

GnRH AND STEROIDS IN CANCER

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1. ABSTRACT

Gonadotropin-releasing-hormone (GnRH) analogues are synthetic compounds derived from decapeptide neurohormones (LHRH; LH/FSH-RH). They have a key role in hormone dependent cancer, particularly breast and prostate cancer. GnRH analogues produce an efficient inhibition of gonadotropins and sex steroid hormones. Their use in cancer therapy result in a, pharmacological castration (i.e. ovariectomy and orchiectomy), providing an androgen and estrogen ablation. GnRH exert an inhibitory action on the growth of hormone-dependent human and canine mammary tumor. Mammary tumors can produce growth factor that potentially could modulate their own proliferation in an autocrine fashion (i.e. TGF- α and TGF- β or with a paracrine mechanism (i.e. EGF, IGF, FGF). The expression of EGF receptors is related in mammary tissues to the action of oestrogen and progesteron and to the presence of functional receptors for oestrogen (ER) and progesteron (PR). The present review elucidate the role of

GnRH receptors in cancer and their connection with steroid hormones. Besides we showed the link between GnRH and signal transductions pathways: Estrogen-receptors, GnRH-receptors, EGF-receptors signal transduction pathways. A very tight link exists between steroid hormones and GnRH analogues both on central pituitary gonadal axis and on tumor receptors peripherically. This last mechanism could be explained either locally activating GnRH receptors or locally interacting with EGF receptor-Intracellular NitricOxide system.

2. INTRODUCTION

LHRH was first isolated, sequenced and synthesized in the early 1970s (1,2,3) and showed that both natural LHRH and the synthetic decapeptide corresponding to its structure possessed major follicle-stimulating hormone (FSH)-releasing as well as LH-

releasing activity (3). It was forecast that chemical substitutions in the molecule would lead to analogues possessing antagonist or increased agonist activity, useful as anti-and pro-fertility agents, respectively. Initial efforts were highly successful in generating highly potent agonist analogues: substitution of only one or two amino acids resulted in analogues with to 200 times the potency of the native molecule (4). However, the causes of this increase in potency, i.e. increased binding affinity and resistance to metabolism, accentuated the ability to shut down rather than simulate reproductive function.

The concept that LHRH regulates the secretion of both FSH and LH from the pituitary gland, is upheld by much experimental and clinical evidence (5,6). Indeed, it was originally suggested that the name LHRH be changed to GnRH for gonadotropin-releasing hormone (3,6).

In the past years, more than 3000 analogs of LHRH have been synthesized (5,7). Agonistic analogs, such as Decapeptyl, Zoladex, Leuprolide and Buserelin, much more active than the LHRH itself, and available in depot preparations, have important clinical application in gynecology and oncology (5). Potent antagonist of LHRH such as Cetrorelix, Ganerelix and Abarelix, suitable for clinical use, have been likewise synthesized (8,9,10).

2.1. GnRH chemical

Luteinizing hormone-releasing hormone (LHRH; LH/FSH-RH, GnRH) is an endogenous decapeptide neurohormone with an obligatory role in reproduction. In normal circumstances it is synthesized in hypothalamic neurones and secreted in a pulsatile fashion from neurones at the median eminence, traverses the hypothalamo-hypophysial portal system and interacts with its receptors on the gonadotrophs in a transient fashion to stimulate release (and synthesis) of the gonadotrophins (11). By binding to and by activating specific receptors on gonadotrophs, the neurohormone stimulates the synthesis and the release of the two gonadotropins LH and FSH (follicle-stimulating hormone) (12); for this reason it is also called GnRH (gonadotropin-releasing hormone). Hypothalamic neurosecretory cells release LHRH in a pulsatile way and LHRH pulses are critical for the maintenance of gonadotropin gene expression and for the physiological pattern of secretion of LH and FSH (13). The two gonadotropins are themselves secreted in a pulsatile way in the systemic circulation and act on the gonads to regulate gametogenesis and steroid synthesis (13).

The synthesis and secretion of both LHRH and gonadotropin are regulated by gonadal steroids which act at the hypothalamus and the pituitary level (14). The LHRH gene has been shown to carry steroid response elements (15,16). Moreover, estrogen (estrogen receptors β mainly) and androgen binding sites have been demonstrated to be expressed in a transgenically derived LHRH neuronal cell line (GT1) (17), which represents a very useful

experimental tool to study the regulation of LHRH gene expression.

The gene encoding hypothalamic LHRH has been identified in several species and is organized into four exons and three introns (16). This gene encodes for a preprohormone consisting of a signal peptide followed by the sequences of LHRH and by GAP (gonadotropin-releasing hormone associated peptide), separated by a canonical cleavage site (18). In vertebrate, the sequence of the different forms of LHRH is similar but not identical: amino acids in positions 5 – 8 show a reduced degree of conservation. Recently, a gene coding for a second form of LHRH (LHRH-II) has been identified in several placental mammals, including humans. LHRH-II encode a peptide identified as [His5Trp7Tyr8]LHRH, indicating that the amino acid sequence presents three substitution in these three position. LHRH-II is mainly present in the midbrain, while peripheral tissues expressing the peptide include the prostate, bone marrow and kidney (18). The existence of the two form of LHRH may indicate specific function for the peptides and suggest the possible presence of multiple receptor subtypes.

2.2. GnRH analogues

The elucidation of the crucial role played by GnRH in the control of the pituitary-gonadal system immediately underlined the relevance of its possible clinical applications for the treatment of several reproductive-related disease. However, GnRH is rapidly degraded at pituitary level and its half-life in the circulation is about 2 – 4 min. (20); moreover, the binding affinity of native GnRH to its pituitary receptor is quite low (K_d - 10-7M) (21). Therefore, efforts have been directed at obtaining GnRH analogues with increased stability against enzymatic attack and with higher affinity for the receptor. These studies led to the synthesis of two different class of GnRH analogues:

- GnRH agonists, binding with high affinity to the GnRH - R and mimicking the action of the neurohormone on pituitary function,
- GnRH antagonist, competing with GnRH for the binding to the receptor but devoid of intrinsic activity.

2.2.1. GnRH agonist

In past years, intensive analyses of the structure-activity relationship of the GnRH molecule have been performed. These studies revealed that both the NH_2 - and $COOH$ -terminal domain plays a crucial role in receptor activation (22). Moreover, although a lack of conservation of amino acids 5 – 8 has been reported in the primary structures of GnRH peptides in different species, ARG8 seems to be important for a high-affinity binding to the mammalian receptor (22). From the enzymatic point of view, it has been widely reported that the degradation of native GnRH occurs mainly at the Gly residue in position 6 (23). Based on these experimental observations, several synthetic GnRH agonists have been developed to be clinically used in the place of GnRH itself. Available GnRH agonist are mainly characterised by the presence of a D-amino acid in position 6; some of them present a

Table 1. Amino acid sequences of GnRH agonist

Peptide	Sequence
• GnRH	pGlu ¹ -His ² -Trp ³ -Ser ⁴ -Tyr ⁵ -Gly ⁶ -Leu ⁷ -Arg ⁸ -Pro ⁹ -Gly ¹⁰ -NH ₂
• Leuprolide	pGlu ¹ -His ² -Trp ³ -Ser ⁴ -Tyr ⁵ -D-Leu ⁶ -Leu ⁷ -Arg ⁸ -Pro ⁹ -NHC ₂ H ₅
• Buserelin	pGlu ¹ -His ² -Trp ³ -Ser ⁴ -Tyr ⁵ -D-Ser(tBu) ⁶ -Leu ⁷ -Arg ⁸ -Pro ⁹ -NHC ₂ H ₅
• Tryptorelin	pGlu ¹ -His ² -Trp ³ -Ser ⁴ -Tyr ⁵ -D-Trp ⁶ -Leu ⁷ -Arg ⁸ -Pro ⁹ -Gly ¹⁰ -NH ₂
• Goserelin	pGlu ¹ -His ² -Trp ³ -Ser ⁴ -Tyr ⁵ -D-Ser(tBu) ⁶ -Leu ⁷ -Arg ⁸ -Pro ⁹ -AzGly ¹⁰ -NH ₂

Table 2. Amino acid sequences of GnRH antagonists

Peptide	Sequence
• Nal-Glu- GnRH	Ac-D-Nal ¹ -D-Cpa ² -D-Pal ³ -Ser ⁴ -Arg ⁵ -D-Glu ⁶ (AA)-Leu ⁷ -Arg ⁸ -Pro ⁹ -D-Ala ¹⁰ -NH ₂
• Cetorelix	Ac-D-Nal ¹ -D-Cpa ² -D-Pal ³ -Ser ⁴ -Tyr ⁵ -D-Cit ⁶ -Leu ⁷ -Arg ⁸ -Pro ⁹ -D-Ala ¹⁰ -NH ₂
• Antide	Ac-D-Nal ¹ -D-Cpa ² -D-Pal ³ -Ser ⁴ -NicLys ⁵ -D-NicLys ⁶ -Leu ⁷ -ILys ⁸ -Pro ⁹ -D-Ala ¹⁰ -NH ₂
• Azaline B	Ac-D-Nal ¹ -D-Cpa ² -D-Pal ³ -Ser ⁴ -Aph ⁵ (Atz)-D-Aph ⁶ (Atz)-Leu ⁷ -ILys ⁸ -Pro ⁹ -D-Ala ¹⁰ -NH ₂
• Ganirelix	Ac-D-Nal ¹ -D-Cpa ² -D-Pal ³ -Ser ⁴ -Tyr ⁵ -D-hArg(Et2) ⁶ -Leu ⁷ -hArg(Et2) ⁸ -Pro ⁹ -D-Ala ¹⁰ -NH ₂

deleted carboxy-terminal Gly10- amide with the addition of an ethylamidic residue to Pro9 (Table 1). These compounds share the same biological activity of the native decapeptide but are more resistant to enzymatic degradation and bind with high affinity (K_d - 10^{-9} M) to the pituitary receptor. This affinity can be further increased by the introduction of hydrophobic groups on the sixth amino acid (24).

At pituitary level, GnRH agonist elicit the same biological activities observed after GnRH stimulation. A low dose or a pulsatile administration of GnRH agonist leads to receptor activation and to stimulation of gonadotropin secretion. On the other hand, a prolonged application of GnRH agonist induces, after an initial release of LH and FSH (the so-called “flare-up phenomenon”), the downregulation and desensitization of the GnRH receptor, resulting in the complete suppression of pituitary-gonadal functions. This latter effect is called “chemical castration” and represent the molecular basis for several important clinical applications of these compounds (24).

2.2.2. GnRH antagonists

The search for antagonistic analogues of GnRH was initially driven by the desire to develop new family of regulatory compounds with higher specificity and lower toxicity than steroidal contraceptives. Antagonists of GnRH generally present multiple amino acid substitutions at positions 1,2,3,6,8 and 10 (Table 2).

By competing with the native hormone, GnRH antagonists were determined to be unsuitable for clinical applications because of their undesired side effects, such as oedematogenic as anaphylactoid reactions, due to concomitant stimulation of histamine release (25). More recently, new generations of GnRH antagonists have been developed; these present a similar complex structure with several amino acid substitutions but appear to be free of anaphylactoid side effects. However, in order to continuously counteract the activity of the endogenous hormone, these compounds must be administered at high doses and in the form of long-acting preparations. Therefore, possible limitations of the use of GnRH antagonists may reside in some of their low solubility and propensity to form gels aqueous solutions. Efforts have been made to develop antagonistic analogues of GnRH devoid of side effects and with increased solubility (26).

Cetorelix, Ganirelix and Abarelix (a short acting antagonist with low histamine-releasing activity) are now available as long acting depot formulations (26,27) and have been already tested in long-term clinical trials (28,29). More recently, more soluble GnRH antagonists have been obtained by increasing the number of hydrogen binding sites on the peptide side chains. Among these compounds, FE200486 has been reported to possess a very long duration of action, likely to be due to the unique physicochemical properties, such as solubility in aqueous milieu, low propensity to form gels and ability to diffuse slowly from the s.c.sites of administration (30). Further studies might optimise the application of these compounds for the treatment of those reproductive-related pathologies which require a complete blockade of the pituitary-gonadal axis.

3 GnRH CLINICAL USE IN CANCER

The action of GnRH and its analogs are mediated by high-affinity receptors for GnRH found on the membranes of the pituitary gonadotrophs. An acute administration of GnRH agonists induces a marked release of LH and FSH. However, continuous stimulation of the pituitary by chronic administration of GnRH agonist produce an inhibition of the hypophyseal-gonadal axis through the process of “down-regulation” of pituitary receptors for GnRH, desensitization of the pituitary gonadotrophs, and a suppression of circulating levels of LH and sex steroids (5,31,32). This down-regulation of GnRH receptors, produced by sustained administration of GnRH agonists, provides the basis for clinical applications in gynaecology and oncology of this class of compounds (32). Antagonists of GnRH exhibit no intrinsic activity, but compete with GnRH for the same receptor sites. GnRH antagonists produce a competitive blockade of GnRH receptors and cause an immediate inhibition of the release of gonadotropins and sex steroids (5,32). The principal mechanism of action of GnRH antagonist was thought to be based only on a competitive occupancy of GnRH receptors, but recently, has been demonstrated that administration of the GnRH antagonist Cetorelix produce down-regulation of pituitary GnRH receptors and a decrease in the levels of mRNA for GnRH receptors (32,33). Other works indicates that GnRH antagonists exert their inhibitory effects on the gene expression of pituitary GnRH receptors by counteracting the stimulatory effect of endogenous GnRH

(34). Specific GnRH receptors are also found on breast, prostatic, ovarian, endometrial and even pancreatic cancers (31,32). These GnRH receptors on tumor cells can mediate direct effect of GnRH analogs (32). Thus, high-affinity binding sites for GnRH and the expression of mRNA for GnRH receptors were detected in human prostate cancer samples, human prostate cancer lines, and Dunning rat prostate cancer (35,36). The presence of GnRH receptors in various human mammary carcinoma cell lines was also reported. GnRH receptors were similarly found in human ovarian epithelial cancer specimens and human ovarian cancer lines (31,32,37,38). The presence of high-affinity membrane receptors for GnRH was also established in nearly 80% of human endometrial carcinomas and in some endometrial cancer lines (32). GnRH receptors on human cancer appear to be similar to pituitary GnRH receptors. The expression GnRH receptors gene in human breast, endometrial, ovarian tumors, and respective cancer cell lines was also demonstrated by RT-PCR (37,38,39,40). The inhibition of growth of human mammary, ovarian, endometrial and prostatic cancer cell lines by GnRH agonists and antagonists in vitro strongly support the concept of their direct effect (5). The evidence for the production of an GnRH-like peptide and/or expression of mRNA for GnRH was also demonstrated in human prostatic, mammary, endometrial and ovarian cancer lines. This suggest that local GnRH may be involved in the growth of these tumors. The existence of functional regulatory system consisting of locally produced GnRH-like peptides and specific GnRH receptors has also been postulated in prostate cancer and ovarian cancer. The authors suggested that this GnRH, produced by tumor cells, might have an inhibitory function. However, the proliferation of various cancer cells in vitro is dose-dependently suppressed by GnRH antagonists and inhibitor effects of agonists might be explained by receptor down-regulation (31). Other studies in ES-2 human ovarian cancer lines suggest that locally produced GnRH is stimulatory. Additional investigations are needed to resolve the role and the action of endogenous GnRH-like peptides produced by various tumors.

3.1. Breast cancer

Breast cancer is the most common malignancy in women. About 30% of women with breast cancer have estrogen-dependent tumors and can be treated by hormonal manipulations such Tamoxifen or oophorectomy (44). Experimental and clinical studies clearly demonstrated that agonists of GnRH can be used for treatment of estrogen-dependent breast cancer. Thus initial investigations in rat and mouse models of breast adenocarcinoma showed that chronic administration of agonist [D-Trp6] GnRH decreased tumor weight and volume (5). This suggested that agonists of GnRH should be considered for a new hormonal therapy for breast cancer in women. Various clinical trials carried out since the early 1980s, demonstrated regression of tumor mass and disappearance of metastases in premenopausal and postmenopausal women with breast cancer treated with [D-Trp6] GnRH, Buserelin, Zoladex or Leuprolide (44,45). These studies showed that GnRH agonists are efficacious for the treatment of premenopausal women with estrogen-

dependent, estrogen-receptors positive breast cancer (45). Recently, our studies, demonstrated that Goserelin is efficacious on such disease producing tumor shrinkage and reducing the incidence of metastases in bitches with hormone-dependent mammary cancer (46).

3.2. Prostate cancer

The greatest therapeutic impact of GnRH analogues was in the field of prostate cancer, which is the most common noncutaneous malignant tumor in men. About 70% of human prostate cancers are testosterone-dependent and the treatment of advanced prostate cancer is based upon androgen deprivation (36). Therapy with agonist of GnRH with or without antiandrogens is currently the preferred treatment for men with advanced prostate cancer and in about 70% of cases GnRH agonist are selected for primary treatment (5,36). Administration of antiandrogens prior to and during early therapy with agonist can prevent the disease flare. Clinical trials in patients with advanced prostate cancer show that the antagonist of GnRH could be beneficial as a monotherapy for patients with prostate cancer and metastases in the brain, spine, liver and bone marrow in whom the GnRH agonist cannot be used as single drug, because of the possibility of flare-up (41,42). GnRH antagonists cause an immediate fall in the levels of gonadotropins and sex steroid and greatly reduce the time onset of therapeutic effects (36,40). In addition, treatment with GnRH antagonists can produce long-term improvement in patients with symptomatic benign prostatic hyperplasia (BPH) (42,43) and offers a therapeutic alternative to patients with BPH who are considered poor surgical risk (43).

3.3. Endometrial cancer

Endometrial cancer is a common gynaecologic malignancy in the Western world (5). Surgery or radiotherapy is successful in 75% of cases, but new methods are needed for advanced or relapsed cancer (31). Endometrial carcinoma is estrogen-dependent and thus it should respond to therapy with GnRH analogs. In addition, high affinity receptors for GnRH are present on nearly 80% of membranes of human endometrial cancer and endometrial cancer cell lines. Bioactive and immunoreactive GnRH and the expression of mRNA for GnRH were also found in these cells.

3.4. Epithelial ovarian cancer

Epithelial ovarian cancer is the fourth most frequent cause of cancer-related deaths in women (5). The treatment based on surgery or chemotherapy is not very effective and new approaches are needed. Ovarian cancer may be dependent on LH and FSH and in experimental cancer models, the suppression of the secretion of gonadotropins produced by GnRH analogs inhibits the growths of ovarian tumors (47). Studies in vivo indicate that GnRH antagonist inhibits growth of human OV-1063 and ES-2 epithelial ovarian cancer which was xenografted into nude mice better than agonist [D-Trp6] GnRH (32). In clinical studies some patients with advanced ovarian carcinoma treated with agonists of GnRH showed stabilization of disease (5), but in a multicenter trial no beneficial effects of therapy with [D-Trp6] GnRH could be found (48).

4 GnRH ANALOGUES ACTION

The role of female hormone, especially estrogen in the development of mammary tumor in mammals including the bitch is well established (51,52). Ovarian hormones act synergistically with pituitary hormones, especially growth hormone and prolactin to promote the development of mammary tumors (53,54). The expression of GnRH-receptors and/or their ligands, as well as other growth factors, is related in mammary tissue to the action of estrogen and progesterone and to the presence of functional receptors for estrogen (ER) and progesterone (PgR) (55,56). Consequently, several therapeutic approaches to this malignancy are aimed at achieving a blockade of ovarian hormone secretion and/or action (57,58). Gonadotropin-releasing hormone (GnRH agonist) have been shown to be effective to suppress ovarian hormones in human and in bitch, through the down-regulation of the pituitary-ovarian axis (59,60).

In addition to its function as a key hormone in the regulation of pituitary-gonadal axis, GnRH2 probably affects human extrapituitary tissues (61,62). GnRH analogues have been used in some of sex hormone-dependent cancer therapy, including breast (63,64,65), prostatic (66,67,68,69), pancreatic (70,71), endometrial (72), and ovarian (73,74) cancers. Although this effect may be mediated by an indirect mechanism based on the reduction in sex hormone secretion, there are indications that GnRH analogues suppress the growth of cancer cells *in vitro* (75,76,77,78). Specific binding sites for GnRH are demonstrated in certain tumors responsive to GnRH analogues (79,80,81). Moreover, GnRH analogues activate GnRH signal transduction pathways in breast cancer cells (82). Despite the fact that surgical castration showed no therapeutic effect on canine mammary tumors (83,84), our results showed that Goserelin, which acts according with a chemical castration, is efficacious on such disease producing tumor shrinkage and reducing the incidence of metastases in bitches with hormone-dependent mammary cancer (46). This suggests that GnRH analogues efficacy is not only due to the suppression of gonadal activity but also to the block of hypothalamus-pituitary axis and to a direct action on tumor cells; such hypothesis is supported by the detection of GnRH receptors in canine mammary tumors (85) as well as by the results of Vincze *et al.* (86), who have reported that the GnRH agonists significantly inhibit the growth of xenografts of the estrogen-receptors-negative of human MDA-MB 231 mammary tumor, and the effect of GnRH analogues.

On murine MXT mammary adenocarcinoma GnRH analogues which reduce significantly the concentration of binding sites (87). Moreover, recent findings showed that GnRH agonist exert both *in vivo* and *in vitro* a direct inhibitory action on the proliferation of human prostate tumor cells by interfering with the stimulatory action of EGF, reducing EGF-R and c-fos expression (88).

All these findings suggest direct inhibitory effects of GnRH analogues on the tumor growth. However, their mechanism of action remain unknown.

4.1. Pituitary action level

GnRH and /or GnRH analogues can be employed for two opposite clinical goals: to restore fertility (pulsatile administration) and to suppress the pituitary-gonadal function (chronic administration). The paradoxical antifertility effect of a chronic treatment with GnRH agonists, also called “chemical castration”, is usually reversible after the cessation of drug administration. Daily injections or monthly depot formulations of GnRH agonists have been successfully used either to interfere with the physiological LH peaks in order to improve the results of *in vitro* fertilisation procedures or to block an abnormal function of the pituitary-gonadal axis.

Chemical castration is also the basis for the clinical employment of GnRH agonists for the treatment of those malignancies whose progression may depend on gonadal steroids, such as breast, prostate, endometrial and ovarian cancer. GnRH agonists represent an established and well-tolerated alternative to surgical castration for some of these tumors, at least during their initial phase of hormone dependency (89). Through the down-regulation of pituitary GnRH receptors, GnRH agonists suppress the secretion of gonadal steroids, thus depriving the tumors of their mitogenic stimulus. For the treatment of these malignancies, GnRH agonists are usually combined with anti-steroidal agents to counteract the initial burst of gonadotropin secretion induced by GnRH agonists themselves.

GnRH antagonists, which have been experimented in clinical trials but are not yet available for routine clinical use, might offer the advantage of inhibiting the pituitary-gonadal axis without exhibiting the “flare up” phenomenon. (Figure 1 A and B).

4.2. Extrapituitary action level

In the last few years, evidence has been accumulated that GnRH agonists can exert a direct antiproliferative action on a number of malignancies related to the reproductive system (90,91). These compounds seem to exert their antimitogenic activity through the binding to GnRH receptors that have been consistently shown to be expressed in cancer cells. Tumor cells have been further demonstrated to express GnRH itself (or a GnRH-like peptide) which, in turn, might regulate cell proliferation in an autocrine/paracrine way by acting as a growth inhibitory factor (90). Therefore, in addition to their action at the pituitary level, GnRH agonists might affect tumor growth by modulating the activity of the GnRH system which is locally expressed in cancer tissues (Figure 2). The intracellular mechanisms mediating the direct growth-inhibiting activity of GnRH agonists have not been fully elucidated so far. A better clarification of these molecular mechanisms might improve the pharmacological treatment.

5 GnRH RECEPTORS IN CANCER CELL LINES

The expression of mRNA coding for the GnRH receptors has been reported in different hormone-related malignancies, such as prostate (92,93) breast (94,95), endometrium (96,97) and ovarian (94,98,99) cancers. The nucleotide sequence of GnRH receptor mRNA in these

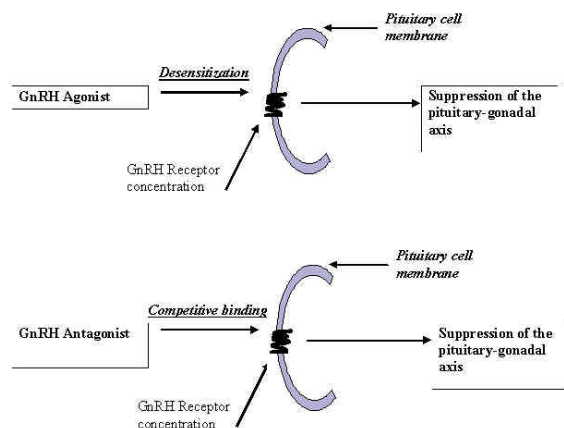


Figure 1. A GnRH agonist pituitary action levels. Receptor desensitization B GnRH antagonist pituitary action levels. Receptor competition.

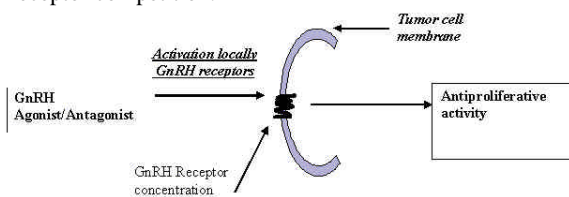


Figure 2. GnRH analogues extrapituitary action levels. Activation GnRH receptors.

tumors appears to be identical to that found in the pituitary gland. In cancer cells, the GnRH receptors is detectable also at the protein level. By Western blot analysis and by using a specific monoclonal antibody raised against the human pituitary receptor (100), Limonta *et al.* has been able to demonstrate that a protein band of approximately 64 kDa is present in membrane preparations of prostate cancer cells (101). This molecular weight is the same as that previously reported for the GnRH receptors protein at pituitary level in humans (100). Although these observations seem to support the hypothesis that the GnRH receptors in cancer tissues corresponds to that of the pituitary, at least in terms of nucleotide sequence and molecular weight; different results have been reported so far for the binding characteristics of GnRH receptors in cancer. Other authors have reported the presence of two types of GnRH binding sites, in gynaecological cancer cells, one of low affinity and the other one of high affinity (102).

In our studies, in GnRH positive canine mammary cancer cells, we observed, in binding competition experiments, that GnRH agonist was able to affect EGF binding reducing binding affinity of [125 I]EGF (103). These data are in agreement with the results obtained in human and murine tumor cells in which a phosphorylation of EGF receptor (86) and a significant reduction of the EGF-binding sites was demonstrated in presence of GnRH agonists (87). Finally, only one class of high affinity receptors has been identified in endometrial as well as in ovarian cancer cells (104,105). Some of these discrepancies might be due to the different experimental conditions adopted for the binding assay, or to

point mutations in the receptor or in the G-protein to which it is coupled or to the different cancer etiology.

Most of the reports so far summarised have been performed in hormone-dependent cancer cells. Interestingly, the observation of the expression of GnRH receptors also in androgen-independent carcinoma of the prostate, indicate that the receptor is still present when the tumor has progressed to the more advanced phase of steroid-unresponsiveness, which is also poorly responsive to standard therapy.

6 GROWTH FACTORS AND STEROID RECEPTORS IN CELLULAR REGULATION

6.1. Growth factor in the normal and malignant gland

The natural secretory products of the mammary epithelial cell, colostrum and milk, are abundant sources of growth factors (131). Growth factors in the normal gland probably are necessary for multiple purposes: in the newborn development, mammary growth, and mammary carcinogenesis. A large amount of literature has shown that estrogens, antiestrogens, progestins, and antiproggestins strongly regulate certain growth factors of the Epidermal Growth Factor (EGF) and Transforming Growth Factor β (TGF- β) families, as well as growth factors and secreted binding proteins for the IGF family (132,133) (Table 3).

EGF, apparently the most abundant milk-derived growth factor, is an important regulator both of the proliferation and differentiation of the mouse mammary gland in vivo and of mouse mammary explant in vitro (134,135). Modulation of signal of transduction pathways of EGF and its family members, as well as other unrelated growth factors are proving to be critical during mammary development. The EGF family consist, at present, of four receptors, a half dozen growth factors, and an additional half dozen growth factors encoded only by certain viruses (135,136).

In human breast cancer lines in vitro, autocrine growth factor dependent on the TGF α -EGF receptor system as also been documented in one of ER-negative breast cancer cell line, the MDA-MB-468 cell line. It is clear in this line that a rare gene amplification of the EGF receptor sensitizes the cells to autocrine function of TGF α . This line, representative of only a small percentage of breast cancers, express more than 10^6 EGF receptors/cell. TGF α and amphireguline are also induced by estrogen in hormone-dependent MCF-7 breast cancer cells; antisense cDNA or anti TGF α antibodies partially block estrogen induced growth in vitro (137,138,139), strongly suggesting a hormone-inducible type of autocrine system. The potential clinical relevance of this type of system is under investigation.

In human breast cancer an inverse correlation was observed between ER and EGF receptor positivity (106,107,108), while a significant direct correlation was observed between the concentration of ER and EGF receptors in canine malignant tumors (109). These findings suggest that the growing mechanism of human hormone-dependent breast cancer are different from those of the canine tumors. Further studies are required to clarify this issue.

Table 3. Hormonal regulation of Growth Factor system in breast cancer

Family	Growth Factor	Receptors	Binding protein
• EGF	EGF, TGF α , amphiregulin	EGF, erB-2	Unknown
• IGF	IGF-II	IGF-I, IGF-II, insulin	BP-3, BP-4
• TGF- β	TGF- β_1 , TGF- β_2 , TGF- β_3	Unknown	Unknown
• PDGF	PDGF-1, PDGF-2	Unknown	Unknown

Paracrine tumor-host interactive functions of this family of factors may then begin to dominate its functions as the disease progresses. Strategies employing toxic, genetically engineered EGF receptors, ligand, bacterial toxin, fusion proteins termed oncotoxins or anti-EGF receptors antibodies coupled to toxin or other therapeutic drugs (immunotoxins) or the closely related c-erbB2 receptor could possibly find future therapeutic utility, since a large portion of hormone-independent breast cancer express significant levels of these receptors (140,141,142,143). Recent studies have addressed the function of the heregulins acting on the EGF receptors-related c-erbB-3 and c-erbB-4 protein in breast cancer. The heregulins appear to act in vitro with biphasic effects: low levels are proliferative, while higher levels may inhibit epithelial proliferation. The heregulins also appear to promote differentiation and to induce casein synthesis in the developing gland and in breast cancer cells (144,145,146).

6.2. Regulation of steroid receptors

The estrogen and progesterone receptors are dimeric, gene-regulatory proteins. In mammary tissue, a single gene encodes an estrogen receptor subunit, and these subunits homodimerize and complex with additional proteins, such as heat shock proteins, to form the complete estrogen receptor (ER) assembly (110). Recent studies have complicated the picture with the identification of alternate spliced and mutated forms of the ER in breast cancer and some normal tissues such as brain and uterus (111). A single gene encodes three different isoforms of the progesterone receptor subunit in mammary tissue; there are two forms of progesterone receptor, homodimeric and heterodimeric (PR) (112,113). The multiplicity of progesterone and estrogen receptor isoforms in breast cancer may allow for significant variations in patterns of dimerization and in resultant variations in specificity of ligand recognition with respect to agonist versus antagonist and differential regulation of target genes (110,114). On top of this complexity, each receptor is able to adopt multiple conformations, depending upon the characteristic of interaction of the steroid (or nonsteroid ligand) with the receptor binding pocket. For example, the estrogen receptor can adopt at least three distinct conformations, depending on the antiestrogen bound (115).

The ER cannot be clearly classified as an oncoprotein or tumor suppressor protein. Although it clearly mediates onset and progression of the disease, unexpected results were obtained when the ER was expressed endogenously in ER-positive breast cancer cell lines and compared with its heterotypical expression in formerly ER-negative cell lines. In striking contrast to

its normal function in ER-positive cell lines, ER expressed in ER-negative cell lines suppresses cell growth, in spite of its normal action in regulating expression of certain hormonally responsive genes (116,117). Thus, the multiple differences between ER+ and ER- breast cancer seem to include incompatible growth-regulatory mechanism. We still do not understand the molecular basis for a lack of expression of PR in certain ER-positive breast cancer. A recent cell hybridization study with an antiestrogen resistant, ER-positive but PR-negative cell subline of MCF-7 has shown that PR expression last is a recessive phenotype in this system (118).

Increased expression and altered isozyme patterns of the cellular enzyme protein kinase C (PKC) family has also been implicated in the malignant progression of breast cancer (119). This enzyme family can act down-modulating ER-mRNA, activating ER function, independently inducing some estrogen-responsive genes with AP-1 sites in their promoters, and allowing more invasive cellular characteristic to be expressed (120). The PKC family contain at least nine cytoplasmic-nuclear enzymes, which possess serine-threonine specificity for phosphate addition to other cellular proteins (119); different isotype serve different cellular functions. The activity of PKC is known to be regulated by hormones and /or growth factors during normal lactational differentiation and to contribute to casein expression regulation. PKC activity has been found to be elevated in ER-negative and drug-resistant breast cancer relative to ER-positive breast cancer. Treatment of ER-positive breast cancer with an activatory of PKC such as the phorbol ester 12-O-tetradecanoyl-phorbol-13-acetic (TPA) leads to rapid down-regulation of ER, destabilization of its mRNA, and phosphorylation of the ER protein, coincident with modulation of its function (121,122,123,124). Phosphorylation of ER and PR, induced by estrogen itself, growth factor pathways (such as insulin-like growth factor-1 (IGF-1)), heregulin, cAMP, dopamine agonists, and other hormones may also constitutively activate the steroid receptors (125,126,127,128). Other current studies have suggested that receptors for other steroids (potential cancer prevention agents, retinoids and vitamin D) may modulate ER/PR function by forming heterodimers with ER or Pr or by modulating chromatin interactions of ER and PR (129,130).

7 SIGNAL TRANSDUCTION

7.1. Signal transduction and nuclear oncogenes

A unifying mechanistic link between the proliferative action of growth-promoting steroid and

Table 4. Genetic Defect in Breast cancer

A. Established Familial Breast Cancer Genes		
Gene	Disease	
• BRCA-1	Femal breast and ovarian cancer	
• p53	Li-Fraumeni syndrome of hereditary cancers	
• BRCA-2	Female and male breast cancer	
B. Established Breast Cancer Progression Genes		
Gene	Class	Function
• Rb-1	Suppressor Gene	Cell cycle G ₁ regulator
• cyclin-D1	Oncogene	Cell cycle G ₁ regulator
• myc	Oncogene	Cell cycle/cell death regulator
• erB-2	Oncogene	Growth factor receptor
• p53	Suppressor gene	Cell Cycle/cell death/DNA repair regulator

growth factor in different tissues are the nuclear protooncogenes (Table 4 A and 4B).

These transcription-regulating proteins mediate convergent pathways of growth regulatory stimuli through direct steroid action, through growth factor-induced mitogen-activated protein (MAP) kinase or phospholipase C-PKC, or through cytokine-induced JAK-STAT pathways (147,148,1150). The most important pathway for the proliferative stimuli exerted through the EGF receptor, the erB-2 receptor, and the insulin receptor families (151,152) appears to be the MAP kinase pathway. Receptors trigger this pathway through autophosphorylation and subsequent binding to PTB domains of signal transduction adaptor proteins (153,154). Following a cascade of protein phosphorylation, the products of the c-fos, c-myc, c-mib and c-jun protooncogenes are commonly observed to be induced shortly following mitogenic growth factor treatment of many types of cells, including normal and malignant breast epithelial cells. The protein products of at least three nuclear protooncogenes, c-fos, c-myc and c-jun, are also induced by both estrogen and progesterone in breast cancer (147). Progestins additionally induce a c-jun related protooncogene known as c-junB (148). Not surprisingly, tamoxifen down modulates c-yic expression during treatment-induced regression of patient tumors, (155) c-myc, c-fos and c-jun induction have also been shown to occur in human mammary epithelial cells in vitro and in the rat uterus in response to estrogen treatment in vivo (156,157,158). The protein products of c-fos, c-myc and c-jun genes form a heterodimeric complex, which interacts to form with a gene promoter consensus sequence termed AP-1. It has also been shown that the Myc protein binds retinoblastoma tumors-suppressor gene product Rb-1 or its partner to block their growth-inhibitory action (159). The Myc protein is also of particular importance in human breast cancer, since its gene is amplified in approximately 30% of the cases. As discussed below, other important signal transduction pathways are induced by oxidative stress, intracellular calcium and nitric oxide.

7.2. Signal transduction pathways of Estrogen receptors

The biological activity of estrogen is mediated by specific high-affinity estrogen receptor (ER) located within target cell nuclei. In the absence of hormone, ER is associated with a host of proteins that prevent it from interacting with the cellular transcription apparatus. Upon

binding estrogen, the receptor undergoes an activating conformational change, facilitating its association with target genes and permitting it to regulate gene transcription (160). In addition to the well established pathway, however, it has been shown that estrogen can induce extremely rapid increase in the concentration of the intracellular second messengers, calcium and cAMP (161,162,163). The time course of these events is similar to those elicited by growth factors and peptide hormones, lending support to the hypothesis that they do not involve the classical genomic action of estrogen through its receptor. These similarities, between the nongenomic actions of steroids and growth factor-signalling pathways converge in such a manner to permit cross-talk. For instance, both estrogen and epidermal growth factor (EGF) are known to act as mitogens in promoting cellular proliferation in the breast cancer and reproductive tract (160). Furthermore, the effects of these two agents sometimes overlap: estrogen has been shown to increase the uterine levels of both EGF and its receptor (164,165,166), and EGF has been shown to mimic the effects of estrogens in the mouse reproductive tract (167). Although the molecular details of this cross-talk remain to be elucidated, it is clear that ER itself is an important point of convergence. Specifically, it has been shown that ER transcriptional activity can be activated by binding to its cognate estrogen ligand but also by a variety of other extracellular signals: EGF, TGF, insulin and dopamine (127,168,169). The activation of ER by EGF has been demonstrated to involve direct phosphorylation by mitogen-activated protein kinase or extracellular-regulated kinase (MAPK/ERK) of serine residue (170,171). A further embodiment of this cross-talk was revealed when it was demonstrated that 17 β -Estradiol (E2) causes rapid activation of MAPK in mammalian cells in an ER-dependent manner (172,173). Thus, we can consider that a feed-forward system exists where E2 activates MAPK, an event that, in turn, enhances the transcriptional activity for ER. Recently, Sica *et al.* (174) showed that Leuprolerin, a GnRH-agonist, inhibits ERK1 and 2 phosphorylation in androgen-sensitive LNCaP cells while a stimulatory activity on ERK phosphorylation in androgen-insensitive PC-3 cells has been demonstrated. Such evidences suggest a different activation of MAPK in different cellular models and that ERK don't necessary induce a cell proliferation. Although the mechanisms underlying estrogen-induced MAPK activation and its physiological significance

remains to be explained, the activation of this signalling pathway may represent a potential mechanism by which estrogens regulate proliferation.

7.3. Signal transduction pathways of GnRH receptors

Individual molecules of the signal transduction cascades turned on by receptor agonist binding can play important roles not only in the intracellular activation process but also in the fine feedback regulation of signalling itself. In cancer cells and tissues, GnRH and GnRH analogues exert an antiproliferative action. This observation stimulated intensive experimental research aimed at identifying the signal transduction pathways coupled to the GnRH receptor at tumor level.

GnRH binds to a G protein-coupled membrane receptor in gonadotropes (177,178,179,180) and results in activation of multiple signaling pathways (181). The initial phase of GnRH action involves G protein-mediated stimulation of phospholipase C, leading to the formation of inositol 1,4,5-triphosphate (Ins-P3) and diacylglycerol.

Although various studies suggested a possible link between the GnRH-R and PLC-mediated phosphoinositide metabolism in mammary and ovarian cancer (82,175), later reports indicated that PLC activation might not represent the crucial mechanism of GnRH receptor activation in tumors (176). In prostate, both androgen-dependent and androgen-independent cancer cells, it been demonstrated that antiproliferative action of GnRH agonists is completely abrogated by pertussis toxin. Moreover, GnRH agonists substantially antagonize the pertussis toxin-catalysed ribosylation of a Gi protein. These data consistently indicate that, at variance with the receptor of the gonadotrophs, prostate cancer GnRH receptor might be coupled to the Gi-cAMP signal transduction pathway.

Inositol 1,4,5-triphosphate induces the release of intracellular calcium, and diacylglycerol activates PKC, resulting in multiple cellular responses to GnRH. Intracellular transmission of extracellular signals is mediated in large part by several groups of sequentially activated protein kinases, which are collectively known as the MAPK cascades (182,183). In the growth factor signaling, the key elucidated MAPK cascade is the ERK (184). Recent evidence indicates that some G protein-coupled receptors can activate ERK cascade (185,186,187,188). The signals transmitted through the ERK cascade lead to activation of a set of regulatory molecules that eventually initiates cellular responses such as growth and differentiation (189,190,191). Recently, it has been shown that GnRH agonist is capable of activating ERK in the pituitary organ culture (192) and the α T3-1 gonadotroph cell line (191,192) and that ERK is involved in regulation of gene expression of the gonadotropin α -subunit (193). However, the ERK cascade is not the only link between membrane receptors and their intracellular targets, and in the past few years, several other ERK-like cascades have been identified (190). One of the most studied is JNK [also known as stress-activated protein kinase (195,196)] cascade, which is known to be activated in response to cellular stress such as apoptosis (197). ERK,

JNK, and p38 (198) are members of the MAPK family. Recent data suggest that GnRH is capable of activating JNK in the α T3-1 gonadotroph cell line (199). It was reported that the signaling cascade by which GnRH acts in peripheral tumors is distinct from that in the anterior pituitary (200,201). In addition, the ERK cascade has been implicated in both cell proliferation (202,203,204) and growth arrest (205,206). In particular, ERK is reported to be involved in G1-specific cell cycle arrest of human breast cancer cells (207), NIH 3T3 murine fibroblasts (208), and human myeloblastic leukemia cells (209). Dephosphorylation of a tumor suppressor gene product, pRB, seems to be a target of the extracellular signals that induce cell cycle arrest and differentiation (210,211). In Go/Gi, pRB is underphosphorylated and complexed to the E2F transcription factor. This prevents the activation of some E2F-regulated genes required for DNA replication (212,213). Phosphorylation of pRB, during mid to late G1 by cyclin D- and cyclin E-associated kinases (214,215), is accompanied by release of E2F and activation of transcription of E2F-regulated genes, resulting in entry into S phase.

Recently Kimura *et al.* (216) showed that although GnRH agonist had no effect on the activation of the Jun N-terminal kinase (JNK), treatment of Caov-3 cells with GnRH agonist activated extracellular signal-regulated protein kinase (ERK), and its effect was more than that induced by GnRH. GnRH agonist also activated ERK kinase (mitogen-activated protein/ERK kinase) and resulted in an increase in phosphorylation of son of sevenless (Sos), and Shc. Both pertussis toxin, which inactivates Gi/Go proteins, and expression of a peptide derived from the carboxyl terminus of the beta-adrenergic receptor kinase I, which specifically blocks signaling mediated by the betagamma subunits of G proteins, blocked the GnRH agonist-induced ERK activation. Phorbol 12-myristate 13-acetate (PMA) also induced the ERK activity, but pretreatment of the cultured cells with PMA to down-regulate protein kinase C did not abolish the activation of ERK by GnRH agonists. Elimination of extracellular Ca^{2+} by EGTA also did not abolish the activation of ERK by GnRH agonist. Inhibition of mitogen-activated protein/ERK kinase by means of PD98059 canceled the antiproliferative effect of GnRH agonist and apparently reversed the GnRH-induced dephosphorylation of the retinoblastoma protein, the hyperphosphorylation of which is a hallmark of G1-S transition in the cell cycle. All these results provide evidence that GnRH agonist stimulation of ERK activity may be mediated by $\beta\gamma$ protein, not by PMA-sensitive protein kinase C nor extracellular Ca^{2+} in the Caov-3 human ovarian cancer cell line, suggesting that this cascade may play an important role in the antiproliferative effect of GnRH agonist. This cascade might have a role in GnRH agonist-induced antiproliferative effect. Furthermore, post-GnRH receptor signaling cascade in ovarian cancer cells seems to be different from that in pituitary cells, suggesting that the postreceptor signaling cascade might be different, depending on the cell, although the receptor is same.

7.4. Signal transduction pathways of Growth factors

Tumor growth factor (TGF α) and Epidermal growth factor (EGF) bind to the same membrane receptor

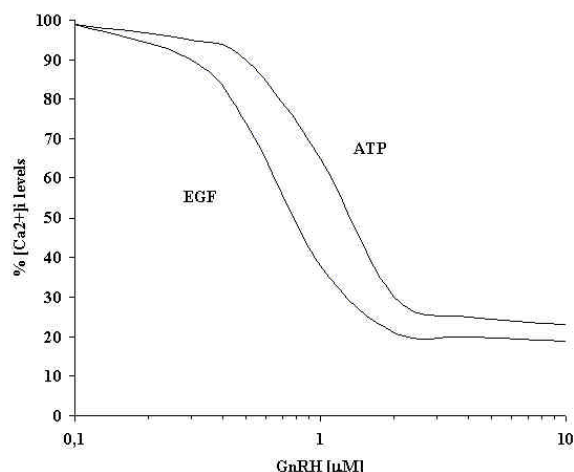


Figure 3. Effect of increasing concentration of GnRH agonist in canine mammary tumor. Cells treated with a single dose of EGF or ATP (1 μ M) in a 1 mM Ca²⁺ Medium (calcium influx).

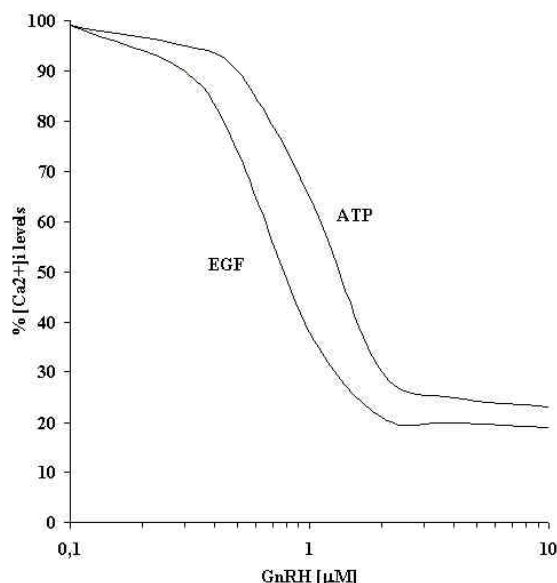


Figure 4. Effect of increasing concentration of GnRH agonist in canine mammary tumor cells treated with a single dose of EGF or ATP (1 μ M) in Medium without a 1 mM Ca²⁺ in presence of 10 μ M EGTA (calcium release from intracellular stores).

(EGF-R), which is a class I receptor tyrosin kinase (244) which induce direct PIP₂ hydrolysis, by a direct phosphorylation of PLCs, producing Inositol triphosphate (InsP₃) and variations in [Ca²⁺]_i sustained by both release from intracellular stores and influx across the plasmalemma. (218,219). The divalent cation calcium (Ca²⁺) is used by cells as a second messengers to control many cellular process including muscle contraction, secretion, metabolism and neuronal excitability (219). Moreover cytosolic Ca²⁺ plays an important role in the regulation of several cell types (220). Importantly, it participates in the regulation of the cell cycle in proliferating cells and in tumor cells in particular (221).

Moreover it has long been thought that Ca²⁺ and Nitric oxide (NO) work together in the control of cell homeostasis and NO could have appeared as a step in the signalling cascade initiated by the cation. However the interaction between the two messengers does not exist as a dependence but as a true, bi-directional cross-talk. In fact currently, almost all aspects of Ca²⁺ homeostasis have been reported to involve modulation by NO (222). Our experiments (103) demonstrated that, in canine mammary tumor cells, the GnRH agonist, Goserelin, was able to reduce calcium proliferative stimuli acting both on a specific proliferative stimulus such as EGF and on an aspecific proliferative stimulus such as ATP. We found that the interaction of GnRH with ATP system in canine mammary tumor cells showed that GnRH agonists was able to reduce both EGF and ATP induced Ca²⁺ rises both from released from internal stores and extracellular calcium entry (Figure 3 and 4). Interestingly, Goserelin, did not induce a typical calcium response in canine mammary tumor cells (93). Finally, our results on GnRH activity on intracellular NO levels EGF or ATP induced (Figure 5), suggest that Nitric oxide may have a role in the chain of intracellular event elicited by activation of epidermal growth factor receptors and in the down-regulation of calcium signalling by GnRH. While the Ca²⁺ storage machinery is unaffected by the treatment with NO, the gaseous messengers is shown to negatively modulate PIP₂ hydrolysis and the ensuing generation of IP₃ (223). An important consequence is the reduction of the growth factor-induced release of Ca²⁺ from the intracellular stores. In fact Goserelin, used in association with proliferating stimuli such as EGF or ATP, significantly increased nitric oxide production and affecting both calcium signal as well as cell proliferation in canine mammary tumor cells (103). This observation in according with fact that NO-induced negative signal modulations were already observed in several types of neuronal cell lines, strongly suggests that it may have a wider meaning (225).

8 DIRECT EFFECT OF GNRH ON TUMOR GROWTH

The observation that GnRH and its receptors are expressed in cancer cells suggest that this GnRH autocrine/paracrine system might be involved in the local control of tumor growth. This hypothesis has been confirmed by a number of reports. Antimitogenic activity of GnRH agonists has been reported for breast cancer (82,226,225), prostatic cancer (226,227), endometrial cancer (75,102,228) and ovarian cancer (26,105,229,230). These observations have been further confirmed by in vivo studies showing that GnRH agonists can significantly counteract the growth of cancer cells xenografted into nude mice (86,231). The GnRH system, which is present in cancer cells, appears then to participate in the local regulation of tumor growth by inhibiting cell proliferation. It has also been suggested that, when used for the treatment of hormone-dependent tumors GnRH agonist might directly reduce cancer growth by activating the locally expressed GnRH receptor, in addition to their main action at the pituitary level. Moreover GnRH analogues seem to interact on canine mammary tumor cells, not only

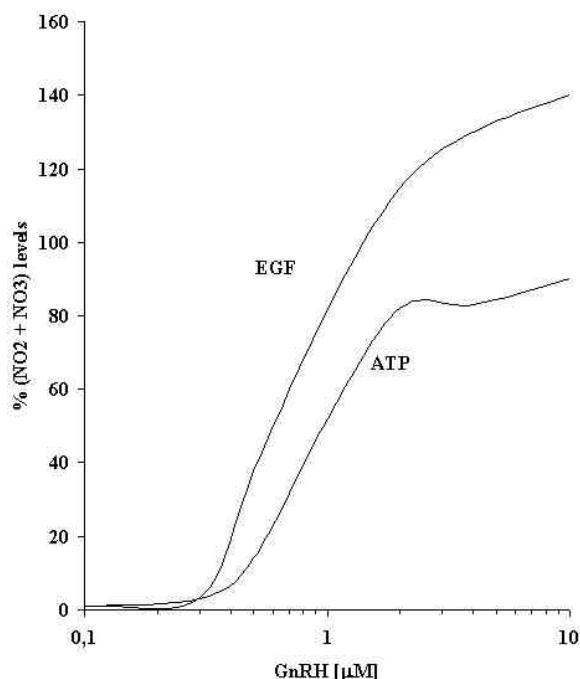


Figure 5. Effect of increasing concentration of GnRH agonist in canine mammary tumor cells treated with a single dose of EGF or ATP (5μM) on Nitric Oxide production.

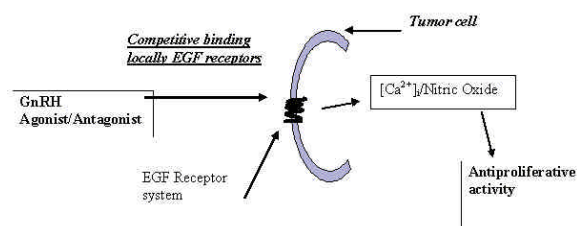


Figure 6. GnRH analogues locally action levels. Competitive binding on EGF receptors and pathways Nitric Oxide System

at pituitary level but also with Nitric Oxide system, directly to esplicate their antiproliferative activity (Figure 6).

GnRH agonists might also be considered as a possible therapy for the treatment of steroid-unresponsive tumors. However, the clinical observation so far available on the efficacy of GnRH agonist in hormone-independent cancers are poor. These observations seem to suggest a low efficacy of these compounds, at least when administrated according to the standard routes of administration. The elucidation of the concentration of GnRH agonists which might be required at the level of the tumor tissue to induce their antiproliferative action might help clarify this issue.

Unexpectedly and interestingly, Cetrorelix, belonging to the previous generation of GnRH antagonists, has been reported to have an antiproliferative activity similar to that of GnRH agonists on cancer cells, either in vitro or in vivo (105,163). Therefore, Cetrorelix seem to

behave as a potent suppressor of the pituitary-gonadal axis at pituitary level and as an activator of the GnRH receptors at tumoral levels. Although the molecular mechanisms making Cetrorelix a GnRH antagonistic analogue at the pituitary level and an agonistic analogue at cancer level are still unknown. This peculiarity of this molecule and structurally related compounds, with a longer half-life, may give an advantage over GnRH agonists for the treatment of hormone-related cancers, especially as GnRH antagonists are completely devoid of the “flare-up” phenomenon. Further studies are necessary to confirm these observations. The presence of GnRH receptors in tumor tissue has recently prompted studies aimed at the development of targeted chemotherapy based on GnRH analogues. Target cytotoxic GnRH conjugates are hybrid molecules composed of a GnRH agonist and a cytotoxic part. By specifically binding to its receptor, the GnRH analogue brings the chemotherapeutic drug directly to the tumor tissue. This might increase the efficacy of standard chemotherapy while reducing its toxicity (232).

9. ACKNOWLEDGMENT

We are grateful to Dr. Ciarcia R. for technical assistance and for helpful discussion. Work supported by grant from MURST 1999.

10. REFERENCES

1. Matsuo H, Y. Baba, R.M.G. Nair, A. Arimura, A.V. Schally: Structure of the porcine LH- and FSH-releasing hormone. I. The proposed amino acid sequence. *Biochem Biophys Res Commun* 43, 1334-1339 (1971)
2. Schally AV, A. Arimura, Y. Baba, R.M.G. Nair, H. Matsuo, T.W. Redding, L. Debeljuk, W.F. White: Isolation and properties of the FSH- and LH-releasing hormone. *Biochem Biophys Res Commun* 43, 393-399 (1971)
3. Schally AV, A. Arimura, A.J. Kastin, H. Matsuo, Y. Baba, T.W. Redding, R.M.G. Nair, L. Debeljuk, W.F. White: Gonadotropin-releasing hormone: One polypeptide regulates secretion of luteinizing and follicle-stimulating hormones. *Science* 173, 1036-1038 (1971)
4. Nestor JJ Jr: Development of agonistic LHRH analogs. In: *LHRH and Its analogs: Contraceptive and Therapeutic Applications*. Eds: Vickery BH, Nestor JJ Jr, Hafez ESE, MTP Press, Lancaster: 3-10.
5. Schally AV, A.M. Comaru-Schally: Hypothalamic and other peptide hormones. In: *Bast RC, Kufe DW, Pollock RE, Weichselbaum RR, Holland JF, Frei III E. Eds: Cancer Medicine*, 5th ed. Lewiston, NY: Decker, 715-729 (2000)
6. Schally AV, A.J. Kastin, A. Arimura: Hypothalamic FSH and LH-regulating hormone, structure, physiology, and clinical studies. *Fertil Steril* 22, 703-721 (1971)
7. Karten MJ, J.E. Rivier: Gonadotropin-releasing hormone analog design. Structure-function studies toward the development of agonists and antagonists: Rationale and perspective. *Endocr Rev* 7, 44-66 (1986)
8. Bajusz S, V.J. Csernus, T. Janaky, L. Bokser, M. Fekete, A.V. Schally: New antagonists of LHRH: II. Inhibition and potentiation of LHRH by closely related analogues. *Int J Peptide Prot Res* 32, 425-435 (1988)

9. Molineaux CJ, P.M. Sluss, M.P. Bree, M.L. Gefer, L.M.Sullivan, M.B. Garnick: Suppression of plasma gonadotropins by Abarelix: A potent new LHRH antagonist. *Mol Urol* 2, 265-268 (1998)
10. Nelson LR, V.Y. Fujimoto, R.B. Jaffe, S.E. Monroe: Suppression of follicular phase pituitary-gonadal function by a potent new gonadotropin-releasing hormone antagonist with reduced histamine-releasing properties (ganirelix) *Fertil Steril* 63, 963-969 (1995)
11. Vickery BH, Nestor J.J. Jr.: LHRH analogues: development and mechanism of action. *Sem Reprod Endocrinol* 5, 353-370 (1987)
12. Stojilkovic SS, K.J. Catt: Expression and signal transduction pathways of gonadotropin releasing hormone receptors. *Rec Prog Horm Res* 30,161-205 (1995)
13. Kalra SP: Mandatory neuropeptide-steroid signaling for the preovulatory luteinizing hormone-releasing hormone discharge. *Endocr Rev* 14, 507-538 (1993)
14. Shupnik MA: Gonadotropin gene modulation by steroids and gonadotropin-releasing hormone. *Biol Reprod* 54, 279-286 (1996)
15. Belsham DD, A. Evangelou, D. Roy, L.E.D. Vinh, T.J. Brown: Regulation of gonadotropin-releasing hormone (GnRH) gene expression by 5 alpha-dihydrotestosterone in GnRH-secreting GT1-7 hypothalamic neurons. *Endocrinology* 139, 1108-1114 (1998)
16. Fernald RD, R.B. White: Gonadotropin-releasing hormone genes: phylogeny, structure and functions. *Front Neuroendocrinol* 20, 224-240 (1999)
- Review on comparative structures of LHRH genes and LHRH peptides in vertebrates.
17. Poletti A, R.C. Melcangi, P. Negri-Cesi, R. Maggi, L. Martini: Steroid binding and metabolism in the luteinizing hormone-releasing hormone producing neuronal cell line GT1-1. *Endocrinology* 135, 2623-2638 (1994)
18. Sherwood N, D. Lovejoy, I. Coe. Origin of mammalian gonadotropin-releasing hormones. *Endocr Rev* 14, 241-254, (1993)
19. White RB, J.A. Eisen, T.L. Kasten , R.D. Fernald: Secon gene for gonadotropin-releasing hormone in humans. *Proc Natl Acad Sci USA* 95, 305-309 (1998)
20. Handelsman DJ, R.S. Swerdloff: Pharmacokinetics of gonadotropin-releasing hormone and its analogs. *Endocr Rev* 7, 95-105 (1986)
21. Clayton RN: Gonadotropin-releasing hormone: its actions and receptors. *J Endocrinol* 120, 11-19 (1989)
22. Sealfon SC, H. Weinstein, R.P. Millar: Molecular mechanisms of ligand interaction with the gonadotropin-releasing hormone receptor. *Endocr Rev* 18, 180-205 (1997)
23. Molineaux CJ, A. Lasdun, C. Michaud, M. Orlowski: Endopeptidase 24.15 is the primary enzyme that degrades LHRH both in vitro and in vivo. *J Neurochem* 51, 624-633 (1988)
24. Conn PM, W.F. Jr. Crowley: Gonadotropin-releasing hormone and its analogs. *Ann Rev Med* 45, 391-405 (1994)
- Review on LHRH: from physiology to pharmacology.
25. JE Rivier: Novel antagonists og GnRH: a compendium of their physicochemical properties, activities, relative potencies and efficacy in humans. In: GnRH Analogues: The State of the Art 1993. Parthenon Publishing Group, Carnforth, UK 13-26 (1993)
26. J Rivier: GnRH analogues towards the next millenium. In: GnRH Analogues: The State of the Art at the Millenium. Parthenon Publishing Group, Carnforth, UK 31-45 (1999)
27. Garnick MB, M. Campion: Abarelix depot, a GnRH antagonist, v LHRH superagonists in prostate cancer: differential effects on follicle-stimulating hormone. *Mol Urol* 4, 275-277 (2000)
28. Rabinovici J, P. Rothman, S.E. Monroe, C. Nerenberg, R.B. Jaffe: Endocrine effects and pharmacokinetics characteristics of a potent new gonadotropin-releasing hormone antagonist (Ganirelix) with minimal histamine-releasing properties: studies in postmenopausal women. *J Clin Endocr Metab* 75, 1220-1225 (1992)
29. Diedrich K, C. Diedrich, E. Santos: Suppression of the endogenous luteinizing hormone surge by the gonadotropin-releasing hormone antagonist Cetorelix during ovarian stimulation. *Hum Reprod* 9, 788-791 (1994)
30. Jiang G, J. Stalewski, R. Galyean: GnRH antagonists: a new generation of long acting analogues incorporating p-ureido-phenylalanines at positions 5 and 6. *J Med Chem* 44, 453-467 (2001)
31. Emons G, A.V. Schally: The use of luteinizing hormone releasing hormone agonists and antagonists in gynecological cancers. *Human Reprod* 9, 1364-1379, (1994)
32. AV Schally, G Halmos, Z Rekasi, JM Arencibia. The actions of LH-RH agonists, antagonists, and cytotoxic analogs on the LH-RH receptors on the pituitary and tumors. In: Devroey P, Ed. Infertility and Reproductive Medicine Clinics of North America. Philadelphia: Saunders 12, 17-44 (2001)
33. Pinsky J, N. Lamharzi, G. Halmos, K. Groot, A. Jungwirth, M. Vadillo-Buenfil, S.S. Kakar, A.V. Schally: Chronic administration of luteinizing hormone-releasing hormone (LHRH) antagonist Cetorelix decreases gonadotrope responsiveness and pituitary LHRH receptor messenger ribonucleic acid levels in rats. *Endocrinology* 137, 3430-3436 (1996)
34. Kovacs M, A.V. Schally, B. Csernus, Z. Rekasi: Luteinizing hormone-releasing hormone (LH-RH) antagonist Cetorelix down-regulates the mRNA expression of pituitary receptors for LHRH by counteracting the stimulatory effect of endogenous LHRH. *Proc Natl Acad Sci USA* 98, 1829-1834 (1998)
35. Koppan M, A. Nagy, A.V. Schally , A. Plonowski, G. Halmos, J.M. Arencibia, K. Groot: Targeted cytotoxic analog of luteinizing hormone-releasing hormone AN-207 inhibits the growth of PC-82 human prostate cancer in nude mice. *Prostate* 38, 151-158 (1999)
36. Schally AV, A.M. Comaru-Schally, A. Plonowski, A. Nagy, G. Halmos, Z. Rekasi: Peptide analogs in the therapy of prostate cancer. *The prostate* 45, 158-166 (2000)
37. Miyazaki M, A. Nagy, A.V. Schally, N. Lamharzi, G. Halmos, K. Szepeshazi, K. Groot, P. Armatis: Growth inhibition of human ovarian cancers by cytotoxic analogues of luteinizing hormone-releasing hormone. *J Natal Cancer Inst* 89, 1803-1809 (1997)
38. Miyazaki M, A.V. Schally, A. Nagy, N. Lamharzi, G. Halmos, K. Szepeshazi, P. Armatis: Target cytotoxic analog of luteinizing hormone-releasing hormone AN-207 inhibits growth of OV-1063 human epithelial ovarian

cancers in nude mice. *Am J Obstet Gynecol* 180, 1095-1103 (1999)

39. Kahan Z, A. Nagy, A.V. Schally, G. Halmos, J.M. Arencibia, K. Groot: Complete regression of MX-1 human breast cancer xenografts after targeted chemotherapy with a cytotoxic analog of luteinizing hormone-releasing hormone, AN-207. *Cancer* 85, 2608-2615 (1999)

40. Kahan Z, A. Nagy, A.V. Schally, G. Halmos, J.M. Arencibia, K. Groot: Administration of a targeted cytotoxic analog of luteinizing hormone-releasing hormone inhibits growth of estrogen-independent MDA-MB-231 human breast cancers in nude mice. *Breast Cancer Res Treat* 59, 255-262 (2000)

41. Gonzalez-Barcena D, M. Vadillo-Buenfil, A. Cortez-Morales, M. Fuentes-Garcia, I. Cardenas-Cornejo, A.M. Comaru-Schally, A.V. Schally: Luteinizing hormone-releasing hormone antagonist SB-75 (Cetorelix) as primary single therapy in patients with advanced prostatic cancer and paraplegia due to metastatic invasion of spinal cord. *Urology* 45, 275-281 (1995)

42. Gonzalez-Barcena D, M. Vadillo-Buenfil, F. Gomez-Orta, M. Fuentes-Garcia, I. Cardenas-Cornejo, A. Graef-Sanchez, A.M. Comaru-Schally, A.V. Schally: Responses to the antagonistic analog of LH-RH (SB-75, Cetorelix) in patients with benign prostatic hyperplasia and prostatic cancer. *The prostate* 24, 84-92 (1994)

43. Comaru-Schally AM, W. Brannan, A.V. Schally, M. Colcolough, M. Monga: Efficacy and safety of luteinizing hormone-releasing hormone antagonist Cetorelix in the treatment of symptomatic benign prostatic hyperplasia. *J Clin Endocrinol Metab* 83, 3826-3831 (1998)

44. Santen RJ, A. Manni, H. Harvey, C. Redmond: Endocrine treatment of breast cancer in women. *Endocr Rev* 11, 221-265 (1990)

45. Kaufmann M, W. Jonat, U. Kleeburg, W. Eirmann, F. Janicke, J. Hilfrick, R. Kreienberg, M. Albrecht, H.K. Weitzel, H. Schmid, P. Strunz, E. Schachner-Wunschmann, G. Bastert, H. Maass: The German Zoladex trial group: Goserelin, a depot gonadotropin releasing hormone agonist in the treatment of premenopausal patients with metastatic breast cancer. *J Clin Oncol* 7, 1113-1119 (1989)

46. Lombardi P, S. Florio, U. Pagnini, A. Crispino, L. Avallone: Ovarian function suppression with a GnRH analogue: D-ser(But[t])[6]-Arzgly[10]-LHRH (Goserelin) in hormone dependent canine mammary cancer. *J Europ Veter Pharmacol and Therap* 22, 56-61 (1999)

47. Yano T, J. Pinski, G. Halmos, K. Szepeshazi, K. Groot, A.V. Schally: Inhibition of growth of OV-1063 human epithelial ovarian cancer xenografts in nude mice by treatment with luteinizing hormone-releasing hormone antagonist SB-75. *Proc Natl Acad Sci USA* 91, 7090-7094 (1994)

48. Emons G, O. Ortmann, H.M. Teichert, H.G. Fassl, U. Lohrs, S. Kullander, A. Kauppila, D. Ayalon, A.V. Schally, F. Oberheuser: Luteinizing hormone-releasing hormone triptorelin in combination with cytotoxic chemotherapy in advanced epithelial ovarian cancer-A prospective double blind randomized trial. *Cancer* 78, 1452-1460 (1996)

49. Szepeshazi K, G. Halmos, A.V. Schally, J.M. Arencibia, K. Groot, M. Vadillo-Buenfil, E. Rodriguez-

Martin: Growth inhibition of experimental pancreatic cancers and sustained reduction in epidermal growth factor receptors during therapy with hormonal peptide analogues. *J Cancer Res Clin Oncol* 125, 444-452 (1999)

50. Szepeshazi K, K. Lapis, A.V. Schally: Effect of combination treatment with analogs of luteinizing hormone-releasing hormone (LH-RH) or somatostatin and 5-fluorouracil on pancreatic cancer in hamsters. *Int J Cancer* 49, 260-266 (1991)

51. Owen LN: A comparative study of canine and human breast cancer. *Investigation in Cell Pathology* 2, 257-275 (1979)

52. Lespagnard L, R. Kiss, A. Danguy, N. Legros, G. Lenglet, N. Devleeschouwer, R. Paridaens. In vitro Studies of Canine Mammary Tumors: Influence of 17-Beta-Estradiol and Progesterone on Cell Kinetics Parameters. *Oncology* 44, 292-301 (1987)

53. Schneider R, C.R. Dorn, D.O. Taylor: Factors influencing canine mammary cancer development and postsurgical survival. *J National Cancer Inst* 43, 1249-1261 (1969)

54. Schneider R: Comparison of age, sex, and incidence rates in human and canine breast cancer. *Cancer* 26, 49-426 (1970)

55. Koenders PG: Epidermal growth factor receptor and its ligand(s): association with prognosis of patients with breast cancer. PhD thesis. Nijmegen, The Netherlands (1992)

56. Stewart AJ, M.D. Johnson, F.E.B. Westley, B.R. Westley: Role of insulin-like growth factors and the type I insulin-like growth factor receptor in the estrogen-stimulated proliferation of human breast cancer cells. *J Biol Chem* 265, 21172-21178 (1990)

57. G Robustelli della Cuna: Principi di endocrinoterapia antitumorale. In: Manuale di oncologia medica. Eds: Bonadonna G & Robustelli della Cuna G. Masson Milan Italy 311-333 (1992)

58. Bajetta E, L. Celio, N. Zilembo, A. Bono, D. Galluzzo, M.G. Zampino, A. Longhi, L. Ferrari, R. Buzzoni: Ovarian function suppression with the gonadotropin-releasing hormone (GnRH) analogue Goserelin in premenopausal advanced breast cancer. *Tumori* 80, 28-32 (1994)

59. Mc Rae GI, B.B. Roberts, A.C. Worden, A. Bajka, B.H. Vickery: Long-term reversible suppression of oestrus in bitches with narfarelin acetate, a potent LHRH agonist. *J Rep Fert* 74, 389-397 (1985)

60. Kawakami E, H. Tonsho, T. Tsutsui, A. Ogasa: Effects of LHRH-analogue treatment of spermatogenic dysfunction in the dog. *Int J Andr* 14, 441-4522 (1991)

61. Imai A, T. Furui, T. Tamaya: Is extrapituitary action of gonadotropin-releasing hormone biologically significant? *Ann Clin Biochem* 29, 477-480 (1992)

62. Schally AV, A.M. Comarus-Schally, T.W. Redding: Antitumor effects of analogs of hypothalamic hormones in endocrine-dependent cancers. *Proc. Soc. Exp. Biol Med* 175, 259-281 (1984)

63. Manni A, R. Santen, H. Harvey, A. Lipton, D. Max: Treatment of breast cancer with gonadotropin-releasing hormone. *Endocr Rev* 7, 89-113 (1986)

64. Harris AL, J. Carmichael, B.M.L. Cantwell, M. Dowsett: Zoladex: endocrine and therapeutic effects in postmenopausal breast cancer. *Br J Cancer* 59, 97-99 (1989)

65. Waxman JH, S.J. Harland, R.C. Coombes, P.F.R. Wrigley, J.S. Malpa, T.A. Lister: The treatment of postmenopausal women with advanced breast cancer with busferlin. *Cancer Chemother Pharmacol* 15, 171-173 (1985)
66. Bèland G, M. Elhilali, Y. Frader, B. Laroche, E.W. Ramsey, J.A. Trachtenberg: controlled trial of castration with and without Nilutamide in metastatic prostatic carcinoma. *Cancer* (Phila.) 66, 1074-1079 (1990)
67. Huben RP, G.P. Murphy: Investigators of the national prostatic cancer project: a comparison of diethylstilbestrol or orchiectomy with busferlin and with methotrexate plus diethylstilbestrol or orchiectomy in newly diagnosed patients with clinical stage D2 cancer of the prostate. *Cancer* (Phila.) 62, 1881-1887 (1988)
68. Iversen P, S. Sucijs, R. Sylvester, I. Christensen, L. Denis: Zoladex and flutamide versus orchiectomy in the treatment of advanced prostatic cancer. *Cancer* (Phila.) 66, 1067-1073 (1990)
69. Schroder FH: Hormonal manipulation of prostatic cancer: too soon for total androgen blockade? *Br Med J* 303, 1489-1490 (1991)
70. Gonzalez-Barcena D, M.A. Ibarro-Olmons., F. Garcia-Carrasco, C. Gutierrez-Samperio, A.M. Comarus-Schally, A.V. Schally: Influence of d-Trp-6-LH-RH on the survival time in patients with advanced pancreatic cancer. *Biomed Pharmacother* 43, 313-317 (1989)
71. Greenway BA: Carcinoma of the exocrine pancreas: a sex hormone responsive tumor? *Br J Surg* 74, 441-442 (1987)
72. Gallagher CJ, R.T. Oliver, D.H. Oram, C.G. Fowler, P.R. Blake, B.S. Mantell, M.L. Slevin, H.F. Hope Stone: A new treatment for endometrial cancer with gonadotropin releasing-hormone analogue. *Br J Obstet Gynaecol* 98, 1037-1041 (1991)
73. Parmar H, G. Rustin, S.L. Lighman, R.H. Philips, I.W. Hanham, A.V. Schally: Response to D-Trp-6-luteinizing hormone releasing hormone (Decapeptyl) microcapsules in advanced ovarian cancer. *Br Med J* (Clin. Res. Ed.) 296, 1229 (1988)
74. Emons G, O. Ortmann, G.P. Pahwa, R. Hackenberg, F. Oberheuser, K.D. Schultz: Intracellular actions of gonadotropic and peptide hormones and the therapeutic value of Gn-RH agonist in ovarian cancer. *Obstet Gynecol Scand* 71 (Suppl. 155), 31-38 (1992)
75. Kleinman D, A. Duvdevani, A.V. Schally, J. Levy, Y. Sharoni: Direct growth inhibition of human endometrial cancer cells by the gonadotropin-releasing hormone antagonist SB-75: role of apoptosis. *Am J Obstet Gynecol* 170, 96-102 (1994)
76. Sharoni Y, E. Bosin, M.J. Levy, A.V. Schally: Inhibition of growth of human mammary tumor cells by potent antagonist of luteinizing hormone-releasing hormone. *Proc Natl Acad Sci USA* 86, 1648-1651 (1989)
77. Szende B, A. Zalatnai, A.V. Schally: Programmed cell death (apoptosis) in pancreatic cancers of hamsters after treatment with analogs of both luteinizing hormone-releasing hormones and somatostatin. *Proc Natl Acad Sci USA* 86, 1613-1617 (1989)
78. Yano T, J. Pinski, S. Radulovic, A.V. Schally: Inhibition of human epithelial ovarian cell growth in vitro by agonistic and antagonistic analogues of luteinizing hormone-releasing hormone. *Proc Natl Acad Sci USA* 91, 1701-1705 (1994)
79. Imai A, T. Ohno, K. Iida, T. Fuseya, T. Tamaya: Gonadotropin-releasing hormone receptor in gynecologic tumors. *Cancer* (Phila.) 74, 2555-2561 (1994)
80. Emons G, B. Schroder, O. Ortmann, S. Westphalen, K.D. Schulz, A.V. Schally: High affinity binding and direct antiproliferative effects of luteinizing hormone-releasing hormone analogs in human endometrial cancer cell lines. *J Clin Endocrinol Metab* 77, 1458-1464 (1993)
81. Eidne KA, C.A. Flanagan, R.P. Milar: Gonadotropin-releasing hormone binding sites in human breast carcinoma. *Science* (Washington DC) 229: 989-991.
82. Kèri G, A. Balogh, B. Szoke, I. Teplàn, O. Csuka: Gonadotropin-releasing hormone analogues inhibit cell proliferation and activate signal transduction pathways in MDA-MB-231 human breast cancer cell line. *Tumor Biol* 12, 61-67 (1991)
83. Yamagami T, T. Kobayashi, K. Takahashi, M. Sugiyama: Influence of ovariectomy at the time of mastectomy on the prognosis for canine malignant mammary tumours. *J Small Animal Practice* 37, 462-464 (1996)
84. Morris JS, J.M. Dobson, D.E. Bostock, E. O'Farrell: Effect of ovariectomy in bitches with mammary neoplasms. *Veterinary Record* 142, 656-658 (1998)
85. Sartin EA, C.H. Rahe, J.C. Wright, J.L. Sartin: Luteinizing Hormone Releasing Hormone, Estrogen, and Progesterone receptors in Canine Mammary lesions and Tumor Cell Lines. *Anticancer Res* 15, 2029-2032 (1995)
86. Vincze B, I. Palyi, D. Daubner, T. Krammer, I. Szamel, I. Bodrogi, J. Sugar, J. Seprodi, I. Mezo, I. Teplan, S. Eckhardt: Influence of luteinizing hormone releasing hormone agonists on human mammary carcinoma cell lines and their xenografts. *J Steroid Biochem Mol Biol* 38, 119-126 (1991)
87. Szende B, G. Srkalovic, K. Groot, A.V. Schally: Growth inhibition of mouse MXT mammary tumour by the luteinizing hormone releasing hormone antagonist SB-75. *J Nat Cancer Inst* 82, 513-517 (1990)
88. Montagnani Marelli M, R.M. Moretti, D. Dondi, P. Limonta, M. Motta: Effects of LHRH agonists on the growth of human prostatic tumour cells: in vitro and in vivo studies. *Archivi Italiani di Urologia e Andrologia* 69, 257-263 (1997)
89. Schally AV: Hypothalamic hormones from neuroendocrinology to cancer therapy. *Anticancer Drugs* 5, 115-130 (1994)
90. Emons G, O. Ortmann, K.D. Schulz, A.V. Schally: Growth-inhibitory actions of analogues of luteinizing hormone-releasing hormone on tumour cells. *Trends Endocrinol Metab* 8, 355-362 (1997)
- Review on the direct antiproliferative action of LHRH analogues at tumour level.
91. Imai A, T. Tamaya: GnRH receptor and apoptotic signaling. *Vit Horm* 59, 1-33 (2000)
- Review on the possible mechanism for LHRH-induced antiproliferative action.
92. Bahk JY, J.S. Hyun, H. Lee: Expression of gonadotropin-releasing hormone (GnRH) and GnRH receptor mRNA in prostate cancer cells and effect of GnRH on the proliferation of prostate cancer cells. *Urol Res* 26, 259-264 (1998)

93. Halmos G, J.M. Arencibia, A.V. Schally, R. Davis, D.G. Bostwick: High incidence of receptors for luteinizing hormone-releasing hormone (LHRH) and LHRH receptor gene expression in human prostate cancers. *J Urol* 163, 623-629 (2000)
94. Kakar SS, W.E. Grizzle, J.D. Neil: The nucleotide sequences of human GnRH receptors in breast and ovarian tumors are identical with that found in pituitary. *Mol Cell Endocrinol* 106, 145-149 (1994)
95. Kottler ML, A. Starzec, M.C. Carre, J.P. Lagarde, A. Martin, R. Counis: The genes for gonadotropin-releasing hormone and its receptor are expressed in human breast with fibrocystic disease and cancer. *Int J Cancer* 71, 595-599 (1997)
96. Imai A, T. Ohno, K. Iida, T. Fuseya, T. Furui, T. Tamaya: Presence of gonadotropin-releasing hormone receptor and its messenger ribonucleic acid in endometrial carcinoma and endometrium. *Gynecol Oncol* 55, 114-118 (1994)
97. Chatzaki E, C.M.R. Bax, K.A. Eidne, L.Anderson, J.G. Grudzinskas, C.J. Gallacher: The expression of gonadotropin-releasing hormone and its receptor in endometrial cancer and its relevance as an autocrine growth factor. *Cancer Res* 56, 2055-2065 (1996)
98. Imai A, T. Ohno, K. Iida, T. Fuseya, T. Furui, T. Tamaya: Gonadotropin-releasing hormone receptors in gynecological tumors. *Cancer* 74, 2555-2561 (1994)
99. Yin H, K. Cheng, H. Hwa, C. Peng, N. Auersperg, P. Leung: Expression of the messenger RNA for gonadotropin-releasing hormone and its receptor in human cancer cell line. *Life Sci* 62, 2015-2023 (1998)
100. Karande AA, K. Rajeshwari, D.J. Schol, J.H.M. Hilgers: Establishment of immunological probes to study human gonadotropin-releasing hormone receptors. *Mol Cell Endocrinol* 114, 51-56 (1995)
101. Limonta P, R.M. Moretti, M. Montagnani Marelli, D. Dondi, M. Parenti, M. Motta: The luteinizing hormone-releasing hormone receptor in human prostate cancer cells: messenger ribonucleic acid expression, molecular size and signal transduction pathway. *Endocrinology* 140, 5250-5256 (1999)
102. G Emons, O Ortmann & KD Schulz: GnRH analogues in ovarian, breast and endometrial cancers. In: GnRH Analogues: The State of the Art. Parthenon Publishing Group, Carnforth, UK 13-26 (1996)
103. Pagnini U, S. Florio, L. Crispino, G. Pagnini, D. Col angelo, D. Rocco, C. Pacilio, M. Pacilio, M. Macaluso, A. Giordano: Direct effect of a gonadotropin-releasing hormone agonist on the growth of canine mammary tumour cells. *Journal of Cellular Biochem* 85, 470-481 (2002)
104. Irmer G, C. Burger, R. Muller: Expression of the messenger RNAs for luteinizing hormone-releasing hormone (LHRH) and its receptor in human ovarian epithelial carcinoma. *Cancer Res* 55, 817-822 (1995)
105. Emons G, O. Ortmann, M. Becker: High affinity binding and direct antiproliferative effects of LHRH analogues in human ovarian cancer cell lines. *Cancer Res* 54, 5439-5446 (1993)
106. Formento JL, M. Francoual, P. Pormento, M. Etienne, J.L. Fischel, M. Namer, M. Frenay, E. Francois, G. Milano: Epidermal growth factor receptor assay: validation of a single point method and application to breast cancer. *Breast Canc Res and Treatment* 17, 211-219 (1990)
107. Koenders PG, L.V.M. Beex, T.J. Benraad: Epidermal growth factor receptor-negative tumours are predominantly confined to the subgroup of estradiol receptor-positive human primary breast cancers. *Cancer Res* 51, 4544-4548 (1991)
108. Blamey RW: Transforming growth factor-alpha and endocrine sensitivity in breast cancer. *Cancer Res* 54, 1684-1689 (1994)
109. Donnay I, N. Develeeschouwer, P. Wouters-Ballman, G. Leclercq, J. Verstegen: Relationship between receptors for epidermal growth factor and steroid hormones in normal, dysplastic and neoplastic canine mammary tissues. *Research in Veterinary Sci* 60, 251-254 (1996)
110. Smith D, D. Toft: Steroid receptors and their associated proteins. *Mol Endocrinol* 7, 4 (1993)
111. Skipper JK, L.J. Young, J.M. Bergeron, M.T. Tetzlaff, C.T. Osborn, D. Crews: Identification of an isoform of the estrogen receptor messenger RNA lacking exon four and present in the brain. *Proc Natl Acad Sci USA* 90, 71-72 (1993)
112. D Horowitz K: Hormone resistance in breast cancer: the role of normal and mutant steroid receptors. In: Dickson R, Lippman M. Eds: Mammary tumorigenesis and malignant progression. Boston: Kluwer 111 (1994)
113. C Beck & D Edwards. Progesterone receptor in breast cancer. In: Dickson R, Lippman M. Eds: Genes, oncogenes, and hormones. Boston: Kluwer 317 (1992)
114. Tung L, M.K. Mohamed, J.P. Hoeffler, G.S. Takimoto, K.B. Horwitz: Antagonist-occupied human progesterone b-receptors activate transcription without binding to progesterone response elements and are dominantly inhibited by a-receptor. *Mol Endocrinol* 7, 1256 (1993)
115. McDonnel DP, D.L. Clemm, T. Hermann: Analysis of the estrogen receptor function in vitro reveals three distinct classes of antiestrogen. *Mol Endocrinol* 9, 659 (1995)
116. Jiang SY, V.C. Jordan: Growth regulation of estrogen receptor-negative breast cancer cells transfected with complementary DNAs for estrogen receptor. *J Natl Cancer Inst* 84, 580 (1992)
117. Touitou I, F. Vignon, V. Cavailles, H. Rochefort: Hormonal regulation of cathepsin d following transfection of the estrogen or progesterone receptor into three sex steroid hormone resistant cancer cell lines. *J Steroid Biochem Mol Biol* 40, 231 (1991)
118. Paik S, D.P. Hartmann, R.B. Dickson, M.E. Lippman: Antiestrogen resistance in er positive breast cancer cells. *Breast Cancer Res Treat* 31, 301 (1994)
119. Dekker LV, P.J. Parker: Protein kinase c: a question of specificity. *Trends Biochem Sci* 19, 73 (1994)
120. Ways DK, C.A. Kukoly, J. de Ventre: MCF-7 breast cancer cells transfected with protein kinase c-alpha exhibit altered expression of other protein kinase isoforms and display a more aggressive neoplastic phenotype. *J Clin Invest* 95, 1906 (1995)
121. Tzukerman M, X.K. Zhang, M. Pfahl: Inhibition of estrogen receptor activity by the tumor promoter 12-o-tetradecanylphorbol-13-acetate: a molecular analysis. *Mol Endocrinol* 5, 1983 (1991)

122. Saceda M, C. Knabbe, R.B. Dickson: Post-transcriptional destabilization of estrogen receptor mRNA in mcf-7 cells by 12-o-tetradecanylphorbol-13-acetate. *J Biol Chem* 266, 17809 (1991)
123. Joel PB, A.M. Traish, D.A. Lannigan: Estradiol and phorbol ester cause phosphorylation of serine 118 in the human estrogen receptor. *Mol Endocrinol* 9, 1041 (1995)
124. Kato S, H. Endoh, Y. Masuhiro: Activation of the estrogen receptor through phosphorylation by mitogen-activated protein kinase. *Science* 1491 (1995)
125. Power RF, O.M. Conneely, B.W. O'Malley: New insights into activation of the steroid hormone receptor superfamily. *Trends Pharmacol Sci* 13, 318 (1992)
126. Power R.F., S.K. Mani, J. Codina, O.M. Conneely, B.W. O'Malley: Dopaminergic and ligand-independent activation of steroid hormone receptors. *Science* 254, 1636 (1991)
127. Aronica SM, B.S. Katzenellenbogen: Stimulation of estrogen receptor-mediated transcription and alteration in the phosphorylation state of the rat uterine estrogen receptor by estrogen, cyclic adenosine monophosphate, and insulin-like growth factor-I. *Mol Endocrinol* 7, 743 (1993)
128. Pietras RJ, J. Arboleda, D.M. Reese: Her-2 tyrosine kinase pathways targets estrogen receptor and promotes hormone-independent growth in human breast cancer cells. *Oncogene* 10, 2435-2446 (1995)
129. Segars JH, M.S. Marks, S. Hirshfeld: Inhibition of estrogen-responsive gene activation by the retinoid x receptor beta: evidence for multiple inhibitory pathways (published erratum appears in *Mol Cell Biol* 13, 2258 (1993))
130. Salbert G, A. Fanjul, F.J. Piedrafita: Retinoic acid receptors and retinoid x receptor-alpha down-regulate the transforming growth factor-beta 1 promoter by antagonizing ap-1 activity. *Mol Endocrinol* 7, 1347 (1993)
131. Grosvenor CE, M.F. Picciano, C.R. Baumrucker: Hormones and growth factors in milk. *Endocr Rev* 14, 710 (1993)
132. Cullen KJ, M.E. Lippman: Stromal-epithelial interactions in breast cancer. *Cancer Treat Res* 61, 413 (1992)
133. Dickson R, M.E. Lippman: Growth factors in breast cancer. *Endocr Rev* 16, 559 (1995)
134. Kurachi H, S. Okamoto, T. Oka: Evidence for the involvement of the submandibular gland epidermal growth factor in mouse mammary tumorigenesis. *Proc Natl Acad Sci USA* 82, 5940 (1985)
135. Vonderhaar BK: Regulation of development of the normal mammary gland by hormones and growth factors. *Cancer Treat Res* 40, 251 (1988)
136. Salomon DS, I. Perroteau, W.R. Kidwell, J. Tam, R. Derynck: Loss of growth responsiveness to epidermal growth factor and enhanced production of alpha-transforming growth factors in rats-transformed mouse mammary epithelial cells. *J Cell Physiol* 130, 397 (1987)
137. Kenney NJ, T. Saeki, M. Gottardis: Expression of transforming growth factor alpha antisense mRNA inhibits the estrogen-induced production of tgf alpha and estrogen-induced proliferation of estrogen-responsive human breast cancer cells. *J Cell Physiol* 156, 497 (1993)
138. Dickson RB, S.E. Bates, M.F. McManaway, M.F. Lippman: Characterization of estrogen responsive transforming activity in human breast cancer cell lines. *Cancer Res* 46, 1707 (1986)
139. Martinez-Lacaci I, M. Saceda, G.D. Plowman: Estrogen and phorbol esters regulate amphiregulin expression by two separate mechanisms in human breast cancer cell lines. *Endocrinology* 136, 3983 (1995)
140. R. Dickson & M Lippman: Control of human breast cancer by estrogen, growth factors, and oncogenes. In: Lippman M, Dickson R. Eds: *Breast Cancer: cellular and molecular biology*. Boston: Kluwer 119 (1988)
141. Hynes NE, D.F. Stern: The biology of erbB-2/neu/her-2 and its role in cancer. *Biochim Biophys Acta* 1198, 165 (1994)
142. Ennis B, E. Valverius, M. Lippman: Anti EGF receptor antibodies inhibit the autocrine stimulated growth of MDA-MB-468 breast cancer cells. *Mol Endocrinol* 3, 1830 (1989)
143. Pastan I, V. Chaudhary, D.J. Fitz Gerald: Recombinant toxins as novel therapeutic agents. *Annu Rev Biochem* 61, 331 (1992)
144. Yang U, E. Spitzer, D. Meyer, M. Sachs, C. Birchmeir, W. Birchmeir: Sequential requirement of scatter factor/hepatocyte growth factor (SF/HGF) and neu differentiation factor/hergulin (NDF/HRG) in the morphogenesis and differentiation of the mammary gland. *J Cell Biol* 131, 215 (1995)
145. Holmes WE, M.X. Sliwkowski, R.W. Akita: Identification of heregulin, a specific activator of p185erbB2. *Science* 256, 1205 (1992)
146. Wen D, E. Peles, R. Cupples: Neu differentiation factor: a transmembrane glycoprotein containing an egf domain and an immunoglobulin homology unit. *Cell* 69, 559 (1992)
147. Roberts T: A signal chain of events. *Nature* 360, 534 (1992)
148. Silvennoinen O, C. Schindler, J. Schlessinger, D.E. Levy: Ras-independent growth factor signaling by transcription factor tyrosine phosphorylation. *Science* 261, 1736 (1993)
149. van der Burg B, R.P. de Groot, L. Isbrucker, W. Kruijer, S.W. de Laat: Oestrogen directly stimulates growth factor signal transduction pathways in human breast cancer cells. *J Steroid Biochem Mol Biol* 40, 215 (1991)
150. Alkhalaf M, L.C. Murphy: Regulation of c-jun and jun-b by progesterins in t-47d human breast cancer cells. *Mol Endocrinol* 6, 1625 (1992)
151. Earp HS, T.L. Dawson, X. Li, H. Yu: Heterodimerization and functional interaction between egf receptor family members: a new signaling paradigm with implications for breast cancer research. *Breast Cancer Res Treat* 35, 115 (1995)
152. Ben-Levy R, H.F. Paterson, C.J. Marshall, Y. Yarden: A single autophosphorylation site confers oncogenicity to the neu/erbB-2 receptor and enables coupling to the map kinase pathway. *EMBO J* 13, 3302 (1994)
153. Seger R, E.G. Krebs: The mapk signaling cascade. *FASEB J* 9, 726 (1995)
154. van der Geer P, T. Pawson: The p1b domain: a new protein module implicated in signal transduction. *Trends Biochem Sci* 20, 277 (1995)
155. Le Roy X, C. Escot, J.P. Brouillet: Decrease of c-erbB-2 and c-myc RNA levels in tamoxifen-treated breast cancer. *Oncogene* 6, 431 (1991)
156. Chiappetta C, J.L. Kirkland, D.S. Loose-Mitchell, L. Murthy, G.M. Stancel: Estrogen regulates expression of the

- jun family of protooncogenes in the uterus. *J Steroid Biochem Mol Biol* 41, 113 (1992)
157. Murphy LJ. Estrogen induction of insuline-like growth factors and myc proto-oncogene expression in the uterus. *J Steroid Biochem Mol Biol* 40, 223 (1991)
158. Leygue E, R. Gol-Winkler, A. Gompel: Estradiol stimulates c-myc proto-oncogene expression in normal human breast epithelial cells in culture. *J Steroid Biochem Mol Biol* 52, 299 (1995)
159. Rustgi AK, N. Dyson, N. Bernards: Amino-terminal domains of c-myc and n-myc proteins mediate binding to the retinoblastoma gene product. *Nature* 352, 541 (1991)
160. O'Malley BW, S.Y. Tsai, M Bagchi, N.L. Weigel, W.T. Schrader, M.J. Tsai: Molecular mechanism of action of a steroid hormone receptor. *Recent Prog Horm Res* 47, 1-26 (1991)
161. Batra S: Effect of estrogen and progesterone treatment on calcium uptake by the myometrium and smooth muscle of the lower urinary tract. *Eur J Pharmacol* 127, 37-42 (1986)
162. Morley P, J.F. Whitfield, B.C. Vanderhyden, B.K. Tsang, J.L. Schwartz: A new, nongenomic estrogen action: the rapid release of intracellular calcium. *Endocrinology* 131, 1305-1312 (1992)
163. Aronica SM, W.L. Kraus, B.S. Katzenellenbogen: Estrogen action via the cAMP signaling pathway: stimulation of adenylate cyclase and cAMP-regulated gene transcription. *Proc Natl Acad Sci USA* 91, 8517-8521 (1994)
164. Di Augustine RP, P. Petruetz, G.I. Bell, C.F. Brown, K.S. Korach, J.A. McLachlan: Influence of estrogens on mouse uterine epidermal growth factor precursor protein and messenger ribonucleic acid. *Endocrinology* 122, 2355-2363 (1988)
165. Mukku VR, C.M. Stancel: Regulation of epidermal growth factor receptor by estrogen. *J Biol Chem* 260, 9820-9824 (1985)
166. Das SK, H. Tsukamura, B.C. Paria, G.K. Andrews, S.K. Dey: Differential expression of epidermal growth factor receptor (EGF-R) gene and regulation of EGF-R bioactivity by progesterone and estrogen in the adult mouse uterus. *Endocrinology* 134, 971-981 (1994)
167. Nelson KG, T. Takahashi, N.L. Bossert, D.K. Walmer, J.A. McLachlan: Epidermal growth factor replaces estrogen in the stimulation of female genital-tract growth and differentiation. *Proc Natl Acad Sci USA* 88, 21-25 (1991)
168. Ignar-Trowbridge DM, C.T. Teng, K.A. Ross, M.G. Parker, K.S. Korach, J.A. McLachlan: Peptide growth factors elicit estrogen receptor-dependent transcriptional activation of an estrogen-responsive element. *Mol Endocrinol* 7, 992-998 (1993)
169. O'Malley BW, W.T. Schrader, S. Mani, C. Smith, N.L. Weigel, O.M. Conneely, J.H. Clark: An alternative ligand-independent pathway for activation of steroid receptors. *Recent Prog Horm Res* 50, 333-347 (1995)
170. Kato S, H. Endoh, Y. Masuhiro, T. Kitamoto, S. Uchiyama, H. Sasaki, S. Masushige, Y. Gotoh, E. Nishida, H. Kawashima: Activation of the estrogen receptor through phosphorylation by mitogen-activated protein kinase. *Science* 270, 1491-1494 (1995)
171. Bunone G, P.A. Briand, R.J. Miksicek, D. Picard: Activation of the unliganded estrogen receptor by EGF involves the MAP kinase pathway and direct phosphorylation. *EMBO J* 15, 2174-2183 (1996)
172. Migliaccio A, M. Di.Domenico, G. Castoria, A. de.Falco, E.Nola, F. Auricchio: Tyrosine kinase/p21ras/MAP- kinase pathway activation by estradiol-receptor complex in MCF-7 cells. *EMBO J* 14, 1292-1300 (1996)
173. Endoh H, H. Sasaki, K. Maruyama, K. Takeyama, I. Waga, T. Shmizu, S. Kato, H. Kawashima: Rapid activation of MAP kinase by estrogen in the bone cell line. *Biochem Biophys Res Commun* 253, 99-102 (1997)
174. Sica G, S. Capucci, C. Angelucci, F. Iacopino: Interferenza del leupororelin acetato con il segnale mitogenico indotto da ormoni e fattori di crescita in linee cellulari di carcinoma della prostata umana. *XI Congresso Nazionale Società Italiana di Urologia Oncologica* 5, 55-56 (2001)
175. Imai A, T. Ohno, T. Furui, K. Takahashi, T. Matsuda, T. Tamaya: Gonadotropin-releasing hormone phospholipase C but not protein phosphorylation/dephosphorylation in plasma membrane from human epithelial ovarian cancer. *Int J Gynecol Cancer* 3, 311-317 (1993)
176. Emons G, V. Muller, O. Ortmann, K. D. Schulz: Effects of LHRH-analogues on mitogenic signal transduction in cancer cells. *J Steroid Biochem Mol Biol* 65, 199-206 (1998)
177. Tsutsumi, M., W. Zhou, R. P. Millar, P. L. Mellon, J. L. Roberts, C. A. Flanagan, K. Dong, K. Gillo, S. C. Sealton: Cloning and functional expression of a mouse gonadotropin-releasing hormone receptor. *Mol Endocrinol* 6, 1163-1169 (1992)
178. Reinhart, J., L. M. Mertz, K. J. Catt: Molecular cloning and expression of cDNA encoding the murine gonadotropin-releasing hormone receptors. *J Biol Chem* 267, 21281-21284 (1992)
179. Stojikovic, S. S., J. Reinhart, K. J. Catt: GnRH receptors: structure and signal transduction pathways. *Endocr Rev* 15, 462-499 (1994)
180. Naor, Z.: Signal transduction mechanisms of Ca²⁺ mobilizing hormones. The case of gonadotropin-releasing hormone. *Endocr Rev* 11, 326-353 (1990)
181. Naor, Z., S. Shacham, D. Harris, R. Seger, N. Reiss: Signal transduction of the gonadotropin releasing hormone (GnRH) receptor: cross-talk of calcium, protein kinase C (PKC) and arachidonic acid. *Cell Mol Neurobiol* 15, 527-544 (1995)
182. Winston, B. W., L. K. Remigio, D. W. Riches: Preferential involvement of MEK1 in the tumor necrosis factor- α -induced activation of p42mapk/erk2 in mouse macrophages. *J Biol Chem* 270, 27391-27394 (1995)
183. Cobb, M. H., E. J. Goldsmith: How MAP kinases are regulated. *J Biol Chem* 270, 14843-14846 (1995)
184. Egan, S. E., B. W. Gidding, M. W. Brooks, L. Buday, A. M. Sizeland, R. A.: Weinberg: Association of Sos Ras exchange protein with Grb2 is implicated in tyrosine kinase signal transduction and transformation. *Nature (Lond.)* 363, 15-16 (1993)
185. Ohmichi, M., T. Sawada, Y. Kanda, K. Koike, K. Hirota, A. Miyake, A. R. Saltiel: Thyrotropin-releasing hormone stimulates MAP kinase activity in GH3 cells by divergent pathways. *J Biol Chem* 269, 3783-3788 (1994)

186. Ohmichi, M., K. Koike, A. Nohara, Y. Kanda, Y. Sakamoto, Z. X. Zhang, K. Hirota, A. Miyake: Oxytocin stimulates mitogen-activated protein kinase activity in cultured human puerperal uterine myometrial cells. *Endocrinology* 136, 2082-2087 (1995)
187. Ohmichi, M., K. Koike, A. Kimura, K. Masuhara, H. Ikegami, Y. Ikebuchi, T. Kanzaki, K. Touhara, M. Sakaue, Y. Kobayashi, M. Akabane, A. Miyake, Y. Murata: The role of MPA kinase pathway in PGF2 α -induced rat puerperal uterine contraction. *Endocrinology* 138, 3103-311 (1997)
188. Sawada, T., M. Ohmichi, K. Koike, Y. Kanda, A. Kimura, K. Masuhara, H. Ikegami, M. Inoue, A. Miyake, Y. Murata: Norepinephrine stimulates mitogen activated protein kinase activity in GT-1 GnRH neuronal cell lines. *Endocrinology* 138, 5275-5281 (1997)
189. Nishida, E., Y. Gotoh: The MAP kinase cascade is essential for diverse signal transduction pathways. *Trends Biochem Sci* 18, 128-131 (1993)
190. Seger, R., E. G. Krebs: The MAPK signaling cascade. *FASEB J* 9, 726-735 (1995)
191. Marshal, C. J.: Specificity of receptor tyrosine kinase signaling: transient vs. sustained extracellular signal-regulated kinase activation. *Cell* 80, 179-185 (1995)
192. Mitchell, R., P. J. Sim, T. Leslie, M. S. Jojnsn, F. J. Thomson: Activation of MAP kinase associated with the probing effect of LHRH. *J Endocrinol* 140, R15-R18, (1994)
193. Sundaresam, S., I. M. Colin, R. G. Pestell, J. L. Jamesson: Stimulation of mitogen-activated protein kinase by gonadotropin releasing hormone: evidence for the involvement of protein kinase C. *Endocrinology* 137, 304-311 (1996)
194. Reiss, N., L. N. Llev, S. Shacham, D. Harris, R. Seger, Z. Naor: Mechanism of mitogen-activated protein kinase activation by gonadotropin-releasing hormone in the pituitary α T3-1 cell line: differential roles of calcium and protein kinase C. *Endocrinology* 138, 1673-1682 (1996)
195. Kyriakis, J. M., P. Banerjee, E. Nikolakaki, T. Dai, E. A. Rubie, M. F. Ahmad, J. Avruch, J. R. Woodgett: The stress-activated protein kinase subfamily of c-jun kinases. *Nature (Lond.)* 369, 156-160 (1994)
196. Derijard, B., M. Hibi, L. H. Wu, T. Barret, B. Su, T. Deng, M. Karin, R. J. Davis: JNK1: a protein kinase stimulated by UV light and Ha-Ras that binds and phosphorylates the c-Jun activation domain. *Cell* 76, 1025-1027 (1994)
197. Minden, A., A. Lin, F. X. Claret, A. Abo, M. Karin: Selective activation of the JNK signaling cascade and c-Jun transcriptional activity by the small GTPases Rac and Cdc42Hs. *Cell* 81, 1147-1157 (1995)
198. Cano, E., L. C. Mahadevan: Parallel signal processing among mammalian MAPKs. *Trends Biochem Sci* 20, 117-122 (1995)
199. Levi, N. L., T. Hanoch, O. Benard, M. Rozenblat, D. Harris, N. Reiss, Z. Naor, R. Seger: Stimulation of Jun N-terminal kinase (JNK) by gonadotropin-releasing hormone in pituitary α T3-1 cell line is mediated by protein kinase C, c-Src, and CDC42. *Mol Endocrinology* 12, 815-824 (1998)
200. Imai, A., H. Takagi, S. Horibe, T. Fuseya, T. Tamaya: Coupling of gonadotropin-releasing hormone receptor to Gi protein in human reproductive tract tumors. *J Clin Endocrinol Metab* 81, 3249-3253 (1996)
201. Imai, A., S. Horibe, H. Takagi, T. Tamaya: Gi protein activation of gonadotropin-releasing hormone-mediated protein dephosphorylation in human endometrial carcinoma. *Am J Obstet Gynecol* 176, 371-376 (1997)
202. Cowley, S., H. Paterson, P. Kemp, C. J. Marshall: Activation of MPA kinase is necessary and sufficient for PC12 differentiation and for transformation of NIH 3T3 cells. *Cell* 77, 841-852 (1994)
203. Pages, G., P. Lenormand, G. L'Allemain, J. Chambard, S. Meloche, J. Pouyssegur: Mitogen-activated protein kinase p42 mapk and p44 mapk are required for fibroblast proliferation. *Proc Natl Acad Sci USA* 90, 8319-8323 (1993)
204. Pang, L., T. Sawada, S. J. Decker, A. R. Saltiel: Inhibition of MAP kinase blocks the differentiation of PC-12 cells induced by nerve growth factor. *J Biol Chem* 270, 13585-13588 (1995)
205. Oehlen, B., F. R. Cross: Signal transduction in the budding yeast *Saccharomyces cerevisiae*. *Curr Opin Cell Biol* 6, 836-841 (1994)
206. Herskowitz, I.: MAP kinase pathways in yeast: for mating and move. *Cell* 80, 187-197 (1995)
207. Alblas, J., R. Slager-Davidov, P. H. Steenbergh, J. S. Sussenbach, B. van der Burg: The role of MAP kinase in TPA-mediated cell cycle arrest of human breast cancer cells. *Oncogene* 16, 131-139 (1998)
208. Sewing, A., B. Wiseman, A. C. Lloyd, H. Land: High-intensity RAF signal causes cell cycle arrest mediated by p21-Cipl. *Mol Cell Biol* 17, 5588-5597 (1997)
209. Yen, A., M. S. Roberson, S. Varvayanis, A. T. Lee: Retinoic acid induced mitogen-activated protein (MPA)/extracellular signal-regulated kinase (ERK) kinase-dependent MAP kinase activation needed to elicit HL-60 cell differentiation and growth arrest. *Cancer Res* 58, 3163-3172 (1998)
210. Laiho, M., J. A. DeCaprio, J. W. Ludlow, D. M. Livingston, J. Massagué: Growth inhibition by TGF- β linked to suppression of retinoblastoma protein phosphorylation. *Cell* 62, 175-185 (1990)
211. Richon, V. M., R. A. Rifkind, P. A. Marks: Expression and phosphorylation of the retinoblastoma protein during induced differentiation of murine erythroleukemia cells. *Cell Growth Differ* 3, 413-420 (1992)
212. Hiebert, S. W., S. P. Chellappan, J. M. Horowitz, J. R. Nevins: The interaction of RB with E2F coincides with an inhibition of the transcriptional activity of E2F. *Genes Dev* 6, 177-185 (1992)
213. Weintraub, S. J., C. A. Prater, D. C. Dean: Retinoblastoma protein switches the E2f site from positive to negative element. *Nature (Lond.)* 358, 259-261 (1992)
214. Akiyama, T., T. Ohuchi, S. Sumida, K. Matsumoto, K. Toyoshima: Phosphorylation of retinoblastoma protein by cdk 2. *Proc Natl Acad Sci USA* 89, 7900-7904 (1992)
215. Kato, J. H., Matsushima, S. W. Hiebert, M. E. Ewen, C. J. Sherr: Direct binding of cyclin D to the retinoblastoma gene product (pRB) and pRB phosphorylation by the cyclin d-dependent kinase CDK4. *Genes Dev* 7, 331-342 (1993)
216. Kimura A., M. Ohmichi, H. Kurachi, H. Ikegami, J. Hayakawa, K. Tasaka, Y. Kanda, Y. Nishio, H. Jikihara, N. Matsuura, Y. Murata: Role of mitogen-activated protein kinase/extracellular signal-regulated cascade in gonadotropin-releasing hormone-induced growth inhibition

- of a human ovarian cancer cell line. *Cancer Res* 59, 5133-5142 (1999)
217. Baselga J, J. Mendelsohn, Y.M. Kim, A. Pandiella: Autocrine regulation of membrane transforming growth factor- α cleavage. *J Biol Chem* 271, 3279-3284 (1996)
218. Fantl WL, D.E. Johnson, L.T. Williams: Signalling by receptor tyrosine kinases. *Annual Review of Biochemistry* 62, 453-481 (1999)
219. Berridge MJ: Inositol triphosphate and calcium signalling. *Nature* 361, 315-325 (1993)
220. Meldolesi J, E. Clementi, C. Fasolato, D. Zacchetti, T. Pozzan: Ca^{2+} influx following receptor activation. *Trends Pharmacol Sci* 12, 289-292 (1991)
221. Dixon CJ, W.B. Bowler, P. Fleetwood, A.F. Gitny, J.A. Gallagher, J.A. Caron : Extracellular nucleotides stimulate proliferation in MCF-7 breast cancer cells via P_2 -purinoceptors. *British J Cancer* 75, 34-39 (1997)
222. Clementi E, J. Meldolesi: The cross-talk between nitric oxide and Ca^{2+} : a story with a complex past and a promising future. *Trends Pharmacol Science* 18, 266-269 (1997)
223. Clementi E, C. Sciorati, M. Riccio, J. Meldolesi, G. Nisticò: Nitric oxide action on Growth Factor-elicited Signals. *J Biol Chem* 22, 22277-82 (1993)
224. Miller WR, W.N. Scott, R. Morris, H.M. Fraser, R.M. Sharpe: Growth of human breast cancer cells inhibited by a luteinizing hormone-releasing hormone agonist. *Nature* 313, 231-233 (1985)
225. Marini L, F. Iacopino, G. Schinzari, F.S. Robustelli Della Cuna, G. Mantovani, G. Sica: Direct antiproliferative effect of Triptorelin on human breast cancer cells. *Anticancer Res* 14, 1881-1886 (1994)
226. Sica G, F. Iacopino, D. Settesoldi, G. Zelano: Effect of Leuporelin Acetate on Cell Growth and Prostate-Specific antigen gene expression in human prostatic cancer cells. *European Urol* 35, 2-8 (1999)
227. Sica G, F. Iacopino, F. Capucci, A. Pistilli, G. Zelano: Leuporelin acetate regulates the expression of apoptosis-related genes in human prostate cells. *European Urol* 38, 485-534 (2000)
228. Sica G, G. Schinzari, C. Angelucci, G. Lama, F. Iacopino: Direct effects of GnRH agonists in human hormone-sensitive endometrial cells. *Molecular Cell Endocrinol* 176, 121-128 (2001)
229. Thompson MA, M.D. Adelson, L.M. Kaufman: Lupron retards proliferation of ovarian tumour cells cultured in serum-free medium. *J Clin Endocrinol Metab* 72, 1036-1041 (1991)
230. Emons G, S. Weiss, O. Ortmann, C. Grundker, K.D. Schulz: LHRH might act as a negative autocrine regulator of proliferation of human ovarian cancer. *Eur J Endocrinol* 142, 665-670 (2000)
231. Dondi D, R.M. Moretti, M. Montagnani Marelli: Growth inhibitory effects of luteinizing hormone-releasing hormone (LHRH) agonists on xenografts of the DU 145 human androgen-independent prostate cancer cell line in nude mice. *Int J Cancer* 76, 506-511 (1998)
232. Schally AV, A. Nagy: Cancer chemotherapy based on targeting of cytotoxic peptide conjugates to their receptors on tumors. *Eur J Endocrinol* 141, 1-14 (1999)

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Key Words: GnRH, Steroids, Cancer, Signal Trasduction, Review