Airborne inflammatory factors: "from the nose to the brain"

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1. ABSTRACT

The intranasal pathway is a direct route of communication between the environment and the brain. This pathway is currently used for the delivery of several experimental therapeutic peptides and vaccines because it bypasses the blood brain barrier. It is also a route of entrance to the brain for several viruses and toxic substances. Airborne infectious, allergic and pollution agents are among the most common inflammatory factors which may affect brain function via the brain-nose interface. The inflammatory processes triggered in the upper respiratory tract by these agents are positioned to influence the immune response of the brain and therefore, influence its function and alter behavior. Several clinical and epidemiological studies find an association between inflammatory factors affecting the intranasal pathway and neurological disorders such as multiple sclerosis, Alzheimer and Parkinson diseases as well as mental disorders including anxiety and mood disorders. However the mechanisms of interaction between the immune response in the nasal epithelium and the brain are poorly understood. This article discusses current evidence about these mechanisms and associations with neurological and mental diseases.

2. INTRODUCTION

Diseases of the airways due to airborne inflammatory factors are highly prevalent and their incidence is increasing. The causes have been attributed to several sources including climate changes, increasing air pollution and population growth (1-5). The major sources of contamination are infectious organisms including viruses and bacteria, allergens and air pollutants. Although they produce different types of inflammatory responses, these processes have overlapping features and influence or may potentiate each other. Moreover, while these diverse etiologies differ on other features such as their mortality rate, disease progression and damage to the respiratory system, they share the characteristics of the route of exposure and massive recruitment of lymphocytes to the respiratory epithelium with the consequent production of inflammatory molecules such as cytokines and chemokines.

The present review will discuss the impact of these inflammatory processes of the airways on the brain and behavior. This is based on the direct communication that exists between the nasal epithelium and the brain and the effects on behavior produced by activation of the brain's immune response. The vast diversity of airborne inflammatory factors will be briefly presented and summarized. Issues related with seasonality *vs* chronic exposure will be considered throughout. The distinct inflammatory processes activated by different pathogenic sources and data about their effects on the immune response of the brain will be discussed. Emerging evidence showing altered emotional responses induced by inflammation of the airways will be put forth. Finally, the implications of these processes on neurological and mental disorders will be considered.

3. THE INTRANASAL PATHWAY

The brain is protected by many barriers that prevent the free access of molecules and cells present in the systemic circulation as well as potential pathogens. One of the most important systems is the blood brain barrier BBB composed of endothelial cells sealed by tight junctions impeding the passage of cells and large molecules into the brain (6). There are few structures in the brain were the BBB is incomplete and allow contact between the main circulation and the brain. These places are known as circumventricular organs (CVOs) and function as sensory organs monitoring the status of the "internal milieu" sending that information to higher order neuronal groups (7,8). They also are the places of release of many neuroactive substances and hormones into the general circulation. Although these places may be a direct route of entry into the brain for pathogens present systemically, these pathogens have to first reach the peripheral circulation and overcome immune surveillance mechanisms. There is however a direct route of access from the external environment into the brain bypassing the systemic circulation and the BBB. The olfactory neuroepithelium is one of the places in which neurons of the central nervous system (CNS) are in direct contact with the environment, in this case the nasal cavities (9). The olfactory neurons send their axons trough perforations in the ethmoid bone called the cribriform plate making synapses into the olfactory bulbs inside the brain. This constitutes a direct pathway of communication between the brain and the nasal epithelium and a direct route of access for pathogens and molecules present in the nasal cavities. In fact, there is currently a significant amount of evidence showing how small and large molecules can access the brain from the nasal epithelium (10-12). For example, intranasal instillation of the cytokine interferon-β results in its rapid distribution in several regions of the brain following two major pathways, one corresponding to the olfactory and another to the trigeminal systems (13). In the first case, the main targets are the olfactory bulbs, olfactory nucleus, and prefrontal cortex. Additional presence of interferon- β in other regions far from the nasal cavities including the amygdala and hippocampus was also detected. The target regions in the brain for the trigeminal pathway are the brainstem including the pons and to a lesser extent the spinal cord. Similar patterns of distribution and transfer from the nasal cavities to the brain was also demonstrated for several other molecules including insulinlike growth factor I (11) nerve growth factor and vasoactive intestinal peptide (14). Two major mechanisms have been proposed for this transfer from the nasal cavities into the brain. A direct extracellular pathway through open

intercellular cleft of the olfactory epithelium into the olfactory bulbs, and an intracellular pathway from the olfactory sensory neurons via anterograde transport. This transfer of molecules have been shown for the bioactive forms of these molecules as determined by the capacity to activate cellular processes in target areas (11) or to produce effects in the brain. Indeed, intranasal instillation of recombinant cytokines including IL-4, IL-6 and IL-12 have been shown to affect the course of experimentally induced neurological disorders (15-17).

Effects on brain function and behavior of molecules administered via the intranasal pathway has also been shown in humans. Insulin administered via the nasal cavities has been reported to improve memory and cognition in humans (18-20). In addition, oxytocin was shown capable of influencing endocrine and sexual function (21,22). Similarly, intranasal atrial natriuretic peptide (ANP) and growth hormone releasing hormone (GHRH) have been shown to influence the hypothalamic-pituitary adrenal (HPA)-axis in humans (23,24). Recently, it was reported that intranasal administration of testosterone increases the reactivity of the amygdala in middle-aged women to a young adult level (25).

It is known that the intranasal pathway is also an important route of entry into the brain for several neurotropic viruses including herpes simple virus (HSV) (26), influenza virus (27) mumps virus (28) measles virus (29), Borna disease virus (BDV) (30) and arenaviruses (31). The distribution in the brain of most of these viral infections also involves regions following the olfactory and trigeminal pathway (26,32,33). In summary, the intranasal pathway is a direct route of access into the brain for molecules present in the nasal epithelium and a direct route of entry for environmental pathogens such as viruses. Either of these processes produces significant changes in brain activity and capable of affecting its function and consequently affecting behavior.

4. EXPOSURE TO AIRBORNE INFLAMMATORY FACTORS

4.1. Infectious agents

Airborne viruses and bacteria are transmitted mainly by moist droplets containing virus or bacteria originated by coughing and sneezing from infected individuals. Although the distance that they can travel has been believed relatively small (few meters), studies suggest that depending on the droplet's size and climate conditions in some cases the potential range for virulent infection may be significantly bigger (34). Some reports suggest that viruses and bacteria can move considerable distances transported in dust originated from wind storms (35). Similarly, zoonotic arenaviruses such as those responsible for hemorrhagic fever and encephalitis spread significant distance from rodents to human by wind-blown dust containing the virus (36,37). This form of transmission makes airborne viruses and bacteria extremely relevant for human health from an epidemiological point of view. They are of high incidence in the population and are among the most common causes of death (5,38-44). For example,

influenza is responsible for 100 to 500 hospitalizations per every 100,000 children and persons over 65 years of age and accounts for 36,000 deaths per year in the U.S (5,40,45,46). For these reasons extensive research has been oriented in developing early detection and preventive strategies of disease epidemic and outbreak.

In addition to mortality, airborne viruses and bacteria causes significant health problems that require interventions. An estimated of 32 million adults in the US suffer from sinusitis annually resulting in about 13 million visits to a health professional (47) with an estimated cost of 2.4 billion (48). The most common causes of sinusitis are viral infections including rhinoviruses, influenza and parainfluenza that may be followed by complications with bacterial infections (49,50). More than 50% of bacterial sinusitis are caused by *Haemophilus influenzae and Streptoccocus pneumoniae* (49,50) eventually requiring the use of antibiotics.

Exposure to airborne viruses and bacteria usually fluctuates during seasons and their incidence varies year to year. Viral respiratory infections including influenza, coronaviruses and respiratory syncytial virus (RSV) are more common during winter with a peak occasionally extending to early spring (5,44,51). In addition, many studies report an association of this peak with a subsequent respiratory bacterial pneumococcal peak of infections (44,52-54). The increased incidence of respiratory viral and bacterial infectious diseases during winter time has been related to increased contact among individuals related to living conditions in cold weather. For example, mechanisms of vector transmission through sneezing and coughs will be facilitated in closed crowded environments (44). In addition, cold air and low humidity typical of winter time will affect the capacity of the respiratory epithelium to clear airborne pathogens and increase virus transmission (55). There is, however, a certain degree of variation in the peak of incidence for each pathogen reported in the referenced studies. There are also variations between the years analyzed and between cities and countries depending on the study. The temporal sequence obtained from the referenced surveys in the northern hemisphere considering the most frequent reported incidences across studies is as follows: RSV infections have been reported early in winter from November to January. Influenza has been more consistently reported between December and February. Pneumonia and pneumococcal infections follow the peak of influenza after a 2 week interval. Finally rhinovirus and coronaviruses, including the recent human version of the bird flu causing severe acute respiratory syndrome peaks during March (5,40,44,51,52).

4.2. Allergens

Exposure to airborne allergens or aeroallergens is responsible for the most common types of allergies including allergic rhinitis or hay fever, allergic asthma and allergic conjunctivitis. These diseases are highly prevalent in developed countries and their incidence has been estimated from 20 to 30 percent in adults and up to 40 percent in children (56-59). There are basically two types

of aeroallergens causing respiratory allergies, indoor allergens responsible for perennial allergic rhinitis and outdoor allergens responsible for most of the cases of seasonal allergic rhinitis. Some individuals suffer from both perennial and seasonal allergic rhinitis and a similar situation is often found for allergic asthma as well. The most common indoor aeroallergens are dust mite, mold, animal dander and cockroach parts while the most prevalent outdoor aeroallergens are pollen and spores (60). In susceptible individuals that are prone for the development of allergies, repeated exposure to any of these allergens may result in sensitization and development of respiratory allergies. Allergic rhinitis accounts for approximately 70% of all cases of allergies and is also linked to the development of allergic asthma. The burden caused by these diseases is costly for society and bothersome to individuals suffering from respiratory allergies causing impairments in daily functioning and quality of life (56-59).

Exposure to airborne allergens may occur chronically depending on housing conditions such as the case of perennial aeroallergens and may lead to chronic allergic disorders. Outdoor aeroallergens are mostly of seasonal nature with a high incidence in specific times of the year such as spring (tree pollen) and fall (weed pollen). One of the most dramatic seasonal changes in incidence of upper respiratory inflammatory disease is represented by the robust increase in pollen-induced allergy during spring (61). Pollen represents the most important and the most seasonal among aeroallergens (http://www.aaaai.org/nab/index.cfm). While pollen levels are detectable throughout the year, they are usually below 100 grains/cubic meter which are the level below the majority of susceptible allergic humans do not report symptoms (61). For example, a study analyzing weekly mean values for total pollen production in the Washington-Baltimore area reported an increase form less than 100 grains/cubic meter to about 400 grains/cubic meter during the first week of April followed by an increase to more than 1000 to 1.300 grains cubic/meter in the second to third week of April (62). The levels of pollen return to below 100 grains/cubic meter by the first week of June. Allergies to pollen represent the most important cause of seasonal allergic rhinitis and inflammatory complications of the respiratory system. It has been reported that individuals suffering from allergic rhinitis are more likely to suffer from bacterial and viral complications of the sinuses and upper respiratory tract and that a history of viral infections worsens the symptoms of allergies (63-66). Thus, respiratory allergies caused by aeroallergens are highly prevalent and viral and bacterial infections more common in people suffering from allergies.

4.3. Pollutants

Exposure to air pollutants is an increasing problem in industrialized countries and is believed to impose a serious health problem in urban populations. The most common air pollutants are particulate matter (PM) of different sizes originated from different sources including power plants, motor vehicles, construction sites, deforestation and industrialization processes (2,49). The most dangerous for human health are fine sizes particles up to 2.5 microns (PM_{2.5}) that can be suspended in the air for large periods of time and transported long distances by wind and climatic variables (4). Among the most problematic are fine carbon particles originated mainly from motor vehicles using diesel fuel. They have been associated with increased mortality rate due to respiratory and cardiovascular diseases believed to be produced or worsened by increased exposure to particulate matter (2,67-69). In addition, several epidemiological studies report the association between diesel exhaust particles and increased incidence or respiratory diseases such as bronchitis and asthma (3,70,71). Perhaps the most remarkable and controversial series of studies are those relating the increase incidence of asthma in Caribbean countries due to dust containing coarse particulate matter (less than 10 microns) originated in the Sahara desert (72,73). Although more research is needed to confirm this association, these studies signify the potential of range of exposure and health hazard that particulate matter has in the world.

Another feature of PM is that they may interact with others pathogens and serve as transporters of viruses, bacteria or molecules with infectious or antigenic properties such as cell walls of bacteria (35). For instance, PM containing bacterial lipopolysaccharides (LPS) has been associated with the development of respiratory diseases and is believed an important occupational hazard (69,74,75). Moreover, the combination of pollen with diesel exhaust particles is believed to increase the occurrence of asthma and the aggravation of symptoms in asthmatics (3).

The pattern of exposure to PM depends from the distance of the sources and the interaction of PM with atmospheric variables including (but not limited to) winds, humidity, precipitations and temperature. In some cases, a seasonal pattern of exposure has been observed for certain PM depending on seasonal weather conditions such as high pressure systems and rainfall (4). Because of the increasing sources of PM, their capacity to reach long distances and ability to serve as "carriers" of other inflammatory and pathogenic factors, PM is a significant airborne inflammatory factor of increasing importance across the world.

5. NEUROIMMUNE PROCESSES TRIGGERED BY AIRBORNE INFLAMMATORY FACTORS

Airborne inflammatory factors including viruses, bacteria, allergens and pollutants have the potential to affect brain function and behavior by two major mechanisms. In the case of infectious agents such as viruses and bacteria one mechanism is by accessing the brain and establishing acute and/or permanent infections. The second mechanism is by inducing an inflammatory process in the respiratory epithelium, in particular the upper respiratory tract, produced in response to non-invading inflammatory factors such as infectious agents, allergens and pollutants. The first case conveys the most dramatic outcomes in which some airborne viruses and bacteria may produce major neurological and systemic symptoms. They also may result in death by meningitis or encephalitis or sub-lethal infections with significant cell loss and neurological sequelae. The second case involves exposure to the most common and prevalent types of viral and bacterial infections, allergens and air pollutants activating an inflammatory response in the upper respiratory epithelium that may interfere with the functioning of certain areas of the brain.

5.1. Invasion of the brain

Several types of airborne viruses including influenza, measles and mumps and others such herpes simple viruses may access the brain from the nasal cavities and produce meningitis and/or meningoencephalitis (27,76-79). Similarly, airborne bacteria including Neisseria menigitidis. Haemophilus influenza and Streptococcus pneumoniae establish infections in the nasopharinx that can spread to the meningeal spaces causing more serious types of meningitis (42,80). The course of these illnesses, treatment and mechanisms of brain damage of these infectious pathogens has been largely studied and there is today significant available information on the effects of acute meningitis and encephalitis (76,79-85). Among the best known effects are the massive recruitment of lymphocytes to the menigeal spaces and production of inflammatory molecules usually resulting in edema formation and intracranial hemorrhage (42). The neurological consequences of these processes depends on the brain region affected and range from loss of function such as impairment in vision and/or the development of seizures (42,81,82,86).

Inflammatory mechanisms of the brain's immunity have a deep impact on brain function with variable outcomes for survivors. One of the most important is the massive production of cytokines, chemokines and reactive oxygen species, mostly by microglial cells of the brain parenchyma, that produce important alterations and/or damage to neurons and nerve cells (85,87-90). The presence of toll-like receptors in glial cells allows the detection of bacterial and viral products which triggers the production of a common set of cytokines including interleukin-1β, interleukin-6, tumor necrosis factoralpha and additionally interferons in the case of viral infections (30,83,84,90-92). The effect that these cytokines may produce in the brain varies according to the intensity of the response as well as the developmental stage of the infected organism. Cytokines at high concentration are cytotoxic and may trigger neuronal cell death by necrosis or apoptosis and that may result in the loss of some function depending on the region affected (84,92,93). Moreover, if the infections occur during early development of the nervous system, they are likely to cause permanent alteration in brain function. In this case, it may not only involve loss of function but also subtle alterations in information (94-96). Cytokines have processing important neurodevelopmental functions and alteration in their expression may have important effects on brain function (97). Indeed, experimental models of maternal and neonatal influenza or BVD infection in rodents show the damaging long-term consequences on memory function and other behavioral traits (98-100).

Airborne neurotropic arenaviruses are less common and limited to endemic regions, mostly in Africa and South America. However, the mortality rate is high ranging between 15 to 35 % (36,37,101,102). These viruses are capable of accessing the brain parenchyma trough the olfactory and trigeminal pathway causing viral encephalitis (31,32,103). Their clinical course is characterized by neurological disorders including ataxia, tremors and seizures. The long term consequences in survivors are neurological disabilities and psychiatric disorders including impaired cognitive abilities and depression (32). Recent data indicates that immune mechanisms of pathogen clearance are mayor players in inducing neuronal loss and apoptosis. Recent data provided by experimental animal models of neonatal arenavirus infections via the intranasal pathway shows that the immune response has a greater role in the process of encephalitis than the virus itself (32). Infected mice deficient in T-cells had a reduced production of interferon- γ and TNF- α but a 100% survival and equal viral load in the brain to that of wild type mice (32). In addition, neutralization of TNF-a with antibodies also protected new born mice from the arenavirus encephalitis (32).

Some viruses may also access the brain by the intranasal pathway although their airborne status is less common or documented. Borna disease virus (BDV) (30,96,104-106) and herpes simplex virus-1 (HSV-1) (26,106-109) have been shown to access the brain through the intranasal pathway following olfactory and trigeminal routes and establish acute and/or permanent slow infections. Permanent asymptomatic infections may affect brain function by mechanisms that interfere with neuronal processes. Experimental models of neonatal BVD infection have shown that asymptomatic BVD infections interfere with several neuronal intracellular processes involving kinases (110-112). The dysfunction in kinase activity and interference with cell-signaling caused by the virus has been related with impaired memory function (108,113).

In summary, airborne infectious agents may access the brain through the intranasal pathway and are potent activators of the immune response of the brain that may lead to neuronal damage and long-term functional sequelae.

5.2. Inflammation in the nasal and respiratory epithelium

The most common and prevalent types of airborne inflammatory factors do not enter the brain, however they produces significant inflammatory processes in the respiratory system and in particular the nasal cavities, nasal sinuses and nasopharinx. Viruses such as influenza induce the activation of the innate immune response in respiratory epithelial cells and the production of type I interferons that initiate the antiviral response (114). Adaptive immune processes involving activation and recruitment of T- lymphocytes and later antibody production are mostly responsible for viral clearance (115,116). Experimental animal models of influenza infection show that the virus induces significant apoptosis in olfactory neurons and that this is a mechanism prevents invasion of the brain (117). On the other side, rhinoviruses

causing the common cold do not induce a strong interferon response (118,119) but produces significant inflammation of the upper respiratory tract and the induction of TNF- α , interleukin-6 and the chemokines interleukin-8 and RANTES (120). Prolonged inflammatory responses caused by rhinovirus have been associated with increased susceptibility to bacterial infections (121). Bacterial complications of the sinus such as infections with nontypeable *Haemophilus influenza* are characterized by the production of several cytokines including TNF- α , interleukin-1 β and interleukin-6 and the presence of several types of T lymphocytes (117,121-124). All together, these inflammatory pathogens produce significant changes in the immunity of the respiratory system and the type of response mounted after successive infections (125-127).

Allergic inflammation is characterized by a massive recruitment of type 2 (T_H2) T-lymphocytes into the nasal epithelium and the production of the cytokines IL-4, IL-5 and IL-13 (60). In individuals with respiratory allergies, the cytokine response of T-cells is biased towards a $T_{\rm H}2$ type characterized by the production of IL-4, IL-5 and IL-13 (128). This T_{H2} biased response drives isotype switching towards the production of IgE, which in turn, sensitizes mast cells to respond to environmental stimuli. Mast cells are resident immune cells involved in many types of defense mechanisms against pathogens and are instrumental in the antibacterial, anti-parasitic and allergic responses. Mast cells are the first responder to pathogens and allergens in the nasal cavity and are involved in both innate and adaptive immune responses (60,129-131). They constitutively and upon activation produce several cytokines including IL-5, IL-13 and TNF- α , and massively release them during degranulation. Moreover, they release several neurotransmitter and inflammatory mediators such as histamine, leukotrienes and proteases responsible for the nasal symptoms of allergies. T-lymphocytes (T-cells) are also important source of T_H2 cytokines in the nasal epithelium (in particular IL-4 and IL-13) during allergic processes (60,130). Although their presence and production of cytokines is limited during early stages of allergic responses, their number and activity increases rapidly thereafter becoming perhaps the major producers of IL-4 and IL-13 during allergic inflammation. Eosinophils and basophils also contribute significantly to the production of cytokines including IL-5 and IL-13 and other immune mediators such as proteases and leukotrienes. A growing number of cytokines have been shown produced and released during allergic inflammation including IL-3, IL-9 and IL-17 with relatively lower titers with respect to those of IL-4, IL-5 and IL-13. Repeated and chronic exposure to allergens may lead to chronic allergic inflammation characterized by significant tissue damage and remodeling with a consequent decline in function.

Pollutants may induce either a pro-inflammatory response in the nasal epithelium or exacerbate allergic inflammatory processes. Most animal models showed pro-inflammatory processes with the production of TNF- α that were induced in response to several contaminants such as carbon ultrafine particles (132-134). On the other hand, the combination of diesel-exhaust particles or particulate

matter with pollen increases specific IgE antibody responses against pollen (135) and IL-4 production (136) resulting in the amplification of allergic inflammation. Therefore, pollutants generally induce a pro-inflammatory response when acting alone or may serve as adjuvant of allergic inflammation when combined with allergens.

Although the inflammatory process and the production of cytokine described above are mainly restricted to the nasal and respiratory epithelium, recent experiments from our laboratory showed that inflammation in the nasal cavities is a stimulus capable of inducing cytokine production in the brain (137). Intranasal administration of bacterial products such as LPS resulted in the expression of the cvtokines TNF- α and IL-6 in the brainstem and hippocampus of rats. These effects were more pronounced in females as compared to males and simultaneous whit the development of depressive-like behavior (see next section). Similarly, studies from other laboratories on experimental animal models showed that exposure to PM are capable of inducing cytokine expression in the brain of mice (134,138). Ultrafine carbon particles intranasally instilled or aerosolized induced the expression of IL-1 α and β , TNF- α and several chemokines including MIP-1a and CXCL9 (134,138). Recently, it has been shown that exposure to PM increased the levels of glial fibrillary acid protein (GFAP) and produced activation of the transcription factors NF-kappa B and AP-1 in the brain (133). Furthermore, intranasal instillations with polyinosinic:polycytidylic acid (poly I:C) that mimics immune processes triggered by viruses were shown to induce cytokine expression in the brain (139). Together, these studies show that bacterial, viral and pollutant products administered in the nasal cavities induces activation of the immune response inside the brain probably reflecting inflammation in the nasal cavities.

Allergic inflammatory processes of the airways such as allergic rhinitis and allergic asthma have been shown to affect brain function (140-143). Exposure to the allergen in sensitized rodents is also capable of inducing the expression of cytokines in the brain. Data from experimentally-induced allergic rhinitis in rats showed that the cytokines IL-4, 5 and 13 are induced in the prefrontal cortex and olfactory bulbs of rats made allergic to tree pollen (144). Expression of these cytokines in the same brain regions were also observed in mice with experimentally-induced allergic rhinitis to albumin from chicken egg (OVA) (144). Moreover, neuronal activation of limbic regions was reported in models of allergic asthma in OVA-sensitized mice intranasally challenged with the allergen (141,142). Using the expression of c-fos as a marker of neuronal activation, several responsive areas in the brain including the amygdala and hypothalamus were observed during allergic processes of the airways (141,142). These studies show that airborne allergies are capable of affecting the brain inducing the expression of $T_{\rm H}2$ cytokines and producing the activation of several brain regions.

In summary, data from different sources shows that intranasal immune challenge resulting in distinct

inflammatory processes of the nasal epithelium is a stimulus capable of activating at least the molecular inflammatory response of the brain.

6. EFFECTS ON BEHAVIOR

6.1. Acute Infections

As discussed earlier, the effects on brain function and behavior of acute meningitis or encephalitis has been extensively studied. The most important consequences are loss of function such as impaired vision and hearing, movement disorders or ataxia and seizures (76,80). Survivors of bacterial or viral meningitis or encephalitis usually present alterations in behavior that relates to the degree of damage caused during the acute phase. Despite extensive research in this subject, the mechanisms on how brain function and behavior are affected after viral clearance and resolution of symptoms are only partially understood (145). Depression, anxiety and cognitive impairments have been described in survivors of acute encephalitis (145). Experimental animal models of prenatal infection with influenza virus reveal the important role that the immune response has on neurodevelopment (76,80,97,99,146). Infections with influenza virus in pregnant dams results in neurological and behavioral alterations in the offspring (97-99,146). The offspring of mice infected during pregnancy display deficits in exploratory behavior, social interaction and recognition of novel objects (99). Moreover, specific deficits in acoustic startle responses similar to those observed in humans with schizophrenia were also reported (99). These studies determined that the virus was not present in the fetus and that the effects on the brain and behavior are likely related to the activation of the maternal immune response. These findings have led to the hypothesis of a maternal immune activation as a possible etiology for cases of schizophrenia (98,99).

6.2. Permanent infections

Permanent infections or slow viral infections of the CNS such as those established by BDV and HSV virus have been strongly implicated with impairment in learning and memory function (96,112,113). For instance, using models of neonatal BVD infection it is possible to detect a progressive impairment in the performance of rats in the Morris Water Maze test (96). This test is usually employed to assess spatial learning and memory function. When BVD infected animals were evaluated for ambulatory activity in an open field, no signs of altered behavior was observed (113) showing that only specific behavioral alterations were caused by the virus. Many of the behavioral effects of permanent or slow BVD infection may not involve activation of the cytokine response of the brain (113). This may be related with mechanism of evasion from immune surveillance and the ability of BDV to establish permanent infections (147). BVD usually affects the hippocampus and cerebral cortex, regions known to process several aspects of memory function (111,148). Using cell culture systems it has been shown that BVD blocks potentiation of synaptic activity of neurons that is believed a neuronal process associated with memory formation and consolidation (110). Moreover, the effects of BVD infection in neurons seem to

be specific for activity dependent changes in synaptic function without affecting other electrical properties (112). These effects relates with the interference of BVD virus of several intracellular phosphorylation processes required for synaptic plasticity.

6.3. Inflammation in the airways

It is known that activation of the peripheral immune response produces behavioral responses of depression and anxiety (149-151). Some features of depressed patients including anhedonia, loss of appetite, sleep disturbances and reduced general activity are similar to the behavioral symptoms of sickness behavior. This has led to hypothesize that the high incidence of mood and anxiety disorders in clinically ill people stems from an immune or neuroimmune origin (149,151,152).

During inflammatory processes, depression and anxiety in humans and depressive-like and anxiety-like behaviors in animals have been shown associated or mediated by the actions of a specific subset of proinflammatory cytokines including interleukin-1ß (IL-1ß) IL-2, IL-6, and tumor necrosis factor- α (TNF- α) and the antiviral cytokines interferons α , β and γ (149-151,153,154). Interferons and IL-2 have been shown to promote depression mainly by acting in the periphery by mechanisms involving the enzyme indolamine 2.3 dyoxigenase (IDO) (149). IL-1 β , IL-6 and TNF- α have been shown to influence depression by peripheral mechanisms and by direct central actions in the CNS (149,154). Central mechanisms of these cytokines involve activation of the HPA-axis and a reduction in neurogenesis. We have also proposed that behaviors of depression and anxiety may result by direct actions of these cytokines at the synaptic level by reducing synaptic plasticity (155).

One known mechanisms of the immune response of the brain is the induction of inflammatory genes in response to peripheral immune challenge (152,155-158). Thus, inflammatory processes of the airways that may result in the expression of cytokines in the brain may result in altered behavioral responses. Indeed, intranasal administration of bacterial LPS in rats induced depressivelike behavior and exaggerated hormonal responses to stress without any overt sign of sickness (137). Rats administered for 2 consecutive days in the nasal cavities with bacterial LPS had reduced swimming time in the forced swim test which is indicative of behavioral despair. These effects were paralleled by the expression of $TNF-\alpha$ and IL-6 in the brain. Although it has not been determined if the depressive-like behavior was produced by the expression of the cytokines in the brain, it is likely that the may play a contributing role for the manifestation of depressive-like behavior. These studies were intended to mimic the activation of the immune system due to a gram negative bacterial infection in the nasal cavities. However, it may also represent inflammatory processes triggered by PM containing LPS.

Similarly, allergic inflammatory processes of the airways have been shown to induce anxiety-like behaviors. Animals sensitized and exposed to an allergen shows

increased anxiety-like behavior as determined by the elevated plus maze test and the open field test (144). Allergic mice spend less time in the open arms of an elevated plus maze or in the center area of an open field which are considered measures of increased anxiety. Moreover, evidence of increased anxiety caused by aerosol challenge with allergen was obtained using a modified version of the passive avoidance test (142). Mice that were sensitized and then exposed to an allergen in a compartment showed avoidance to that compartment when re-exposed a day later in the absence of the allergen. Thus, allergic mice developed an aversion to the context in which the allergic reaction occurred suggesting that neuroimmune mechanisms were capable of behaviorally conditioning allergic mice (142).

7. IMPLICATIONS FOR NEUROLOGICAL AND MENTAL DISEASES

It has been hypothesized that neurological and mental disorders of unknown etiology may have an infectious or immunological origin (98,100,159). The data supporting an inflammatory component originates from epidemiological, clinical, postmortem and basic animal models. However, attempts to evaluate and establish a direct relationship between these disorders and specific infectious agents or inflammatory diseases have been elusive. In this regard, it has to be considered that while studies exist that lead to these hypotheses, this area of research is on its beginning. Moreover, taking into account the likelihood that the clinical manifestation of a specific neurological or mental disorder may originate from different etiologies, attempts to relate complex disorders of the brain to a single infectious or inflammatory source are likely to produce mixed or inconclusive results. Recently, evidence of the presence of a specific infectious agent associated with a specific disease started to emerge supporting the more indirect epidemiological analysis encountered often in the literature. Together with cases of effectiveness of response to treatment using immunomodulatory drugs, we will briefly present some of the evidence suggesting a relationship between some neurological and mental disorders with immune mediated processes affecting the brain involving the intranasal pathway and possibly airborne exposure to inflammatory factors.

7.1. Neurological disorders

Alzheimer's disease (AD), multiple sclerosis (MS) and Parkinson's disease (PD) have been proposed to have an inflammatory etiology or at least, that inflammatory processes are a contributing factor or a factor of risk (98,160-166). In the case of AD and MS, specific viruses have been studied in association with these diseases while PD has been associated with exposure to environmental toxins and the occurrence of rhinorrhea. Perhaps, the most compelling evidence for an association between a neurological disorder and a virus is the case of MS (167-172). Human Herpes viruses (HHVs) have been found with more frequency in MS patients with respect to other neurological diseases and controls. Varicella zoster virus (HHV-3), Epstein-Barr virus (HHV-4) and HHV-6 in

conjunction with a strong T-cell responses against these viruses have been consistently found in MS patients (168,171-176). With the exception of HHV-6 which an airborne transmission is not known, HHV-3 and HHV-4 can be transmitted by moist droplets containing the virus and infect the nasopharinx and also display neurotrophism accessing the brain via the intranasal pathway. Measles virus has also been associated with MS and a similar body of evidence supports a role of measles virus with MS (164,172). Evidence of antibody titers against measles virus in the cerebrospinal fluid of MS patients has been reported (172). Also, epidemiological evidence of an association between MS and measles virus is available (164). Other than the form of transmission, the initial infection site in the nasopharinx and access to the brain via the intranasal pathway, these two groups of viruses do not share many similarities.

The pattern of functional loss and degeneration from the olfactory sensory group in both AD and PD has lead to the olfactory degenerative hypothesis for these diseases (159,177-179). In these cases, a causative environmental factor is assumed to initiate in the olfactory bulbs the series of neurological alterations that lead to the progressive degeneration along the olfactory pathway. In the case of AD, one of such potential factors is herpes simplex virus type 1 (HSV-1) (160,161,165,180-183). Although the occurrence of aerosolized HSV-1 infection is rare, there is evidence that the spread of infection to the brain occurs trough the intranasal pathway (26,33,139,184). Experimental animal models of intranasal HSV virus infection shows that the virus spreads through the olfactory pathway reaching the limbic regions where degeneration occurs in AD. Amyloid deposition in neurons in response to HSV infection has also been demonstrated using cell culture (182). Data demonstrating the presence of HSV virus in the amyloid plaques of AD patients was reported by two studies (180,183). These studies also reported that the interaction between genetic susceptibility for amyloid deposition and a viral presence was associated with increased neurodegeneration.

The evidence of a viral involvement in PD is only circumstantial and limited (185). However, epidemiological data points to a different inflammatory process to that of viral or bacterial infections. Studies have reported increased susceptibility for PD associated with exposure to environmental contaminants and in particular with pesticides (165,180,183,186). In addition to these factors, PD has been associated with the presence of rhinorrhea and immediate-hypersensitive reactions (187). These studies suggest that inflammatory processes in the nasal epithelium may be of importance if there is in fact a relationship between PD and environmental contaminants. In this regard, an involvement of inflammatory processes in PD has been demonstrated by the presence of elevated cytokines in the brain of PD patients and in animal models of PD (166,188,189). This is corroborated by the protection to PD conferred by the use of anti-inflammatory drugs and the reductions of symptoms in animal models (190,191). Furthermore, specific markers of the adaptive immunity involving T-lymphocytes have been found in the brain of postmortem PD patients and in animal models of PD (192).

While the type of inflammatory agent or pathogen that has been associated with these diseases differs, the unifying point that represents the route of exposure involving inflammatory processes initiated in the nasal epithelium and regions of the brain affected in relation with the intranasal pathway highlights the potential role of neuroimmune mechanisms at the environment-brain interface. If research continues to produce evidence of an inflammatory role in these diseases for airborne infectious, toxic and allergic factors, understanding of neuroimmune mechanisms of the intra nasal pathway will be crucial for a better approach for the prevention and treatment of these diseases.

7.2. Mental Disorders

The concept that immune function may play a role in mental illness and in particular depression has been reviewed several times (149,193-196). The concept has been built based on several lines of evidence including a) depression and anxiety occurring in a large number of patients receiving cytokine therapy for hepatitis-C or melanoma (197) b) the presence of markers of inflammation in depressed patient and the normalization after antidepressant treatment (198) c) the occurrence of depression in autoimmune and chronically ill patients (199) d) animal studies showing that challenge with bacterial and viral products as well as cytokines induces features of depressive-like behavior (149,150,193). There is today general consensus on some mechanisms of neuroimmune interaction that leads to the occurrence or worsening of depression (see section 6.2). However, the possibility that mood disorders and other psychiatry diseases may be related (at least in part) with inflammatory processes involving the intranasal pathway has never been considered. One observation in favor of this connection is that the most consistent findings of anomalies in the brain of patients with mood disorders and animal models of depression involve the fronto-temporal regions (200). The frontal cortex and hippocampus that are regions closely related with the intranasal pathway consistently show neurochemical and cellular alterations that reflects in functional deficits in mood disorders patients (200-203). A genetic predisposition and exposure to stress have been identified as the major contributors to alterations on these brain regions. Specifically, the role of the stress response and the effects of glucocorticoids on brain architecture and neurochemistry are proposed as a major mechanism leading to the neuropathology of mood disorders in fronto-temporal regions (200). While inflammatory processes of the airways and the intranasal pathway are unlikely to be the cause of these neurochemical alterations and the development of depression, they are positioned to greatly influence its outcome. We have recently shown that in rats, subsyndromal inflammatory processes of the airway mimicking bacterial or toxic exposure are capable of enhancing the glucocrticoid response to stress (137). Moreover, they were capable of inducing transcription of pro-inflammatory cytokines in the hippocampus. The simultaneous effects of cytokines in the brain with increased hormonal responses to stress are likely to result in increased deleterious effects to a stressor on hippocampal neurons.

Epidemiological and clinical studies report a higher incidence of depression and anxiety in individuals suffering from respiratory allergies including allergic rhinitis and allergic asthma (56,204-212). Increased state and trait anxiety were identified using the state and trait anxiety inventory (STAI) in individuals suffering from allergic rhinitis compared with matched controls (208). These findings were recently replicated by an independent study including atopic dermatitis and seasonal allergic rhinitis with the use of the STAI (209). Comparisons of allergic vs non-allergic asthmatic patients revealed and association with anxiety only for allergic asthmatics independent of age, sex, smoking status and asthma severity (210). This study adds evidence for a role of allergies to the vast literature reporting increased anxiety in asthmatic patients (206,211,213). As discussed in section 6.2, experimentally induced allergic rhinitis and asthma in rodents induces anxiety-like behavior, yet the mechanisms linking respiratory allergies with anxiety and depression are poorly understood.

It has been recently hypothesized that a changing microbial environment is responsible for a faulty mechanism of the immune response in terminating inflammatory processes (214). This has been related with the increased incidence of chronic inflammatory diseases in developed countries such as allergic rhinitis and asthma, type I diabetes and MS and inflammatory bowel diseases among others (214,215). The so-called "hygiene hypothesis" or "old friends hypothesis" has been recently proposed in relation with the increased incidence of psychiatric diseases, in particular anxiety and mood disorders (216). Rook and Lowry proposes that a low ratio of regulatory to effector T cells possibly related with diminished harmless organisms and almost absent parasitic Helminth infections in developed countries is the cause of the high incidence of chronic inflammatory diseases. Moreover, the authors propose that T_H1- driven chronic inflammatory processes and the production of IFNy will be associated with increased incidence of depression in individuals with autoimmune diseases. Conversely, T_H2driven chronic inflammatory processes and the production of IL-4, 5 and 13 will be associated with a higher incidence of anxiety disorders in allergic individuals. In relation with inflammatory processes of the airways, this view is consistent with our findings that intranasal immune challenge with LPS induces depressive-like behavior (137) and that intranasal challenge with allergens induces anxiety-like behavior (144).

In summary, activation of the immune system and elevation in cytokines has been shown to promote depression and anxiety. The high rate of exposure to airborne inflammatory factors and the characteristics of the intranasal pathway may play a contributing role by triggering, worsening or perpetuating mood and anxiety disorders.

8. SUMMARY AND PERSPECTIVES

The concept that infectious diseases may play a role in neurological and mental disorders has been

proposed several times, nevertheless studies have produced mixed results and conclusive evidence is still lacking. New evidence linking infectious agents with diseases of the brain have emerged recently and this subject of research may continue to provide information about a potential environmental role on neurological and mental disorders. However, it is acknowledged that these diseases are highly heterogeneous on their clinical manifestations and that they may result from the interaction between genetic susceptibility and several environmental variables. Therefore, if there is an inflammatory origin in the etiology of any of these disorders, it is likely that more than one single infectious or inflammatory environmental factor is involved and capable of producing a group of symptoms leading to a specific clinical manifestation. We have proposed that the intranasal pathway represents one unifying mechanism by which environmental inflammatory factors may initiate, maintain or influence the progression of neurological and mental disorders. The constant or repetitive nature of exposure to airborne inflammatory factors that covers the lifespan of an individual may play an important role in the magnitude of the inflammatory response mounted by the brain. Importantly, we suggest that these inflammatory factors influence each other as evidenced by the increased co-occurrence of infectious and allergic diseases and the link between air pollution and airway inflammation. Potential neuroimmune mechanisms at the nasal epithelium and activation of the immune response of the brain are likely candidates to affect specific brain-regions related with the intranasal pathway involving the olfactory-frontotemporal regions and the trigeminal target areas. However, these mechanisms have to be yet characterized and explored in more detail to understand how exposure of environmental inflammatory factors may play a role in brain function and disease.

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9. REFERENCES

1. Langmuir AD. Changing concepts of airborne infection of acute contagious diseases: a reconsideration of classic epidemiologic theories *Ann N Y Acad Sci* 353: 35-44 (1980)

2. Kinney PL. Climate change, air quality, and human health *Am J Prev Med* 35: 459-67 (2008)

3. Bartra J, Mullol J, del Cuvillo A, Davila I, Ferrer M, Jauregui I, Montoro J, Sastre J, Valero A. Air pollution and allergens *J Investig Allergol Clin Immunol* 17 Suppl 2: 3-8 (2007)

4. Ebi KL, McGregor G. Climate change, tropospheric ozone and particulate matter, and health impacts *Environ Health Perspect* 116: 1449-55 (2008)

5. Glezen WP. The changing epidemiology of respiratory syncytial virus and influenza: impetus for new control measures Pediatr Infect Dis J 2004; 23: S202-6.

6. Weiss N, Miller F, Cazaubon S, Couraud PO. The bloodbrain barrier in brain homeostasis and neurological diseases *Biochim Biophys Acta* (2008)

7. Quan N, Banks WA. Brain-immune communication pathways *Brain Behav Immun* 21: 727-35 (2007)

8. Fry M, Ferguson AV. The sensory circumventricular organs: brain targets for circulating signals controlling ingestive behavior *Physiol Behav* 91: 413-23 (2007)

9. Harkema JR, Carey SA, Wagner JG. The nose revisited: a brief review of the comparative structure, function, and toxicologic pathology of the nasal epithelium *Toxicol Pathol* 34: 252-69 (2006)

10. Ross TM, Zuckermann RN, Reinhard C, Frey WH 2nd. Intranasal administration delivers peptoids to the rat central nervous system *Neurosci Lett* 439: 30-3 (2008)

11. Thorne RG, Pronk GJ, Padmanabhan V, Frey WH 2nd. Delivery of insulin-like growth factor-I to the rat brain and spinal cord along olfactory and trigeminal pathways following intranasal administration *Neuroscience* 127: 481-96 (2004)

12. Hanson LR, Frey WH 2nd. Intranasal delivery bypasses the blood-brain barrier to target therapeutic agents to the central nervous system and treat neurodegenerative disease *BMC Neurosci* 9 Suppl 3: S5 (2008)

13. Ross TM, Martinez PM, Renner JC, Thorne RG, Hanson LR, Frey WH 2nd. Intranasal administration of interferon beta bypasses the blood-brain barrier to target the central nervous system and cervical lymph nodes: a non-invasive treatment strategy for multiple sclerosis *J Neuroimmunol* 151: 66-77 (2004)

14. Gozes I, Giladi E, Pinhasov A, Bardea A, Brenneman DE. Activity-dependent neurotrophic factor: intranasal administration of femtomolar-acting peptides improve performance in a water maze *J Pharmacol Exp Ther* 293:1091-8 (2000)

15. Deretzi G, Zou LP, Pelidou SH, Nennesmo I, Levi M, Wahren B, Mix E, Zhu J. Nasal administration of recombinant rat IL-4 ameliorates ongoing experimental autoimmune neuritis and inhibits demyelination *J Autoimmun* 12: 81-9 (1999)

16. Kalueff AV, Lehtimaki KA, Ylinen A, Honkaniemi J, Peltola J. Intranasal administration of human IL-6 increases the severity of chemically induced seizures in rats *Neurosci Lett* 365: 106-10 (2004)

17. Pelidou SH, Zou LP, Deretzi G, Nennesmo I, Wei L, Mix E, Van Der Meide PH, Zhu J. Intranasal administration of recombinant mouse interleukin-12 increases inflammation and demyelination in chronic experimental autoimmune neuritis in Lewis rats *Scand J Immunol* 51: 29-35 (2000)

18. Benedict C, Hallschmid M, Schmitz K, Schultes B, Ratter F, Fehm HL, Born J, Kern W. Intranasal insulin improves memory in humans: superiority of insulin aspart *Neuropsychopharmacology* 32: 239-43 (2007)

19. Benedict C, Hallschmid M, Hatke A, Schultes B, Fehm HL, Born J, Kern W. Intranasal insulin improves memory in humans *Psychoneuroendocrinology* 29: 1326-34 (2004)

20. Reger MA, Watson GS, Frey WH 2nd, Baker LD, Cholerton B, Keeling ML, Belongia DA, Fishel MA, Plymate SR, Schellenberg GD, Cherrier MM, Craft S. Effects of intranasal insulin on cognition in memoryimpaired older adults: modulation by APOE genotype *Neurobiol Aging* 27: 451-8 (2006)

21. Burri A, Heinrichs M, Schedlowski M, Kruger TH. The acute effects of intranasal oxytocin administration on endocrine and sexual function in males *Psychoneuroendocrinology* 33: 591-600 (2008)

22. Ditzen B, Schaer M, Gabriel B, Bodenmann G, Ehlert U, Heinrichs M. Intranasal Oxytocin Increases Positive Communication and Reduces Cortisol Levels During Couple Conflict *Biol Psychiatry* (2008)

23. Perras B, Schultes B, Behn B, Dodt C, Born J, Fehm HL. Intranasal atrial natriuretic peptide acts as central nervous inhibitor of the hypothalamo-pituitary-adrenal stress system in humans *J Clin Endocrinol Metab* 89: 4642-8 (2004)

24. Perras B, Marshall L, Kohler G, Born J, Fehm HL. Sleep and endocrine changes after intranasal administration of growth hormone-releasing hormone in young and aged humans *Psychoneuroendocrinology* 24: 743-57 (1999)

25. van Wingen GA, Zylicz SA, Pieters S, Mattern C, Verkes RJ, Buitelaar JK, Fernandez G. Testosterone increases amygdala reactivity in middle-aged women to a young adulthood level *Neuropsychopharmacology* 34: 539-47 (2009)

26. Mori I, Goshima F, Ito H, Koide N, Yoshida T, Yokochi T, Kimura Y, Nishiyama Y. The vomeronasal chemosensory system as a route of neuroinvasion by herpes simplex virus *Virology* 334: 51-8 (2005)

27. Iwasaki T, Itamura S, Nishimura H, Sato Y, Tashiro M, Hashikawa T, Kurata T. Productive infection in the murine central nervous system with avian influenza virus A (H5N1) after intranasal inoculation *Acta Neuropathol* 108: 485-92 (2004)

28. Sauder CJ, Vandenburgh KM, Iskow RC, Malik T, Carbone KM, Rubin SA. Changes in mumps virus neurovirulence phenotype associated with quasispecies heterogeneity *Virology* 350: 48-57 (2006)

29. Sellin CI, Davoust N, Guillaume V, Baas D, Belin MF, Buckland R, Wild TF, Horvat B. High pathogenicity of

wild-type measles virus infection in CD150 (SLAM) transgenic mice *J Virol* 80: 6420-9 (2006)

30. Shankar V, Kao M, Hamir AN, Sheng H, Koprowski H, Dietzschold B. Kinetics of virus spread and changes in levels of several cytokine mRNAs in the brain after intranasal infection of rats with Borna disease virus *J Virol* 66: 992-8 (1992)

31. Pedras-Vasconcelos JA, Goucher D, Puig M, Tonelli LH, Wang V, Ito S, Verthelyi D. CpG oligodeoxynucleotides protect newborn mice from a lethal challenge with the neurotropic Tacaribe arenavirus *J Immunol* 176: 4940-9 (2006)

32. Pedras-Vasconcelos JA, Puig M, Sauder C, Wolbert C, Ovanesov M, Goucher D, Verthelyi D. Immunotherapy with CpG oligonucleotides and antibodies to TNF-alpha rescues neonatal mice from lethal arenavirus-induced meningoencephalitis *J Immunol* 180: 8231-40 (2008)

33. Soucy G, Boivin G, Labrie F, Rivest S. Estradiol is required for a proper immune response to bacterial and viral pathogens in the female brain *J Immunol* 174: 6391-8 (2005)

34. Morawska L. Droplet fate in indoor environments, or can we prevent the spread of infection? Indoor Air 16: 335-47 (2006)

35. Griffin DW. Atmospheric movement of microorganisms in clouds of desert dust and implications for human health Clin Microbiol Rev 20: 459-77, table of contents. (2007)

36. Enria DA, Pinheiro F. Rodent-borne emerging viral zoonosis. Hemorrhagic fevers and hantavirus infections in South America Infect Dis Clin North Am 14: 167-84, x. (2000)

37. Krautkramer E, Zeier M. Hantavirus causing hemorrhagic fever with renal syndrome enters from the apical surface and requires decay-accelerating factor (DAF/CD55) J Virol 82: 4257-64 (2008)

38. Simonsen L, Clarke MJ, Schonberger LB, Arden NH, Cox NJ, Fukuda K. Pandemic versus epidemic influenza mortality: a pattern of changing age distribution J Infect Dis 178: 53-60 (1998)

39. Brammer L, Fukuda K, Arden N, Schmeltz LM, Simonsen L, Khan A, Regnery HL, Schonberger LB, Cox NJ. Influenza surveillance--United States, 1992-93 and 1993-94 MMWR CDC Surveill Summ 46: 1-12 (1997)

40. Simonsen L, Fukuda K, Schonberger LB, Cox NJ. The impact of influenza epidemics on hospitalizations J Infect Dis 181: 831-7 (2000)

41. Griffin MR, Coffey CS, Neuzil KM, Mitchel EF Jr, Wright PF, Edwards KM. Winter viruses: influenza- and

respiratory syncytial virus-related morbidity in chronic lung disease *Arch Intern Med* 162: 1229-36 (2002)

42. de Souza AL, Seguro AC. Two centuries of meningococcal infection: from Vieusseux to the cellular and molecular basis of disease *J Med Microbiol* 57: 1313-21 (2008)

43. Noakes CJ, Beggs CB, Sleigh PA, Kerr KG. Modelling the transmission of airborne infections in enclosed spaces *Epidemiol Infect* 134: 1082-91 (2006)

44. Dowell SF, Ho MS. Seasonality of infectious diseases and severe acute respiratory syndrome-what we don't know can hurt us *Lancet Infect Dis* 4: 704-8 (2004)

45. Barker WH, Mullooly JP. "A study of excess mortality during influenza epidemics in the United States, 1968-1976" *Am J Epidemiol* 115: 479-80 (1982)

46. Glezen WP, Payne AA, Snyder DN, Downs TD. Mortality and influenza *J Infect Dis* 146: 313-21 (1982)

47. Sande MA, Gwaltney JM. Acute community-acquired bacterial sinusitis: continuing challenges and current management *Clin Infect Dis* 39 Suppl 3: S151-8 (2004)

48. Kaliner MA, Osguthorpe JD, Fireman P, Anon J, Georgitis J, Davis ML, Naclerio R, Kennedy D. Sinusitis: bench to bedside. Current findings, future directions *Otolaryngol Head Neck Surg* 116: S1-20 (1997)

49. Nadadur SS, Miller CA, Hopke PK, Gordon T, Vedal S, Vandenberg JJ, Costa DL. The complexities of air pollution regulation: the need for an integrated research and regulatory perspective *Toxicol Sci* 100: 318-27 (2007)

50. Gwaltney JM Jr. Acute community-acquired sinusitis *Clin Infect Dis* 23: 1209-23; quiz 1224-5 (1996)

51. Nakaji S, Parodi S, Fontana V, Umeda T, Suzuki K, Sakamoto J, Fukuda S, Wada S, Sugawara K. Seasonal changes in mortality rates from main causes of death in Japan (1970--1999) *Eur J Epidemiol* 19: 905-13. (2004)

52. Talbot TR, Poehling KA, Hartert TV, Arbogast PG, Halasa NB, Edwards KM, Schaffner W, Craig AS, Griffin MR. Seasonality of invasive pneumococcal disease: temporal relation to documented influenza and respiratory syncytial viral circulation *Am J Med* 118: 285-91 (2005)

53. Cooper DL, Smith GE, Edmunds WJ, Joseph C, Gerard E, George RC. The contribution of respiratory pathogens to the seasonality of NHS Direct calls *J Infect* 55: 240-8 (2007)

54. Dowell SF, Whitney CG, Wright C, Rose CE Jr, Schuchat A. Seasonal patterns of invasive pneumococcal disease *Emerg Infect Dis* 9: 573-9 (2003)

55. Lowen AC, Mubareka S, Steel J, Palese P. Influenza virus transmission is dependent on relative humidity and temperature *PLoS Pathog* 3: 1470-6 (2007)

56. Blaiss MS. Allergic rhinoconjunctivitis: burden of disease *Allergy Asthma Proc* 28: 393-7 (2007)

57. Meltzer EO. The prevalence and medical and economic impact of allergic rhinitis in the United States *J Allergy Clin Immunol* 99: S805-28 (1997)

58. Meltzer EO, Szwarcberg J, Pill MW. Allergic rhinitis, asthma, and rhinosinusitis: diseases of the integrated airway *J Manag Care Pharm* 10: 310-7 (2004)

59. Nathan RA. The burden of allergic rhinitis *Allergy Asthma Proc* 28: 3-9 (2007)

60. Galli SJ, Tsai M, Piliponsky AM. The development of allergic inflammation *Nature* 454: 445-54 (2008)

61. Frenz DA. Interpreting atmospheric pollen counts for use in clinical allergy: allergic symptomology *Ann Allergy Asthma Immunol* 86: 150-7, quiz 158 (2001)

62. Kosisky SE, Carpenter GB. Predominant tree aeroallergens of the Washington, DC area: a six year survey (1989-1994) *Ann Allergy Asthma Immunol* 78: 381-92 (1997)

63. Mallia P, Johnston SL. How viral infections cause exacerbation of airway diseases *Chest* 130: 1203-10 (2006)

64. Cirillo I, Marseglia G, Klersy C, Ciprandi G. Allergic patients have more numerous and prolonged respiratory infections than nonallergic subjects *Allergy* 62: 1087-90 (2007)

65. Zeldin Y, Weiler Z, Cohen A, Kalinin M, Schlesinger M, Kidon M, Magen E. Efficacy of nasal Staphylococcus aureus eradication by topical nasal mupirocin in patients with perennial allergic rhinitis *Ann Allergy Asthma Immunol* 100: 608-11 (2008)

66. Tamay Z, Akcay A, Ones U, Guler N, Kilic G, Zencir M. Prevalence and risk factors for allergic rhinitis in primary school children *Int J Pediatr Otorhinolaryngol* 71: 463-71 (2007)

67. Wold LE, Simkhovich BZ, Kleinman MT, Nordlie MA, Dow JS, Sioutas C, Kloner RA. *In vivo* and *in vitro* models to test the hypothesis of particle-induced effects on cardiac function and arrhythmias *Cardiovasc Toxicol* 2006; 6: 69-78.

68. Simkhovich BZ, Kleinman MT, Kloner RA. Air pollution and cardiovascular injury epidemiology, toxicology, and mechanisms *J Am Coll Cardiol* 52: 719-26 (2008)

69. Michel O, Nagy AM, Schroeven M, Duchateau J, Neve J, Fondu P, Sergysels R. Dose-response relationship to inhaled endotoxin in normal subjects *Am J Respir Crit Care Med* 156: 1157-64 (1997)

70. Davila I, Mullol J, Bartra J, Del Cuvillo A, Ferrer M, Jauregui I, Montoro J, Sastre J, Valero A. Effect of

pollutants upon patients with respiratory allergies *J Investig Allergol Clin Immunol* 17 Suppl 2: 9-20 (2007)

71. Mundandhara SD, Becker S, Madden MC. Effects of diesel exhaust particles on human alveolar macrophage ability to secrete inflammatory mediators in response to lipopolysaccharide *Toxicol In vitro* 20: 614-24 (2006)

72. Gyan K, Henry W, Lacaille S, Laloo A, Lamsee-Ebanks C, McKay S, Antoine RM, Monteil MA. African dust clouds are associated with increased paediatric asthma accident and emergency admissions on the Caribbean island of Trinidad *Int J Biometeorol* 49: 371-6 (2005)

73. Monteil MA. Saharan dust clouds and human health in the English-speaking Caribbean: what we know and don't know *Environ Geochem Health* 30: 339-43 (2008)

74. Sigsgaard T, Bonefeld-Jorgensen EC, Hoffmann HJ, Bonlokke J, Kruger T. Microbial cell wall agents as an occupational hazard *Toxicol Appl Pharmacol* 207: 310-9 (2005)

75. Michel O. Role of lipopolysaccharide (LPS) in asthma and other pulmonary conditions *J Endotoxin Res* 9: 293-300 (2003)

76. McCarthy M. Viral infections. In: Richard T. Johnson JWGJCM, ed.

77. Watson-Creed G, Saunders A, Scott J, Lowe L, Pettipas J, Hatchette TF. Two successive outbreaks of mumps in Nova Scotia among vaccinated adolescents and young adults *CMAJ* 175: 483-8 (2006)

78. Boyd M, Clezy K, Lindley R, Pearce R. Pandemic influenza: clinical issues *Med J Aust* 185: S44-7 (2006)

79. Norrby E, Kristensson K. Measles virus in the brain *Brain Res Bull* 44: 213-20. (1997)

80. Aksamit Jr A . Nonviral Infectious Disease. In: Johnson R, Griffin J, McArthur Justin, eds. Current therapy in neurologic diseases.

81. Toovey S. Influenza-associated central nervous system dysfunction: a literature review *Travel Med Infect Dis* 6: 114-24. (2008)

82. Misra UK, Tan CT, Kalita J. Viral encephalitis and epilepsy Epilepsia 49 Suppl 6: 13-8 (2008)

83. Chauhan VS, Sterka DG Jr, Gray DL, Bost KL, Marriott I. Neurogenic exacerbation of microglial and astrocyte responses to Neisseria meningitidis and Borrelia burgdorferi *J Immunol* 180: 8241-9 (2008)

84. Konat GW, Kielian T, Marriott I. The role of Toll-like receptors in CNS response to microbial challenge *J Neurochem* 99: 1-12 (2006)

85. Mitchell L, Smith SH, Braun JS, Herzog KH, Weber JR, Tuomanen EI. Dual phases of apoptosis in pneumococcal meningitis *J Infect Dis* 190: 2039-46 (2004)

86. Urbanska EM, Chambers BJ, Ljunggren HG, Norrby E, Kristensson K. Spread of measles virus through axonal pathways into limbic structures in the brain of TAP1 -/- mice *J Med Virol* 52: 362-9 (1997)

87. Marques CP, Cheeran MC, Palmquist JM, Hu S, Lokensgard JR. Microglia are the major cellular source of inducible nitric oxide synthase during experimental herpes encephalitis *J Neurovirol* 14: 229-38 (2008)

88. Bright RA, Cho DS, Rowe T, Katz JM. Mechanisms of pathogenicity of influenza A (H5N1) viruses in mice *Avian Dis* 47: 1131-4 (2003)

89. Nimmerjahn A, Kirchhoff F, Helmchen F. Resting microglial cells are highly dynamic surveillants of brain parenchyma *in vivo Science* 308: 1314-8 (2005)

90. Aloisi F. Immune function of microglia Glia 36: 165-79 (2001)

91. Neher JJ, Brown GC. Neurodegeneration in models of Gram-positive bacterial infections of the central nervous system *Biochem Soc Trans* 35: 1166-7 (2007)

92. Klein M, Obermaier B, Angele B, Pfister HW, Wagner H, Koedel U, Kirschning CJ. Innate immunity to pneumococcal infection of the central nervous system depends on toll-like receptor (TLR) 2 and TLR4 *J Infect Dis* 198: 1028-36 (2008)

93. Klein M, Koedel U, Pfister HW. Oxidative stress in pneumococcal meningitis: a future target for adjunctive therapy? *Prog Neurobiol* 80: 269-80 (2006)

94. Hornig M, Weissenbock H, Horscroft N, Lipkin WI. An infection-based model of neurodevelopmental damage *Proc Natl Acad Sci* U S A 96: 12102-7 (1999)

95. Hornig M, Solbrig M, Horscroft N, Weissenbock H, Lipkin WI. Borna disease virus infection of adult and neonatal rats: models for neuropsychiatric disease *Curr Top Microbiol Immunol* 253: 157-77 (2001)

96. Rubin SA, Sylves P, Vogel M, Pletnikov M, Moran TH, Schwartz GJ, Carbone KM. Borna disease virusinduced hippocampal dentate gyrus damage is associated with spatial learning and memory deficits *Brain Res Bull* 48: 23-30 (1999)

97. Bauer S, Kerr BJ, Patterson PH. The neuropoietic cytokine family in development, plasticity, disease and injury *Nat Rev Neurosci* 8: 221-32 (2007)

98. Shi L, Smith SE, Malkova N, Tse D, Su Y, Patterson PH. Activation of the maternal immune system alters cerebellar development in the offspring *Brain Behav Immun* 23: 116-23 (2009)

99. Shi L, Fatemi SH, Sidwell RW, Patterson PH. Maternal influenza infection causes marked behavioral and pharmacological changes in the offspring *J Neurosci* 23: 297-302 (2003)

100. Lipkin WI, Hornig M. Psychotropic viruses Curr Opin Microbiol 7: 420-5 (2004)

101. Peters CJ. Emerging infections: lessons from the viral hemorrhagic fevers *Trans Am Clin Climatol Assoc* 117: 189-97 (2006)

102. Carballal G, Videla CM, Merani MS. Epidemiology of Argentine hemorrhagic fever *Eur J Epidemiol* 4: 259-74 (1988)

103. Kenyon RH, McKee KT Jr, Zack PM, Rippy MK, Vogel AP, York C, Meegan J, Crabbs C, Peters CJ. Aerosol infection of rhesus macaques with Junin virus *Intervirology* 33: 23-31 (1992)

104. Hornig M, Briese T, Lipkin WI. Borna disease virus J Neurovirol 2003; 9: 259-73.

105. Morales JA, Herzog S, Kompter C, Frese K, Rott R. Axonal transport of Borna disease virus along olfactory pathways in spontaneously and experimentally infected rats *Med Microbiol Immunol* 177: 51-68 (1988)

106. Mori I, Nishiyama Y, Yokochi T, Kimura Y. Olfactory transmission of neurotropic viruses J Neurovirol 11: 129-37 (2005)

107. Mori I, Goshima F, Mizuno T, Imai Y, Kohsaka S, Ito H, Koide N, Yoshida T, Yokochi T, Kimura Y, Nishiyama Y. Axonal injury in experimental herpes simplex encephalitis *Brain Res* 1057: 186-90 (2005)

108. Ando Y, Kitayama H, Kawaguchi Y, Koyanagi Y. Primary target cells of herpes simplex virus type 1 in the hippocampus *Microbes Infect* 10: 1514-1523. (2008)

109. Boivin G, Coulombe Z, Rivest S. Intranasal herpes simplex virus type 2 inoculation causes a profound thymidine kinase dependent cerebral inflammatory response in the mouse hindbrain *Eur J Neurosci* 16: 29-43 (2002)

110. Volmer R, Monnet C, Gonzalez-Dunia D. Borna disease virus blocks potentiation of presynaptic activity through inhibition of protein kinase C signaling *PLoS Pathog* 2: e19. (2006)

111. Hans A, Bajramovic JJ, Syan S, Perret E, Dunia I, Brahic M, Gonzalez-Dunia D. Persistent, noncytolytic infection of neurons by Borna disease virus interferes with ERK 1/2 signaling and abrogates BDNF-induced synaptogenesis *FASEB J* 18: 863-5 (2004)

112. Volmer R, Prat CM, Le Masson G, Garenne A, Gonzalez-Dunia D. Borna disease virus infection impairs synaptic plasticity *J Virol* 81: 8833-7 (2007)

113. Sauder C, Wolfer DP, Lipp HP, Staeheli P, Hausmann J. Learning deficits in mice with persistent Borna disease virus infection of the CNS associated with elevated chemokine expression *Behav Brain Res* 120: 189-201 (2001)

114. Guillot L, Le Goffic R, Bloch S, Escriou N, Akira S, Chignard M, Si-Tahar M. Involvement of toll-like receptor 3 in the immune response of lung epithelial cells to double-stranded RNA and influenza A virus *J Biol Chem* 280: 5571-80 (2005)

115. Sant AJ, Chaves FA, Krafcik FR, Lazarski CA, Menges P, Richards K, Weaver JM. Immunodominance in CD4 T-cell responses: implications for immune responses to influenza virus and for vaccine design *Expert Rev Vaccines* 6: 357-68 (2007)

116. Grayson MH, Holtzman MJ. Emerging role of dendritic cells in respiratory viral infection *J Mol Med* 85: 1057-68 (2007)

117. Mori I, Goshima F, Imai Y, Kohsaka S, Sugiyama T, Yoshida T, Yokochi T, Nishiyama Y, Kimura Y. Olfactory receptor neurons prevent dissemination of neurovirulent influenza A virus into the brain by undergoing virus-induced apoptosis *J Gen Virol* 83: 2109-16 (2002)

118. Peng T, Kotla S, Bumgarner RE, Gustin KE. Human rhinovirus attenuates the type I interferon response by disrupting activation of interferon regulatory factor 3 *J Virol* 80: 5021-31 (2006)

119. Kotla S, Peng T, Bumgarner RE, Gustin KE. Attenuation of the type I interferon response in cells infected with human rhinovirus *Virology* 374: 399-410 (2008)

120. Dreschers S, Dumitru CA, Adams C, Gulbins E. The cold case: are rhinoviruses perfectly adapted pathogens? *Cell Mol Life Sci* 64: 181-91 (2007)

121. Sajjan U, Wang Q, Zhao Y, Gruenert DC, Hershenson MB. Rhinovirus disrupts the barrier function of polarized airway epithelial cells *Am J Respir Crit Care Med* 178: 1271-81 (2008)

122. Foxwell AR, Kyd JM, Cripps AW. Nontypeable Haemophilus influenzae: pathogenesis and prevention *Microbiol Mol Biol Rev* 62: 294-308 (1998)

123. Foxwell AR, Kyd JM, Cripps AW. Kinetics of inflammatory cytokines in the clearance of non-typeable Haemophilus influenzae from the lung *Immunol Cell Biol* 76: 556-9 (1998)

124. Zhang N, Van Zele T, Perez-Novo C, Van Bruaene N, Holtappels G, DeRuyck N, Van Cauwenberge P, Bachert C. Different types of T-effector cells orchestrate mucosal inflammation in chronic sinus disease *J Allergy Clin Immunol* 122: 961-8 (2008) 125. Didierlaurent A, Goulding J, Hussell T. The impact of successive infections on the lung microenvironment *Immunology* 122: 457-65 (2007)

126. Didierlaurent A, Goulding J, Patel S, Snelgrove R, Low L, Bebien M, Lawrence T, van Rijt LS, Lambrecht BN, Sirard JC, Hussell T. Sustained desensitization to bacterial Toll-like receptor ligands after resolution of respiratory influenza infection *J Exp Med* 205: 323-9 (2008)

127. Goulding J, Snelgrove R, Saldana J, Didierlaurent A, Cavanagh M, Gwyer E, Wales J, Wissinger EL, Hussell T. Respiratory infections: do we ever recover? *Proc Am Thorac Soc* 4: 618-25 (2007)

128. Gleich GJ, Kita H. Bronchial asthma: lessons from murine models *Proc Natl Acad Sci* U SA 94: 2101-2 (1997)

129. Galli SJ, Nakae S, Tsai M. Mast cells in the development of adaptive immune responses *Nat Immunol* 6: 135-42 (2005)

130. Finkelman FD. Anaphylaxis: lessons from mouse models *J Allergy Clin Immunol* 120: 506-15; quiz 516-7 (2007)

131. Pawankar R, Lee KH, Nonaka M, Takizawa R. Role of mast cells and basophils in chronic rhinosinusitis *Clin Allergy Immunol* 20: 93-101 (2007)

132. Finnerty K, Choi JE, Lau A, Davis-Gorman G, Diven C, Seaver N, Linak WP, Witten M, McDonagh PF. Instillation of coarse ash particulate matter and lipopolysaccharide produces a systemic inflammatory response in mice *J Toxicol Environ Health A* 70: 1957-66 (2007)

133. Kleinman MT, Araujo JA, Nel A, Sioutas C, Campbell A, Cong PQ, Li H, Bondy SC. Inhaled ultrafine particulate matter affects CNS inflammatory processes and may act via MAP kinase signaling pathways *Toxicol Lett* 178: 127-30 (2008)

134. Campbell A, Oldham M, Becaria A, Bondy SC, Meacher D, Sioutas C, Misra C, Mendez LB, Kleinman M. Particulate matter in polluted air may increase biomarkers of inflammation in mouse brain *Neurotoxicology* 26: 133-40 (2005)

135. Steerenberg PA, Dormans JA, van Doorn CC, Middendorp S, Vos JG, van Loveren H. A pollen model in the rat for testing adjuvant activity of air pollution components *Inhal Toxicol* 11: 1109-22 (1999)

136. Steerenberg PA, Withagen CE, Dormans JA, van Dalen WJ, van Loveren H, Casee FR. Adjuvant activity of various diesel exhaust and ambient particles in two allergic models *J Toxicol Environ Health A* 66: 1421-39 (2003)

137. Tonelli LH, Holmes A, Postolache TT. Intranasal immune challenge induces sex-dependent depressive-like behavior and cytokine expression in the brain *Neuropsychopharmacology* 33: 1038-48 (2008) 138. Tin-Tin-Win-Shwe, Yamamoto S, Ahmed S, Kakeyama M, Kobayashi T, Fujimaki H. Brain cytokine and chemokine mRNA expression in mice induced by intranasal instillation with ultrafine carbon black *Toxicol Lett* 163: 153-60 (2006)

139. Boivin N, Sergerie Y, Rivest S, Boivin G. Effect of pretreatment with toll-like receptor agonists in a mouse model of herpes simplex virus type 1 encephalitis *J Infect Dis* 198: 664-72 (2008)

140. Costa-Pinto FA, Basso AS, Russo M. Role of mast cell degranulation in the neural correlates of the immediate allergic reaction in a murine model of asthma *Brain Behav Immun* 21: 783-90 (2007)

141. Costa-Pinto FA, Basso AS, De Sa-Rocha LC, Britto LR, Russo M, Palermo-Neto J. Neural correlates of IgEmediated allergy *Ann N Y Acad Sci* 1088: 116-31 (2006)

142. Costa-Pinto FA, Basso AS, Britto LR, Malucelli BE, Russo M. Avoidance behavior and neural correlates of allergen exposure in a murine model of asthma *Brain Behav Immun* 19: 52-60 (2005)

143. Rosenkranz MA, Busse WW, Johnstone T, Swenson CA, Crisafi GM, Jackson MM, Bosch JA, Sheridan JF, Davidson RJ. Neural circuitry underlying the interaction between emotion and asthma symptom exacerbation *Proc Natl Acad Sci* U S A 102: 13319-24 (2005)

144. Tonelli LH, Katz M, Kovacsis C, Gould TD, Joppy B, Hoshino A, Hoffman GE, Komarow H, Postolache TT. Allergic rhinitis induces anxiety-like behavior and reduced social interaction in rodents. *Brain Behav Immun* (in press)

145. Pewter SM, Williams WH, Haslam C, Kay JM. Neuropsychological and psychiatric profiles in acute encephalitis in adults *Neuropsychol Rehabil* 17: 478-505 (2007)

146. Fatemi SH, Earle J, Kanodia R, Kist D, Emamian ES, Patterson PH, Shi L, Sidwell R. Prenatal viral infection leads to pyramidal cell atrophy and macrocephaly in adulthood: implications for genesis of autism and schizophrenia *Cell Mol Neurobiol* 22: 25-33 (2002)

147. Danaher RJ, McGarrell BS, Stromberg AJ, Miller CS. Herpes simplex virus type 1 modulates cellular gene expression during quiescent infection of neuronal cells *Arch Virol* 153: 1335-45 (2008)

148. Ovanesov MV, Moldovan K, Smith K, Vogel MW, Pletnikov MV. Persistent Borna Disease Virus (BDV) infection activates microglia prior to a detectable loss of granule cells in the hippocampus *J Neuroinflammation* 5: 16 (2008)

149. Dantzer R, O'Connor JC, Freund GG, Johnson RW, Kelley KW. From inflammation to sickness and depression: when the immune system subjugates the brain *Nat Rev Neurosci* 9: 46-56 (2008)

150. Anisman H, Gibb J, Hayley S. Influence of continuous infusion of interleukin-1beta on depression-related processes in mice: corticosterone, circulating cytokines, brain monoamines, and cytokine mRNA expression *Psychopharmacology (Berl)* 199: 231-44 (2008)

151. Anisman H, Merali Z, Hayley S. Neurotransmitter, peptide and cytokine processes in relation to depressive disorder: comorbidity between depression and neurodegenerative disorders *Prog Neurobiol* 85: 1-74 (2008)

152. Tonelli LH, Postolache TT. Tumor necrosis factor alpha, interleukin-1 beta, interleukin-6 and major histocompatibility complex molecules in the normal brain and after peripheral immune challenge *Neurol Res* 27: 679-84 (2005)

153. Goshen I, Kreisel T, Ben-Menachem-Zidon O, Licht T, Weidenfeld J, Ben-Hur T, Yirmiya R. Brain interleukin-1 mediates chronic stress-induced depression in mice via adrenocortical activation and hippocampal neurogenesis suppression *Mol Psychiatry* 13: 717-28 (2008)

154. Goshen I, Yirmiya R, Iverfeldt K, Weidenfeld J. The role of endogenous interleukin-1 in stress-induced adrenal activation and adrenalectomy-induced adrenocorticotropic hormone hypersecretion *Endocrinology* 144: 4453-8 (2003)

155. Tonelli LH, Postolache TT, Sternberg EM. Inflammatory genes and neural activity: involvement of immune genes in synaptic function and behavior *Front Biosci* 10: 675-80 (2005)

156. Tonelli LH, Maeda S, Rapp KL, Sternberg EM. Differential induction of interleukin-I beta mRNA in the brain parenchyma of Lewis and Fischer rats after peripheral injection of lipopolysaccharides *J Neuroimmunol* 140: 126-36 (2003)

157. Foster JA, Quan N, Stern EL, Kristensson K, Herkenham M. Induced neuronal expression of class I major histocompatibility complex mRNA in acute and chronic inflammation models *J Neuroimmunol* 131: 83-91 (2002)

158. Quan N, Stern EL, Whiteside MB, Herkenham M. Induction of pro-inflammatory cytokine mRNAs in the brain after peripheral injection of subseptic doses of lipopolysaccharide in the rat *J Neuroimmunol* 93: 72-80 (1999)

159. Doty RL. The olfactory vector hypothesis of neurodegenerative disease: is it viable? *Ann Neurol* 63: 7-15. (2008)

160. Itzhaki RF, Cosby SL, Wozniak MA. Herpes simplex virus type 1 and Alzheimer's disease: the autophagy connection *J Neurovirol* 14: 1-4. (2008)

161. Ball MJ. The essential lesion of Alzheimer disease: a surprise in retrospect *J Alzheimers Dis* 9: 29-33 (2006)

162. Letenneur L, Peres K, Fleury H, Garrigue I, Barberger-Gateau P, Helmer C, Orgogozo JM, Gauthier S, Dartigues JF. Seropositivity to herpes simplex virus antibodies and risk of Alzheimer's disease: a population-based cohort study *PLoS ONE*; 3: e3637 (2008)

163. Ascherio A, Munger K. Epidemiology of multiple sclerosis: from risk factors to prevention *Semin Neurol* 28: 17-28 (2008)

164. Tucker WG, Andrew Paskauskas R. The MSMV hypothesis: measles virus and multiple sclerosis, etiology and treatment *Med Hypotheses* 71: 682-9 (2008)

165. Elbaz A, Tranchant C. Epidemiologic studies of environmental exposures in Parkinson's disease *J Neurol Sci* 262: 37-44 (2007)

166. Nagatsu T, Mogi M, Ichinose H, Togari A. Cytokines in Parkinson's disease *J Neural Transm Suppl* 143-51 (2000)

167. Pugliatti M, Harbo HF, Holmoy T, Kampman MT, Myhr KM, Riise T, Wolfson C. Environmental risk factors in multiple sclerosis *Acta Neurol Scand Suppl* 188: 34-40 (2008)

168. Posnett DN. Herpesviruses and autoimmunity Curr Opin Investig Drugs 9: 505-14. (2008)

169. Alvarez-Lafuente R, Garcia-Montojo M, De Las Heras V, Dominguez-Mozo MI, Bartolome M, Benito-Martin MS, Arroyo R. Herpesviruses and human endogenous retroviral sequences in the cerebrospinal fluid of multiple sclerosis patients *Mult Scler* 14: 595-601 (2008)

170. Soldan SS, Berti R, Salem N, Secchiero P, Flamand L, Calabresi PA, Brennan MB, Maloni HW, McFarland HF, Lin HC, Patnaik M, Jacobson S. Association of human herpes virus 6 (HHV-6) with multiple sclerosis: increased IgM response to HHV-6 early antigen and detection of serum HHV-6 DNA *Nat Med* 3: 1394-7 (1997)

171. Berti R, Brennan MB, Soldan SS, Ohayon JM, Casareto L, McFarland HF, Jacobson S. Increased detection of serum HHV-6 DNA sequences during multiple sclerosis (MS) exacerbations and correlation with parameters of MS disease progression *J Neurovirol* 8: 250-6 (2002)

172. Jarius S, Eichhorn P, Jacobi C, Wildemann B, Wick M, Voltz R. The intrathecal, polyspecific antiviral immune response: Specific for MS or a general marker of CNS autoimmunity? *J Neurol Sci* (2008)

173. Ascherio A, Munger KL, Lennette ET, Spiegelman D, Hernan MA, Olek MJ, Hankinson SE, Hunter DJ. Epstein-Barr virus antibodies and risk of multiple sclerosis: a prospective study *JAMA* 286: 3083-8 (2001)

174. Jilek S, Schluep M, Meylan P, Vingerhoets F, Guignard L, Monney A, Kleeberg J, Le Goff G, Pantaleo G, Du Pasquier RA. Strong EBV-specific CD8+ T-cell

response in patients with early multiple sclerosis *Brain* 131: 1712-21 (2008)

175. Lindsey J, Hatfield L, Crawford M, Patel S. Quantitative PCR for Epstein-Barr virus DNA and RNA in multiple sclerosis *Mult Scler* (2008)

176. Rodriguez-Violante M, Ordonez G, Bermudez JR, Sotelo J, Corona T. Association of a history of varicella virus infection with multiple sclerosis *Clin Neurol Neurosurg* 111: 54-6 (2009)

177. Hawkes C. Olfaction in neurodegenerative disorder *Mov Disord* 18: 364-72 (2003)

178. Hawkes C. Olfaction in neurodegenerative disorder *Adv Otorhinolaryngol* 63: 133-51 (2006)

179. Mesholam RI, Moberg PJ, Mahr RN, Doty RL. Olfaction in neurodegenerative disease: a meta-analysis of olfactory functioning in Alzheimer's and Parkinson's diseases *Arch Neurol* 55: 84-90 (1998)

180. Wozniak MA, Mee AP, Itzhaki RF. Herpes simplex virus type 1 DNA is located within Alzheimer's disease amyloid plaques *J Pathol* 217: 131-8 (2009)

181. Itzhaki RF, Wozniak MA. Herpes simplex virus type 1 in Alzheimer's disease: the enemy within *J Alzheimers Dis* 13: 393-405 (2008)

182. Wozniak MA, Itzhaki RF, Shipley SJ, Dobson CB. Herpes simplex virus infection causes cellular betaamyloid accumulation and secretase upregulation *Neurosci Lett* 429: 95-100 (2007)

183. Wozniak MA, Shipley SJ, Combrinck M, Wilcock GK, Itzhaki RF. Productive herpes simplex virus in brain of elderly normal subjects and Alzheimer's disease patients *J Med Virol* 75: 300-6 (2005)

184. Mori I, Goshima F, Watanabe D, Ito H, Koide N, Yoshida T, Liu B, Kimura Y, Yokochi T, Nishiyama Y. Herpes simplex virus US3 protein kinase regulates virus-induced apoptosis in olfactory and vomeronasal chemosensory neurons *in vivo Microbes Infect* 8: 1806-12 (2006)

185. Hawkes CH, Del Tredici K, Braak H. Parkinson's disease: a dual-hit hypothesis *Neuropathol Appl Neurobiol* 33: 599-614 (2007)

186. Rojo AI, Montero C, Salazar M, Close RM, Fernandez-Ruiz J, Sanchez-Gonzalez MA, de Sagarra MR, Jackson-Lewis V, Cavada C, Cuadrado A. Persistent penetration of MPTP through the nasal route induces Parkinson's disease in mice *Eur J Neurosci* 24: 1874-84 (2006)

187. Friedman JH, Amick MM. Rhinorrhea is increased in Parkinson's disease *Mov Disord* 23: 452-4 (2008) 188. Meredith GE, Sonsalla PK, Chesselet MF. Animal models of Parkinson's disease progression *Acta Neuropathol* 115: 385-98 (2008)

189. Nagatsu T, Mogi M, Ichinose H, Togari A. Changes in cytokines and neurotrophins in Parkinson's disease *J Neural Transm Suppl* 277-90 (2000)

190. Bower JH, Maraganore DM, Peterson BJ, Ahlskog JE, Rocca WA. Immunologic diseases, anti-inflammatory drugs, and Parkinson disease: a case-control study *Neurology* 67: 494-6 (2006)

191. McCoy MK, Ruhn KA, Martinez TN, McAlpine FE, Blesch A, Tansey MG. Intranigral lentiviral delivery of dominant-negative TNF attenuates neurodegeneration and behavioral deficits in hemiparkinsonian rats *Mol Ther* 16: 1572-9 (2008)

192. Brochard V, Combadiere B, Prigent A, Laouar Y, Perrin A, Beray-Berthat V, Bonduelle O, Alvarez-Fischer D, Callebert J, Launay JM, Duyckaerts C, Flavell RA, Hirsch EC, Hunot S. Infiltration of CD4+ lymphocytes into the brain contributes to neurodegeneration in a mouse model of Parkinson disease *J Clin Invest* 119: 182-92 (2009)

193. Hayley S, Poulter MO, Merali Z, Anisman H. The pathogenesis of clinical depression: stressor- and cytokine-induced alterations of neuroplasticity *Neuroscience* 135: 659-78 (2005)

194. Pollak Y, Yirmiya R. Cytokine-induced changes in mood and behaviour: implications for 'depression due to a general medical condition', immunotherapy and antidepressive treatment *Int J Neuropsychopharmacol* 5: 389-99 (2002)

195. Raison CL, Capuron L, Miller AH. Cytokines sing the blues: inflammation and the pathogenesis of depression *Trends Immunol* 27: 24-31 (2006)

196. Yirmiya R. Depression in medical illness: the role of the immune system *West J Med* 173: 333-6 (2000)

197. Raison CL, Broadwell SD, Borisov AS, Manatunga AK, Capuron L, Woolwine BJ, Jacobson IM, Nemeroff CB, Miller AH. Depressive symptoms and viral clearance in patients receiving interferon-alpha and ribavirin for hepatitis C *Brain Behav Immun* 19: 23-7 (2005)

198. Castanon N, Leonard BE, Neveu PJ, Yirmiya R. Effects of antidepressants on cytokine production and actions *Brain Behav Immun* 16: 569-74 (2002)

199. Yirmiya R, Pollak Y, Morag M, Reichenberg A, Barak O, Avitsur R, Shavit Y, Ovadia H, Weidenfeld J, Morag A, Newman ME, Pollmacher T. Illness, cytokines, and depression *Ann N Y Acad Sci* 917: 478-87 (2000)

200. Belmaker RH, Agam G. Major depressive disorder *N Engl J Med* 358: 55-68 (2008)

201. Drevets WC, Price JL, Furey ML. Brain structural and functional abnormalities in mood disorders: implications for neurocircuitry models of depression *Brain Struct Funct* 213: 93-118 (2008)

202. Drevets WC, Savitz J, Trimble M. The subgenual anterior cingulate cortex in mood disorders *CNS Spectr* 13: 663-81 (2008)

203. Drevets WC. Orbitofrontal cortex function and structure in depression Ann N Y Acad Sci 1121: 499-527 (2007)

204. Postolache TT, Lapidus M, Sander ER, Langenberg P, Hamilton RG, Soriano JJ, McDonald JS, Furst N, Bai J, Scrandis DA, Cabassa JA, Stiller JW, Balis T, Guzman A, Togias A, Tonelli LH. Changes in allergy symptoms and depression scores are positively correlated in patients with recurrent mood disorders exposed to seasonal peaks in aeroallergens *ScientificWorldJournal* 7: 1968-77 (2007)

205. Guzman A, Tonelli LH, Roberts D, Stiller JW, Jackson MA, Soriano JJ, Yousufi S, Rohan KJ, Komarow H, Postolache TT. Mood-worsening with high-pollen-counts and seasonality: a preliminary report *J Affect Disord* 101: 269-74 (2007)

206. Goodwin RD, Buka SL. Childhood respiratory disease and the risk of anxiety disorder and major depression in adulthood *Arch Pediatr Adolesc Med* 162: 774-80 (2008)

207. Roy-Byrne PP, Davidson KW, Kessler RC, Asmundson GJ, Goodwin RD, Kubzansky L, Lydiard RB, Massie MJ, Katon W, Laden SK, Stein MB. Anxiety disorders and comorbid medical illness *Gen Hosp Psychiatry* 30: 208-25 (2008)

208. Addolorato G, Ancona C, Capristo E, Graziosetto R, Di Rienzo L, Maurizi M, Gasbarrini G. State and trait anxiety in women affected by allergic and vasomotor rhinitis *J Psychosom Res* 46: 283-9 (1999)

209. Buske-Kirschbaum A, Ebrecht M, Kern S, Gierens A, Hellhammer DH. Personality characteristics in chronic and non-chronic allergic conditions *Brain Behav Immun* 22: 762-8 (2008)

210. Barone S, Bacon SL, Campbell TS, Labrecque M, Ditto B, Lavoie KL. The association between anxiety sensitivity and atopy in adult asthmatics *J Behav Med* 31: 331-9 (2008)

211. Deshmukh VM, Toelle BG, Usherwood T, O'Grady B, Jenkins CR. The association of comorbid anxiety and depression with asthma-related quality of life and symptom perception in adults *Respirology* 13: 695-702 (2008)

212. Timonen M, Jokelainen J, Hakko H, Silvennoinen-Kassinen S, Meyer-Rochow VB, Herva A, Rasanen P. Atopy and depression: results from the Northern Finland 1966 Birth Cohort Study Mol Psychiatry 8: 738-44 (2003)

213. Goodwin RD. Self-reported hay fever and panic attacks in the community *Ann Allergy Asthma Immunol* 88: 556-9 (2002)

214. Rook GA. The hygiene hypothesis and the increasing prevalence of chronic inflammatory disorders *Trans R Soc Trop Med Hyg* 101: 1072-4 (2007)

215. Bach JF. Infections and autoimmune diseases J Autoimmun 25 Suppl: 74-80 (2005)

216. Rook GA, Lowry CA. The hygiene hypothesis and psychiatric disorders *Trends Immunol* 29: 150-8 (2008)

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