rat model made by 5/6 nephrectomy. In this study, troglitazone treatment reduced albuminuria, serum creatinine level, and glomerulosclerosis through decreasing glomerular cell proliferation, in parallel with decreased mRNA expression of p21 and p27 (50). The renoprotective effect of PPARγ agonists on renal cell carcinoma and glomerular fibrosis seems to share a similar pathway involving cell cycle arrest, thereby inhibiting cell proliferation.

Interestingly, endotoxin (lipopolysaccharide, LPS) can protect the kidney against ischemia/reperfusion-induced renal injury by inducing endogenous ligands of PPARγ such as lysophosphatidic acid and 15d-PGJ2, which could be abolished by the selective PPARγ antagonist GW9662 (112). The PPARγ endogenous ligand 15d-PGJ2 can protect renal function in acute renal failure caused by ischemia/reperfusion (113) or in multiple organ failure caused by endotoxin (114). As well, pretreating rats with TZD decreased cell apoptosis in injured kidney induced by ischemia-reperfusion by inducing hepatocyte growth factor (51). In addition, PPARγ mRNA and protein levels were reduced in rats with glycerol-induced acute renal failure. When PPARγ expression was restored by the PPARγ inducer ciglitazone, the renal dysfunction was markedly ameliorated (115, 116).

Finally, it should be mentioned that TZD treatment can cause severe side effects, such as weight gain, fluid retention, and increased cardiovascular risk (13, 14, 48). Fluid retention has been found to be caused by the

up-regulation of one PPARγ target gene, epithelial Na⁺ channel which is located in the collecting duct and mediates Na⁺ reabsorption. The collecting duct-specific diuretic amiloride can block this pathway and might provide one potential specific therapy (13, 117). In addition, extrarenal mechanisms are also involved in TZD-induced fluid retention. As discussed above, PPARγ can lower blood pressure, which may contribute to reduced water excretion (118). The vasodepressor action of TZD demonstrated by using human arterial resistance vessels could cause fluid retention as well (119). Moreover, the altered endothelial permeability, interstitial ion transport, and sympathetic nervous system activity have been reported to be associated with the development of edema following TZD treatment (120).

7. CONCLUSION

PPARs are transcription factors and nuclear receptors. They are widely expressed throughout the body and differentially located in the kidney. Activation of the three PPAR isoforms can result in distinct but overlapping biological processes. Through both indirect systemic effects and direct renal actions, agonists of PPARs hold great promise for treatment of diabetic nephropathy, glomerulonephritis, acute renal failure and chronic renal disease. PPARγ could also represent a therapeutic target for renal cell carcinoma. However, before considering a translational approach, the benefits/risks of using PPAR agonists should be carefully evaluated. Increasing reports suggest that PPARα and PPARγ agonists may cause severe undesirable effects. Thus, caution should be taken in use of these agonists in clinical therapy for diabetes and renal diseases.

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9. REFERENCES

1. Pisoni R., C. Aros, P. Ruggenenti and G. Remuzzi: Mechanisms of progression of chronic renal disease. *Saudi J Kidney Dis Transpl*, 13, 250-6 (2002)

2. Guan Y.: Peroxisome proliferator-activated receptor family and its relationship to renal complications of the metabolic syndrome. *J Am Soc Nephrol*, 15, 2801-15 (2004)
3. Guan Y. and M. D. Breyer: Peroxisome proliferator-activated receptors (PPARs): novel therapeutic targets in renal disease. *Kidney Int*, 60, 14-30 (2001)
4. Committee N. R. N.: A unified nomenclature system for the nuclear receptor superfamily. *Cell*, 97, 161-3 (1999)
5. Mukherjee R., L. Jow, D. Noonan and D. P. McDonnell: Human and rat peroxisome proliferator activated receptors (PPARs) demonstrate similar tissue distribution but different responsiveness to PPAR activators. *J Steroid Biochem Mol Biol*, 51, 157-66 (1994)
6. Bookout A. L., Y. Jeong, M. Downes, R. T. Yu, R. M. Evans and D. J. Mangelsdorf: Anatomical profiling of nuclear receptor expression reveals a hierarchical transcriptional network. *Cell*, 126, 789-99 (2006)
7. Di Paola R. and S. Cuzzocrea: Peroxisome proliferator-activated receptors ligands and ischemia-reperfusion injury. *Naumyn Schmiedebergs Arch Pharmacol*, 375, 157-75 (2007)
8. Peters J. M., N. Hennuyer, B. Staels, J. C. Fruchart, C. Fievet, F. J. Gonzalez and J. Auwerx: Alterations in lipoprotein metabolism in peroxisome proliferator-activated receptor alpha-deficient mice. *J Biol Chem*, 272, 27307-12 (1997)
9. Wang Y. X., C. H. Lee, S. Tiep, R. T. Yu, J. Ham, H. Kang and R. M. Evans: Peroxisome-proliferator-activated receptor delta activates fat metabolism to prevent obesity. *Cell*, 113, 159-70 (2003)
10. Hao C. M., R. Redha, J. Morrow and M. D. Breyer: Peroxisome proliferator-activated receptor delta activation promotes cell survival following hypertonic stress. *J Biol Chem*, 277, 21341-5 (2002)
11. Gurnell M., J. M. Wentworth, M. Agostini, M. Adams, T. N. Collingwood, C. Provenzano, P. O. Browne, O. Rajanayagam, T. P. Burris, J. W. Schwabe, M. A. Lazar and V. K. Chatterjee: A dominant-negative peroxisome proliferator-activated receptor gamma (PPARgamma) mutant is a constitutive repressor and inhibits PPARgamma-mediated adipogenesis. *J Biol Chem*, 275, 5754-9 (2000)
12. Nissen S. E., S. J. Nicholls, K. Wolski, D. C. Howey, E. McErlean, M. D. Wang, E. V. Gomez and J. M. Russo: Effects of a potent and selective PPAR-alpha agonist in patients with atherogenic dyslipidemia or hypercholesterolemia: two randomized controlled trials. *Jama*, 297, 1362-73 (2007)
13. Guan Y., C. Hao, D. R. Cha, R. Rao, W. Lu, D. E. Kohan, M. A. Magnuson, R. Redha, Y. Zhang and M. D. Breyer: Thiazolidinediones expand body fluid volume through PPARgamma stimulation of ENaC-mediated renal salt absorption. *Nat Med*, 11, 861-6 (2005)
14. Nissen S. E. and K. Wolski: Effect of rosiglitazone on the risk of myocardial infarction and death from cardiovascular causes. *N Engl J Med*, 356, 2457-71 (2007)
15. Guan Y., Y. Zhang, L. Davis and M. D. Breyer: Expression of peroxisome proliferator-activated receptors in urinary tract of rabbits and humans. *Am J Physiol*, 273, F1013-22 (1997)
16. Ruan X. Z., J. F. Moorhead, R. Fernando, D. C. Wheeler, S. H. Powis and Z. Varghese: PPAR agonists protect mesangial cells from interleukin 1beta-induced intracellular lipid accumulation by activating the ABCA1 cholesterol efflux pathway. *J Am Soc Nephrol*, 14, 593-600 (2003)
17. Yang T., D. E. Michele, J. Park, A. M. Smart, Z. Lin, F. C. Brosius, 3rd, J. B. Schnermann and J. P. Briggs: Expression of peroxisomal proliferator-activated receptors and retinoid X receptors in the kidney. *Am J Physiol*, 277, F966-73 (1999)
18. Guan Y., Y. Zhang, A. Schneider, L. Davis, R. M. Breyer and M. D. Breyer: Peroxisome proliferator-activated receptor-gamma activity is associated with renal microvasculature. *Am J Physiol Renal Physiol*, 281, F1036-46 (2001)
19. Park C. W., Y. Zhang, X. Zhang, J. Wu, L. Chen, D. R. Cha, D. Su, M. T. Hwang, X. Fan, L. Davis, G. Striker, F. Zheng, M. Breyer and Y. Guan: PPARalpha agonist fenofibrate improves diabetic nephropathy in db/db mice. *Kidney Int*, 69, 1511-7 (2006)
20. Kasiske B. L., M. P. O'Donnell, W. J. Garvis and W. F. Keane: Pharmacologic treatment of

hyperlipidemia reduces glomerular injury in rat 5/6 nephrectomy model of chronic renal failure. *Circ Res*, 62, 367-74 (1988)

21. Li S., N. Gokden, M. D. Okusa, R. Bhatt and D. Portilla: Anti-inflammatory effect of fibrate protects from cisplatin-induced ARF. *Am J Physiol Renal Physiol*, 289, F469-80 (2005)

22. Portilla D., G. Dai, T. McClure, L. Bates, R. Kurten, J. Megyesi, P. Price and S. Li: Alterations of PPARalpha and its coactivator PGC-1 in cisplatin-induced acute renal failure. *Kidney Int*, 62, 1208-18 (2002)

23. Idzior-Walus B., J. Sieradzki, W. Rostworowski, A. Zdzenicka, E. Kawalec, J. Wojcik, A. Zarnecki and G. Blane: Effects of comiconised fenofibrate on lipid and insulin sensitivity in patients with polymetabolic syndrome X. *Eur J Clin Invest*, 30, 871-8 (2000)

24. Koh E. H., M. S. Kim, J. Y. Park, H. S. Kim, J. Y. Youn, H. S. Park, J. H. Youn and K. U. Lee: Peroxisome proliferator-activated receptor (PPAR)-alpha activation prevents diabetes in OLETF rats: comparison with PPAR-gamma activation. *Diabetes*, 52, 2331-7 (2003)

25. Lee W. J., M. Kim, H. S. Park, H. S. Kim, M. J. Jeon, K. S. Oh, E. H. Koh, J. C. Won, M. S. Kim, G. T. Oh, M. Yoon, K. U. Lee and J. Y. Park: AMPK activation increases fatty acid oxidation in skeletal muscle by activating PPARalpha and PGC-1. *Biochem Biophys Res Commun*, 340, 291-5 (2006)

26. Chalkley S. M., M. Hettiarachchi, D. J. Chisholm and E. W. Kraegen: Five-hour fatty acid elevation increases muscle lipids and impairs glycogen synthesis in the rat. *Metabolism*, 47, 1121-6 (1998)

27. Crabb D. W., A. Galli, M. Fischer and M. You: Molecular mechanisms of alcoholic fatty liver: role of peroxisome proliferator-activated receptor alpha. *Alcohol*, 34, 35-8 (2004)

28. Mishra R., S. N. Emancipator, C. Miller, T. Kern and M. S. Simonson: Adipose differentiation-related protein and regulators of lipid homeostasis identified by gene expression profiling in the murine db/db diabetic kidney. *Am J Physiol Renal Physiol*, 286, F913-21 (2004)

29. Park C. W., H. W. Kim, S. H. Ko, H. W. Chung, S. W. Lim, C. W. Yang, Y. S. Chang, A.

Sugawara, Y. Guan and M. D. Breyer: Accelerated diabetic nephropathy in mice lacking the peroxisome proliferator-activated receptor alpha. *Diabetes*, 55, 885-93 (2006)

30. Tordjman K., C. Bernal-Mizrachi, L. Zemani, S. Weng, C. Feng, F. Zhang, T. C. Leone, T. Coleman, D. P. Kelly and C. F. Semenkovich: PPARalpha deficiency reduces insulin resistance and atherosclerosis in apoE-null mice. *J Clin Invest*, 107, 1025-34 (2001)

31. Guerre-Millo M., C. Rouault, P. Poulain, J. Andre, V. Poitout, J. M. Peters, F. J. Gonzalez, J. C. Fruchart, G. Reach and B. Staels: PPAR-alpha-null mice are protected from high-fat diet-induced insulin resistance. *Diabetes*, 50, 2809-14 (2001)

32. Cha D. R., J. Y. Han, D. M. Su, Y. Zhang, X. Fan, M. D. Breyer and Y. Guan: Peroxisome proliferator-activated receptor-alpha deficiency protects aged mice from insulin resistance induced by high-fat diet. *Am J Nephrol*, 27, 479-82 (2007)

33. Li S., P. Wu, P. Yarlagadda, N. M. Vadjunec, A. D. Proia, R. A. Harris and D. Portilla: PPAR alpha ligand protects during cisplatin-induced acute renal failure by preventing inhibition of renal FAO and PDC activity. *Am J Physiol Renal Physiol*, 286, F572-80 (2004)

34. Li S., A. Basnakian, R. Bhatt, J. Megyesi, N. Gokden, S. V. Shah and D. Portilla: PPAR-alpha ligand ameliorates acute renal failure by reducing cisplatin-induced increased expression of renal endonuclease G. *Am J Physiol Renal Physiol*, 287, F990-8 (2004)

35. Baud L. and E. Letavernier: PPARalpha contributes to tubular protection. *J Am Soc Nephrol*, 18, 3017-8 (2007)

36. Kamijo Y., K. Hora, K. Kono, K. Takahashi, M. Higuchi, T. Ehara, K. Kiyosawa, H. Shigematsu, F. J. Gonzalez and T. Aoyama: PPARalpha protects proximal tubular cells from acute fatty acid toxicity. *J Am Soc Nephrol*, 18, 3089-100 (2007)

37. Portilla D., G. Dai, J. M. Peters, F. J. Gonzalez, M. D. Crew and A. D. Proia: Etomoxir-induced PPARalpha-modulated enzymes protect during acute renal failure. *Am J Physiol Renal Physiol*, 278, F667-75 (2000)

38. Sivarajah A., P. K. Chatterjee, Y. Hattori, P. A. Brown, K. N. Stewart, Z. Todorovic, H. Mota-Filipe

and C. Thiemermann: Agonists of peroxisome-proliferator activated receptor-alpha (clofibrate and WY14643) reduce renal ischemia/reperfusion injury in the rat. *Med Sci Monit*, 8, BR532-9 (2002)

39. Murai T., T. Yamada, T. Miida, K. Arai, N. Endo and T. Hanyu: Fenofibrate inhibits reactive amyloidosis in mice. *Arthritis Rheum*, 46, 1683-8 (2002)

40. Maruyama S., K. Kato, M. Kodama, S. Hirano, K. Fuse, O. Nakagawa, M. Nakazawa, T. Miida, T. Yamamoto, K. Watanabe and Y. Aizawa: Fenofibrate, a peroxisome proliferator-activated receptor alpha activator, suppresses experimental autoimmune myocarditis by stimulating the interleukin-10 pathway in rats. *J Atheroscler Thromb*, 9, 87-92 (2002)

41. Saga D., M. Sakatsume, A. Ogawa, Y. Tsubata, Y. Kaneko, T. Kuroda, F. Sato, J. Ajiro, D. Kondo, T. Miida, I. Narita and F. Gejyo: Bezafibrate suppresses rat antglomerular basement membrane crescentic glomerulonephritis. *Kidney Int*, 67, 1821-9 (2005)

42. Kamijo Y., K. Hora, T. Nakajima, K. Kono, K. Takahashi, Y. Ito, M. Higuchi, K. Kiyosawa, H. Shigematsu, F. J. Gonzalez and T. Aoyama: Peroxisome proliferator-activated receptor alpha protects against glomerulonephritis induced by long-term exposure to the plasticizer di- (2-ethylhexyl)phthalate. *J Am Soc Nephrol*, 18, 176-88 (2007)

43. Letavernier E., J. Perez, E. Joye, A. Bellocq, B. Fouqueray, J. P. Haymann, D. Heudes, W. Wahli, B. Desvergne and L. Baud: Peroxisome proliferator-activated receptor beta/delta exerts a strong protection from ischemic acute renal failure. *J Am Soc Nephrol*, 16, 2395-402 (2005)

44. Bach P. H. and T. K. Nguyen: Renal papillary necrosis--40 years on. *Toxicol Pathol*, 26, 73-91 (1998)

45. Wang Y. X., C. L. Zhang, R. T. Yu, H. K. Cho, M. C. Nelson, C. R. Bayuga-Ocampo, J. Ham, H. Kang and R. M. Evans: Regulation of muscle fiber type and running endurance by PPARdelta. *PLoS Biol*, 2, e294 (2004)

46. Escher P., O. Braissant, S. Basu-Modak, L. Michalik, W. Wahli and B. Desvergne: Rat PPARs: quantitative analysis in adult rat tissues and

regulation in fasting and refeeding. *Endocrinology*, 142, 4195-202 (2001)

47. Proctor G., T. Jiang, M. Iwahashi, Z. Wang, J. Li and M. Levi: Regulation of renal fatty acid and cholesterol metabolism, inflammation, and fibrosis in Akita and OVE26 mice with type 1 diabetes. *Diabetes*, 55, 2502-9 (2006)

48. Miyazaki Y., E. Cersosimo, C. Triplitt and R. A. DeFronzo: Rosiglitazone decreases albuminuria in type 2 diabetic patients. *Kidney Int*, 72, 1367-73 (2007)

49. Yang F. G., Z. W. Zhang, D. Q. Xin, C. J. Shi, J. P. Wu, Y. L. Guo and Y. F. Guan: Peroxisome proliferator-activated receptor gamma ligands induce cell cycle arrest and apoptosis in human renal carcinoma cell lines. *Acta Pharmacol Sin*, 26, 753-61 (2005)

50. Ma L. J., C. Marcantoni, M. F. Linton, S. Fazio and A. B. Fogo: Peroxisome proliferator-activated receptor-gamma agonist troglitazone protects against nondiabetic glomerulosclerosis in rats. *Kidney Int*, 59, 1899-910 (2001)

51. Doi S., T. Masaki, T. Arakawa, S. Takahashi, T. Kawai, A. Nakashima, T. Naito, N. Kohno and N. Yorioka: Protective effects of peroxisome proliferator-activated receptor gamma ligand on apoptosis and hepatocyte growth factor induction in renal ischemia-reperfusion injury. *Transplantation*, 84, 207-13 (2007)

52. Barroso I., M. Gurnell, V. E. Crowley, M. Agostini, J. W. Schwabe, M. A. Soos, G. L. Maslen, T. D. Williams, H. Lewis, A. J. Schafer, V. K. Chatterjee and S. O'Rahilly: Dominant negative mutations in human PPARgamma associated with severe insulin resistance, diabetes mellitus and hypertension. *Nature*, 402, 880-3 (1999)

53. Way J. M., W. W. Harrington, K. K. Brown, W. K. Gottschalk, S. S. Sundseth, T. A. Mansfield, R. K. Ramachandran, T. M. Willson and S. A. Kliewer: Comprehensive messenger ribonucleic acid profiling reveals that peroxisome proliferator-activated receptor gamma activation has coordinate effects on gene expression in multiple insulin-sensitive tissues. *Endocrinology*, 142, 1269-77 (2001)

54. Peraldi P., M. Xu and B. M. Spiegelman: Thiazolidinediones block tumor necrosis factor-

alpha-induced inhibition of insulin signaling. *J Clin Invest*, 100, 1863-9 (1997)

55. Zhao S. P. and D. Q. Zhang: Atorvastatin reduces interleukin-6 plasma concentration and adipocyte secretion of hypercholesterolemic rabbits. *Clin Chim Acta*, 336, 103-8 (2003)

56. Steppan C. M., S. T. Bailey, S. Bhat, E. J. Brown, R. R. Banerjee, C. M. Wright, H. R. Patel, R. S. Ahima and M. A. Lazar: The hormone resistin links obesity to diabetes. *Nature*, 409, 307-12 (2001)

57. Yang W. S., C. Y. Jeng, T. J. Wu, S. Tanaka, T. Funahashi, Y. Matsuzawa, J. P. Wang, C. L. Chen, T. Y. Tai and L. M. Chuang: Synthetic peroxisome proliferator-activated receptor-gamma agonist, rosiglitazone, increases plasma levels of adiponectin in type 2 diabetic patients. *Diabetes Care*, 25, 376-80 (2002)

58. Choi K. C., O. H. Ryu, K. W. Lee, H. Y. Kim, J. A. Seo, S. G. Kim, N. H. Kim, D. S. Choi, S. H. Baik and K. M. Choi: Effect of PPAR-alpha and -gamma agonist on the expression of visfatin, adiponectin, and TNF-alpha in visceral fat of OLETF rats. *Biochem Biophys Res Commun*, 336, 747-53 (2005)

59. Kim E. K., K. B. Kwon, B. S. Koo, M. J. Han, M. Y. Song, E. K. Song, M. K. Han, J. W. Park, D. G. Ryu and B. H. Park: Activation of peroxisome proliferator-activated receptor-gamma protects pancreatic beta-cells from cytokine-induced cytotoxicity via NF kappaB pathway. *Int J Biochem Cell Biol*, 39, 1260-75 (2007)

60. Jiang C., A. T. Ting and B. Seed: PPAR-gamma agonists inhibit production of monocyte inflammatory cytokines. *Nature*, 391, 82-6 (1998)

61. Ricote M., A. C. Li, T. M. Willson, C. J. Kelly and C. K. Glass: The peroxisome proliferator-activated receptor-gamma is a negative regulator of macrophage activation. *Nature*, 391, 79-82 (1998)

62. Cai D., M. Yuan, D. F. Frantz, P. A. Melendez, L. Hansen, J. Lee and S. E. Shoelson: Local and systemic insulin resistance resulting from hepatic activation of IKK-beta and NF-kappaB. *Nat Med*, 11, 183-90 (2005)

63. Duan S. Z., C. Y. Ivashchenko, S. E. Whitesall, L. G. D'Alecy, D. C. Duquaine, F. C. Brosius, 3rd, F. J. Gonzalez, C. Vinson, M. A. Pierre, D. S. Milstone

and R. M. Mortensen: Hypotension, lipodystrophy, and insulin resistance in generalized PPARgamma-deficient mice rescued from embryonic lethality. *J Clin Invest*, 117, 812-22 (2007)

64. Todorov V. T., M. Desch, N. Schmitt-Nilsson, A. Todorova and A. Kurtz: Peroxisome proliferator-activated receptor-gamma is involved in the control of renin gene expression. *Hypertension*, 50, 939-44 (2007)

65. Diep Q. N., M. El Mabrouk, J. S. Cohn, D. Endemann, F. Amiri, A. Virdis, M. F. Neves and E. L. Schiffrin: Structure, endothelial function, cell growth, and inflammation in blood vessels of angiotensin II-infused rats: role of peroxisome proliferator-activated receptor-gamma. *Circulation*, 105, 2296-302 (2002)

66. Benkirane K., E. C. Viel, F. Amiri and E. L. Schiffrin: Peroxisome proliferator-activated receptor gamma regulates angiotensin II-stimulated phosphatidylinositol 3-kinase and mitogen-activated protein kinase in blood vessels *in vivo*. *Hypertension*, 47, 102-8 (2006)

67. Chen K., J. Chen, D. Li, X. Zhang and J. L. Mehta: Angiotensin II regulation of collagen type I expression in cardiac fibroblasts: modulation by PPAR-gamma ligand pioglitazone. *Hypertension*, 44, 655-61 (2004)

68. Kubota N., Y. Terauchi, H. Miki, H. Tamemoto, T. Yamauchi, K. Komeda, S. Satoh, R. Nakano, C. Ishii, T. Sugiyama, K. Eto, Y. Tsubamoto, A. Okuno, K. Murakami, H. Sekihara, G. Hasegawa, M. Naito, Y. Toyoshima, S. Tanaka, K. Shiota, T. Kitamura, T. Fujita, O. Ezaki, S. Aizawa, T. Kadowaki and *et al.*: PPAR gamma mediates high-fat diet-induced adipocyte hypertrophy and insulin resistance. *Mol Cell*, 4, 597-609 (1999)

69. Miles P. D., Y. Barak, W. He, R. M. Evans and J. M. Olefsky: Improved insulin-sensitivity in mice heterozygous for PPAR-gamma deficiency. *J Clin Invest*, 105, 287-92 (2000)

70. Yamauchi T., H. Waki, J. Kamon, K. Murakami, K. Motojima, K. Komeda, H. Miki, N. Kubota, Y. Terauchi, A. Tsuchida, N. Tsuboyama-Kasaoka, N. Yamauchi, T. Ide, W. Hori, S. Kato, M. Fukayama, Y. Akanuma, O. Ezaki, A. Itai, R. Nagai, S. Kimura, K. Tobe, H. Kagechika, K. Shudo and T. Kadowaki: Inhibition of RXR and PPARgamma ameliorates

diet-induced obesity and type 2 diabetes. *J Clin Invest*, 108, 1001-13 (2001)

71. Bakris G., G. Viberti, W. M. Weston, M. Heise, L. E. Porter and M. I. Freed: Rosiglitazone reduces urinary albumin excretion in type II diabetes. *J Hum Hypertens*, 17, 7-12 (2003)

72. Nakamura T., C. Ushiyama, S. Suzuki, N. Shimada, K. Sekizuka, L. Ebihara and H. Koide: Effect of troglitazone on urinary albumin excretion and serum type IV collagen concentrations in Type 2 diabetic patients with microalbuminuria or macroalbuminuria. *Diabet Med*, 18, 308-13 (2001)

73. Lebovitz H. E., J. F. Dole, R. Patwardhan, E. B. Rappaport and M. I. Freed: Rosiglitazone monotherapy is effective in patients with type 2 diabetes. *J Clin Endocrinol Metab*, 86, 280-8 (2001)

74. Nakamura T., C. Ushiyama, N. Shimada, K. Hayashi, I. Ebihara and H. Koide: Comparative effects of pioglitazone, glibenclamide, and voglibose on urinary endothelin-1 and albumin excretion in diabetes patients. *J Diabetes Complications*, 14, 250-4 (2000)

75. Imano E., T. Kanda, Y. Nakatani, T. Nishida, K. Arai, M. Motomura, Y. Kajimoto, Y. Yamasaki and M. Hori: Effect of troglitazone on microalbuminuria in patients with incipient diabetic nephropathy. *Diabetes Care*, 21, 2135-9 (1998)

76. Wolffenbuttel B. H., R. Gomis, S. Squatrito, N. P. Jones and R. N. Patwardhan: Addition of low-dose rosiglitazone to sulphonylurea therapy improves glycaemic control in Type 2 diabetic patients. *Diabet Med*, 17, 40-7 (2000)

77. Fujii M., R. Takemura, M. Yamaguchi, G. Hasegawa, H. Shigeta, K. Nakano and M. Kondo: Troglitazone (CS-045) ameliorates albuminuria in streptozotocin-induced diabetic rats. *Metabolism*, 46, 981-3 (1997)

78. Buckingham R. E., K. A. Al-Barazani, C. D. Toseland, M. Slaughter, S. C. Connor, A. West, B. Bond, N. C. Turner and J. C. Clapham: Peroxisome proliferator-activated receptor-gamma agonist, rosiglitazone, protects against nephropathy and pancreatic islet abnormalities in Zucker fatty rats. *Diabetes*, 47, 1326-34 (1998)

79. Isshiki K., M. Haneda, D. Koya, S. Maeda, T. Sugimoto and R. Kikkawa: Thiazolidinedione

compounds ameliorate glomerular dysfunction independent of their insulin-sensitizing action in diabetic rats. *Diabetes*, 49, 1022-32 (2000)

80. Baylis C., E. A. Atzpodien, G. Freshour and K. Engels: Peroxisome proliferator-activated receptor (gamma) agonist provides superior renal protection versus angiotensin-converting enzyme inhibition in a rat model of type 2 diabetes with obesity. *J Pharmacol Exp Ther*, 307, 854-60 (2003)

81. McCarthy K. J., R. E. Routh, W. Shaw, K. Walsh, T. C. Welbourne and J. H. Johnson: Troglitazone halts diabetic glomerulosclerosis by blockade of mesangial expansion. *Kidney Int*, 58, 2341-50 (2000)

82. Ohga S., K. Shikata, K. Yozai, S. Okada, D. Ogawa, H. Usui, J. Wada, Y. Shikata and H. Makino: Thiazolidinedione ameliorates renal injury in experimental diabetic rats through anti-inflammatory effects mediated by inhibition of NF-kappaB activation. *Am J Physiol Renal Physiol*, 292, F1141-50 (2007)

83. Matsui T., S. Yamagishi, S. Ueda, K. Nakamura, T. Imaizumi, M. Takeuchi and H. Inoue: Telmisartan, an angiotensin II type 1 receptor blocker, inhibits advanced glycation end-product (AGE)-induced monocyte chemoattractant protein-1 expression in mesangial cells through downregulation of receptor for AGEs via peroxisome proliferator-activated receptor-gamma activation. *J Int Med Res*, 35, 482-9 (2007)

84. Tang S. C., J. C. Leung, L. Y. Chan, A. W. Tsang and K. N. Lai: Activation of tubular epithelial cells in diabetic nephropathy and the role of the peroxisome proliferator-activated receptor-gamma agonist. *J Am Soc Nephrol*, 17, 1633-43 (2006)

85. Zheng F., A. Fornoni, S. J. Elliot, Y. Guan, M. D. Breyer, L. J. Striker and G. E. Striker: Upregulation of type I collagen by TGF-beta in mesangial cells is blocked by PPARgamma activation. *Am J Physiol Renal Physiol*, 282, F639-48 (2002)

86. Guo B., D. Koya, M. Isono, T. Sugimoto, A. Kashiwagi and M. Haneda: Peroxisome proliferator-activated receptor-gamma ligands inhibit TGF-beta 1-induced fibronectin expression in glomerular mesangial cells. *Diabetes*, 53, 200-8 (2004)

87. Asano T., M. Wakisaka, M. Yoshinari, K. Iino, K. Sonoki, M. Iwase and M. Fujishima: Peroxisome

- proliferator-activated receptor gamma1 (PPARgamma1) expresses in rat mesangial cells and PPARgamma agonists modulate its differentiation. *Biochim Biophys Acta*, 1497, 148-54 (2000)
88. Chang P. C., T. H. Chen, C. J. Chang, C. C. Hou, P. Chan and H. M. Lee: Advanced glycosylation end products induce inducible nitric oxide synthase (iNOS) expression via a p38 MAPK-dependent pathway. *Kidney Int*, 65, 1664-75 (2004)
89. Onozaki A., S. Midorikawa, H. Sanada, Y. Hayashi, T. Baba, T. Katoh and T. Watanabe: Rapid change of glucose concentration promotes mesangial cell proliferation via VEGF: inhibitory effects of thiazolidinedione. *Biochem Biophys Res Commun*, 317, 24-9 (2004)
90. Okada T., J. Wada, K. Hida, J. Eguchi, I. Hashimoto, M. Baba, A. Yasuhara, K. Shikata and H. Makino: Thiazolidinediones ameliorate diabetic nephropathy via cell cycle-dependent mechanisms. *Diabetes*, 55, 1666-77 (2006)
91. Panchapakesan U., C. A. Pollock and X. M. Chen: The effect of high glucose and PPAR-gamma agonists on PPAR-gamma expression and function in HK-2 cells. *Am J Physiol Renal Physiol*, 287, F528-34 (2004)
92. Zafiriou S., S. R. Stanners, S. Saad, T. S. Polhill, P. Poronnik and C. A. Pollock: Pioglitazone inhibits cell growth and reduces matrix production in human kidney fibroblasts. *J Am Soc Nephrol*, 16, 638-45 (2005)
93. Panchapakesan U., S. Sumual, C. A. Pollock and X. Chen: PPARgamma agonists exert antifibrotic effects in renal tubular cells exposed to high glucose. *Am J Physiol Renal Physiol*, 289, F1153-8 (2005)
94. Li Y., X. Wen, B. C. Spataro, K. Hu, C. Dai and Y. Liu: hepatocyte growth factor is a downstream effector that mediates the antifibrotic action of peroxisome proliferator-activated receptor-gamma agonists. *J Am Soc Nephrol*, 17, 54-65 (2006)
95. Calkin A. C., S. Giunti, K. A. Jandeleit-Dahm, T. J. Allen, M. E. Cooper and M. C. Thomas: PPAR-alpha and -gamma agonists attenuate diabetic kidney disease in the apolipoprotein E knockout mouse. *Nephrol Dial Transplant*, 21, 2399-405 (2006)
96. Cha D. R., X. Zhang, Y. Zhang, J. Wu, D. Su, J. Y. Han, X. Fang, B. Yu, M. D. Breyer and Y. Guan: Peroxisome proliferator activated receptor alpha/gamma dual agonist tesaglitazar attenuates diabetic nephropathy in db/db mice. *Diabetes*, 56, 2036-45 (2007)
97. Buse J. B., C. J. Rubin, R. Frederich, K. Viraswami-Appanna, K. C. Lin, R. Montoro, G. Shockey and J. A. Davidson: Muraglitazar, a dual (alpha/gamma) PPAR activator: a randomized, double-blind, placebo-controlled, 24-week monotherapy trial in adult patients with type 2 diabetes. *Clin Ther*, 27, 1181-95 (2005)
98. Kendall D. M., C. J. Rubin, P. Mohideen, J. M. Ledezine, R. Belder, J. Gross, P. Norwood, M. O'Mahony, K. Sall, G. Sloan, A. Roberts, F. T. Fiedorek and R. A. DeFronzo: Improvement of glycemic control, triglycerides, and HDL cholesterol levels with muraglitazar, a dual (alpha/gamma) peroxisome proliferator-activated receptor activator, in patients with type 2 diabetes inadequately controlled with metformin monotherapy: A double-blind, randomized, pioglitazone-comparative study. *Diabetes Care*, 29, 1016-23 (2006)
99. Brand C. L., J. Sturis, C. F. Gotfredsen, J. Fleckner, C. Fledelius, B. F. Hansen, B. Andersen, J. M. Ye, P. Sauerberg and K. Wassermann: Dual PPARalpha /gamma activation provides enhanced improvement of insulin sensitivity and glycemic control in ZDF rats. *Am J Physiol Endocrinol Metab*, 284, E841-54 (2003)
100. Harrity T., D. Farrelly, A. Tieman, C. Chu, L. Kunselman, L. Gu, R. Ponticello, M. Cap, F. Qu, C. Shao, W. Wang, H. Zhang, W. Fenderson, S. Chen, P. Devasthale, Y. Jeon, R. Seethala, W. P. Yang, J. Ren, M. Zhou, D. Ryono, S. Biller, K. A. Mookhtiar, J. Wetterau, R. Gregg, P. T. Cheng and N. Hariharan: Muraglitazar, a novel dual (alpha/gamma) peroxisome proliferator-activated receptor activator, improves diabetes and other metabolic abnormalities and preserves beta-cell function in db/db mice. *Diabetes*, 55, 240-8 (2006)
101. Pickavance L. C., C. L. Brand, K. Wassermann and J. P. Wilding: The dual PPARalpha/gamma agonist, ragaglitazar, improves insulin sensitivity and metabolic profile equally with pioglitazone in diabetic and dietary obese ZDF rats. *Br J Pharmacol*, 144, 308-16 (2005)

102. Ye J. M., M. A. Iglesias, D. G. Watson, B. Ellis, L. Wood, P. B. Jensen, R. V. Sorensen, P. J. Larsen, G. J. Cooney, K. Wassermann and E. W. Kraegen: PPARalpha /gamma ragaglitazar eliminates fatty liver and enhances insulin action in fat-fed rats in the absence of hepatomegaly. *Am J Physiol Endocrinol Metab*, 284, E531-40 (2003)
103. Mamnoor P. K., P. Hegde, S. R. Datla, R. K. Damarla, R. Rajagopalan and R. Chakrabarti: Antihypertensive effect of ragaglitazar: a novel PPARalpha and gamma dual activator. *Pharmacol Res*, 54, 129-35 (2006)
104. Chakrabarti R., P. Misra, R. K. Vikramadithyan, M. Premkumar, J. Hiriyani, S. R. Datla, R. K. Damarla, J. Suresh and R. Rajagopalan: Antidiabetic and hypolipidemic potential of DRF 2519--a dual activator of PPAR-alpha and PPAR-gamma. *Eur J Pharmacol*, 491, 195-206 (2004)
105. Verreth W., J. Ganame, A. Mertens, H. Bernar, M. C. Herregods and P. Holvoet: Peroxisome proliferator-activated receptor-alpha,gamma-agonist improves insulin sensitivity and prevents loss of left ventricular function in obese dyslipidemic mice. *Arterioscler Thromb Vasc Biol*, 26, 922-8 (2006)
106. Hegarty B. D., S. M. Furler, N. D. Oakes, E. W. Kraegen and G. J. Cooney: Peroxisome proliferator-activated receptor (PPAR) activation induces tissue-specific effects on fatty acid uptake and metabolism *in vivo*--a study using the novel PPARalpha/gamma agonist tesaglitazar. *Endocrinology*, 145, 3158-64 (2004)
107. Inoue K., Y. Kawahito, Y. Tsubouchi, M. Kohno, R. Yoshimura, T. Yoshikawa and H. Sano: Expression of peroxisome proliferator-activated receptor gamma in renal cell carcinoma and growth inhibition by its agonists. *Biochem Biophys Res Commun*, 287, 727-32 (2001)
108. Yang F. G., Z. W. Zhang, D. Q. Xin, C. J. Shi, X. Q. Wu, W. J. Liu, Y. L. Guo and J. P. Wu: (Peroxisome proliferator-activated receptor-gamma ligand troglitazone induces apoptosis in renal cell carcinoma). *Beijing Da Xue Xue Bao*, 36, 173-6 (2004)
109. Yuan J., A. Takahashi, N. Masumori, N. Itoh and T. Tsukamoto: Peroxisome proliferator-activated receptor gamma is frequently underexpressed in renal cell carcinoma. *Int J Urol*, 13, 265-70 (2006)
110. Gupta R. A., D. Wang, S. Katkuri, H. Wang, S. K. Dey and R. N. DuBois: Activation of nuclear hormone receptor peroxisome proliferator-activated receptor-delta accelerates intestinal adenoma growth. *Nat Med*, 10, 245-7 (2004)
111. Haraguchi K., H. Shimura and T. Onaya: Suppression of experimental crescentic glomerulonephritis by peroxisome proliferator-activated receptor (PPAR)gamma activators. *Clin Exp Nephrol*, 7, 27-32 (2003)
112. Collino M., N. S. Patel, K. M. Lawrence, M. Collin, D. S. Latchman, M. M. Yaqoob and C. Thiemermann: The selective PPARgamma antagonist GW9662 reverses the protection of LPS in a model of renal ischemia-reperfusion. *Kidney Int*, 68, 529-36 (2005)
113. Chatterjee P. K., N. S. Patel, S. Cuzzocrea, P. A. Brown, K. N. Stewart, H. Mota-Filipe, D. Britti, W. Eberhardt, J. Pfeilschifter and C. Thiemermann: The cyclopentenone prostaglandin 15-deoxy-Delta (12,14)-prostaglandin J2 ameliorates ischemic acute renal failure. *Cardiovasc Res*, 61, 630-43 (2004)
114. Collin M., N. S. Patel, L. Dugo and C. Thiemermann: Role of peroxisome proliferator-activated receptor-gamma in the protection afforded by 15-deoxydelta12,14 prostaglandin J2 against the multiple organ failure caused by endotoxin. *Crit Care Med*, 32, 826-31 (2004)
115. Newaz M., Z. Yousefipour and A. Oyekan: Role of PPAR-gamma on the pathogenesis and vascular changes in glycerol-induced acute renal failure. *Pharmacol Res*, 54, 234-40 (2006)
116. Yousefipour Z., H. Hercule, L. Truong, A. Oyekan and M. Newaz: Ciglitazone, a peroxisome proliferator-activated receptor gamma inducer, ameliorates renal preglomerular production and activity of angiotensin II and thromboxane A2 in glycerol-induced acute renal failure. *J Pharmacol Exp Ther*, 322, 461-8 (2007)
117. Zhang H., A. Zhang, D. E. Kohan, R. D. Nelson, F. J. Gonzalez and T. Yang: Collecting duct-specific deletion of peroxisome proliferator-activated receptor gamma blocks thiazolidinedione-induced fluid retention. *Proc Natl Acad Sci U S A*, 102, 9406-11 (2005)

118. Buchanan T. A., W. P. Meehan, Y. Y. Jeng, D. Yang, T. M. Chan, J. L. Nadler, S. Scott, R. K. Rude and W. A. Hsueh: Blood pressure lowering by pioglitazone. Evidence for a direct vascular effect. *J Clin Invest*, 96, 354-60 (1995)

119. Walker A. B., E. K. Naderali, P. D. Chattington, R. E. Buckingham and G. Williams: Differential vasoactive effects of the insulin sensitizers rosiglitazone (BRL 49653) and troglitazone on human small arteries *in vitro*. *Diabetes*, 47, 810-4 (1998)

120. Hosokawa M., H. Tsukada, K. Fukuda, M. Oya, M. Onomura, H. Nakamura, M. Kodama, Y. Yamada and Y. Seino: Troglitazone inhibits bicarbonate secretion in rat and human duodenum. *J Pharmacol Exp Ther*, 290, 1080-4 (1999)

Key Words

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