

## Contribution of PKB/AKT signaling to thyroid cancer

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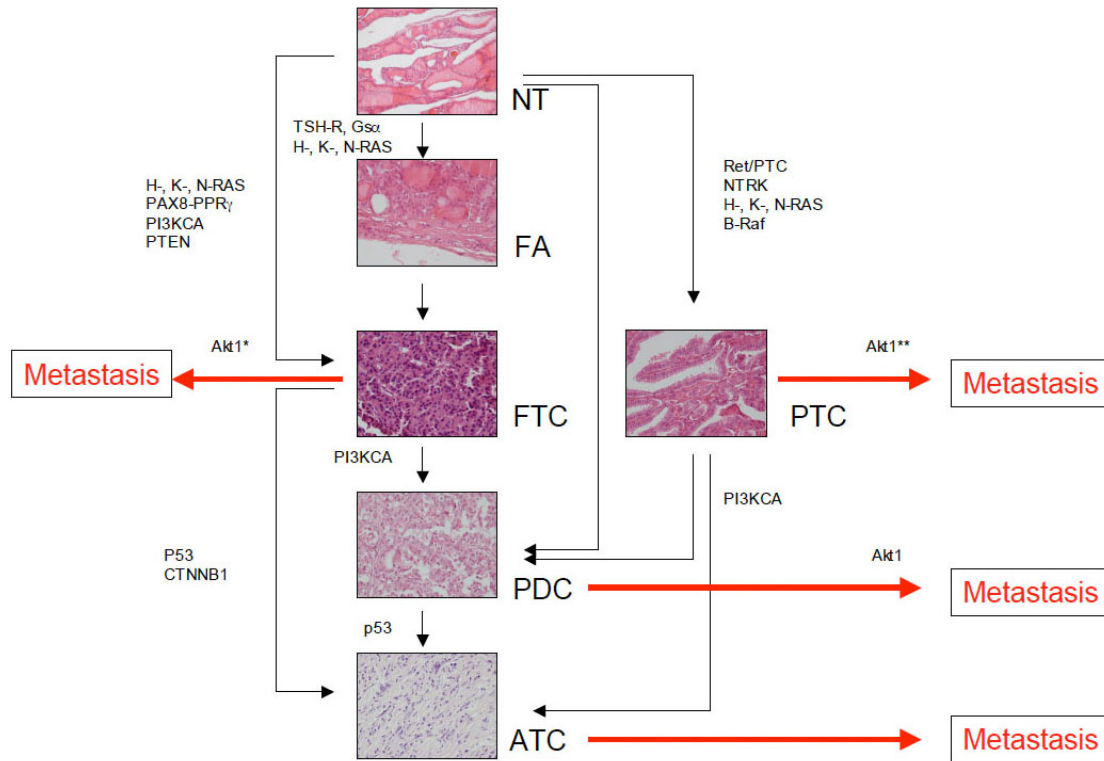
## 1. ABSTRACT

The family of serine/threonine kinases B/Akt (hereafter Akt) represents a central node in signalling pathways downstream of growth factors, cytokines, and other cellular stimuli. In mammalian cells the Akt family comprises three highly homologous members -known as Akt1/PKB $\alpha$ , Akt2/PKB $\beta$ , and Akt3/PKB $\gamma$ - that regulate several processes including cell proliferation and survival, growth and response to nutrient availability, migration, tissue invasion and angiogenesis. Aberrant activation of Akt is involved in a variety of human cancers including those arising in the thyroid gland. Here, we review the contribution of Akt-dependent pathway in the proliferation of normal thyrocytes, the different pathogenic mechanisms underlying aberrant Akt signalling in thyroid malignancies as well as the relative roles of Akt substrates that most likely contribute to the onset and/or progression of thyroid cancer. Finally, we discuss the current therapeutic strategies targeting the components of the PI3K/Akt pathway in the context of thyroid malignancy.

## 2. INTRODUCTION

Thyroid cancer is the most common malignant tumour of the endocrine system and accounts for approximately 1% of all newly diagnosed cancer cases (1, 2). The major histological types of thyroid cancer are derived from the epithelial cells that line follicles and are classified into papillary thyroid carcinoma (PTC), follicular thyroid carcinoma (FTC), poorly differentiated thyroid carcinoma (PDC) and anaplastic thyroid carcinoma (ATC) (3, 4). Medullary thyroid cancer derived from para-follicular cells is a relatively rare malignancy that will not be discussed here.

PTC is the most frequent type of thyroid malignancy, and accounts for approximately 80% of all cases (3), FTC accounts for approximately 15% of all thyroid malignancies whereas PDC and ATC are rare (<2% of all thyroid cancer). PDC and ATC can develop directly or from pre-existing well-differentiated PTC or FTC, with



**Figure 1.** Schematic stepwise model for the origin and the progression of human thyroid cancer. PTC is characterized by genetic alterations that lead to the constitutive activation of the MAPK signalling pathway: these include rearrangement of Ret/PTC, rearrangement of TRKN1, activating gain-of-function mutations of B-Raf or activating gain-of-function mutations in one of the three Ras genes. Ret rearrangements are specific for PTC. Ret/PTC rearrangements are found on average in ~20% of sporadic PTC in adult patients and is more common in patients with a history of radiation exposure (50–80%). The TRK oncogenes occurs in PTC, with a frequency of <10%. Mutations of the B-Raf gene are commonly found in PTC, PDC and ATC (45% of cases) but are rare in FTC. Virtually all mutations involve nucleotide 1799 that results in a valine-to-glutamate substitution at residue 600 (V600E) with consequent activation of B-Raf kinase. Ras mutations are relatively rare in PTC (10–20%) and are most common in FA (20–40%) and conventional FTC (40–50%). Ras mutations have been described also in 18%–27% of PDC and in 50%–60% of ATC. The PAX8/PPAR $\gamma$  rearrangement, typical of FTC, results from t(2;3) (q13,p25) translocation and leads to the fusion of the PAX8 gene, a member of the paired box (PAX) family of transcription factors, with the PPAR $\gamma$  gene that encodes a nuclear receptor protein. PAX8/PPAR $\gamma$  rearrangement occurs in in 2%–10% FA and 35% FTC. Alterations involving the PI3K/Akt signalling pathway, as well as those involving the p53 and CTNNB1 gene, are common in PDC and ATC and thus play a role particularly in later stages of tumour progression. In particular, PIK3CA mutations have been observed in PDC and ATC whereas Akt1 mutations have been observed exclusively in recurrence of PDC, Hürthle cell carcinoma (HCC) and in the follicular variant of PTC (FV-PTC) and/or in the metastatic lesions thereof. Akt1\*, mutations in the gene encoding Akt1 in HCC; Akt1\*\*, mutations in the gene encoding Akt1 in FV-PTC. It is of note that the stepwise progression of thyroid cancer presented here is essentially a hypothetical model.

PDC that can further progress into ATC (4) (Figure 1). PTC and FTC are well differentiated, indolent tumours with rather good prognosis that are generally curable with current treatments. In contrast PDC and ATC represent aggressive partially or completely undifferentiated form of thyroid cancer, respectively.

However, aggressive variants of well-differentiated thyroid carcinomas have been identified such as the tall cell variant, columnar cell variant, diffuse sclerosing variant, insular carcinoma, and Hürthle cell (oncocytic, oxyphilic) carcinomas (HCC). HCC are rare entities comprising ~5% of epithelial thyroid tumors that were previously classified as follicular carcinomas by the

World Health Organization, though they are now recognized as a distinct clinicopathological entity (5, 6). The Tall Cell Variant of papillary thyroid carcinoma (TCV-PTC), named for its distinct morphology, is characterized by tumour cells being twice as tall as they are wide (5, 6).

Patients with ATC have a mean life expectancy of few months, representing the major therapeutic challenge for thyroid cancer therapy (4, 7, 8). The main cause of thyroid cancer-related mortality is due to the surgical inoperability at diagnosis of many patients and to the frequent insensitivity exhibited by PDC and ATC patients to radioiodine treatment. Therefore, there is a compelling need for ameliorating the comprehension of

**Table 1.** Cellular models of human thyroid cancer

HISTOTYPE	CELL LINE	SEX	MUTATIONS IN ONCOGENES				
			HRAS	BRAF	PI3K	RET	TP53
	BCPAP	F		V600E		WT	Asp259Tyr
PTC	KTC-1	M	WT	V600E		WT	
	K1	M	His27His	V600E/WT	Glu549Lys	WT	Arg213Arg
	TPC1	F	His27His	WT		RET/PTC1	WT
	TT2609-CO2	M					
	FT133	M					
FTC	ML1	F					
	WRO82-1	F		V600E/WT			Pro23Leu
	8505C	F	His27His	V600E		WT	Arg248Gln
	SW1736			V600E		WT	
	Cal-62	F		WT			
	T235	F					
	T238	F					
ATC	Uth-104	F		V600E			
	ACT-1			WT			
	HTh74	F	His27His	WT		WT	WT
	KAT18			WT		WT	
	TTA1			WT			
	FRO81-2			V600E/WT		WT	
	HTh7			WT		WT	
	C643	M	Gly13Arg	WT		WT	Arg248Gln
	BHT101	F					
	KTC-2	F					

Mutations and rearrangements reported in oncogenes and tumour suppressor genes in human thyroid cancer-derived cell lines (12-14).

**Table 2.** Animal models of thyroid cancer

	MOUSE LINE	STRAIN	PATHOLOGY	METASTASIS	THYROID FUNCTION
	<i>RET/PTC1</i>	CS57BL/6J	22% with PTC	No	NA
	<i>RET/PTC1</i>	FVB/N	100% with multifocal PTC	No	Normal/hypothyroidism
	<i>RET/PTC3</i>	C3H/He	31% with PTC by 3 months	Lymph Nodes	NA
PTC	<i>BRAF<sup>V600E</sup>(Tg-BRAF2)</i>	FVB/N	93% with PTC by 3 months	Lymph Nodes	TSH increased
	<i>BRAF<sup>V600E</sup>(Tg-BRAF3)</i>	FVB/N	25% PTC at 3 months	No	TSH increased
	<i>TRK-T1</i>	B6C3F1	23% with PTC-like cancer	No	NA
	<i>H-Ras<sup>G12V</sup></i>	Mixed	3 out of 4 mice developed PTC	Unclear	Normal T4 level
	<i>Prkar1a<sup>A2/+</sup></i>	Mixed	95% with PTC-like cancer	No	NA
	<i>K-Ras<sup>G12V</sup></i>	C57BL/6J,DBA/2	2% with FTC	No	Normal
	<i>Pten<sup>-/-</sup></i>	FVB/N crossed with 129Sv	FTC in females	No	Normal TSH and T4 levels
	<i>N-Ras<sup>Glu61Lys</sup></i>	Founders crossed with C57BL/6J	11% adenoma, 29,5% FTC/mixed	Liver, lung, bone	Elevated TSH
FTC	<i>K-Ras<sup>G12D</sup> Pten<sup>-/-</sup></i>	129SV	100% FTC with local invasion	Lung	Low TSH/High T4
	<i>TRβ<sup>PPV</sup></i>	C57BL/6Jx129SV	100% FTC	Lung	Elevated TSH, T3 and T4
	<i>1b-adrenergic receptor</i>	Founders crossed with C57BL/6J	3 out of 6 lines with goiter and FTC	Lung	Elevated T4
	<i>Rap1b<sup>G12V</sup></i>	FVB(Taconic)	FTC after treatment with a goitrogen	No	Normal

Activation of TSHR or PKA is not sufficient to initiate tumourigenesis in mice. Ret/PTC rearrangement and B-Raf V600E mutation are able to initiate thyroid cancer *in vivo*. Ras activation contributes to the development of FTC and PTC. Conditional loss of Pten in the thyroid gland renders the thyrocytes highly susceptible to neoplastic transformation (255). Pten mutant mice develop diffuse goiter characterized by enlarged follicles, in the presence of normal TSH and T4 hormone levels and by 10 months of age, develop follicular adenomas. The concurrent activation of K-Ras (mutation G12D) and PI3K (Pten deletion) in thyroid cells led to aggressive, invasive and metastatic FTC (73).

thyroid tumourigenesis and for improving the treatment of patients with PDC and ATC.

The past decade has witnessed significant expansion in the understanding of the molecular basis of thyroid cancer (9). Cytogenetic and molecular analysis of human thyroid tumours have allowed the identification of several genes that contribute to the development of thyroid carcinomas and have uncovered the association between certain molecular alterations and a specific type of thyroid cancer (9, 10). The use of *in vitro* cellular models of thyroid cancer has provided invaluable tools to study the molecular mechanisms responsible for aberrant activation of intracellular signalling pathways that occurs during the malignant conversion of human thyrocytes (11, 12). More

than 30 cell lines derived from human thyroid carcinomas have been established so far (Table 1), though recent studies have reported cross-contaminations and/or misidentifications of some of the most used cell lines (13, 14) that has led to a critical revision of the literature (15). In parallel, multiple genetically modified mice that harbour thyroid-specific activated oncogenes or inactivated tumour suppressor genes have been generated with the aim to model the different types of human thyroid cancer (16). These mouse strains have significantly enhanced our understanding of the mechanisms of thyroid tumour development and dissemination (Table 2). From the use of human and experimental models, it has become apparent that the malignant transformation of the normal thyroid follicular cell involves multiple genetic events that

sequentially activate certain oncogenes (i.e. Ras, Ret/PTC, NTRK1, B-Raf, PIK3CA, Akt1) and inactivate specific tumour suppressors (i.e. p53, PTEN) (9, 10, 12). See Figure 1 for a detailed summary of the known molecular alterations found in thyroid cancer.

This review -updated to may 2010- is focused on the issues related to Akt signalling that are most relevant to thyroid tumourigenesis: 1) the role of Akt signalling in normal and transformed follicular thyroid cells; 2) the molecular mechanisms that dysregulate Akt signalling in thyroid cancer; 3) the role of Akt substrates in the malignant transformation of thyrocytes; 4) the current therapeutic strategies targeting the components of the PI3K/Akt pathway in the contest of thyroid malignancies.

## 2. PROTEIN KINASE B/Akt

The PI3K-Akt pathway is a fundamental signalling cascade through which different proliferative, survival and/or differentiative signals are funnelled from tyrosine kinase receptors in multiple cell types, including thyrocytes (17, 18) (Figures 2 and 3). The Akt kinase represents the primary downstream mediator of the effects of the PI3K pathway, and plays a central role in both normal and pathological signalling. In mammalian cells Akt comprises three highly homologous members (>80% protein sequence identity) termed Akt1/PKB $\alpha$ , Akt2/PKB $\beta$  and Akt3/PKB $\gamma$ , encoded by three different genes located on chromosomes 14q32, 19q13 and 1q43, respectively (Figure 4). Akt kinases have been included into the AGC kinase family, because they show extensive homology within their kinase domains to PKA and PKC (18, 19). Akt kinases share the same structural organization, containing an N-terminal pleckstrin homology (PH) domain, a central catalytic domain and a C-terminal regulatory region. The PH domain of Akt can bind specifically to D3-phosphorylated phosphoinositides with high affinity and mediates Akt activation (20, 21). See Figure 4 for a schematic model of Akt kinases.

Despite their sequence similarity however, Akt isoforms are functionally distinct (22), as suggested by the different phenotypes of Akt1 $^{-/-}$  (23, 24), Akt2 $^{-/-}$  (25, 26), Akt3 $^{-/-}$  (27), as well as double mutant Akt1 $^{-/-}$ /Akt2 $^{-/-}$  (28), Akt1 $^{-/-}$ /Akt3 $^{-/-}$  (29) and Akt2 $^{-/-}$ /Akt3 $^{-/-}$  mice (30). Akt1 $^{-/-}$  mice are smaller and have shorter life span than wild type littermates and exhibit increased spontaneous apoptosis in the testis and thymus (23, 24). In contrast, akt2 $^{-/-}$  mice show insulin resistance and a diabetes mellitus-like syndrome (24) and akt3 $^{-/-}$  mice show brain defects (27).

More relevant in the context of the oncogenic properties ascribed to Akt signalling are recent studies showing that Akt1 promotes proliferation and inhibits migration and invasiveness of breast cancer cells in response to oncogenic signals, while Akt2 has apparently the opposite effects (31-34). However, the three Akt isoforms apparently have opposite roles in different cellular contexts, which may have important therapeutic implications. In fact, Akt1 positively stimulates migration

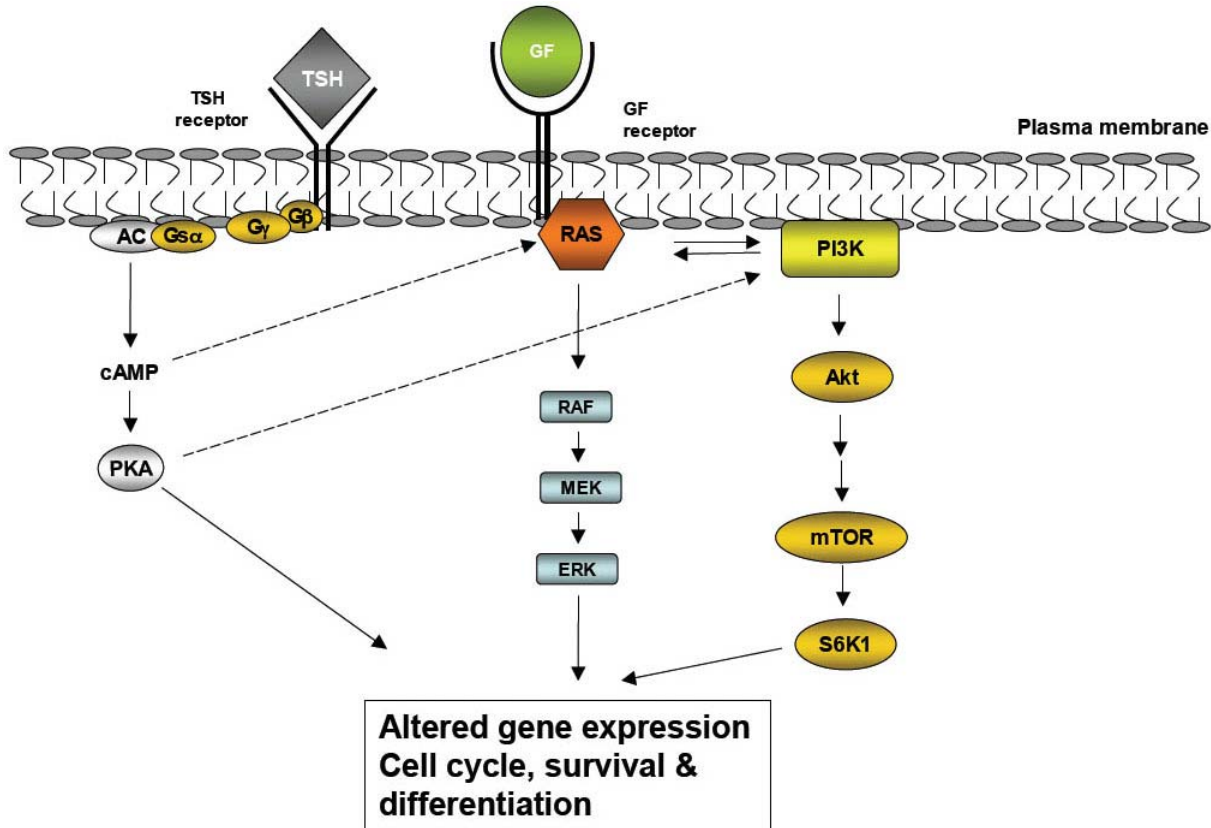
in MEF and lung epithelial cells (35, 36). Moreover, nuclear translocation of activated Akt1 is associated with cell invasion and migration in thyroid cancer cells (37).

In this regard, it is of note that the three Akt isoforms differ in their ability to transduce certain oncogenic signals *in vivo* such as those initiated by the Neu and PyMT oncogenes in mammary epithelium (38). Accordingly, ablation of Akt1 inhibits whereas ablation of Akt2 accelerates mammary tumour development initiated by Neu and PyMT oncogenes, while ablation of Akt3 is phenotypically neutral (34). Also the expression of Akt1, Akt2 and Akt3 apparently contribute to the different roles of Akt isoforms. Akt1 and Akt2 are widely expressed while tissue distribution of Akt3 seems to be more restricted, being primarily expressed in brain and testis (39). Accordingly, Akt1 and Akt2 are the principal isoforms expressed in the thyroid gland (40).

Growth factors and cytokines activate Akt through stimulation of PI3K activity (20). Accordingly, PI3K inhibition by chemical inhibitors (wortmannin, LY294002) or PTEN expression results in inhibition of tyrosine kinase receptor-mediated Akt activation (41). Activation of Akt is a multi-step process that involves membrane binding and phosphorylation. Upon activation, PI3K produces increased levels of phosphatidylinositol-3,4,5-trisphosphate (PtdIns-3,4,5-P $_3$ ) and phosphatidylinositol-3,4-trisphosphate (PtdIns-3,4-P $_2$ ), which contribute to recruit Akt to the plasma membrane where it binds to the phosphoinositides through its PH domain (41). Once at the cell membrane, Akt activation is obtained through phosphorylation on two critical aminoacids. One such residue lies within the kinase domain activation loop (Thr 308 in Akt1) and is phosphorylated by a PH-domain containing protein, PDK1 (42). This is thought to be the major activating phosphorylation event. Full Akt activity requires the subsequent phosphorylation of a second amino acid in the C-terminus (Ser 473 in Akt1). The identity of the serine 473 kinase is not completely established but this phosphorylation event may result from the rapamycin-insensitive mTOR complex (mTORC2) (43). Upon activation, Akt leaves the plasma membrane and phosphorylates a number of substrates both in the cytoplasm and in the nucleus, which mediates Akt-dependent regulation of cell growth and survival, mitogenesis, migration, glucose metabolism and protein translation (44). Akt targets include: Glycogen Synthase Kinase-3 (GSK-3 $\alpha$  and  $\beta$ ), tuberous sclerosis complex 2 (TSC-2), the pro-apoptotic Bcl-2 family members Bad and Bim, procaspase-9, I $\kappa$ B Kinases (I $\kappa$ B $\alpha$  and  $\beta$ ), the Forkhead family of transcription factors (FOXO), the ubiquitin ligases Mdm2 and Skp2, the CDK inhibitors p21 and p27, the kinases Raf and B-Raf, and others (20, 44). Akt signalling is counteracted by phosphatases PHLPP1 and PP2A (45, 46).

## 3. AKT ACTIVATION: UPSTREAM PATHWAYS

The central core of the Akt pathway is composed of upstream regulators of Akt function that include PI3K and the Phosphatase Tensin Homolog Deleted on

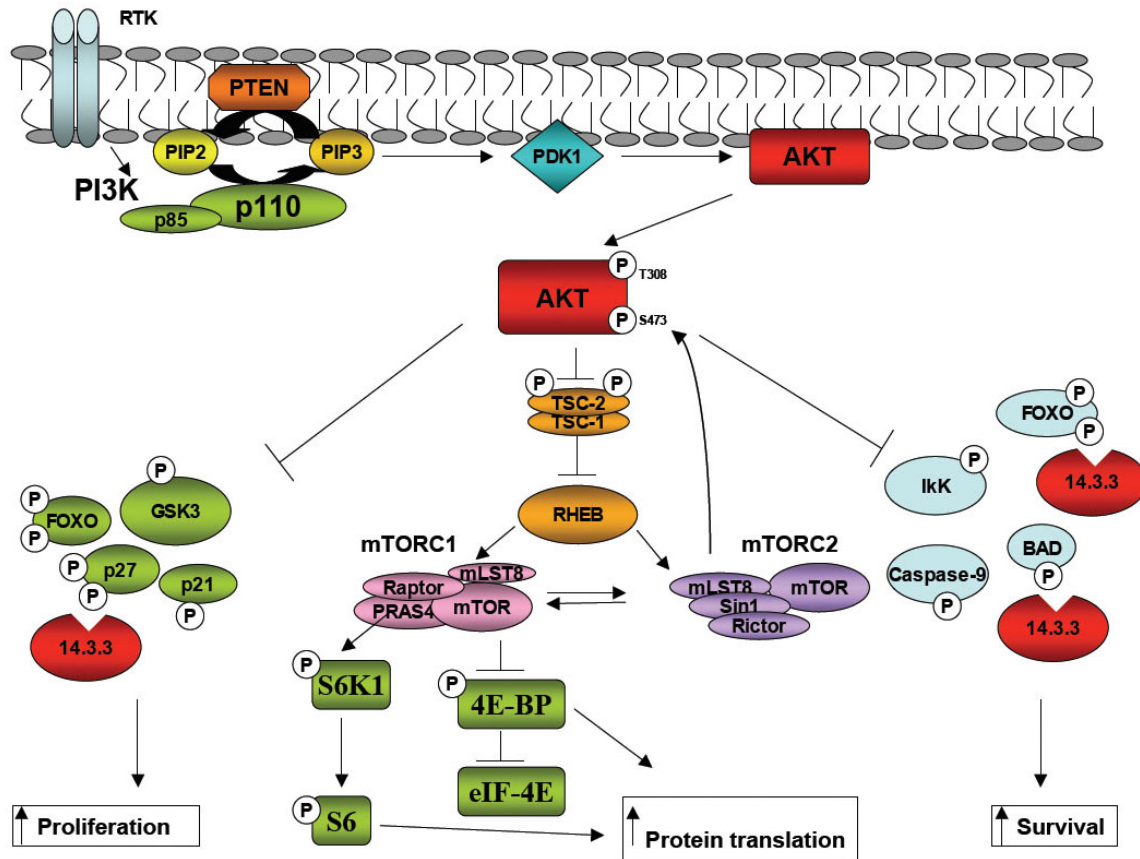


**Figure 2.** Proliferative pathways in thyroid cells. The cellular *in vitro* models that have been used to investigate the mechanisms involved in the proliferation of thyroid cells include rat thyroid cells lines (FRTL-5, WRT, and PC Cl3 cells) and short-term primary cultures of dog and human thyrocytes. DNA synthesis in canine thyrocytes requires the simultaneous presence of TSH and insulin/IGF1. Insulin or IGF1 alone have minimal effects on DNA replication, but they support DNA synthesis and cell cycle progression induced by TSH, EGF, bFGF, or phorbol. HGF is the only growth factor that acts as a full mitogen in dog thyrocytes, stimulating proliferation also in the absence of insulin/IGF-I. In rat thyroid cells, TSH and insulin/IGF-I cooperates to synergistically promote proliferation; insulin/IGF1 represent powerful mitogens whereas TSH is not able to induce DNA synthesis in the absence of insulin. The role of TSH in the G1 progression is to make cells competent to respond to insulin/IGF-I, which leads to the activations of MAPK and PI3K. In human thyroid cells TSH is able to induce DNA synthesis in serum-free primary cultures of adult. The mitogenic effect of TSH is increased by the presence of IGF-I or insulin, which alone weakly stimulate DNA synthesis (11). TSH is by far the most important physiological regulator of growth and function of thyrocytes. By binding to its cognate heterotrimeric G protein-coupled receptor, the TSH receptor (TSH-R), TSH causes dissociation of the G protein into  $\alpha$  and  $\beta\gamma$  subunits and activates the adenylyl cyclase/cAMP cascade. cAMP activates protein kinase A (PKA), which in turn, is required for thyroid cell differentiation and proliferation. In dog and human thyroid cells TSH does not activate Ras, PI3K, Akt or MAPK, and thus requires insulin- and/or IGF1-dependent activation of PI3K and its effectors, Akt and p70S6 kinase (p70S6K). In rat cells, at difference with dog and human thyrocytes, TSH-induced proliferation apparently requires Ras, Akt and PI3K. Ras activity appears to be necessary for TSH to induce the transition from quiescence to G1. Growth factors (i.e. EGF, HGF) regulate cell cycle progression of dog thyrocytes with different mechanisms that differ from those elicited by TSH. EGF and HGF activate Ras, the MAPK cascade but differ in their ability to activate the parallel PI3K/Akt pathway. EGF is not a full mitogen for dog thyrocytes because it potently activates the MAPK pathway but only weakly Akt; HGF is unique in triggering proliferation of thyrocytes in the absence of insulin because it is the only growth factor that can strongly activate both Ras-MAPK and PI3K-Akt. Akt apparently mediates the effects exerted by insulin/IGF1 as well as those exerted by serum on cell cycle progression of thyrocytes.

Chromosome Ten (PTEN) as well as downstream effectors including kinases [i.e. the mammalian Target of Rapamycin (mTOR), GSK3 $\alpha$  and  $\beta$ ], transcription factors (i.e. FOXO, NF- $\kappa$ B), and CDK inhibitors (p21 and p27) (20, 44). See Figure 3 for a detailed description. In this section we will review those molecules that may play a role in the activation of Akt in thyroid cancer.

### 3.1. The phosphatidylinositol 3-kinase

PI3Ks are a family of intracellular lipid kinases that generate the lipid second messenger PtdIns-3,4,5-P3 and PtdIns-3,4-P2. To date, several members of the PI3K family have been isolated in mammalian cells and they are grouped into three classes according to structure and substrate specificity (47). Class I PI3Ks phosphorylate

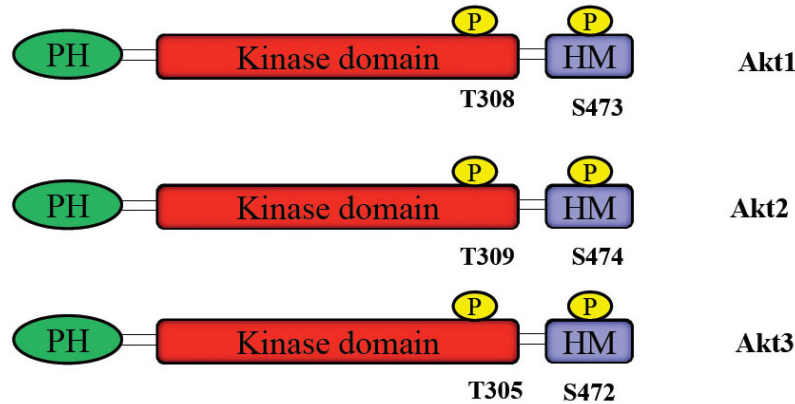


**Figure 3.** The PI3K-Akt pathway. Upstream activation of Akt by the growth factor receptor (RTK) pathway and cellular functions of some Akt substrates. Once activated, Akt phosphorylates a number of substrates in the cytoplasm and in the nucleus. Akt targets include: glycogen synthase kinase-3 (GSK-3 $\alpha$  and  $\beta$ ), tuberous sclerosis complex 2 (TSC-2), the pro-apoptotic Bcl-2 family members Bad and Bim, procaspase-9, I $\kappa$ B Kinases (I $\kappa$ B $\alpha$  and  $\beta$ ), the forkhead family of transcription factors (FOXO), the ubiquitin ligases Mdm2 and Skp2, the CDK inhibitors p21 and p27, the kinases Raf and B-Raf, and others. Nuclear proteins: FOXO, p27, p21; cytoplasmic proteins: GSK3, I $\kappa$ B, TSC2, caspase-9, S6K, 4E-BP; mitochondrial proteins: BAD.

PtdIns-3,4-P3 whereas class II and III PI3Ks use phosphatidylinositol as a substrate. Class I PI3Ks are further subdivided according to the signalling receptors that activate them: usually class IA PI3Ks are activated by growth factor receptor tyrosine kinases (RTKs); class IB PI3Ks are activated by G protein-coupled receptors (GPCRs) (48). Class I PI3Ks are heterodimeric molecules composed of a catalytic subunit known as p110 and a regulatory subunit denoted p85, which contains two SH2 (Src homology) domains that allow interaction with phosphotyrosines on activated tyrosine kinase receptors. This results in recruitment of the protein to the plasma membrane and activation of the enzymatic activity. There are three variants of the p110 catalytic subunit designated p110 $\alpha$ ,  $\beta$ , or  $\delta$ , expressed by separate genes (PIK3CA, PIK3CB, and PIK3CD for p110 $\alpha$ , p110 $\beta$ , and p110 $\delta$ , respectively). By contrast, there are five variants of the p85 regulatory subunit, designated p85 $\alpha$ , p55 $\alpha$ , p50 $\alpha$ , p85 $\beta$ , or p55 $\gamma$ ; the first three regulatory subunits represent splice variants of the same gene (PIK3R1), the other two encoded by different genes (PIK3R2 and PIK3R3, respectively) (48-50).

The  $\alpha$ -type (PIK3CA) and  $\beta$ -type (PIK3CB) p110 subunits are widely expressed in different tissues, including thyroid whereas the other type of p110 subunits has a limited expression. The most highly expressed regulatory subunit is p85 $\alpha$ . So far a central role in cancer has been demonstrated only for class IA PI3Ks, which transduce signals downstream of oncogenic tyrosine kinase receptors (51, 52). In fact, PIK3CA, encoding the class IA PI3K catalytic subunit p110 $\alpha$ , is the only PI3K gene identified with common mutations and gene amplification in human cancer (51). Most of these mutations are gain-of-function mutations that are located in hot spot regions in the helical and kinase domains of the gene encoding p110 $\alpha$  (E542K, E545K, and H1047R) (52). These mutations result in a p110 $\alpha$  variant that is active independent of the p85 regulatory subunit and leads to increased cell proliferation, invasiveness, resistance to apoptosis, and malignant transformation (51, 52). Several reports indicate that, *in vitro*, cellular transformation activity exerted by mutant PIK3CA is dependent on mTOR, which suggest that the major downstream signal for tumour promoting activity of constitutively active PI3K $\alpha$  is funnelled through the Akt-





**Figure 4.** Schematic representation of the three Akt isoforms. In mammalian cells Akt comprises three highly homologous members (>80% protein sequence identity) termed Akt1/PKB $\alpha$ , Akt2/PKB $\beta$  and Akt3/PKB $\gamma$ , encoded by three different genes located on chromosomes 14q32, 19q13 and 1q43, respectively. Activation of Akt is a multi-step process that involves membrane binding and phosphorylation. Akt kinases share the same structural organization, containing an N-terminal pleckstrin homology (PH) domain, a central catalytic domain and a C-terminal regulatory region. The PH domain of Akt can bind specifically to D3-phosphorylated phosphoinositides with high affinity and mediates Akt activation. Full Akt activation is obtained through phosphorylation on two critical aminoacids: one such residue lies within the kinase domain activation loop (Thr 308 in Akt1) and the second lies in the C-terminus (Ser 473 in Akt1).

mTOR axis (53). However, recent studies have also suggested that PI3K might promote cancer also through an Akt-independent pathway (54).

### 3.2. The phosphatase PTEN

PTEN (phosphatase and tensin homolog deleted on chromosome 10)/MMAC1 (mutated in multiple advanced cancers)/TEP-1 (TGF $\beta$ -regulated and epithelial cell-enriched phosphatase) (hereafter referred as to PTEN) is a tumour suppressor gene localized to chromosome 10q23 (55-57). The PTEN protein has been shown to have protein and lipid phosphatase activity (58, 59). The lipid phosphatase activity of PTEN can dephosphorylate the D3 position of PtdIns-3,4-P2 and PtdIns-3,4,5-P3, the lipid products of the PI3K lipid kinase activity (60), thus antagonizing signalling through the PI3K pathway. Indeed, cells lacking PTEN function exhibit a marked increase in the intracellular levels of PtdIns-3,4,5-P3 and Akt activation (61, 62). PTEN represents a pivotal regulator of critical cellular functions such as proliferation and survival. A large body of evidence indicate that PTEN functions as a tumour suppressor (63). Inactivating germ-line mutations in the gene encoding PTEN are the cause of the so-called PTEN hamartoma tumour syndrome (PHTS), a tumour susceptibility syndrome that includes Cowden syndrome (CS), Bannayan-Riley-Ruvalcaba syndrome (BRRS), Proteus syndrome (PS), and Proteus-like syndrome (64). PHTS includes benign and malignant thyroid neoplasias as part of the phenotype. In addition, loss of PTEN function is a frequent finding in sporadic tumours. Several mechanisms have been reported to account for the reduced levels of PTEN observed in cancer cells, including gene mutations and LOH, reduced transcription caused by gene promoter hypermethylation, reduced translation via microRNA (miR21) overexpression or increased protein degradation (65-67).

Targeted disruption of *Pten* in the mice leads to embryonic lethality (68). Mutant embryos show abnormal proliferation but not significant differences in apoptosis. Interestingly, *Pten*<sup>+/-</sup> mice, as the Cowden's syndrome patients, develop a variety of neoplasms including thyroid tumours (69-71). The majority of these tumours exhibit loss of the wild type allele, underscoring the importance of loss of PTEN function in tumour formation.

### 3.3. Ras GTPases

The Ras genes (H-Ras, K-Ras, and N-Ras) encode highly related proteins with GTPase activity that are located at the inner surface of the cell membrane and play a central role in the intracellular transduction of signals arising from cell membrane tyrosine kinase receptors and G-protein-coupled receptors (GPCRs) (72). In its inactive state, Ras is bound to guanosine diphosphate (GDP); upon activation, it releases GDP and binds guanosine triphosphate (GTP), thus transiently activating downstream signalling and terminates signalling by hydrolyzing GTP (72). Ras proteins convey signals from tyrosine kinase receptors and GPCRs to a cascade of mitogen-activated kinases (MAPK), which activate the transcription of target genes resulting in cell proliferation and/or to the PI3K/Akt pathway that contribute to cell growth and proliferation, migration and survival (72).

Ras mutations play an important role in malignant transformation and progression, representing the most common type of abnormality of dominant oncogenes in human cancer including thyroid carcinoma (9, 10). Oncogenic activation of Ras proteins (H-, K-, and N-Ras) is due to point mutations affecting the GTP-binding domain (codons 12 or 13) in Exon 1 or the GTPase domain (codon 61) in Exon 2, which lock the protein in the active GTP-bound form. Ras mutants are able to activate both the PI3K-Akt and MAPK signalling cascades and, conversely,

oncogenic transformation by mutant K-Ras requires activation of both MAPK and PI3K-Akt pathways (73). Consistent with this observation is the relative resistance of thyroid cancer cells harbouring Ras mutations to MEK inhibitors at difference of cell lines with B-Raf-V600E mutations (74, 75).

### 3.4. Tyrosine Kinase Receptors

Activation of tyrosine kinase cascades in thyroid cells regulates cell proliferation and differentiation (76). In 1998, Tanaka *et al* identified at least 21 tyrosine kinases expressed in thyrocytes, including 16 receptor-type and 5 non-receptor type, that in addition to TSH, may play a role in regulating the growth and differentiated functions of thyroid cells (77). In particular, since IGF1, FGF, EGF and the corresponding receptors are expressed by thyroid cells, it is likely that autocrine and/or paracrine loops involving such growth factors regulates growth and differentiation of normal thyrocytes (78, 79). IGF1 and EGF have been shown to stimulate proliferation of rat and dog thyroid cells synergistically with TSH (80, 81); HGF has been reported to have the strongest mitogenic activity on dog thyroid cells (82) (see legend to Figure 2). Moreover, certain subtypes of thyroid carcinomas are characterized by the aberrant expression of activated receptor-type tyrosine kinases such as Ret and NTRK1 proto-oncogenes consequent to chromosomal rearrangements (9, 10) or by overexpression of tyrosine kinase receptors such as MET [the receptor for hepatocyte growth factor (HGF)] or the EGF receptor (EGFR).

The Ret/PTC oncogene is generated by chromosomal rearrangements resulting in the fusion of the Ret tyrosine-kinase domain to the 5'-terminal region of heterologous genes. All RET-fused genes provide putative dimerization domains to the chimeric Ret/PTC genes, allowing dimerization and ligand-independent activation of Ret tyrosine kinase, which is essential for the transformation of thyroid cells. Among human tumours, Ret rearrangements are apparently restricted to the thyroid gland and are specific for PTC (81). Ret/PTC rearrangements are found on average in ~20% of adult patients, though is more common in patients with a history of radiation exposure (50–80%) (84–86). However, the reported prevalence of RET/PTC from different geographic regions varies widely, with the highest prevalence reported in the pediatric population. In particular, increased prevalence of Ret/PTC rearrangements was observed in children from the areas affected by nuclear disaster at Chernobyl (3, 4).

The NTRK1 (or TRKA) gene located on 1q22 encodes a tyrosine kinase receptor for the nerve growth factor (NGF). Similar to Ret, NTRK1 is activated in thyrocytes by chromosomal rearrangements that fuse the NTRK1 tyrosine kinase domain to the 5'-terminal region of heterologous genes. The recombination events that cause the oncogenic activation of NTRK1 include an inversion fusing NTRK1 to non-muscular tropomyosin (TPM3) gene located at 1q31, a different intra-chromosomal rearrangement that juxtaposes NTRK1 to the 5'-end of a translocated promoter region (TPR) gene localized at 1q25

or to the 5'-sequence of a TRK-fused gene (TFG) localized on chromosome 3 (TRK-T1, TRK-T2 and TRK-T3 oncogenes, respectively) (reviewed in 87). In all cases the resulting chimeric proteins exhibit constitutively tyrosine kinase activity. The TRK oncogenes also appear restricted to PTC, with a frequency of 10% of cases (88).

Conceivably, all these activated tyrosine kinase receptors convey their mitogenic signals through the PI3K pathway. Indeed PI3K signalling has been shown to be required for the stimulation of mitogenesis by Ret/PTC1 or Ret/PTC3 oncogenes (89). Similarly, *Met* also converges on Akt activation (90).

## 4. PERTURBATIONS OF AKT SIGNALING IN THYROID CANCER

Combining all the data from the existing literature, it appears that in thyroid cancer Akt activation, as determined by S473 phosphorylation, is frequent and is associated with more aggressive disease. Active Akt is observed more frequently in patients with undifferentiated cancer (40–50% of PTC and FTC; ~93% of ATC, respectively) (37, 91–97). Moreover, recent studies have suggested that cellular compartmentalization of activated Akt may be important in determining its cellular effects (37). In particular, it was proposed that nuclear localization of activated Akt1 promotes invasion and migration in thyroid cancer cells (37). In invasive FTC phospho-Akt localizes primarily to the nucleus, whereas in PTC, it localizes to the cytoplasm, except for the cells at the invasive edge or in metastatic regions where it is localized also in cell nuclei (37).

Different mechanisms that may account for increased Akt signalling in thyroid cancer cells have been proposed. These include amplification or activating mutation in two of the genes encoding Akt kinases, genetic abnormalities in the molecules upstream in the PI3K pathway such as mutation and/or amplification of the catalytic subunit of PI3K (PIK3CA) or loss of Pten (See 9, 10 for review). See Figure 4 for a summary of the molecular alterations in the members of the PI3K-Akt pathway in thyroid cancer. Gain-of-function mutations of two different Akt isoforms, namely Akt1 and Akt3, have been reported to occur in human cancer (98, 99). A unique mutation at nucleotide 49 of the gene encoding Akt1 that results in the substitution of a lysine for glutamic acid at the amino acid 17 (Akt1-E17K) within the PH has been recently discovered (98). The E17K substitution allows membrane recruitment of Akt1 independent of PtdIns binding, increases activity of Akt1 kinase, and confers to Akt1 the capability to transform fibroblasts *in vitro* and induce leukaemia in mice (98). Akt1-E17K mutant has been detected in multiple cancers such as breast, colon, ovarian, lung, endometrial, bladder and prostate carcinomas (98, 100–103). More recently, a mutation homologous to the E17K in Akt1 has been identified also in the PH domain of Akt3 in malignant melanoma (99). In thyroid cancer, the presence of a heterozygote E17K mutation in the Akt1 gene was observed at a relatively high frequency (9/55, 16%) in metastatic lesions of advanced cancer but



not in the corresponding primary tumours, which suggested that Akt1 mutations were acquired during tumour progression (94). Akt1 mutations were most common in metastasis of tall cell variant PTC (17%), Hürthle cell carcinoma (33%), and poorly differentiated PTC (19%) (94). Conversely, no mutation in the genes encoding Akt2 and Akt3 has been reported in thyroid cancer so far. Surprisingly, the knock-in of E17K mutant into the AKT1 gene had minimal phenotypic consequences and did not recapitulate the biochemical and growth characteristics observed with somatic cell knock in of PIK3CA mutations, suggesting that mutations in critical genes within the PI3K pathway are not functionally equivalent (104).

In addition to mutations, an increase in the gene copy number of Akt1 in FTC (8%) and ATC (~19%) and of Akt2 in FTC (~22%), respectively, has also been reported (97). It is not yet known whether amplified Akt differs from mutated Akt in its capability of to activate downstream Akt signalling.

Constitutive activation of Akt can occur also through mutations of other genes upstream in the pathway. Increased copy number of the PIK3CA gene, that encodes p110 $\alpha$ , has been frequently detected in thyroid cancer. Gene amplification at 3q26.3, where the PIK3CA gene is located, is detected in 12-13% of FA, 5-16% of PTC, 24-30% of FTC and 50% of ATC (105-109), though ethnic variation between Middle Eastern, Western or Asian populations exists (96). In addition to PIK3CA, amplification of the gene encoding PI3K p110 $\beta$  has been reported to occur in 46% of FTC and 38% of ATC (97). Constitutive activation of p110 $\alpha$  is also caused by the occurrence of activating mutations in the PIK3CA gene. Several studies have reported the presence of activating mutations within the helical (exon 9) and/or the kinase (exon 20) domains of PIK3CA in primary thyroid cancer and cancer-derived cell lines (91-93, 105, 107). PIK3CA mutations are rare in primary well-differentiated PTC (0-3%) (93, 106, 107), more common in well-differentiated FTC (6-13%) (93, 106) and consistently frequent in ATC (12-23%) (93, 106). PIK3CA p110 $\alpha$  mutations are particularly common in the metastatic lesions of patients with radioactive-iodine refractory disease, similarly to Akt1 (94). So far, PIK3CA and Akt1 mutations are apparently mutually exclusive, suggesting that they may have equivalent roles (94).

Accordingly, it is not yet known whether PIK3CA mutations or amplification are sufficient to cause thyroid cancer *in vivo*. Mutant PIK3CA alleles are transforming in MCF-10 breast cells *in vitro* and in the chorioallantoic membrane of the chicken *in vivo* (110). However, transgenic mouse models indicated that activated PIK3CA mutant is able to induce fully malignant cancer in the lung but not in the ovary (111, 110). Therefore, further studies will be required to fully characterize the role of this oncogene in thyroid cancer development and progression.

In tumours where ATC coexisted with a better-differentiated tumour component, PIK3CA and Akt1 mutations were exclusively observed in the ATC

component (93, 94, 109). This finding suggests an exclusive role for oncogenic PIK3CA and Akt1 mutants in promoting progression from more differentiated to less differentiated or completely undifferentiated cancer (93, 94). Consistently, mutant Akt1 and PIK3CA are found in the metastatic lesions but not in the corresponding primary tumours (94). Conversely, Ras and B-Raf are considered early events in thyroid carcinogenesis that are able to initiate tumours, since Ras mutations are observed both in FA and FTC and B-Raf mutations are generally found in both differentiated PTC and undifferentiated ATC. Accordingly, the generation of murine models has shown that B-Raf and N-Ras can initiate thyroid cancerogenesis *in vivo* (16 and references therein). These results suggest that, at difference with genes involved in the MAPK pathways (i.e. B-Raf) the constitutive activation of PI3K signalling is probably insufficient by itself to initiate the growth of a malignant thyroid cancer, since loss of PTEN results in follicular adenoma; conversely, aberrant PI3K signalling may facilitate progression and dedifferentiation of tumour cells (113).

Mutation analysis of human thyroid cancer indicates also that the MAPK and PI3K-Akt pathways cooperate in the pathogenesis and progression of advanced or metastatic thyroid cancer (93, 94, 106). The great majority of tumours carrying the Akt1-E17K (76%) also displayed concomitant B-Raf mutation, but not Ret/PTC rearrangement or Ras mutations, indicating that simultaneous signalling from Akt and B-Raf efficiently contribute to the development of dedifferentiated cancer (94). Similarly, most tumours (73%) with PIK3CA mutations also showed mutations in Ras and/or in B-Raf genes (93, 96). Likewise, the presence of PIK3CA amplification was observed in approximately half PTC harbouring Ras mutation (96) and in an even higher percentage of PDC (50%) and ATC (40-100%) with concurrent B-Raf mutations (93, 94). Whatever the explanation for coexisting mutations, the putative cooperation of PI3K/Akt signalling and Ras, or B-Raf signalling has already been observed in colon cancer and might have important implications also in the behaviour of thyroid carcinoma (114).

Expectedly, Akt activation is observed in almost all ATC harbouring PIK3CA mutations (>90%) (93). However, because Akt activation can occur regardless of the presence of PIK3CA or Akt1 mutations, other molecular mechanisms that account for Akt activation in undifferentiated thyroid cancer cases, such as PTEN down-regulation or Ras activation, have been proposed. Genetic alterations that inactivate the PTEN gene, including mutations and deletions, are uncommon in sporadic thyroid tumours (90, 106, 107, 115-117). Moreover, allelic losses of the PTEN locus at 10q23.3, though frequent in FA and FTC (up to 25%), are not coupled with mutations in the second allele (118, 119). Conversely, thyroid carcinoma frequently shows decreased expression of PTEN, at both mRNA and protein levels in ~40% of the well-differentiated thyroid carcinomas and lost in most ATC (90, 116-119), in many cases through methylation of the PTEN gene promoter (120, 121). In a limited subset of

thyroid carcinomas, PTEN inactivation has been associated with Akt activation (90).

Additional mechanisms that account for the activation of the PI3K/Akt signalling pathway in thyroid cancer involve aberrant signalling from mutant Ras or tyrosine kinase receptors (See 9, 10 for review). Conceivably, Ret/PTC or NTRK1 rearrangements, Met overexpression and Ras mutations play an important role in activating the PI3K/Akt signalling in thyroid cancer. Ras plays a role in aberrant Akt signalling in thyroid cancer, both directly or indirectly. Activation of the PI3K/Akt pathway by mutant Ras proteins has been shown to occur in FTC, which commonly harbours activating Ras mutations (122). Accordingly, PI3K is a well-characterized direct downstream effector of Ras (123). The p110 catalytic subunit of PI3K possesses a Ras binding domain that mediates binding and direct Ras-dependent activation. Expression of mutated Ras protein in thyroid cells induces activation of PI3K, and this causes an increase in the dependence of thyrocytes on PI3K signalling for survival (124). On the other hand, PI3K is an essential anti-apoptotic effector in the proliferative response of human thyroid cells to mutant Ras (125, 126).

The PI3K-Akt pathway can be efficiently activated also through signalling from aberrant tyrosine kinase receptors. This may be a relevant mechanism particularly in PTC, which most commonly harbours Ret/PTC and NTRK1 rearrangements or Met overexpression (9, 10). Different mechanisms have been proposed for Ret/PTC-dependent Akt activation in thyroid cells. As most tyrosine kinase receptors Ret/PTC can activate PI3K signalling, possibly through phosphorylation of Insulin Receptor Substrate 1 (IRS-1) (127) and XB130 (128), or through RAI(ShcC/N-Shc)-dependent recruitment of GAB1 to phosphorylated tyrosine 1062 (129), which was shown to be essential for Ret-mediated activation of PI3K. Accordingly, PI3K signalling is necessary to Akt activation by Ret/PTC, since PI3K-specific inhibitor LY294002 has been shown to reduce Akt activation in Ret/PTC-transfected rat thyroid cells (92). Alternatively, Ret/PTC can directly phosphorylate and activate Akt1 by direct phosphorylation of Y315, which facilitate Akt phosphorylation of T308 and S473 and activation (130, 131). Signalling of Ret/PTC1 and hepatocyte growth factor receptor (Met) also converges on Akt activation. Met overexpression alone strongly activates Akt signalling in thyroid cells (90). On the other hand, a strong correlation between expression of Met and Akt activation has been demonstrated in PTC (132).

## 5. SIGNALLING DOWNSTREAM AKT IN THYROID CANCER

A recent search of the literature has allowed the identification of over 100 non-redundant Akt substrates, for ~20 of which there have been multiple independent published reports (Figure 3) (44). Expectedly, it appears that each function downstream Akt –proliferation, survival, metabolism- is mediated by multiple targets and is frequently dependent on the cell context. It is worth noting

that Akt can either cause the activation (*i.e.*, Mdm2, I $\kappa$ B $\alpha$ / $\beta$ ) or the inactivation (*i.e.*, p21, p27, Bad, procaspase-9, FOXO3a, and GSK3 $\alpha$ / $\beta$ ) of specific substrates. In most cases Akt-dependent phosphorylation usually results in the change of stability, activity and/or localization of the substrate proteins, which contribute to increase cell proliferation, motility, protein synthesis and gluconeogenesis as well as inhibit apoptosis (44). One common mechanism whereby Akt-mediated phosphorylation results in substrate inhibition is the regulation of substrates' cellular localization as in the case of Bad, FOXOs, p27. Akt substrates' localization is mediated through the interaction with 14-3-3 proteins, cytoplasmic anchors that bind specifically to phosphoproteins and retain them in the cytoplasm (133) away from their targets (Figure 3).

### 5.1. The mTOR pathway

Among the numerous downstream effectors of Akt, activation of the mammalian target of rapamycin (mTOR) kinase [also known as FK506 binding protein 12-rapamycin associated protein (FRAP)1] has recently taken center stage, thanks to the increasing evidence that, in different cell types, it is directly responsible for growth and proliferation associated with PI3K activation (134, 135). mTOR is a serine/threonine protein kinase in the PI3K cascade that regulates protein synthesis and plays an important role in multiple biological processes such as cell growth and survival (136-138). By regulating ribosomal biogenesis and protein translation, mTOR controls cell growth and can restrict cell cycle progression in the presence of suboptimal growth conditions (134-137).

In mammalian cells, mTOR exists in two functionally distinct complexes, namely mTORC1 and mTORC2 (Figure 3). mTORC1 is a critical regulator of translation initiation and ribosome biogenesis and plays an evolutionarily conserved role in cell growth control (reviewed in 134). mTORC1 is composed of mTOR, Raptor, mLST8, and PRAS40 (proline-rich Akt substrate 40kDa), and is sensitive to inhibition by the macrolide antibiotic rapamycin. Conversely, mTORC2 is composed of mTOR, Rictor, mLST8 and Sin1. mTORC2 was originally reported to be insensitive to rapamycin, though recent studies indicate that it may be indirectly inhibited by long-term treatment with rapamycin (134).

mTORC1 activates p70 ribosomal protein S6 kinase (S6K1) and, at the same time, releases eukaryotic translation initiation factor 4E (eIF-4E) by phosphorylating inhibitory eIF-4E binding proteins 1-3 (4E-BPs) (134). In turn, activated S6K1 phosphorylates the ribosomal protein S6 to increase translation of mRNAs with 5'-terminal oligopolypyrimidine tracts whereas mTOR-dependent phosphorylation of 4E-BPs releases the initiation factor eIF4E to promote cap-dependent translation. This results in increased synthesis of growth-related proteins such as cyclin D1, Myc, and vascular endothelial growth factor (VEGF) (135-139).

The dynamic interplay between Akt and mTOR is remarkably complex (Figure 3). On one hand, Akt

activates the downstream mTOR kinase by inhibiting the complex formed by the tumour suppressor proteins TSC-1 and TSC-2 (also known as hamartin and tuberlin) through direct phosphorylation of TSC-2 at S939 and T1462 (140). TSC-2 forms a heterodimeric complex with TSC-1 that inhibits the GTPase Rheb, a selective activator of mTORC1. Akt phosphorylation of TSC2 suppresses the GTPase-activating activity exerted by the TSC1/TSC2 complex on Rheb, thus resulting in the activation of mTORC1 (134, 140). Akt can activate mTORC1 also through a TSC-2-independent mechanism. Akt directly phosphorylates PRAS40, a protein that associates with mTORC1, thus relieving PRAS40 inhibitory effect on mTORC1 (134). However, the other cellular mTOR-containing complex, mTORC2, acts upstream Akt since it has been identified as the kinase that phosphorylates Akt at S473, thus contributing to its activation (134).

Mechanisms whereby mTOR promotes tumorigenesis include Akt activation either by mTORC2 or mTORC1-dependent increase of the synthesis of growth-related proteins. So far, activating mutations in the gene encoding mTOR have not been identified. However, many human cancers including thyroid carcinomas are characterized by aberrant activation of mTOR, possibly through deregulation of upstream components that regulate mTOR (141-143). Accordingly, the enhanced sensitivity of cancer cells exhibiting oncogenic activation of the PI3K-Akt pathway to mTORC1 inhibitors illustrates the importance of mTORC1 activation downstream Akt (139-141). However, the existence of a negative-feedback inhibition exerted by mTORC1 on PI3K-Akt signalling in cells with activated mTOR may limit the therapeutic use of mTOR inhibitors (134).

mTOR has been shown to be a key effector of TSH-mediated proliferative signals in thyroid follicular cells. Brewer *et al.* showed that the *in vivo* proliferative response to chronic TSH stimulation of mice treated with sodium perchlorate and methimazole relies on the activation of the mTOR/S6K1 axis, and that mTOR inhibition abrogates the hyperplastic responses to increased TSH levels (144). In this model, goitre development is apparently independent of Akt activity, underlying the existence of an Akt-independent pathway leading to mTOR activation upon TSH stimulation. This is consistent with recent works showing that Akt-independent activation of mTOR is recognized as a mechanism of control of cell proliferation in luteal cells (145), B lymphocytes and endometrial stromal cells (146, 147). mTOR regulates also the function of normal thyrocytes both *in vitro* and *in vivo*, since it participates in the control of thyroid iodide uptake by regulating NIS protein expression (148). Interestingly, the finding that mTOR inhibition avoids the activation of both S6K1 and Akt suggests the involvement of mTORC1 and mTORC2 in NIS regulation.

mTOR has been shown to be a key effector of the proliferative signals funnelled through PI3K in thyroid cells. Recent studies indicated that the *in vivo* proliferative response to chronic PI3K activation relies profoundly on the activation of the mTOR/S6K1 axis (149). Conditional loss of Pten in the mouse thyroid follicular cells stimulates

autonomous growth leading to the development of adenoma, possibly driven by increased cyclins D1 and D3 (149). mTOR inhibition in Pten mutant mice restores virtually normal proliferation rates, despite the presence of still elevated Akt activity (149). These data extend independent findings obtained in the TR $\beta$ PV/PV knock-in mouse model of thyroid follicular carcinoma (148). TR $\beta$ PV/PV mice harbour a mutant thyroid hormone receptor beta gene that promotes thyroid cancer and distant metastasis similar to human FTC (150). In this strain, inhibition of PI3K signaling reduces tumour cell proliferation by down-regulating TOR activity and cyclin D1 levels. In addition, mTOR inhibition potently suppresses proliferation of human thyroid cancer cells harbouring genetic alterations in the PI3K/Akt pathway (151-153).

### 5.2. Apoptosis-related pathways

Akt enhances cell survival by regulating expression and function of several pro-apoptotic genes (Figure 3) (44). The Bcl-2 family of proteins is a major intracellular regulator of apoptotic signalling, with at least 20 members in mammalian cells (154). Proapoptotic members including Noxa, Puma, Bim and Bad are transcriptionally or post-translationally activated by extracellular pro-apoptotic signals and/or by intracellular damage and are inhibited by intracellular pro-survival proteins such as Akt (154, 155). Similarly, Akt regulates expression and/or function of anti-apoptotic members such as Bcl-2, Bcl-xL, Bcl-w and Mcl-1 that protect cells from a wide range of apoptotic stimuli, including chemotherapeutic drugs and irradiation (154, 155).

Akt directly phosphorylates and inhibits Bad (156, 157). Survival factors stimulate Akt-mediated phosphorylation of Bad on S136, and this creates a binding site for 14-3-3 proteins, which triggers release of Bad from the mitochondrion (157). In the unphosphorylated state, Bad is targeted to the mitochondria where it forms a complex with Bcl-2 or BclXL, inhibiting their anti-apoptotic activity. Conversely, when phosphorylated, Bad associates with 14-3-3 proteins in the cytoplasm allowing Bcl-2 or BclXL binding to Bax and Bak, multidomain Bcl-2 family members that function as an obligate gateway for the activation of apoptosis via the mitochondrial and endoplasmic reticular pathways presumably as pore-forming complexes, to release holocytochrome c in the cytoplasm.

Akt suppresses apoptosis also by inhibiting the activity of pro-apoptotic transcription factors, such as FOXO and p53. The FOXO (Forkhead box-containing protein, O subfamily) transcription factors belong to the winged helix/forkhead family and are the closest homolog of Daf-16 in *C. Elegans*. Mammalian cells express three FOXO isoforms: FOXO1 (FKHR), FOXO3a (FKHRL1) and FOXO4 (AFX) (reviewed in 158 and 159). These transcription factors are negatively regulated through Akt-dependent phosphorylation (160-163). Akt phosphorylates FOXO1 on T24, S256, and S319, and FOXO3a and FOXO4 on equivalent sites (161, 162). Akt phosphorylation of FOXO proteins occurs in the nucleus.

14-3-3 proteins displace phosphorylated FOXO transcription factors from the promoters of target genes and trigger their export from the nucleus and consequent degradation. Through this mechanism, Akt blocks FOXO-mediated transcription of target genes that promote apoptosis and cell-cycle arrest (164). In cancer cell lines lacking functional PTEN, FOXO1 and FOXO3a are constitutively phosphorylated, cytoplasmic and unable to activate transcription. Reconstitution of PTEN expression restores nuclear localization of FOXO1 and restores its ability to activate FOXO responsive elements in the promoters (163). FOXOs can induce apoptosis through the up-regulation of FasL and of the pro-apoptotic Bcl-2 interacting mediator (Bim1) (161, 166). Thyroid cells carrying an activated Akt allele become resistant to programmed cell death induced by starvation because Akt inhibits the induction of proapoptotic genes (Fas, Fas ligand, Bad) in starved cells (167). Notably, activated Akt was found to correlate with phosphorylated Bad in a subset of thyroid cancer specimens (95).

A third Akt target that promotes survival is Mdm2 (or Hdm2 in human), an E3 ubiquitin ligase that catalyzes p53 degradation. Akt phosphorylates Mdm2 on S166 and S186, and this promotes translocation of Mdm2 to the nucleus, where it negatively regulates p53 function (168, 169). Two transcriptional targets of p53 that are essential for p53-induced apoptosis are Puma and Noxa (170). However, the relative importance of downregulation of these p53 targets to Akt-mediated cell survival in thyroid cancer cells has not been thoroughly examined.

In addition to the inhibition of pro-apoptotic factors, Akt can also induce the expression of anti-apoptotic genes through the activation of the transcription factor Nuclear Factor kappa B (NF- $\kappa$ B) (171-174). Akt activate the I $\kappa$ B kinases (I $\kappa$ Ks), which, in turn, phosphorylate I $\kappa$ B, an inhibitory protein that sequesters NF- $\kappa$ B in the cytoplasm, targeting it for degradation by the proteasome (175-177). This allows NF- $\kappa$ B to translocate to the nucleus and activate transcription of a variety of anti-apoptotic genes including the cellular inhibitors of apoptosis (c-IAP1, c-IAP2 and XIAP), TNF receptor-associated factors (TRAF1 and TRAF2), Gadd45b, the Bcl-2 homologue A1/Bfl-1, IEX-IL and Bcl-xL (178, 179). Much of the pro-survival ability of Akt is mediated through NF- $\kappa$ B activation, being one of the major culprits of resistance to chemotherapy (180-182). Several studies have demonstrated that Akt mediates NF- $\kappa$ B activation by tumour necrosis factor (TNF $\alpha$ ) and PDGF (173, 183). Although it is likely that there are multiple levels of cross-talk between the PI3K-Akt and NF- $\kappa$ B pathways, one mechanism has been attributed to direct phosphorylation of the amino acid residue T23 on I $\kappa$ B kinase  $\alpha$  (I $\kappa$ K $\alpha$ ) by Akt, thereby leading to activation of this kinase upstream NF- $\kappa$ B and release of I $\kappa$ B-mediated inhibition of NF- $\kappa$ B (173). Activation of I $\kappa$ K $\alpha$  by Akt may occur also through phosphorylation of NF- $\kappa$ B subunit p65 at S534 (184). Whatever the mechanism, the inhibition of endogenous Akt by overexpression of PTEN results in decreased NF- $\kappa$ B transcriptional activity and sensitization of cells to TNF-

induced apoptosis (185). Therefore, the findings suggest that loss of PTEN or activation of Akt in thyroid cancer cells might contribute to carcinogenesis by activation of the NF- $\kappa$ B pathway. Similarly, Ret/PTC and B-Raf-V600E oncoproteins induce degradation of I $\kappa$ B $\alpha$  and activation of NF- $\kappa$ B signaling in thyroid cancer cells (186, 187).

Several studies have suggested that NF- $\kappa$ B is strongly activated in thyroid cancer specimens, especially in ATC. Thus, NF- $\kappa$ B inhibition may represent an attractive therapeutic target for the treatment of advanced thyroid cancer (188). p65 expression and activity are significantly increased in thyroid cancer cell lines (189, 190) and in tumour specimens (191, 192). By contrast, inhibition of NF- $\kappa$ B activity in human ATC cell lines leads to reduced invasion associated with differential expression of MMP-13 and MMP-9 (193), increased susceptibility to chemotherapeutic drug-induced apoptosis and inhibition of tumours growth in nude mice. Importantly, the combined treatment of thyroid cancer cells with NF- $\kappa$ B inhibitors [i.e. Dehydroxymethylepoxyquinomicin (DHMEQ), R-Roscovitin] and taxanes strongly synergize *in vitro* and show a significantly greater inhibitory effect on tumour growth in nude mice (194, 195). Another NF- $\kappa$ B inhibitor, triptolide, functions as an effective apoptotic inducer in a p53-independent, but NF- $\kappa$ B-dependent mechanism, thus providing a promising agent for tumour types with p53 mutation/deletion (196).

### 5.3. Proliferation-related pathways

Although best known for promoting cell survival and growth, Akt represents also a main regulator of cell proliferation through the phosphorylation of multiple downstream targets impinging on cell-cycle regulation such as Cdk inhibitors and G1 cyclins (Figure 3) (44).

Expression of constitutively active Akt promotes proliferation and survival of thyroid cells without affecting the expression of the differentiated phenotype (92, 167). Thyroid cells carrying an activated Akt allele proliferate in the absence of TSH and insulin (167, 197).

In thyroid cells Akt increased the levels of the G1 cyclins (i.e. cyclins D3 and E) (167) and induced cytoplasmic displacement of p27 (92). By contrast, pharmacological inhibition of PI3K (i.e. with LY294002, wortmannin) is sufficient to inhibit proliferation of human thyroid cancer cells, and this apparently occurs through regulation of the subcellular localization of p27 (37, 92). Transient expression of PTEN inhibits Akt activity in thyroid cancer cell lines and induces cell cycle arrest or cell death in cell contest-dependent manner: PTEN induced G1 cell cycle arrest in PTC-derived cells and/or both G1 arrest and cell death in FTC-, PDC- and ATC-derived cells (91, 198). More recently the direct Akt inhibitor perifosine was shown to potently inhibit the proliferation of cells that harboured PI3K/Akt-activating genetic alterations but to have modest responses in cells that harboured no genetic alterations (151).

p27 inactivation is a critical step in growth regulation exerted by PI3K-Akt in thyroid cancer cells (91).

The PI3K/Akt pathway contributes to inactivation of p27 in thyroid cancer through several mechanisms. Akt can induce phosphorylation-dependent cytoplasmic sequestration of p27 (92), inhibition of p27 gene transcription by targeting the FOXO transcription factors or degradation of p27 through up-regulation of the E3 ubiquitin ligase Skp2. Work from different labs has demonstrated that Akt phosphorylates the p27 cyclin-dependent kinase inhibitor at T157 (199, 200) and T198 (201, 202). By phosphorylating p27 at T157 and T198, Akt induces binding of p27 to 14.3.3, prevents interaction with importin- $\alpha$ , impairs nuclear import, and overcomes p27-induced growth (203, 204). However, although impaired import of p27 into cell nuclei may affect its ability to inhibit cell cycle progression, several experimental evidences supports the idea that p27 exerts additional cytoplasmic functions that foster carcinogenesis (205). Cytoplasmic p27 may suppress apoptosis or regulate migration (206), or increase the number of tissue stem cells and/or progenitors (207), thus allowing cancer cells to dysregulate multiple cellular functions with one hit.

Different mechanisms have been proposed to explain PI3K- and Akt-dependent down-regulation of p27 in thyroid cancer. Akt may inhibit p27 expression in thyroid cancer cells through phosphorylation and cytoplasmic displacement of FOXO3 (208). Accumulation of FOXO3a correlated with increased phospho-Akt staining and with reduced transcription from the p27 gene in thyroid cancer but not in normal thyroid tissue (208). Alternatively, increased turnover of p27 protein in thyroid cancer has also been reported to occur following increased PI3K-dependent expression of Skp2 (209). The increase in Skp2 levels is induced by expression of oncogenic Ret/PTC and Ras proteins (208), and results in PI3K-dependent p27 loss in human normal thyrocytes (210). Accordingly, pharmacological inhibition of endogenous or transfected Ret/PTC restored p27 expression in rat and human thyroid cells (211). Finally, the recent finding that double mutant mice carrying an activated NTRK1 oncogene in a background of a p27 null allele [TRK-T1/p27(-/-)] displayed a higher incidence of PTC, with a shorter latency period and increased proliferation index, compared with p27 wild-type mice [TRK-T1/p27(+/-)], demonstrated the critical role of this Cdk inhibitor in the contest of tyrosine kinase receptor-driven thyroid carcinogenesis (212).

In addition to the inactivation of Cdk inhibitors, Akt can stimulate cell cycle progression through up-regulation of G1 cyclins. Akt phosphorylates the GSK3 isoforms  $\alpha$  and  $\beta$  on a highly conserved N-terminal regulatory site (S21 for GSK3 $\alpha$ , S9 for GSK3 $\beta$ ) (213). Akt-dependent phosphorylation of GSK3 inactivates the kinase and relieves the constraint imposed by GSK3 on the synthesis and the stability of proteins involved in cell-cycle entry. In particular, GSK3 phosphorylates cyclin D and cyclin E, which play a central role in the G1-to-S-phase cell cycle transition, targeting them for nuclear export and proteasomal degradation (214, 215). Phosphorylation and inhibition of GSK3 by Akt enhance the stability of cyclin D, cyclin E and Myc, promoting cell-cycle entry.

Accordingly, cyclins D and E and Myc are frequently overexpressed in the different types of thyroid carcinoma, though it is not known whether this is correlated to increased signalling through the PI3K pathway.

Finally, Akt may also have a role in the deregulation of mitotic checkpoint. Constitutively active Akt is able to promote progression into mitosis, even in the presence of DNA damage (216, 217). One mechanism explaining this observation is that Akt directly phosphorylates the DNA damage checkpoint kinase Chk1 on S280 (218). S280 phosphorylation blocks checkpoint function by stimulating Chk1 translocation to the cytosol, where it is sequestered from the DNA damage-sensing kinases ATM and ATR (219). Accordingly, expression of activated Ras in rat thyroid cells, which activate Akt signalling, induces chromosomal instability, as a consequence of defects in the processing of DNA damage (220).

## 6. THERAPEUTIC STRATEGIES OF TARGETING AKT SIGNALLING IN THYROID CANCER

The major therapeutic challenge for thyroid cancer therapy is represented by the high rate of mortality of patients with PDC and ATC due to surgical inoperability at diagnosis or subsequent insensitivity to radioiodine treatment. Whereas most patients affected by differentiated thyroid cancer are successfully treated with thyroidectomy, radioiodine treatment and/or external beam radiotherapy, cytotoxic systemic chemotherapy for advanced, metastatic thyroid carcinomas has limited effectiveness, with response rates typically in the range of 25% or less (221). Therefore, the development of novel, molecularly-based compounds will improve disease outcomes especially in patients with aggressive thyroid cancers. As noted above, aberrant activation of the PI3K/Akt pathway is an essential step for the initiation and maintenance of thyroid cancer. Almost every single node of this pathway can be subject to pathway-activating genetic alterations, which result from the distinct and often mutually exclusive events that include (i) translocation of receptor tyrosine kinases (for example Ret/PTC or NTRK-1) leading to constitutive recruitment and activation of PI3K and downstream effectors; (ii) amplification of PI3K; (iii) presence of activating mutations in the PIK3CA gene encoding the p110 $\alpha$  catalytic subunit; (iv) amplification of the downstream kinase Akt; (v) presence of activating mutations in the Akt1 gene (vi) loss or inactivating mutations of the tumor suppressor gene PTEN, or (vi) constitutive recruitment and activation by mutant forms of the Ras oncogenes (10). See Figure 5 for a summary of molecular alterations observed in thyroid carcinoma.

To date, most of the compounds tested in treating thyroid cancer are competitive inhibitors of tyrosine kinases (reviewed in 221). The demonstrated oncogenic roles of mutant Ret/PTC, Ras and B-Raf and the contribution of VEGF angiogenic growth factor receptors to development of thyroid cancer, have lead to different clinical trials with small molecule inhibitors (222, 223). These compounds include sunitinib, sorafenib, motesanib

and axitinib and, consistent with the oncogene addiction hypothesis, have been reported to induce either tumor stabilization or partial remission in a certain percentage of patients with advanced thyroid carcinomas (224-226). These orally administered drugs partially inhibit multiple kinases that include Ret/PTC, B-Raf, VEGF receptors, cMet, and PDGF receptors, at nanomolar concentrations and thus affect multiple downstream signaling pathways including the MAPK and PI3K pathways (222). However, if Akt is activated by loss of PTEN or mutations in PIK3CA or Akt1 itself, the inhibition of upstream receptor activity may be ineffectual. In these cases, patients may benefit of PI3K or Akt inhibitors. In particular, both PI3K and Akt may represent potentially relevant therapeutic targets for advanced thyroid cancer (227, 228). However, since PI3K-Akt signaling plays a critical role in essential processes such as insulin signaling, neuron function, and endothelial activity that may be disrupted with systemic administration of a PI3K inhibitor, a careful analysis of safety parameters in clinical trials will have to be taken into account, especially in patients with thyroid cancer, many of whom have excellent quality of life.

The available data indicate that genotype-based targeting of the PI3K/Akt pathway using Akt and mTOR inhibitors may offer an effective therapeutic strategy for thyroid cancer (151-153). Available Akt inhibitors may be of three types: ATP competitive inhibitors that target the kinase domain, the PtdIns analogs that block binding of PH domain to PtdIns and allosteric inhibitors that stabilize Akt in a "close conformation" that is not capable to be activated by PDK1 (227). Several compounds that inhibit all Akt isoforms have been recently developed and are now in phase I clinical trials (reviewed in 225). A series of potent, ATP competitive Akt inhibitors (IC<sub>50</sub>=20 nM for Akt versus IC<sub>50</sub>=1900 nM for PKA) were developed by exploiting the X-ray crystal structure of (-)-balanol, a potent inhibitor of AGC-kinases, in complex with PKA (228-232). Promising Akt inhibitors have also been obtained from indazole-pyridine-based derivatives such as A-443654 (K<sub>i</sub>=160 pM for Akt1) (228-231). These compounds are reversible, ATP competitive inhibitors, which decrease the phosphorylation of Akt downstream targets in cells (for example, GSK3 $\alpha/\beta$ , FOXO3, TSC2 and mTOR) and *in vivo* in a dose-dependent manner. In xenograft experiments, A-443654 showed antitumor activity both as a single agent and in combination regimens. However, in these preclinical experiments, treatment had to be discontinued because of multiple side effects in treated animals, raising concerns that the therapeutic application of Akt inhibitors can be limited by inherent metabolic toxicity (232).

Recently, compounds with improved potency, selectivity and safety have been reported (233, 234). For example, GSK690693, an aminofurazan derivative, is an ATP-competitive, pan-Akt kinase inhibitor (IC<sub>50</sub>=2 nM for Akt1) that has recently entered phase I clinical trials (234). The compound was well tolerated and showed significant antitumour activity *in vivo*. At the moment, the first phase I study in humans to investigate the safety, tolerability, pharmacokinetics and pharmacodynamics of GSK690693 is ongoing.

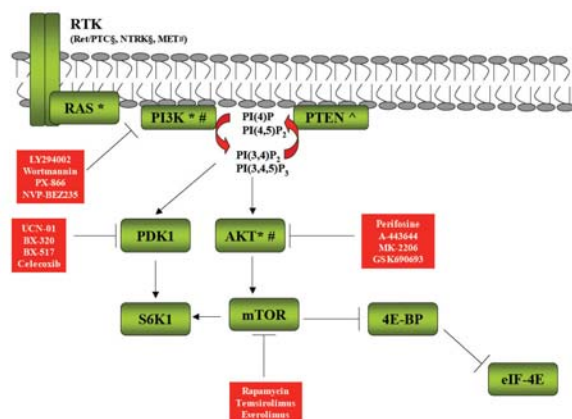
The allosteric Akt inhibitor perifosine (octadecyl-(1,1-dimethyl-piperidino-4-yl)-phosphate) (235) has been shown to inhibit *in vitro* proliferation and the *in vivo* growth of xenografted thyroid cancer cells that carried PI3K/Akt-activating genetic alterations (151-153). However, other studies have shown that cells that harbor p85 mutations, PTEN loss or HER2 amplification also show increased sensitivity to perifosine, which suggests that there is not yet available a good biomarker that predict sensitivity to Akt inhibitors (235). In addition, a number of phase II trials with perifosine as single agent have provided disappointing results in patients with different types of carcinoma, melanoma or sarcoma (236 and references therein). An alternative strategy that has been implemented is the use of perifosine in combination with chemotherapy or radiation therapy, with the aim to overcome the development of Akt-dependent drug resistance. However, definitive results from different phase I and II trials have not been disclosed yet.

PI3K, the most important upstream activator of Akt, also represents a therapeutic target for thyroid cancer. Two well-known PI3K inhibitors are the fungal metabolite wortmanin or LY294002. Wortmannin is an irreversible inhibitor (IC<sub>50</sub>  $\approx$  2 nM) that forms a covalent bond with a conserved lysine residue in the catalytic site (237); LY294002 is a classical reversible, ATP-competitive PI3K inhibitor (IC<sub>50</sub>=1.40  $\mu$ M) (238). Wortmanin or LY294002 suppress proliferation of thyroid cancer cells *in vitro* (91, 92), though these inhibitors show limited use *in vivo*. More recently, PI3K inhibitors with improved pharmaceutical properties and therapeutic indexes such as PWT-458 (239) and PX-866 (240, 241) have been developed for preclinical studies (224, 242). PX-866 has shown antitumor activity in multiple xenograft models with PIK3CA mutations or PTEN loss, though it was not active in cells harbouring simultaneous PIK3CA and Ras mutations (243).

In parallel high-throughput screenings have led to the discovery and development of novel selective PI3K inhibitors such as NVP-BE-Z235, an imidazo[4,5-c]quinoline derivative that inhibits both PI3K (IC<sub>50</sub>=4 nM against p110 $\alpha$ ) and mTOR (IC<sub>50</sub>=21 nM) (242). This compound potently inhibits proliferation of cancer cell lines with PIK3CA mutations or PTEN loss, though, also in this case, activity was not restricted to cells that harbour these alterations (243). Similarly, lung cancer initiated by mutant K-Ras was resistant to NVP-BE-Z235 (243, 244). Therefore, the picture that emerges from the available preclinical studies is that the presence of PIK3CA mutations or the loss of PTEN confers sensitivity to PI3K inhibitors (though also non-mutant cells may be sensitive) and that the presence of K-Ras mutations predicts resistance. Moreover, it remains to be seen whether p110 $\alpha$  mutation-specific inhibitors for cancer therapeutics can be identified.

As with PI3K, also the direct activator of Akt kinases, PDK1, has recently been proved to be a therapeutic target to block this oncogenic pathway. One of the most potent, but nonselective PDK1 kinase inhibitor reported so far is the staurosporine-based compound UCN





**Figure 5.** The PI3K-Akt pathway in cancer. Molecular alterations reported in the members of the PI3K-Akt pathway.\* Activating mutation; # gene amplification; ^ gene rearrangements (translocation/inversion); § inactivating mutations, LOH, promoter methylation. In the red boxes are indicated the novel compounds developed to inhibit the different members of the PI3K pathway.

01 (IC<sub>50</sub>=6–33 nM) (245). Originally developed as an inhibitor of protein kinase C, UCN-01 inhibits a broad array of kinases, including Akt and other members of the AGC family of enzymes such as PDK1 (IC<sub>50</sub>=491 nM for Akt) (246). Inhibition of of tumour growth through UCN-01-dependent inhibition of PDK1 has been observed in murine and human tumor xenografts (245). However, no significant antitumor activity has been reported when UCN-01 was tested in advanced cancer patients in phase I/II clinical trials, both as single agent and in combination with conventional chemotherapeutic drugs (247). In addition, toxic side effects as pulmonary toxicity, nausea/vomiting, lactic acidosis, insulin resistance after treatment with UCN-01 were reported. The aminopyrimidine derivate BX-320, an ATP-competitive inhibitor has also been reported to inhibit PDK1 kinase activity (IC<sub>50</sub>=39 nM) with good selectivity (248). BX-320 blocks the growth in soft agar of a wide range of tumor cell lines (IC<sub>50</sub>=0.093–1.32 μM), and is efficacious in a metastasis mouse model when administered orally. More recently, the identification and the optimization of an indoline-based compound series of PDK1 inhibitors have been reported (BX-517; IC<sub>50</sub>=6 nM) (249, 250). Finally, inhibitory activity against PDK1 has also been reported for compounds originally designed to antagonize different therapeutic targets (i.e. celecoxib, a cyclooxygenase-2 inhibitor) in a variety of cancer cells through inhibition of PDK1 (251). Celecoxib itself is currently being investigated in phase II/III clinical studies as a single agent or in combination therapy.

Another attractive target in the PI3K-Akt pathway is mTOR, one of the most relevant downstream effector of Akt in thyroid cancer. Rapamycin, a well known mTOR inhibitor, has been shown to inhibit cell proliferation of thyroid cancer cells *in vitro* (252) as well as to inhibit proliferation of thyroid cells in thyroid targeted *Pten*-null mice *in vivo* (144). The mTOR inhibitors CCI-

779 (temsirolimus) and RAD-001 (everolimus) are potent inhibitors of mTORC1 complex and thus suppress ribosomal protein and cap-dependent translation (252). mTOR inhibitors have been tested as single agents in phase II studies in a variety of cancer types with promising results in breast carcinoma, glioblastoma, renal cell carcinoma and lymphoma (236 and references therein). At the moment both everolimus and temsirolimus have been approved in the United States as front-line therapy of resistant renal cell carcinoma (236).

Finally, the available data from different preclinical studies provide compelling evidence that the combination of PI3K and MEK inhibitors would be a rational approach in patients showing simultaneous activation of these pathways. The combined treatment with MEK/MAPK and PI3K inhibitors are particularly efficient in inhibiting K-Ras driven lung cancer in mouse (253) and in xenografted human basal-like breast cancer cells (254). Importantly, a collaborative effort has been launched to study the antitumor effects of the combination of MK-2206 (an allosteric Akt inhibitor) and AZD6244 (a MEK inhibitor), two early-stage compounds from Merck and AstraZeneca, respectively (227 and references therein).

Because PI3K activation in advanced thyroid cancer can occur in combination with increased signalling through the MAPK pathway, there has been a growing interest in defining a combination regimen that inhibits both pathways for patients with aggressive thyroid cancers. In particular this approach may be particularly relevant for tumours harbouring genetic alterations of both pathways (B-Raf and PIK3CA or B-Raf and Akt1, respectively). Accordingly, preclinical studies with compound [K-rasG12D/*pten*(-/-)] mice showed that combined pharmacological inhibition of PI3K (LY294002) and MEK/MAPK (UO-126) completely inhibited the growth of double mutant cancer cell lines, providing a compelling rationale for the simultaneous targeting of these pathways in thyroid cancer (73). See Figure 5 for a summary of the compounds targeted to the different members of the PI3K-Akt pathway.

## 7. CONCLUSIONS

In conclusion, genetic alterations that activate the PI3K-Akt pathway, including rearrangement or amplification of tyrosine kinase receptors, Ras mutations, PTEN loss and mutations and amplification of PIK3CA and Akt1, are common in thyroid cancer. Importantly these genetic alterations are preferentially observed in aggressive cancers such as PDC and ATC, which account for most of the incurable cases of thyroid cancer. Therefore it appears that targeting PI3K-Akt pathway in patients with advanced thyroid cancer may represent a rationale approach to improve survival. For this reason, several specific PI3K, Akt or mTOR inhibitors have been developed and many of these inhibitors currently under clinical evaluation represent a promising approach for the treatment of thyroid cancer patients. However, evidence continues to accumulate that the different Akt isoforms have diverse and sometimes even opposing functions in different cell settings. Therefore, to develop effective new compounds

that limit relevant Akt signalling in thyroid cancer cells, it will be important to gain a more complete picture of the precise roles of the PI3K/Akt pathway and, in particular, of the different kinase isoforms in the specific context of PDC and ATC. Moreover, the complex regulation of the PI3K-Akt pathway poses practical issues concerning the design of clinical trials, potential toxicities and criteria for patient selection. Further studies are needed to develop more effective single agents or to identify rational combinations of therapeutic targets that have synergistic effectiveness without enhanced toxicities.

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## 9. REFERENCES

1. L Leenhardt, P Grosclaude, L Chérié-Challine; Thyroid Cancer Committee: Increased incidence of thyroid carcinoma in France: a true epidemic or thyroid nodule management effects? Report from the French Thyroid Cancer Committee. *Thyroid* 14, 1056-60 (2004)
2. L Davies, HG Welch: Increasing incidence of thyroid cancer in the United States, 1973-2002. *JAMA* 295, 2164-7 (2006)
3. RA DeLellis, RV Lloyd, PU Heitz, C Eng (Eds): Pathology and Genetics of Tumours of Endocrine Organs. World Health Organization Classification of Tumours. *IARC Press*, 137-46 (2004)
4. S Cooper, GM Doherty, BR Haugen, RT Kloos, SL Lee, SJ Mandel, EL Mazzaferri, B McIver, SI Sherman, RM Tuttle; American Thyroid Association Guidelines Taskforce: Management guidelines for patients with thyroid nodules and differentiated thyroid cancer. *Thyroid* 16, 109-42 (2006)
5. A Stojadinovic, R Ghossein, A Hoos, MJ Urist, RH Spiro, JP Shah, MF Brennan, AR Shaha, B Singh: Hürthle cell carcinoma: a critical histopathological appraisal. *J Clin Oncol* 19, 2616-2625 (2001)
6. J Rosai, M Caracangui, R DeLellis: Tumors of the thyroid gland. In: Atlas of Tumor Pathology, 3rd series, fascicle. Eds: J. Rosai and L. H. Sobin (1992)
7. KB Ain: Management of undifferentiated thyroid cancer. *Baillieres Best Pract Res Clin Endocrinol Metab* 14, 615-29 (2000)
8. WR Cornett, AK Sharma, TA Day, MS Richardson, RS Hoda, JA van Heerden, J K Fernandes: Anaplastic thyroid carcinoma: an overview. *Curr Oncol Rep* 9, 152-8 (2007)
9. MN Nikiforova, YE Nikiforov: Molecular genetics of thyroid cancer: implications for diagnosis, treatment and prognosis. *Expert Rev Mol Diagn* 8, 83-95 (2008)
10. M Xing: Recent advances in molecular biology of thyroid cancer and their clinical implications. *Otolaryngol Clin North Am* 41, 1135-46 (2008)
11. PP Roger, WC van Staveren, K Coulonval, JE Dumont, C Maenhaut: Signal transduction in the human thyrocyte and its perversion in thyroid tumors. *Mol Cell Endocrinol* 321, 3-19 (2010)
12. T Pilli, KV Prasad, S Jayarama, F Pacini, BS Prabhakar: Potential utility and limitations of thyroid cancer cell lines as models for studying thyroid cancer. *Thyroid* 19, 1333-42 (2009)
13. RE Schweppe, JP Kloppe, C Korch, U Pugazhenth, M Benezra, JA Knauf, JA Fagin, LA Marlow, JA Copland, RC Smallridge, BR Haugen: Deoxyribonucleic acid profiling analysis of 40 human thyroid cancer cell lines reveals cross-contamination resulting in cell line redundancy and misidentification. *J Clin Endocrinol Metab* 93, 4331-41 (2008)
14. WC van Staveren, DW Solís, L Delys, L Duprez, G Andry, B Franc, G Thomas, F Libert, JE Dumont, V Detours, C Maenhaut: Human thyroid tumor cell lines derived from different tumor types present a common dedifferentiated phenotype. *Cancer Res* 67, 8113-20 (2007)
15. MD Ringel: "Thyroid cancer" cell line misidentification: a time for proactive change. *J Clin Endocrinol Metab* 93, 4226-7 (2008)
16. CS Kim, X Zhu: Lessons from mouse models of thyroid cancer. *Thyroid* 9, 1317-31 (2009)
17. B Vanhaesebroeck, MD Waterfield: Signaling by distinct classes of phosphoinositide 3-kinases. *Exp Cell Res* 253, 239-54 (1999)
18. SR Datta, A Brunet, ME Greenberg: Cellular survival: a play in three Acts. *Genes Dev* 13, 2905-27 (1999)
19. I Vivanco, CL Sawyers: The phosphatidylinositol 3-Kinase AKT pathway in human cancer. *Nat Rev Cancer* 2, 489-501 (2002)
20. JR Testa, A Bellacosa: AKT plays a central role in tumorigenesis. *Proc Natl Acad Sci U S A* 98, 10983-5 (2001)
21. DP Brazil, ZZYang, BA Hemmings: Advances in protein kinase B signalling: AKTion on multiple fronts. *Trends Biochem Sci* 29, 233-42 (2004)
22. J Okano, I Gaslightwala, MJ Birnbaum, AK Rustgi, H Nakagawa: Akt/protein kinase B isoforms are

differentially regulated by epidermal growth factor stimulation. *J Biol Chem* 275, 30934-42 (2000)

23. WS Chen, PZ Xu, K Gottlob, ML Chen, K Sokol, T Shiyanova, I Roninson, W Weng, R Suzuki, K Tobe, T Kadowaki, N Hay: Growth retardation and increased apoptosis in mice with homozygous disruption of the Akt1 gene. *Genes Dev* 15, 2203-8 (2001)

24. H Cho, JL Thorvaldsen, Q Chu, F Feng, MJ Birnbaum: Akt1/PKBalpha is required for normal growth but dispensable for maintenance of glucose homeostasis in mice. *J Biol Chem* 276, 38349-52 (2001)

25. H Cho, J Mu, JK Kim, JL Thorvaldsen, Q Chu, EB Crenshaw 3rd, KH Kaestner, MS Bartolomei, GI Shulman, MJ Birnbaum: Insulin resistance and a diabetes mellitus-like syndrome in mice lacking the protein kinase Akt2 (PKB beta). *Science* 292, 1728-31 (2001)

26. RS Garofalo, SJ Orena, K Rafidi, AJ Torchia, JL Stock, AL Hildebrandt, T Coskran, SC Black, DJ Brees, JR Wicks, JD McNeish, KG Coleman: Severe diabetes, age-dependent loss of adipose tissue, and mild growth deficiency in mice lacking Akt2/PKB beta. *J Clin Invest* 112, 197-208 (2003)

27. O Tschopp, ZZ Yang, D Brodbeck, BA Dümmler, M Hemmings-Mieszczak, T Watanabe, T Michaelis, J Frahm, BA Hemmings: Essential role of protein kinase B gamma (PKB gamma/Akt3) in postnatal brain development but not in glucose homeostasis. *Development* 132, 2943-54 (2005)

28. XD Peng, PZ Xu, ML Chen, A Hahn-Windgassen, J Skeen, J Jacobs, D Sundararajan, WS Chen, SE Crawford, KG Coleman, N Hay: Dwarfism, impaired skin development, skeletal muscle atrophy, delayed bone development, and impeded adipogenesis in mice lacking Akt1 and Akt2. *Genes Dev* 17, 1352-65 (2003)

29. ZZ Yang, O Tschopp, N Di-Poï, E Bruder, A Baudry, B Dümmler, W Wahli, BA Hemmings: Dosage-dependent effects of Akt1/protein kinase Balpha (PKBalpha) and Akt3/PKBgamma on thymus, skin, and cardiovascular and nervous system development in mice. *Mol Cell Biol* 25, 10407-18 (2005)

30. B Dümmler, O Tschopp, D Hynx, ZZ Yang, S Dirnhofer, BA Hemmings: Life with a single isoform of Akt: mice lacking Akt2 and Akt3 are viable but display impaired glucose homeostasis and growth deficiencies. *Mol Cell Biol* 26, 8042-51 (2006)

31. RM Easton, H Cho, K Roovers, DW Shineman, M Mizrahi, MS Forman, VM Lee, M Szabolcs, R de Jong, T Oltersdorf, L Ludwig, A Efstratiadis, MJ Birnbaum: Role for Akt3/protein kinase Bgamma in attainment of normal brain size. *Mol Cell Biol* 25, 1869-78 (2005)

32. HY Irie, RV Pearline, D Grueneberg, M Hsia, P Ravichandran, N Kothari, S Natesan, JS Brugge: Distinct roles of Akt1 and Akt2 in regulating cell migration and epithelial-mesenchymal transition. *J Cell Biol* 171, 1023-34 (2005)

33. IG Maroulakou, W Oemler, SP Naber, PN Tschlis: Akt1 ablation inhibits, whereas Akt2 ablation accelerates, the development of mammary adenocarcinomas in mouse mammary tumor virus (MMTV)-ErbB2/neu and MMTV-polyoma middle T transgenic mice. *Cancer Res* 67, 167-77 (2007)

34. M Yoeli-Lerner, GK Yiu, I Rabinovitz, P Erhardt, S Jauliac, A Toker: Akt blocks breast cancer cell motility and invasion through the transcription factor NFAT. *Mol Cell* 20, 539-50 (2005)

35. E Bousquet, J Mazieres, M Privat, V Rizzati, A Casanova, A Ledoux, E Mery, B Couderc, G Favre, A Pradines: Loss of RhoB Expression Promotes Migration and Invasion of Human Bronchial Cells Via Activation of AKT1. *Cancer Res* 69, 6092-6099 (2009)

36. EK Kim, SJ Yun, KH Do, MS Kim, D Suh, CD Kim, JH Kim, MJ Birnbaum, SS Bae: Lysophosphatidic acid induces cell migration through the selective activation of AKT1. *Exp Mol Med* 40, 445-452 (2008)

37. V Vasko, M Saji, E Hardy, M Kruhlak, A Larin, V Savchenko, M Miyakawa, O Isozaki, H Murakami, T Tsushima, KD Burman, C De Micco, MD Ringel: Akt activation and localisation correlate with tumour invasion and oncogene expression in thyroid cancer. *J Med Genet* 41, 161-70 (2004)

38. IG Maroulakou, W Oemler, SP Naber, I Klebba, C Kuperwasser, PN Tschlis: Distinct roles of the three Akt isoforms in lactogenic differentiation and involution. *J Cell Physiol* 217, 468-77 (2008)

39. H Konishi, S Kuroda, M Tanaka, H Matsuzaki, Y Ono, K Kameyama, T Haga, U Kikkawa: Molecular cloning and characterization of a new member of the RAC protein kinase family: association of the pleckstrin homology domain of three types of RAC protein kinase with protein kinase C subspecies and beta gamma subunits of G proteins. *Biochem Biophys Res Commun* 216, 526-34 (1995)

40. MD Ringel, N Hayre, J Saito, B Saunier, F Schuppert, H Burch, V Bernet, KD Burman, LD Kohn, M Saji: Overexpression and overactivation of Akt in thyroid carcinoma. *Cancer Res* 61, 6105-11 (2001)

41. TF Franke, DR Kaplan, LC Cantley, A Toker: Direct regulation of the Akt proto-oncogene product by phosphatidylinositol-3,4-bisphosphate. *Science* 275, 665-8 (1997)

42. DR Alessi: Discovery of PDK1, one of the missing links in insulin signal transduction. Colworth Medal Lecture. *Biochem Soc Trans* 29, 1-14 (2001)
43. DD Sarbassov, DA Guertin, SM Ali, DM Sabatini: Phosphorylation and regulation of Akt/PKB by the rictor-mTOR complex. *Science* 307, 1098–101 (2005)
44. BD Manning, LC Cantley: AKT/PKB signaling: navigating downstream. *Cell* 29, 1261-74 (2007)
45. T Gao, F Furnari, AC Newton: PHLPP: a phosphatase that directly dephosphorylates Akt, promotes apoptosis, and suppresses tumor growth. *Mol Cell* 18, 13-24 (2005)
46. LC Trotman, A Alimonti, PP Scaglioni, JA Koutcher, C Cordon-Cardo, PP Pandolfi: Identification of a tumour suppressor network opposing nuclear Akt function. *Nature* 441, 523-7 (2006)
47. JA Engelman, J Luo, LC Cantley: The evolution of phosphatidylinositol 3-kinases as regulators of growth and metabolism. *Nat Rev Genet* 7, 606-19 (2006)
48. H Lempiäinen, TD Halazonetis: Emerging common themes in regulation of PIKKs and PI3Ks. *EMBO J* 28, 3067-73 (2009)
49. R Williams, A Berndt, S Miller, WC Hon, X Zhang: Form and flexibility in phosphoinositide 3-kinases. *Biochem Soc Trans* 37, 615-26 (2009)
50. RJ Cain, AJ Ridley: Phosphoinositide 3-kinases in cell migration. *Biol Cell* 101, 13-29 (2009)
51. L Shayesteh, Y Lu, WL Kuo, R Baldocchi, T Godfrey, C Collins, D Pinkel, B Powell, GB Mills, JW Gray: PIK3CA is implicated as an oncogene in ovarian cancer. *Nat Genet* 21, 99-102 (1999)
52. PK Vogt, S Kang, MA Elsliger, M Gymnopoulos: Cancer-specific mutations in phosphatidylinositol 3-kinase. *Trends Biochem Sci* 32, 342-9 (2007)
53. JP Gustin, B Karakas, MB Weiss, AM Abukhdeir, J Lauring, JP Garay, D Cosgrove, A Tamaki, H Konishi, Y Konishi, M Mohseni, G Wang, DM Rosen, SR Denmeade, MJ Higgins, MI Vitolo, KE Bachman, BH Park: Knockin of mutant PIK3CA activates multiple oncogenic pathways. *Proc Natl Acad Sci U S A* 106, 2835-40 (2009)
54. KM Vasudevan, DA Barbie, MA Davies, R Rabinovsky, CJ McNear, JJ Kim, BT Hennessy, H Tseng, P Pochanard, SY Kim, IF Dunn, AC Schinzler, P Sandy, S Hoersch, Q Sheng, PB Gupta, JS Boehm, JH Reiling, S Silver, Y Lu, K Stemke-Hale, B Dutta, C Joy, AA Sahin, AM Gonzalez-Angulo, A Lluch, LE Rameh, T Jacks, DE Root, ES Lander, GB Mills, WC Hahn, WR Sellers, LA Garraway: AKT-independent signaling downstream of oncogenic PIK3CA mutations in human cancer. *Cancer Cell* 16, 21-32 (2009)
55. DM Li, H Sun: TEP1, encoded by a candidate tumor suppressor locus, is a novel protein tyrosine phosphatase regulated by transforming growth factor beta. *Cancer Res* 57, 2124-9 (1997)
56. J Li, C Yen, D Liaw, K Podsypanina, S Bose, SI Wang, J Puc, C Miliareis, L Rodgers, R McCombie, SH Bigner, BC Giovanella, M Ittmann, B Tycko, H Hibshoosh, MH Wigler, R Parsons: PTEN, a putative protein tyrosine phosphatase gene mutated in human brain, breast, and prostate cancer. *Science* 275, 1943-7 (1997)
57. PA Steck, MA Pershouse, SA Jasser, WK Yung, H Lin, AH Ligon, LA Langford, ML Baumgard, T Hattier, T Davis, C Frye, R Hu, B Swedlund, DH Teng, SV Tavtigian: Identification of a candidate tumour suppressor gene, MMAC1, at chromosome 10q23.3 that is mutated in multiple advanced cancers. *Nat Genet* 15, 356-62 (1997)
58. LC Cantley, BG Neel: New insights into tumor suppression: PTEN suppresses tumor formation by restraining the phosphoinositide 3-kinase/AKT pathway. *Proc Natl Acad Sci U S A* 96, 4240-5 (1999)
59. F Vazquez, WR Sellers: The PTEN tumor suppressor protein: an antagonist of phosphoinositide 3-kinase signaling. *Biochim Biophys Acta* 1470, M21-35 (2000)
60. T Maehama, JE Dixon: The tumor suppressor, PTEN/MMAC1, dephosphorylates the lipid second messenger, phosphatidylinositol 3,4,5-trisphosphate. *J Biol Chem* 273, 13375-8 (1998)
61. V Stambolic, A Suzuki, JL de la Pompa, GM Brothers, C Mirtsos, T Sasaki, J Ruland, JM Penninger, DP Siderovski, TW Mak: Negative regulation of PKB/Akt-dependent cell survival by the tumor suppressor PTEN. *Cell* 95, 29-39 (1998)
62. H Sun, R Lesche, DM Li, J Liliental, H Zhang, J Gao, N Gavrilova, B Mueller, X Li, H Wu: PTEN modulates cell cycle progression and cell survival by regulating phosphatidylinositol 3,4,5-trisphosphate and Akt/protein kinase B signaling pathway. *Proc Natl Acad Sci U S A* 96, 6199-204 (1999)
63. A Di Cristofano and PP Pandolfi: The multiple roles of PTEN in tumor suppression. *Cell* 100, 387-90 (2000)
64. JA Hobert, C Eng: PTEN hamartoma tumor syndrome: an overview. *Genet Med* 11, 687-94 (2009)
65. TD Bunney, M Katan: Phosphoinositide signalling in cancer: beyond PI3K and PTEN. *Nat Rev Cancer* 10, 342-52 (2010)
66. F Meng, R Henson, H Wehbe-Janek, K Ghoshal, ST Jacob, T Patel: MicroRNA-21 regulates expression of the PTEN tumor suppressor gene in human hepatocellular cancer. *Gastroenterology* 133, 647-58 (2007)

67. N Amodio, M Scrima, L Palaia, A Naeem Salman, A Quintiero, R Franco, G Botti, P Pirozzi, G Rocco, N De Rosa, G Viglietto: Oncogenic role of NEDD4-1, a PTEN-negative regulator, in non small lung carcinomas (NSCLC). *Am J Pathol*. In press (2010)
68. A Di Cristofano, B Pesce, C Cordon-Cardo, PP Pandolfi: Pten is essential for embryonic development and tumour suppression. *Nat Genet* 19, 348-55 (1998)
69. K Podsypanina, LH Ellenson, A Nemes, J Gu, M Tamura, KM Yamada, C Cordon-Cardo, G Catoletti, PE Fisher, R Parsons: Mutation of Pten/Mmac1 in mice causes neoplasia in multiple organ systems. *Proc Natl Acad Sci U S A* 96, 1563-8 (1999)
70. A Suzuki, JL de la Pompa, V Stambolic, AJ Elia, T Sasaki, I del Barco Barrantes, A Ho, A Wakeham, A Itie, W Khoo, M Fukumoto, TW Mak: High cancer susceptibility and embryonic lethality associated with mutation of the PTEN tumor suppressor gene in mice. *Curr Biol* 8, 1169-78 (1998)
71. V Stambolic, MS Tsao, D Macpherson, A Suzuki, WB Chapman, TW Mak: High incidence of breast and endometrial neoplasia resembling human Cowden syndrome in pten<sup>+/−</sup> mice. *Cancer Res* 60, 3605-11 (2000)
72. L Buday, J Downward: Many faces of Ras activation. *Biochim Biophys Acta* 1786, 178-87 (2008)
73. KA Miller, N Yeager, K Baker, XH Liao, S Refetoff, A Di Cristofano: Oncogenic Kras requires simultaneous PI3K signaling to induce ERK activation and transform thyroid epithelial cells *in vivo*. *Cancer Res* 69, 3689-94 (2009)
74. SJ Cohen, L Ho, S Ranganathan, JL Abbruzzese, RK Alpaugh, M Beard, NL Lewis, S McLaughlin, A Rogatko, JJ Perez-Ruixo, AM Thistle, T Verhaeghe, H Wang, LM Weiner, JJ Wright, GR Hudes, NJ Meropol: Phase II and pharmacodynamic study of the farnesyltransferase inhibitor R115777 as initial therapy in patients with metastatic pancreatic adenocarcinoma. *J Clin Oncol* 21, 1301-6 (2003)
75. DW Ball, N Jin, DM Rosen, A Dackiw, D Sidransky, M Xing, BD Nelkin: Selective growth inhibition in BRAF mutant thyroid cancer by the mitogen-activated protein kinase kinase 1/2 inhibitor AZD6244. *J Clin Endocrinol Metab* 92, 4712-8 (2007)
76. JE Dumont, F Lamy, P Roger, C Maenhaut: Physiological and pathological regulation of thyroid cell proliferation and differentiation by thyrotropin and other factors. *Physiol Rev* 72, 667-697 (1992)
77. K Tanaka, Y Nagayama, T Nakano, N Takamura, H Namba, S Fukada, K Kuma, S Yamashita, M Niwa: Expression profile of receptor-type protein tyrosine kinase genes in the human thyroid. *Endocrinology* 139, 852-8 (1998)
78. MC Eggo, MC Sheppard: Autocrine growth factors produced in the thyroid. *Mol Cell Endocrinol* 100, 97-102 (1994)
79. BF Van der Laan, JL Freeman, SL Asa: Expression of growth factors and growth factor receptors in normal and tumorous human thyroid tissues. *Thyroid* 5, 67-73 (1995)
80. XP Pang, JM Hershman: Differential effects of growth factors on [3H] thymidine incorporation and [125I] iodine uptake in FRTL-5 rat thyroid cells. *Proc Soc Exp Biol Med* 194, 240- 244 (1990)
81. T Kimura, JE Dumont, A Fusco, J Golstein: Insulin and TSH promote growth in size of PC Cl3 rat thyroid cells, possibly via a pathway different from DNA synthesis: comparison with FRTL-5 cells. *Eur J Endocrinol* 140, 94-103 (1999)
82. S Dremier, M Taton, K Coulonval, T Nakamura, K Matsumoto, JE Dumont: Mitogenic, dedifferentiating, and scattering effects of hepatocyte growth factor on dog thyroid cells. *Endocrinology* 135, 135-140 (1994)
83. M Santoro, F Carlomagno, ID Hay, MA Herrmann, M Grieco, R Melillo, MA Pierotti, I Bongarzone, G Della Porta, N Berger, JL Peix, C Paulin, N Fabien, G Vecchio, RB Jenkins, A Fusco: Ret oncogene activation in human thyroid neoplasms is restricted to the papillary cancer subtype. *J Clin Invest* 89, 1517-22 (1992)
84. YE Nikiforov, JM Rowland, KE Bove, H Monforte-Munoz, JA Fagin: Distinct pattern of ret oncogene rearrangements in morphological variants of radiation-induced and sporadic thyroid papillary carcinomas in children. *Cancer Res* 57, 1690-4 (1997)
85. CL Fenton, Y Lukes, D Nicholson, CA Dinuer, GL Francis, RM Tuttle: The ret/PTC mutations are common in sporadic papillary thyroid carcinoma of children and young adults. *J Clin Endocrinol Metab* 85, 1170-5 (2000)
86. P Soares, E Fonseca, D Wynford-Thomas, M Sobrinho-Simões: Sporadic ret-rearranged papillary carcinoma of the thyroid: a subset of slow growing, less aggressive thyroid neoplasms? *J Pathol* 18, 71-8 (1998)
87. MA Pierotti: Chromosomal rearrangements in thyroid carcinomas: a recombination or death dilemma. *Cancer Lett* 166, 1-7 (2001)
88. I Bongarzone, P Vigneri, L Mariani, P Collini, S Pilotti, MA Pierotti: RET/NTRK1 rearrangements in thyroid gland tumors of the papillary carcinoma family: correlation with clinicopathological features. *Clin Cancer Res* 4, 223-8 (1998)
89. E Miyagi, M Braga-Basaria, E Hardy, V Vasko, KD Burman, S Jhiang, M Saji, MD Ringel: Chronic expression of RET/PTC 3 enhances basal and insulin-stimulated PI3 kinase/AKT signaling and increases IRS-2 expression in FRTL-5 thyroid cells. *Mol Carcinog* 41, 98-107 (2004)

90. R Mineo, A Costantino, F Frasca, L Sciacca, S Russo, R Vigneri, A Belfiore: Activation of the hepatocyte growth factor (HGF)-Met system in papillary thyroid cancer: biological effects of HGF in thyroid cancer cells depend on Met expression levels. *Endocrinology* 145, 4355-65 (2004)
91. P Bruni, A Boccia, G Baldassarre, F Trapasso, M Santoro, G Chiappetta, A Fusco, G Viglietto: PTEN expression is reduced in a subset of sporadic thyroid carcinomas: evidence that PTEN-growth suppressing activity in thyroid cancer cells mediated by p27kip1. *Oncogene* 19, 3146-55 (2000)
92. ML Motti, D Califano, G Troncone, C De Marco, I Migliaccio, E Palmieri, L Pezzullo, L Palombini, A Fusco, G Viglietto: Complex regulation of the cyclin-dependent kinase inhibitor p27kip1 in thyroid cancer cells by the PI3K/AKT pathway: regulation of p27kip1 expression and localization. *Am J Pathol* 166, 737-49 (2005)
93. G García-Rostán, AM Costa, I Pereira-Castro, G Salvatore, R Hernandez, MJ Hermsem, A Herrero, A Fusco, J Cameselle-Teijeiro, M Santoro: Mutation of the PIK3CA gene in anaplastic thyroid cancer. *Cancer Res* 65, 10199-207 (2005)
94. JC Ricarte-Filho, M Ryder, DA Chitale, M Rivera, A Heguy, M Ladanyi, M Janakiraman, D Solit, JA Knauf, RM Tuttle, RA Ghossein, JA Fagin: Mutational profile of advanced primary and metastatic radioactive iodine-refractory thyroid cancers reveals distinct pathogenetic roles for BRAF, PIK3CA, and AKT1. *Cancer Res* 69, 4885-93 (2009)
95. M Miyakawa, T Tsushima, H Murakami, K Wakai, O Isozaki, K Takano: Increased expression of phosphorylated p70S6 kinase and Akt in papillary thyroid cancer tissues. *Endocr J* 50, 77-83 (2003)
96. J Abubaker, J Jehan, P Bavi, M Sultana, S Al-Harbi, M Ibrahim, A Al-Nuaim, M Ahmed, T Amin, M Al-Fehaily, O Al-Sanea, F Al-Dayel, S Uddin, KS Al-Kuraya: Clinicopathological analysis of papillary thyroid cancer with PIK3CA alterations in a Middle Eastern population. *J Clin Endocrinol Metab* 93, 611-8 (2008)
97. Z Liu, P Hou, M Ji, H Guan, K Studeman, K Jensen, V Vasko, AK El-Naggar, M Xing: Highly prevalent genetic alterations in receptor tyrosine kinases and phosphatidylinositol 3-kinase/akt and mitogen-activated protein kinase pathways in anaplastic and follicular thyroid cancers. *J Clin Endocrinol Metab* 93, 3106-16 (2008)
98. JD Carpten, AL Faber, C Horn, GP Donoho, SL Briggs, CM Robbins, G Hostetter, S Boguslawski, TY Moses, S Savage, K Uhlik, A Lin, J Du, YW Qian, DJ Zeckner, G Tucker-Kellogg, J Touchman, K Patel, S Mousses, M Bittner, R Schevitz, MH Lai, KL Blanchard, JE Thomas: A transforming mutation in the pleckstrin homology domain of AKT1 in cancer. *Nature* 448, 439-44 (2007)
99. MA Davies, K Stemke-Hale, C Tellez, CL Calderone, W Deng, VG Prieto, AJ Lazar, JE Gershenwald, GB Mills: A novel AKT3 mutation in melanoma tumours and cell lines. *Br J Cancer* 99, 1265-8 (2008)
100. D Malanga, M Scrima, C De Marco, F Fabiani, N De Rosa, S De Gisi, N Malara, R Savino, G Rocco, G Chiappetta, R Franco, V Tirino, G Pirozzi, G Viglietto: Activating E17K mutation in the gene encoding the protein kinase AKT1 in a subset of squamous cell carcinoma of the lung. *Cell Cycle* 7, 665-9 (2008)
101. FE Bleeker, L Felicioni, F Buttitta, S Lamba, L Cardone, M Rodolfo, A Scarpa, S Leenstra, M Frattini, M Barbareschi, MD Grammasio, MG Sciarrotta, C Zanon, A Marchetti, A Bardelli: AKT1(E17K) in human solid tumours. *Oncogene* 27, 5648-50 (2008)
102. K Shoji, K Oda, S Nakagawa, S Hosokawa, G Nagae, Y Uehara, K Sone, Y Miyamoto, H Hiraie, O Hiraie-Wada, T Nei, K Kawana, H Kuramoto, H Aburatani, T Yano, Y Taketani: The oncogenic mutation in the pleckstrin homology domain of AKT1 in endometrial carcinomas. *Br J Cancer* 101, 145-8 (2009)
103. JM Askham, F Platt, PA Chambers, H Snowden, CF Taylor, MA Knowles: AKT1 mutations in bladder cancer: identification of a novel oncogenic mutation that can co-operate with E17K. *Oncogene* 29, 150-5 (2010)
104. J Lauring, DP Cosgrove, S Fontana, JP Gustin, H Konishi, AM Abukhdeir, JP Garay, Mohseni, GM Wang, MJ Higgins, D Gorkin, M Reis, B Vogelstein, K Polyak, Cowherd, PJ Buckhaults, BH Park: Knock in of the AKT1 E17K mutation in human breast epithelial cells does not recapitulate oncogenic PIK3CA mutations. *Oncogene* 29, 2337-2345 (2010)
105. G Wu, E Mambo, Z Guo, S Hu, X Huang, SM Gollin, B Trink, PW Ladenson, D Sidransky, M Xing: Uncommon mutation, but common amplifications, of the PIK3CA gene in thyroid tumors. *J Clin Endocrinol Metab* 90, 4688-93 (2005)
106. P Hou, D Liu, Y Shan, S Hu, K Studeman, S Condouris, Y Wang, A Trink, AK El-Naggar, G Tallini, V Vasko, M Xing: Genetic alterations and their relationship in the phosphatidylinositol 3-kinase/Akt pathway in thyroid cancer. *Clin Cancer Res* 13, 1161-1170 (2007)
107. Y Wang, P Hou, H Yu, W Wang, M Ji, S Zhao, S Yan, X Sun, D Liu, B Shi, G Zhu, S Condouris, M Xing: High prevalence and mutual exclusivity of genetic alterations in the phosphatidylinositol-3-kinase/akt pathway in thyroid tumors. *J Clin Endocrinol Metab* 92, 2387-2390 (2007).
108. P Hou, M Ji, M Xing: Association of PTEN gene methylation with genetic alterations in the phosphatidylinositol 3-kinase/AKT signaling pathway in thyroid tumors. *Cancer* 113, 2440-7 (2008)
109. L Santarpia, AK El-Naggar, GJ Cote, JN Myers, SI Sherman: Phosphatidylinositol 3-kinase/akt and ras/raf-



mitogen-activated protein kinase pathway mutations in anaplastic thyroid cancer. *J Clin Endocrinol Metab* 93, 278-84 (2008)

110. AG Bader, S Kang, PK Vogt: Cancer-specific mutations in PIK3CA are oncogenic *in vivo*. *Proc Natl Acad Sci U S A* 103, 1475-9 (2006)

111. JA Engelman, L Chen, X Tan, K Crosby, AR Guimaraes, R Upadhyay, M Maira, K McNamara, SA Perera, Y Song, LR Chirieac, R Kaur, A Lightbown, J Simendinger, T Li, RF Padera, C Garcia-Echeverria, R Weissleder, U Mahmood, LC Cantley, KK Wong: Effective use of PI3K and MEK inhibitors to treat mutant Kras G12D and PIK3CA H1047R murine lung cancers. *Nat Med* 14, 1351-6 (2008)

112. S Liang, N Yang, Y Pan, S Deng, X Lin, X Yang, D Katsaros, KF Roby, TC Hamilton, DC Connolly, G Coukos, L Zhang: Expression of activated PIK3CA in ovarian surface epithelium results in hyperplasia but not tumor formation. *PLoS One* 4, e4295 (2001)

113. CJ Guigon, L Zhao, MC Willingham, SY Cheng: PTEN deficiency accelerates tumour progression in a mouse model of thyroid cancer. *Oncogene* 28, 509-517 (2009)

114. Y Samuels, Z Wang, A Bardelli, N Silliman, J Ptak, S Szabo, Y Yan, A Gazdar, SM Powell, GJ Riggins, JK Willson, S Markowitz, KW Kinzler, B Vogelstein, VE Velculescu: High frequency of mutations of the PIK3CA gene in human cancers. *Science* 304, 554 (2004)

115. N Halachmi, S Halachmi, E Evron, P Cairns, K Okami, M Saji, WH Westra, MA Zeiger, J Jen, D Sidransky: Somatic mutations of the PTEN tumor suppressor gene in sporadic follicular thyroid tumors. *Genes Chromosomes Cancer* 23, 239-43 (1998)

116. PL Dahia, DJ Marsh, Z Zheng, J Zedenius, P Komminoth, T Frisk, G Wallin, R Parsons, M Longy, C Larsson, C Eng: Somatic deletions and mutations in the Cowden disease gene, PTEN, in sporadic thyroid tumors. *Cancer Res* 57, 4710-3 (1997)

117. JJ Yeh, DJ Marsh, J Zedenius, T Dwight, L Delbridge, BG Robinson, C Eng: Fine-structure deletion mapping of 10q22-24 identifies regions of loss of heterozygosity and suggests that sporadic follicular thyroid adenomas and follicular thyroid carcinomas develop along distinct neoplastic pathways. *Genes Chromosomes Cancer* 26, 322-8 (1999)

118. O Gimm, A Perren, LP Weng, DJ Marsh, JJ Yeh, U Ziebold, E Gil, R Hinze, L Delbridge, JA Lees, GL Mutter, BG Robinson, P Komminoth, H Dralle, C Eng: Differential nuclear and cytoplasmic expression of PTEN in normal thyroid tissue, and benign and malignant epithelial thyroid tumors. *Am J Pathol* 156, 1693-700 (2000)

119. LP Weng, WM Smith, JL Brown, C Eng: PTEN inhibits insulin-stimulated MEK/MAPK activation and cell

growth by blocking IRS-1 phosphorylation and IRS-1/Grb-2/Sos complex formation in a breast cancer model. *Hum Mol Genet* 10, 605-16 (2001)

120. F Alvarez-Nuñez, E Bussaglia, D Mauricio, J Ybarra, M Vilar, E Lerma, A de Leiva, X Matias-Guiu; Thyroid Neoplasia Study Group: PTEN promoter methylation in sporadic thyroid carcinomas. *Thyroid* 16, 17-23 (2006)

121. U Schagdarsurengin, O Gimm, H Dralle, C Hoang-Vu, R Dammann: CpG island methylation of tumor-related promoters occurs preferentially in undifferentiated carcinoma. *Thyroid* 16(7), 633-42 (2006)

122. V Vasko, M Ferrand, J Di Cristofaro, P Carayon, JF Henry, C de Micco: Specific pattern of RAS oncogene mutations in follicular thyroid tumors. *J Clin Endocrinol Metab* 88, 2745-52 (2003)

123. JM Shields, K Pruitt, A McFall, A Shaub, CJ Der: Understanding Ras: 'it ain't over 'til it's over'. *Trends Cell Biol* 10, 147-54 (2000)

124. G Cheng, JL Meinkoth: Enhanced sensitivity to apoptosis in Ras-transformed thyroid cells. *Oncogene* 20, 7334-41 (2001)

125. V Gire and D Wynford-Thomas: RAS oncogene activation induces proliferation in normal human thyroid epithelial cells without loss of differentiation. *Oncogene* 9, 737-44 (2000)

126. V Gire, C Marshall, D Wynford-Thomas: PI-3-kinase is an essential anti-apoptotic effector in the proliferative response of primary human epithelial cells to mutant RAS. *Oncogene* 19, 2269-76 (2000)

127. RM Melillo, F Carlomagno, G De Vita, P Formisano, G Vecchio, A Fusco, M Billaud, M Santoro: The insulin receptor substrate (IRS)-1 recruits phosphatidylinositol 3-kinase to Ret: evidence for a competition between Shc and IRS-1 for the binding to Ret. *Oncogene* 20, 209-18 (2001)

128. M Lodyga, V De Falco, XH Bai, A Kapus, RM Melillo, M Santoro, M Liu: XB130, a tissue-specific adaptor protein that couples the RET/PTC oncogenic kinase to PI 3-kinase pathway. *Oncogene* 28, 937-49 (2009)

129. V De Falco, V Guarino, L Malorni, AM Cirafici, F Troglio, M Erreni, G Pelicci, M Santoro, RM Melillo: RAI(ShcC/N-Shc)-dependent recruitment of GAB 1 to RET oncoproteins potentiates PI 3-K signalling in thyroid tumors. *Oncogene* 24, 6303-13 (2005)

130. DW Kim, JH Hwang, JM Suh, H Kim, JH Song, ES Hwang, IY Hwang, KC Park, HK Chung, JM Kim, J Park, BA Hemmings, M Shong: RET/PTC (rearranged in transformation/papillary thyroid carcinomas) tyrosine kinase phosphorylates and activates phosphoinositide-dependent kinase 1 (PDK1): an alternative phosphatidylinositol 3-kinase-independent pathway to activate PDK1. *Mol Endocrinol* 17, 1382-94 (2003)

131. HS Jung, DW Kim, YS Jo, HK Chung, JH Song, JS Park, KC Park, SH Park, JH Hwang, KW Jo, M. Shong: Regulation of protein kinase B tyrosine phosphorylation by thyroid-specific oncogenic RET/PTC kinases. *Mol Endocrinol* 19, 2748-59 (2005)
132. AK Siraj, P Bavi, J Abubaker, Z Jehan, M Sultana, F Al-Dayel, A Al-Nuaim, A Alzahrani, M Ahmed, O Al-Sanea, S Uddin, KS Al-Kuraya: Genome-wide expression analysis of Middle Eastern papillary thyroid cancer reveals c-MET as a novel target for cancer therapy. *J Pathol* 213, 190-9 (2007)
133. MB Yaffe, K Rittinger, S Volinia, PR Caron, A Aitken, H Leffers, SJ Gamblin, SJ Smerdon, LCCantley: The structural basis for 14-3-3:phosphopeptide binding specificity. *Cell* 91, 961-71 (1997)
134. S Wullschleger, R Loewith, MN Hall: TOR signaling in growth and metabolism. *Cell* 124, 471-84 (2006)
135. D Ruggero, N Sonenberg: The Akt of translational control. *Oncogene* 24, 7426-34 (2005)
136. MA Bjornsti, PJ Houghton: Lost in translation: dysregulation of cap-dependent translation and cancer. *Cancer Cell* 5,19-23 (2004)
137. D Ruggero, PP Pandolfi: Does the ribosome translate cancer? *Nat Rev Cancer* 3, 179-92 (2003)
138. DR Plas, CB Thompson: Akt-dependent transformation: there is more to growth than just surviving. *Oncogene* 24, 7435-42 (2005)
139. H Seeliger, M Guba, A Kleespies, KW Jauch, CJ Bruns: Role of mTOR in solid tumor systems: a therapeutic target against primary tumor growth, metastases, and angiogenesis. *Cancer Metastasis Rev* 26, 611-21 (2007)
140. A Astrinidis, EP Henske: Tuberous sclerosis complex: linking growth and energy signaling pathways with human disease. *Oncogene* 24, 7475-81 (2005)
141. RT Abraham, JJ Gibbons: The mammalian target of rapamycin signaling pathway: twists and turns in the road to cancer therapy. *Clin Cancer Res* 13, 3109-14 (2007)
142. DA Guertin, DM Sabatini: Defining the role of mTOR in cancer. *Cancer Cell* 12, 9-22 (2007)
143. DD Sarbassov, SM Ali, S Sengupta, JH Sheen, PP Hsu, AF Bagley, AL Markhard, DM Sabatini: Prolonged rapamycin treatment inhibits mTORC2 assembly and Akt/PKB. *Mol Cell* 22, 159- 68 (2006)
144. C Brewer, N Yeager, A Di Cristofano: Thyroid-stimulating hormone initiated proliferative signals converge *in vivo* on the mTOR kinase without activating AKT. *Cancer Res* 67, 8002-6 (2007)
145. EW Arvisais, A Romanelli, X Hou, JS Davis: AKT-independent phosphorylation of TSC2 and activation of mTOR and ribosomal protein S6 kinase signaling by prostaglandin F2{alpha}. *J Biol Chem* 281, 26904-13 (2006)
146. P Wlodarski, M Kasprzycka, X Liu, M Marzec, ES Robertson, A Slupianek, MA Wasik: Activation of mammalian target of rapamycin in transformed B lymphocytes is nutrient dependent but independent of Akt, mitogen-activated protein kinase/extracellular signal-regulated kinase kinase, insulin growth factor-I, and serum. *Cancer Res* 65, 7800-8 (2005)
147. LY Wing, HM Chen, PC Chuang, MH Wu, SJ Tsai: The mammalian target of rapamycin-p70 ribosomal S6 kinase but not phosphatidylinositol 3-kinase-Akt signaling is responsible for fibroblast growth factor-9-induced cell proliferation. *J Biol Chem* 280, 19937-47 (2005)
148. EC Souza, AS Figueiredo, WM Braga, BM Andrade, M Vaisman, LE Nasciutti, AC Ferreira, DP Carvalho: Mammalian target of rapamycin (mTOR) down-regulates iodide uptake in thyrocytes. *J Endocrinol* 206, 113-20 (2010)
149. N Yeager, C Brewer, KQ Cai, XX Xu, A Di Cristofano: Mammalian target of rapamycin is the key effector of phosphatidylinositol-3-OH-initiated proliferative signals in the thyroid follicular epithelium. *Cancer Res*, 68(2), 444-9 (2008)
150. F Furuya, C Lu, MC Willingham, SY Cheng: Inhibition of phosphatidylinositol 3-kinase delays tumor progression and blocks metastatic spread in a mouse model of thyroid cancer. *Carcinogenesis* 28, 2451-8 (2007)
151. D Liu, P Hou, Z Liu, G Wu, M Xing: Genetic alterations in the phosphoinositide 3-kinase/Akt signaling pathway confer sensitivity of thyroid cancer cells to therapeutic targeting of Akt and mammalian target of rapamycin. *Cancer Res* 69, 7311-9 (2009)
152. N Jin, T Jiang, DM Rosen, BD Nelkin, DW Ball: Dual inhibition of mitogen-activated protein kinase kinase and mammalian target of rapamycin in differentiated and anaplastic thyroid cancer. *J Clin Endocrinol Metab* 94, 4107-4112 (2009)
153. C Papewalis, M Wuttke, S Schinner, HS Willenberg, AM Baran, WA Scherbaum, M Schott: Role of the novel mTOR inhibitor RAD001 (everolimus) in anaplastic thyroid cancer. *Horm Metab Res* 41, 752-6 (2009)
154. S Cory, DC Huang, JM Adams: The Bcl-2 family: roles in cell survival and oncogenesis. *Oncogene* 22, 8590-8607 (2003)
155. NN Danial, SJ Korsmeyer: Cell death: critical control points. *Cell* 116, 205- 219 (2004)
156. SR Datta, A Katsov, L Hu, A Petros, SW Fesik, MB Yaffe, ME Greenberg: 14-3-3 proteins and survival kinases

cooperate to inactivate BAD by BH3 domain phosphorylation. *Mol Cell* 6, 41-51 (2000)

157. L del Peso, M González-García, C Page, R Herrera, G Nuñez: Interleukin-3-induced phosphorylation of BAD through the protein kinase Akt. *Science* 278, 687-9 (1997)

158. GJ Kops, BM Burgering: Forkhead transcription factors: new insights into protein kinase B (c-akt) signaling. *J Mol Med* 77, 656-65 (1999)

159. GJ Kops, ND de Ruiter, AM De Vries-Smits, DR Powell, JL Bos, BM Burgering: Direct control of the Forkhead transcription factor AFX by protein kinase B. *Nature* 398, 630-4 (1999)

160. WH Biggs, J Meisenhelder, T Hunter, WK Cavenee, KC Arden: Protein kinase B/Akt-mediated phosphorylation promotes nuclear exclusion of the winged helix transcription factor FKHR1. *Proc Natl Acad Sci U S A* 96, 7421-6 (1999)

161. A Brunet, A Bonni, MJ Zigmond, MZ Lin, P Juo, LS Hu, MJ Anderson, KC Arden, J Blenis, ME Greenberg: Akt promotes cell survival by phosphorylating and inhibiting a Forkhead transcription factor. *Cell* 96, 857-68 (1999)

162. G Rena, S Guo, SC Cichy, TG Unterman, P Cohen: Phosphorylation of the transcription factor forkhead family member FKHR by protein kinase B. *J Biol Chem* 274, 17179-83 (1999)

163. H Tran, A Brunet, EC Griffith, ME Greenberg: The many forks in FOXO's road. *Sci STKE* 2003, RE5 (2003)

164. N Nakamura, S Ramaswamy, F Vazquez, S Signoretti, M Loda, WR Sellers: Forkhead transcription factors are critical effectors of cell death and cell cycle arrest downstream of PTEN. *Mol Cell Biol* 20, 8969-82 (2000)

165. RH Medema, GJ Kops, JL Bos, BM Burgering: AFX-like Forkhead transcription factors mediate cell-cycle regulation by Ras and PKB through p27kip1. *Nature* 404, 782-7 (2000)

166. PF Dijkers, RH Medema, JW Lammers, L Koenderman, PJ Coffey: Expression of the pro-apoptotic Bcl-2 family member Bim is regulated by the forkhead transcription factor FKHR-L1. *Curr Biol* 10, 1201-1204 (2000)

167. G De Vita, MT Berlingieri, R Visconti, MD Castellone, G Viglietto, G Baldassarre, M Zannini, A Bellacosa, PN Tschlis, A Fusco, M Santoro: Akt/protein kinase B promotes survival and hormone-independent proliferation of thyroid cells in the absence of dedifferentiating and transforming effects. *Cancer Res* 60, 3916-20 (2000)

168. LD Mayo, DB Donner: A phosphatidylinositol 3-kinase/Akt pathway promotes translocation of Mdm2 from

the cytoplasm to the nucleus. *Proc Natl Acad Sci U S A* 98, 11598-603 (2001)

169. BP Zhou, Y Liao, W Xia, Y Zou, B Spohn, MC Hung: HER-2/neu induces p53 ubiquitination via Akt-mediated MDM2 phosphorylation. *Nat Cell Biol* 4, 736 (2001)

170. A Villunger, EM Michalak, L Coultas, F Müllauer, G Böck, MJ Ausserlechner, JM Adams, A Strasser: p53- and drug-induced apoptotic responses mediated by BH3-only proteins puma and noxa. *Science* 302, 1036-8 (2003)

171. LP Kane, VS Shapiro, D Stokoe, A Weiss: Induction of NF-kappaB by the Akt/PKB kinase. *Curr Biol* 9, 601-4 (1999)

172. A Khwaja: Akt is more than just a Bad kinase. *Nature* 401, 33-4 (1999)

173. ON Ozes, LD Mayo, JA Gustin, SR Pfeffer, LM Pfeffer, DB Donner: NF-kappaB activation by tumour necrosis factor requires the Akt serine-threonine kinase. *Nature* 401, 82-5 (1999)

174. JA Romashkova, SS Makarov: NF-kappaB is a target of AKT in anti-apoptotic PDGF signalling. *Nature* 401, 86-90 (1999)

175. S Ghosh, MJ May, EB Kopp: NF-kB and Rel proteins: evolutionarily conserved mediators of immune responses. *Annu Rev Immunol* 16, 225-60 (1998)

176. E Zandi, M Karin: Bridging the gap: composition, regulation, and physiological function of the IκB kinase complex. *Mol Cell Biol* 19, 4547-51 (1999)

177. M Karin, M Delhase: The IκB kinase (IKK) and NF-κB: key elements of proinflammatory signalling. *Semin Immunol* 12, 85-98 (2000)

178. CY Wang, MW Mayo, RG Korneluk, DV Goeddel, AS Baldwin: NF-κB antiapoptosis: induction of TRAF1 and TRAF2 and c-IAP1 and c-IAP2 to suppress caspase-8 activation. *Science* 281, 1680-3 (1998)

179. MX Wu, Z Ao, KV Prasad, R Wu, SF Schlossman: IEX-1L, an apoptosis inhibitor involved in NF-κB-mediated cell survival. *Science* 281, 998-1001 (1998)

180. RC Bargou, F Emmerich, D Krappmann, K Bommert, MY Mapara, W Arnold, HD Royer, E Grinstein, A Greiner, C Scheidereit, B Dörken: Constitutive nuclear factor-κB-RelA activation is required for proliferation and survival of Hodgkins disease tumor cells. *J Clin Invest* 100, 2961-9 (1997)

181. DC Duffey, Z Chen, G Dong, FG Ondrey, JS Wolf, K Brown, U Siebenlist, C van Waes: Expression of a dominant-negative mutant inhibitor-κBa of nuclear factor-κB in human head and neck squamous cell carcinoma

inhibits survival, proinflammatory cytokine expression, and tumor growth *in vivo*. *Cancer Res* 59, 3468-74 (1999)

182. S Huang, CA Pettaway, H Uehara, CD Bucana and IJ Fidler: Blockade of NF- $\kappa$ B activity in human prostate cancer cells is associated with suppression of angiogenesis, invasion, and metastasis. *Oncogene* 20, 4188-97 (2001)

183. JA Romashkova, SS Makarov: NF- $\kappa$ B is a target of AKT in anti-apoptotic PDGF signalling. *Nature* 401, 86-90 (1999)

184. D Bai, L Ueno, PK Vogt: Akt-mediated regulation of NF $\kappa$ B and the essentialness of NF $\kappa$ B for the oncogenicity of PI3K and Akt. *Int J Cancer* 125, 2863-70 (2009)

185. S Vasudevan, N Garneau, D Tu Khounh, SW Peltz: p38 mitogen-activated protein kinase/Hog1p regulates translation of the AU-rich-element-bearing MFA2 transcript. *Mol Cell Biol* 25, 9753-63 (2005)

186. CY Wang, WB Zhong, TC Chang, SM Lai, YF Tsai: Tumor necrosis factor alpha induces three-dimensional cytomorphologic differentiation of human anaplastic thyroid carcinoma cells through activation of nuclear factor kappaB. *Cancer* 95, 1827-33 (2002)

187. I Palona, H Namba, N Mitsutake, D Starenki, A Podtcheko, I Sedliarou, A Ohtsuru, V Saenko, Y Nagayama, K Umezawa, S Yamashita: BRAFV600E promotes invasiveness of thyroid cancer cells through nuclear factor kB activation. *Endocrinology* 147, 5699-707 (2006)

188. H Namba, V Saenko, S Yamashita: Nuclear factor-kB in thyroid carcinogenesis and progression: a novel therapeutic target for advanced thyroid cancer. *Arq Bras Endocrinol Metab* 51, 843-851 (2007)

189. R Visconti, J Cerutti, S Battista, M Fedele, F Trapasso, K Zeki, MP Miano, F de Nigris, L Casalino, F Curcio, M Santoro, A Fusco: Expression of the neoplastic phenotype by human thyroid carcinoma cell lines requires NF $\kappa$ B p65 protein expression. *Oncogene* 15, 1987-94 (1997)

190. D Starenki, H Namba, V Saenko, A Ohtsuru, S Yamashita: Inhibition of nuclear factor-kB cascade potentiates the effect of a combination treatment of anaplastic thyroid cancer cells. *J Clin Endocrinol Metab* 89, 410-8 (2004)

191. F Pacifico, C Mauro, C Barone, E Crescenzi, S Mellone, M Monaco, G Chiappetta, G Terrazzano, D Liguoro, P Vito, E Consiglio, S Formisano, A Leonardi: Oncogenic and anti-apoptotic activity of NF- $\kappa$ B in human thyroid carcinomas. *J Biol Chem* 279, 54610-9 (2004)

192. CS Mitsiades, V Kotoula, V Poulaki, E Sozopoulos, J Negri, E Charalambous, G Fanourakis, G Voutsinas, S Tseloni-Balafouta, N Mitsiades: Epidermal growth factor

receptor as a therapeutic target in human thyroid carcinoma: mutational and functional analysis. *J Clin Endocrinol Metab* 91, 3662-6 (2006)

193. T Bauerle, RE Schweppe, BR Haugen: Inhibition of nuclear factor-kappa B differentially affects thyroid cancer cell growth, apoptosis, and invasion. *Mol Cancer* 9, 117 (2010)

194. M Festa, A Petrella, S Alfano, L Parente: R-roscovitine sensitizes anaplastic thyroid carcinoma cells to TRAIL-induced apoptosis via regulation of IKK/NF-kappaB pathway. *Int J Cancer* 124, 2728-36 (2009)

195. Z Meng, N Mitsutake, M Nakashima, D Starenki, M Matsuse, S Takakura, H Namba, V Saenko, K Umezawa, A Ohtsuru, S Yamashita: Dehydroxymethylepoxyquinomicin, a novel nuclear Factor-kappaB inhibitor, enhances antitumor activity of taxanes in anaplastic thyroid cancer cells. *Endocrinology* 149, 5357-65 (2008)

196. W Zhu, Y Ou, Y Li, R Xiao, M Shu, Y Zhou, J Xie, S He, P Qiu, G Yan. A small-molecule triptolide suppresses angiogenesis and invasion of human anaplastic thyroid carcinoma cells via down-regulation of the nuclear factor-kappa B pathway. *Mol Pharmacol* 75, 812-9 (2009)

197. J Saito, AD Kohn, RA Roth, Y Noguchi, I Tatsumo, A Hirai, K Suzuki, LD Kohn, M Saji, MD Ringel: Regulation of FRTL-5 thyroid cell growth by phosphatidylinositol (OH) 3 kinase-dependent Akt-mediated signaling. *Thyroid* 11, 339-351 (2001)

198. LP Weng, O Gimm, JB Kum, WM Smith, XP Zhou, D Wynford-Thomas, G Leone, C Eng: Transient ectopic expression of PTEN in thyroid cancer cell lines induces cell cycle arrest and cell type-dependent cell death. *Hum Mol Genet*, 10(3), 251-8 (2001)

199. J Liang, J Zubovitz, T Petrocelli, R Kotchetkov, MK Connor, K Han, JH Lee, S Ciarallo, S Catzavelos, R Beniston, E Franssen, JM Slingerland: PKB/Akt phosphorylates p27, impairs nuclear import of p27 and opposes p27-mediated G1 arrest. *Nat Med* 8, 1153-60 (2002)

200. G Viglietto, ML Motti, P Bruni, RM Melillo, A D'Alessio, D Califano, F Vinci, G Chiappetta, P Tschlis, A Bellacosa, A Fusco, M Santoro: Cytoplasmic relocation and inhibition of the cyclin-dependent kinase inhibitor p27(Kip1) by PKB/Akt-mediated phosphorylation in breast cancer. *Nat Med* 8, 1136-44 (2002)

201. N Fujita, S Sato, K Katayama, T Tsuruo: Akt-dependent phosphorylation of p27Kip1 promotes binding to 14-3-3 and cytoplasmic localization. *J Biol Chem* 277, 28706-13 (2002)

202. ML Motti, C De Marco, D Califano, A Fusco, G Viglietto: Akt-dependent T198 phosphorylation of

- cyclin-dependent kinase inhibitor p27kip1 in breast cancer. *Cell Cycle* 3, 1074-80 (2004)
203. I Shin, J Rotty, FY Wu, CL Arteaga: Phosphorylation of p27Kip1 at Thr-157 interferes with its association with importin alpha during G1 and prevents nuclear re-entry. *J Biol Chem* 280, 6055-63 (2005)
204. T Sekimoto, M Fukumoto, Y Yoneda: 14-3-3 suppresses the nuclear localization of threonine 157-phosphorylated p27(Kip1). *EMBO J* 23, 1934-42 (2004)
205. G Viglietto, ML Motti, A Fusco: Understanding p27(kip1) deregulation in cancer: down-regulation or mislocalization. *Cell Cycle* 1, 394-400 (2002)
206. A Besson, HC Hwang, S Cicero, SL Donovan, M Gurian-West, D Johnson, BE Clurman, MA Dyer, JM Roberts: Discovery of an oncogenic activity in p27Kip1 that causes stem cell expansion and a multiple tumor phenotype. *Genes Dev* 21, 1731-46 (2007)
207. A Besson, M Gurian-West, A Schmidt, A Hall, JM Roberts: p27Kip1 modulates cell migration through the regulation of RhoA activation. *Genes Dev* 18, 862-76 (2004)
208. S Karger, C Weidinger, K Krause, SY Sheu, T Aigner, O Gimm, KW Schmid, H Dralle, D Fuhrer: FOXO3a: a novel player in thyroid carcinogenesis? *Endocr Relat Cancer* 16, 189-99 (2009)
209. G Chiappetta, C De Marco, A Quintiero, D Califano, S Gherardi, D Malanga, M Scrima, C Montero-Conde, L Cito, M Monaco, ML Motti, R Pasquinelli, V Agosti, M Robledo, A Fusco, G Viglietto: Overexpression of the S-phase kinase-associated protein 2 in thyroid cancer. *Endocr Relat Cancer* 14, 405-20 (2007)
210. ML Motti, C De Marco, D Califano, S De Gisi, D Malanga, G Troncone, A Persico, S Losito, F Fabiani, M Santoro, G Chiappetta, A Fusco, G Viglietto: Loss of p27 expression through RAS->BRAF->MAP kinase-dependent pathway in human thyroid carcinomas. *Cell Cycle* 6, 2817-25 (2007)
211. D Vitagliano, F Carlomagno, ML Motti, G Viglietto, YE Nikiforov, MN Nikiforova, JM Hershman, AJ Ryan, A Fusco, RM Melillo, M Santoro: Regulation of p27Kip1 protein levels contributes to mitogenic effects of the RET/PTC kinase in thyroid carcinoma cells. *Cancer Res* 64, 3823-9 (2004)
212. M Fedele, D Palmieri, G Chiappetta, R Pasquinelli, I De Martino, C Arra, G Palma, T Valentino, GM Pierantoni, G Viglietto, JL Rothstein, M Santoro, A Fusco: Impairment of the p27kip1 function enhances thyroid carcinogenesis in TRK-T1 transgenic mice. *Endocr Relat Cancer* 16, 483-90 (2009)
213. DA Cross, DR Alessi, P Cohen, M Andjelkovich, BA Hemmings: Inhibition of glycogen synthase kinase-3 by insulin mediated by protein kinase B. *Nature* 378, 785-9 (1995)
214. JA Diehl, M Cheng, MF Roussel, CJ Sherr: Glycogen synthase kinase-3beta regulates cyclin D1 proteolysis and subcellular localization. *Genes Dev* 12, 3499-511 (1998)
215. M Welcker, J Singer, KR Loeb, J Grim, A Bloecher, M Gurien-West, BE Clurman, JM Roberts: Multisite phosphorylation by Cdk2 and GSK3 controls cyclin E degradation. *Mol Cell* 12, 381-92 (2003)
216. ES Kandel, J Skeen, N Majewski, A Di Cristofano, PP Pandolfi, CS Feliciano, A Gartel, N Hay: Activation of Akt/protein kinase B overcomes a G(2)/M cell cycle checkpoint induced by DNA damage. *Mol Cell Biol* 22, 7831-41 (2002)
217. E Shtivelman, J Sussman, D Stokoe: A role for PI 3-kinase and PKB activity in the G2/M phase of the cell cycle. *Curr Biol* 12, 919-24 (2002)
218. FW King, J Skeen, N Hay, E Shtivelman: Inhibition of Chk1 by activated PKB/Akt. *Cell Cycle* 3, 34-7 (2004)
219. J Puc, M Keniry, HS Li, TK Pandita, AD Choudhury, L Memeo, M Mansukhani, VV Murty, Z Gaciong, SE Meek, H Piwnicka-Worms, H Hibshoosh, R Parsons: Lack of PTEN sequesters CHK1 and initiates genetic instability. *Cancer Cell* 7, 193-204 (2005)
220. A Abulaiti, AJ Fikaris, OM Tsygankova, JL Meinkoth: Ras induces chromosome instability and abrogation of the DNA damage response. *Cancer Res* 66, 10505-12 (2006)
221. AD Laird, JM Cherrington: Small molecule tyrosine kinase inhibitors: clinical development of anticancer agents. *Expert Opin Investig Drugs* 12, 51-64 (2003)
222. SI Sherman: Tyrosine kinase inhibitors and the thyroid. *Best Pract Res Clin Endocrinol Metab*, 713-722 (2009)
223. SI Sherman: Targeted therapy of thyroid cancer. *Biochem Pharmacol* 80, 592-601 (2010)
224. C Garcia-Echeverria, WR Sellers: Drug discovery approaches targeting the PI3K/Akt pathway in cancer. *Oncogene* 27, 5511-5526 (2008)
225. SI Sherman, LJ Wirth, JP Droz, M Hofmann, L Bastholt, RG Martins, L Licita, MJ Eschenberg, YN Sun, T Juan, DE Stepan and MJ Schlumberger: Motesanib diphosphate in progressive differentiated thyroid cancer. *N Engl J Med* 359, 31-42 (2008)
226. EE Cohen, LS Rosen, EE Vokes, MS Kies, AA Forastiere, FP Worden, MA Kane, E Sherman, S Kim, P

- Bycott, M Tortorici, DR Shalinsky, KF Liao, RB Cohen: Axitinib is an active treatment for all histologic subtypes of advanced thyroid cancer: results from a phase II study. *J Clin Oncol* 26, 4708–4713 (2008)
227. CW Lindsley: The Akt/PKB family of protein kinases: a review of small molecule inhibitors and progress towards target validation: a 2009 update. *Curr Top Med Chem* 10, 458–77 (2010)
228. Q Li, GD Zhu: Targeting serine/threonine protein kinase B/Akt and cell-cycle checkpoint kinases for treating cancer. *Curr Top Med Chem* 2, 939–971 (2002)
229. M Gassel, CB Breitenlechner, P Ruger, U Jucknischke, T Schneider, R Huber, D Bossemeyer, RA Engh: Mutants of protein kinase A that mimic the ATP-binding site of protein kinase B (Akt). *J Mol Biol* 329, 1021–1034 (2003)
230. CB Breitenlechner, WG Friebe, E Brunet, G Werner, K Graul, U Thomas, KP Kunkele, W Schafer, M Gassel, D Bossemeyer, R Huber, RA Engh, B Masjost: Design and crystal structures of protein kinase B-selective inhibitors in complex with protein kinase A and mutants. *J Med Chem* 48, 163–170 (2005)
231. CB Breitenlechner, T Wegge, L Berillon, K Graul, K Marzenell, WG Friebe, U Thomas, R Schumacher, R Huber, RA Engh, B. Masjost: Structure-based optimization of novel azepane derivatives as PKB inhibitors. *J Med Chem* 47, 1375–1390 (2004)
232. Y Luo, AR Shoemaker, X Liu, KW Woods, SA Thomas, R de Jong, EK Han, T Li, VS Stoll, JA Powlas, A Oleksijew, MJ Mitten, Y Shi, R Guan, TP McGonigal, V Klinghofer, EF Johnson, JD Levenson, JJ Bouska, M Mamo, RA Smith, EE Gramling-Evans, BA Zinker, AK Mika, PT Nguyen, T Oltersdorf, SH Rosenberg, Q Li, VL Giranda: Potent and selective inhibitors of Akt kinases slow the progress of tumors *in vivo*. *Mol Cancer Res* 4, 977–986 (2005)
233. Y Shi, X Liu, E Han, R Guan, AR Shoemaker, A Oleksijew, KW Woods, JP Fisher, V Klinghofer, L Lasko, T McGonigal, Q Li, SH Rosenberg, VL Giranda, Y Luo: Optimal classes of chemotherapeutic agents sensitized by specific small-molecule inhibitors of akt *in vitro* and *in vivo*. *Neoplasia* 7, 992–1000 (2005)
234. N Rhodes, DA Heerding, DR Duckett, DJ Eberwein, VB Knick, TJ Lansing, RT McConnell, TM Gilmer, SY Zhang, K Robell, JA Kahana, RS Geske, EV Kleymenova, AE Choudhry, Z Lai, JD Leber, EA Minthorn, SL Strum, ER Wood, PS Huang, RA Copeland, R Kumar: Characterization of an Akt kinase inhibitor with potent pharmacodynamic and antitumor activity. *Cancer Res* 68, 2366–2374 (2008)
235. BT Hennessy, Y Lu, E Poradosu, Q Yu, S Yu, H Hall, MS Carey, M Ravoory, AM Gonzalez-Angulo, R Birch, IC Henderson, V Kundra, GB Mills: Pharmacodynamic markers of perifosine efficacy. *Clin Cancer Res* 13, 7421–31 (2007)
236. J LoPiccolo, GM Blumenthal, WB Bernstein, PA Dennis: Targeting the PI3K/Akt/mTOR pathway: effective combinations and clinical considerations. *Drug Resist Updat* 11, 32–50 (2008)
237. MP Wymann, G Bulgarelli-Leva, MJ Zvelebil, L Pirola, B Vanhaesebroeck, MD Waterfield, G Panayotou: Wortmannin inactivates phosphoinositide 3-kinase by covalent modification of Lys-802, a residue involved in the phosphate transfer reaction. *Mol Cell Biol* 16, 1722–1733 (1996)
238. CJ Vlahos, WF Matter, KY Hui, RF Brown: A specific inhibitor of phosphatidylinositol 3-kinase, 2-(4-morpholinyl)-8-phenyl-4H-1-benzopyran-4-one (LY294002). *J Biol Chem* 269, 5241–5248 (1994)
239. K Yu, J Lucas, T Zhu, A Zask, C Gaydos, L Toral-Barza, J Gu, F Li, I Chaudhary, P Cai, J Lotvin, R Petersen, M Ruppen, M Fawzi, S Ayral-Kaloustian, J Skotnicki, T Mansour, P Frost, J Gibbons: PWT-458, a novel pegylated-17-dihydroxywortmannin, inhibits phosphatidylinositol 3-kinase signaling and suppresses growth of solid tumors. *Cancer Biol Ther* 4, 538–545 (2005)
240. NT Ihle, G Paine-Murrieta, MI Berggren, A Baker, WRTate, P Wipf, RT Abraham, DL Kirkpatrick, G Powis: The phosphatidylinositol-3-kinase inhibitor PX-866 overcomes resistance to the epidermal growth factor receptor inhibitor gefitinib in A-549 human non-small cell lung cancer xenografts. *Mol Cancer Ther* 4, 1349–1357 (2005)
241. AL Howes, GG Chiang, ES Lang, CB Ho, G Powis, K Vuori, RT Abraham: The phosphatidylinositol 3-kinase inhibitor, PX-866, is a potent inhibitor of cancer cell motility and growth in three-dimensional cultures. *Mol Cancer Ther* 6, 2505–2514 (2007)
242. S Jia, TM Roberts, JJ Zhao: Should individual PI3 kinase isoforms be targeted in cancer? *Curr Opin Cell Biol* 21, 199–208 (2009)
243. NT Ihle, R Lemos, P Wipf, A Yacoub, C Mitchell, D Siwak, GB Mills, P Dent, DL Kirkpatrick, G Powis: Mutations in the phosphatidylinositol-3-kinase pathway predict for antitumor activity of the inhibitor PX-866 whereas oncogenic Ras is a dominant predictor for resistance. *Cancer Res* 69, 143–50 (2009)
244. V Serra, B Markman, M Scaltriti, PJ Eichhorn, V Valero, M Guzman, ML Botero, E Llouch, F Atzori, S Di Cosimo, M Maira, C Garcia-Echeverria, JL Parra, J Arribas, J Baselga: NVP-BEZ235, a dual PI3K/mTOR inhibitor, prevents PI3K signaling and inhibits the growth of cancer cells with activating PI3K mutations. *Cancer Res* 68, 8022–30 (2008)



245. S Sato, N Fujita, T Tsuruo: Interference with PDK1-Akt survival signaling pathway by UCN-01 (7-hydroxystaurosporine). *Oncogene* 21, 1727-1738 (2002)

246. D Komander, GS Kular, J Bain, M Elliott, DR Alessi, DM Van Aalten: Structural basis for UCN-01 (7-hydroxystaurosporine) specificity and PDK1 (3-phosphoinositide-dependent protein kinase-1) inhibition. *Biochem J* 375, 255-262 (2003)

247. S Welch, HW Hirte, MS Carey, SJ Hotte, MS Tsao, S Brown, GR Pond, JE Dancey, AM Oza: UCN-01 in combination with topotecan in patients with advanced recurrent ovarian cancer: a study of the Princess Margaret Hospital Phase II consortium. *Gynecol Oncol* 106, 305-310 (2007)

248. RI Feldman, JM Wu, MA Polokoff, MJ Kochanny, H Dinter, D Zhu, SL Biroc, B Alicke, J Bryant, S Yuan, BO Buckman, D Lentz, M Ferrer, M Whitlow, M Adler, S Finster, Z Chang, DO Arnaiz: Novel small molecule inhibitors of 3-phosphoinositide-dependent kinase-1. *J Biol Chem* 280, 19867-19874 (2005)

249. I Islam, G Brown, J Bryant, P Hrvatin, MJ Kochanny, GB Phillips, S Yuan, M Adler, M Whitlow, D Lentz, MA Polokoff, J Wu, J Shen, J Walters, E Ho, B Subramanyam, D Zhu, RI Feldman, DO Arnaiz: Indolinone based phosphoinositide-dependent kinase-1 (PDK1) inhibitors. Part 2: optimization of BX-517. *Bioorg Med Chem Lett* 17, 3819-3825 (2007)

250. I Islam, J Bryant, YL Chou, MJ Kochanny, W Lee, GB Phillips, H Yu, M Adler, M Whitlow, E Ho, D Lentz, MA Polokoff, B Subramanyam, JM Wu, D Zhu, RI Feldman, DO Arnaiz: Indolinone based phosphoinositide-dependent kinase-1 (PDK1) inhibitors. Part 1: design, synthesis and biological activity. *Bioorg Med Chem Lett* 17, 3814-3818 (2007)

251. S Arico, S Pattingre, C Bauvy, P Gane, A Barbat, P Codogno, E Ogier-Denis: Celecoxib induces apoptosis by inhibiting 3-phosphoinositide-dependent protein kinase-1 activity in the human colon cancer HT-29 cell line. *J Biol Chem* 277, 27613-27621 (2002)

252. A Fasolo, C Sessa: mTOR inhibitors in the treatment of cancer. *Expert Opin Investig Drugs* 17, 1717-34 (2008)

253. JA Engelman, L Chen, X Tan, K Crosby, AR Guimaraes, R Upadhyay, M Maira, K McNamara, SA Perera, Y Song, LR Chirieac, R Kaur, A Lightbown, J Simendinger, T Li, RF Padera, C Garcia-Echeverria, R Weissleder, U Mahmood, LC Cantley, KK Wong. Effective use of PI3K and MEK inhibitors to treat mutant Kras G12D and PIK3CA H1047R murine lung cancers. *Nat Med* 14, 1351-6 (2008)

254. KP Hoeflich, C O'Brien, Z Boyd, G Cavet, S Guerrero, K Jung, T Januario, H Savage, E Punnoose, T Truong, W Zhou, L Berry, L Murray, L Amler, M Belvin, LS Friedman, MR Lackner: *In vivo* antitumor activity of

MEK and phosphatidylinositol 3-kinase inhibitors in basal-like breast cancer models. *Clin Cancer Res* 15, 4649-64 (2009)

255. N Yeager, A Klein-Szanto, S Kimura, A Di Cristofano: Pten loss in the mouse thyroid causes goiter and follicular adenomas: insights into thyroid function and Cowden disease pathogenesis. *Cancer Res* 67, 959-66 (2007)

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