

Normal distribution of alkaline phosphatase levels during pregnancy

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Background: Previous small-scale studies have reported alkaline phosphatase (ALP) distribution throughout gestation. This retrospective, observational analysis aimed to establish a large-scale, week-specific nomogram of total ALP during pregnancy, and to address physiological parameters that may impact normal ALP secretion throughout pregnancy. Methods: We analyzed ALP levels during pregnancy among 5285 healthy women from various ethnicities, with uncomplicated, term, singleton gestations, who delivered in a single tertiary medical center, from August 2007 to December 2012. Results: We found that normal gestational ALP is significantly elevated during pregnancy compared to a non-pregnant state (132.2 \pm 3.0 IU/Lvs. 116.7 \pm 2.3 IU/L, p < 0.001), and during the third and second trimesters compared to the first trimester (166.4 \pm 1.66, 123.7 \pm 2.4 and 95.7 \pm 1.6 IU/L, respectively; p < 0.001). ALP levels remained unaffected by the modifiers we explored: neonatal sex, neonatal birthweight and maternal age. Conclusion: ALP levels during pregnancy are higher than in the non-pregnant state, gradually increase from the end of the first trimester to term, peaking just prior to delivery. The nomogram presented here can serve as the basis for comparison between a normal population and those with various pregnancy complications, to determine how ALP is associated with adverse pregnancy outcomes.

Keywords

Alkaline phosphatase; Distribution; Female; Nomogram; Pregnancy

1. Introduction

Alkaline phosphatase (ALP) is an enzyme that catalyzes the hydrolysis of organic phosphate-esters in the extracellular space [1]. Several isoenzymes are produced by the hepatobiliary system, skeletal system, gastrointestinal tract, kidneys and placenta [2]. The placental isoenzyme, a heat stable homo-dimeric protein [3], is produced by syncytiotrophoblasts. It is the primary contributor to the corresponding rise in total ALP during gestation [4], along with elevated bone-related ALP, secondary to amplified bone formation [5, 6]. These two studies were the first to validate that

ALP gradually increases during gestation, but failed to reveal associations with adverse outcomes [5,6]. Nevertheless, several case reports demonstrated associations between elevated ALP with hypertensive disorders [7], gestational diabetes [7–9], preterm birth [10], placental abruption and stillbirth [11]; as well as in uncomplicated pregnancies [12, 13]. Additionally, elevated mid-trimester ALP, may predict preterm birth [14, 15] and low birth weight [16–18].

It was also suggested to be associated with arterial stiffness as a predictor of cardiovascular disease in non-pregnant individuals [19] and multiparous women [20].

Previous ALP nomograms demonstrated a gradual increase in total ALP, starting early in the second trimester, and peaking in the third trimester just prior to delivery, with maximum levels approximately 2–3 times higher than nonpregnant references [21–27]. Other than their small size, these studies were limited by lack of consideration of possible effect modifiers, such as body mass index, mode of conception, neonatal sex and the foe to-placental mass.

In our clinical practice, total ALP is often measured during biochemistry serum analysis, but is seldom used as a potential biomarker for adverse pregnancy outcomes. As maternal physiological adaptation confers substantial alterations in several laboratory tests, it is imperative to have high-quality pregnancy-specific distributions of ALP during pregnancy, on which to base pathological deviations. Therefore, we aimed to establish a large-scale ALP nomogram, and to address the role of possible physiological modifiers, and of total ALP levels during gestation — all as a foundation for further research, to explore associations of divergent ALP with adverse pregnancy outcomes.

2. Materials and methods

This study was a retrospective analysis of ALP values during pregnancy, among healthy women from a variety of ethnicities, with uncomplicated singleton gestations, who deliv-

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ered in a single tertiary medical center, from August 2007 to December 2012.

2.1 Study population

Eligibility was limited to healthy women, at least 18 years of age, with a singleton gestation, delivering a liveborn, non-anomalous, appropriately grown, term fetus, with at least one total ALP serum measurement during pregnancy.

Only women delivering at term were included, excluding deliveries before 37 + 0 and after 41 + 6 weeks. Gestational age at delivery was determined based on maternally-reported last menstrual period and confirmed by first trimester crownrump length sonographic measurement [28]. We also excluded women with pre-existing chronic diseases, such as essential hypertension, type 1 or 2 diabetes mellitus, thrombophilia, hypo- or hyperthyroidism, inflammatory bowel disease and any other liver, kidney and/or heart disease - all were identified according to ICD-9 diagnosis codes in the maternal electronic health record. Women with the following gestational complications were also excluded: placental complications (abruption, placenta accreta or placenta previa); hypertensive disorders during pregnancy, classified according to American College of Obstetrics and Gynecology guidelines [29]; gestational diabetes mellitus, defined according to Carpenter and Coustan's criteria [30]; oligohydramnios, defined as amniotic fluid index <5 cm; polyhydramnios, defined as amniotic fluid index >25 cm [28]; small or large for gestational age, defined as birthweight below the 10th or above the 90th percentile, respectively, according to nationally validated, sex- and week-specific growth curves [31].

2.2 Data collection

Data were retrieved from maternal and neonatal electronic medical records, derived from the comprehensive computerized databases of the Maternal-Fetal Unit, Obstetrical Emergency Room, delivery ward and post-partum hospitalization. Collected data included maternal demographics; medical and obstetrical variables (gravidity, parity, coexisting maternal chronic diseases and gestational-obstetrical complications); labor and delivery outcomes (gestational age at delivery, birth weight, mode of delivery and fetal sex); as well as the results of laboratory studies, specifically alkaline phosphatase, as collected periodically throughout gestation and up to one year before conception.

2.3 Alkaline phosphate measurements

Blood samples were collected for each woman during routine pregnancy follow-up, from the last menstrual period to delivery. As biochemistry analysis is not routinely collected at fixed intervals during prenatal follow-up, the exact timing and number of ALP samples per woman varied and they were available according to physician discretion.

We documented a minimum of one ALP measurement per participant and up to three measurements - one per trimester; first trimester (0 to 13 + 6 gestational weeks), second trimester (14 to 27 + 6 weeks) and third trimester (28 to 41 + 6 weeks), as available. If more than one ALP mea-

surement was obtained in a trimester, the first was used for analysis.

Also, one pre-conception measurement was collected, the closest available prior to the date of conception.

All blood samples were collected in serum separator gel tubes (BD, Mississauga, Ontario, CA) and ALP measurements were evaluated at the central laboratory of Clalit Health Services, under standard laboratory protocols, utilizing the same technique and equipment, in accordance with the recommendations of the International Federation for Clinical Chemistry [32]. Serum samples were processed by an Olympus AU5430 analyzer (Beckman Coulter, Atlanta, GA, USA), using manufacturer's standardized reagents (OSR6204). ALP results were reported as units/liter (U/L).

Per manufacturers specification, the reference interval for reporting the result (in adults >17 years) is 30–120 U/L, and test being linear within an enzyme activity range of 5–1500 U/L.

Table 1. Demographic, clinical and obstetrical characteristics of the study group (included population) and the women with unavailable ALP (excluded population).

Parameter	Included population	p-value		
	n = 5285	n = 23,292	p varue	
Maternal age, years	ernal age, years 32.4 ± 5.1		0.199	
Gravidity	2.7 ± 1.8	2.7 ± 1.7	0.025	
Parity	2.2 ± 1.3	2.3 ± 1.3	0.001	
Nulliparous	1906 (36.1%)	7409 (31.8%)	< 0.001	
Fetal sex				
Male	2674 (50.6%)	11,883 (51.0%)	0.580	
Female	2611 (49.4%)	11,409 (49.0%)	0.580	
Mode of delivery				
Vaginal	3990 (75.5%)	17,940 (77.0%)		
Assisted	477 (9.0%)	1866 (8.0%)	0.024	
Cesarean	818 (15.5%)	3486 (15.0%)		
Neonatal birthweight, grams	3213 ± 313	3225 ± 309	0.015	

Data presented as mean \pm standard error for continuous variables and n (%) for categorical variables.

Coefficients of variability (%CV) are 2.1, 1.3, 1.3 (intra assay) and 4.2, 2.3, 1.8 (inter assay) at 38, 360, 1254 U/L, respectively.

2.4 Data analysis

Statistical analysis was performed using the SPSS Software (version 21.0, IBM Corp., Chicago, IL, USA). Data were calculated and presented week-by-week and per-trimester of pregnancy, providing means (standard errors), medians (ranges) and percentiles. Data were presented as mean \pm standard error for continuous variables, evaluated using dependent of independent t-tests, as appropriate; while categorical variables were presented as number and percentage and evaluated using Pearson's chi-square test.

1394 Volume 48, Number 6, 2021

Table 2. Alkaline phosphatase (IU/L) measurements per gestational week, across gestation.

Week	Week n Mean ± SE		Range	Percentile								
WCCK	11	Wican ± 0L	Range	2.5th	5th	10th	25th	50th	75th	90th	95th	97.5th
4	231	84.21 ± 4.1	29.0-415.8	32.00	37.60	42.00	50.00	63.00	83.00	164.92	230.00	289.38
5	300	83.10 ± 3.4	27.0-425.7	35.53	40.00	43.00	51.25	63.00	81.00	179.21	231.48	259.74
6	325	$\textbf{75.08} \pm \textbf{3.1}$	23.0-433.0	32.15	36.30	40.00	47.00	57.00	72.00	150.48	218.80	246.60
7	306	$\textbf{81.11} \pm \textbf{4.3}$	26.0-779.3	32.00	34.35	39.00	45.00	57.00	74.25	183.78	247.88	290.30
8	267	83.05 ± 4.3	20.0-470.8	32.70	36.00	39.00	47.00	59.00	79.00	192.60	245.52	295.41
9	251	$\textbf{97.74} \pm \textbf{5.9}$	27.0-794.0	33.00	35.00	40.00	47.00	62.00	93.00	219.80	272.32	334.55
10	224	103.23 ± 5.7	24.0-544.7	34.00	36.25	40.75	49.00	64.00	144.50	216.85	248.43	346.10
11	190	119.26 ± 7.5	31.0-804.3	33.00	35.55	41.10	51.75	68.00	182.23	261.99	314.34	372.03
12	138	105.52 ± 8.3	21.0-766.8	33.00	37.90	41.00	50.00	60.50	144.05	220.11	289.44	367.49
13	116	115.20 ± 9.2	32.0-516.4	33.93	37.85	40.70	49.25	70.00	154.38	257.25	334.69	394.33
14	128	115.92 ± 8.6	27.0-533.1	36.00	37.00	41.90	49.25	68.50	172.5	255.21	297.84	398.30
15	133	116.74 ± 7.3	24.3-418.5	36.70	40.00	43.40	53.50	72.00	178.15	235.58	286.54	328.65
16	140	115.95 ± 8.4	33.0-658.0	35.05	41.05	47.00	53.00	68.50	161.43	269.70	322.06	368.85
17	137	125.17 ± 7.9	32.0-418.8	39.35	41.90	47.00	53.00	75.00	196.15	248.88	331.67	373.70
18	133	127.23 ± 7.7	32.0-402.2	37.05	39.70	44.00	57.00	78.00	194.45	256.16	302.89	326.27
19	129	125.76 ± 7.8	36.0-411.0	39.25	42.50	49.00	56.00	77.00	185.85	269.60	299.35	338.58
20	122	134.06 ± 9.4	33.0-545.0	36.23	44.15	46.00	56.00	89.00	191.73	278.40	346.43	438.89
21	132	126.72 ± 8.2	4.0-520.1	40.00	44.65	48.00	58.25	78.00	176.08	254.29	317.83	387.22
22	117	138.53 ± 10.0	14.0-498.0	37.70	42.90	50.80	59.00	79.00	196.35	296.64	391.49	422.40
23	171	136.77 ± 7.2	6.0-476.8	34.30	42.20	47.20	60.00	94.00	210.8	260.18	320.20	374.00
24	110	146.58 ± 10.6	41.0-541.0	46.88	52.10	56.10	64.00	105.60	187.15	294.16	397.24	510.07
25	102	139.10 ± 8.6	42.0-410.6	47.45	52.00	56.30	65.00	100.00	200.18	267.12	293.40	370.64
26	91	134.96 ± 8.9	4.0-454.0	43.60	48.20	59.00	72.00	103.00	189.00	241.22	306.20	397.53
27	102	135.77 ± 8.2	39.0-420.1	46.88	53.15	62.00	71.75	105.50	193.70	249.44	283.85	383.11
28	113	143.09 ± 7.4	42.0-367.0	49.55	54.50	65.00	78.50	111.00	193.95	257.50	302.94	348.10
29	108	166.88 ± 9.6	44.0-493.0	57.18	62.00	72.90	89.50	135.00	225.10	316.10	377.79	443.81
30	124	156.32 ± 7.91	59.0-690.8	62.25	66.75	74.00	96.00	132.55	195.90	254.30	319.50	376.80
31	139	154.50 ± 6.0	49.0-356.3	63.30	69.00	81.00	98.00	141.00	195.00	255.60	295.20	339.50
32	129	162.47 ± 6.2	52.0-464.0	59.75	77.00	88.00	111.50	147.00	211.35	248.70	282.00	359.05
33	146	163.85 ± 7.3	62.0-655.1	65.68	79.70	87.40	104.08	134.00	199.20	289.68	320.96	438.63
34	159	163.81 ± 5.59	17.0-495.8	72.00	79.00	87.00	119.00	152.00	192.00	244.00	291.90	362.60
35	157	178.80 ± 5.8	77.0-525.0	84.00	101.90	109.00	129.50	160.30	212.20	266.40	322.02	379.35
36	207	177.7 ± 5.8	77.0-702.0	84.00	90.80	100.00	126.00	158.00	208.00	273.48	335.08	354.80
37	249	178.79 ± 4.6	43.0–657.0	80.55	90.50	104.00	130.00	172.70	210.00	249.00	315.00	360.78
38	243	182.07 ± 5.3	75.0–765.0	91.30	101.00	112.00	134.00	165.70	201.00	259.04	331.16	415.69
39	198	186.00 ± 5.23	64.0–704.0	76.95	99.00	116.99	135.75	170.00	221.23	271.55	313.50	354.15
40	94	192.98 ± 8.2	44.0–551.0	72.55	103.50	113.00	142.00	189.00	229.80	261.00	350.98	451.29
41	3	204.30 ± 0.2	180.7-245.6	180.70	180.70	180.70	180.70	186.60	245.60	245.60	245.60	245.60
Total	6610	128.92 ± 91.5	130 2.3.0	100.70	100.70	1000	100., 0	100.00	2.0.00	2.0.00	2.0.00	2 .5.00

Differences in means ALP values according to fetal sex (male vs. female), birthweight (below or above 2500 g, as a reflection of the feto-placental mass) were also calculated and compared. ALP values during pregnancy and in a non-pregnant state were compared for a subgroup of women with available ALP values. For all comparisons, a p-value < 0.05 was considered statistically significant.

3. Results

Overall, 44,153 women gave birth to singleton, non-anomalous, liveborn neonates at our center during the study period. Among them, 8718 had at least one available ALP

measurement during pregnancy. We excluded women with any pre-existing comorbidity (n = 546), preterm or post-term delivery (n = 803), large or small for gestational age (n = 1553), hypertensive disorders (n = 296), gestational diabetes (n = 474), oligo- or polyhydramnios (n = 575), placental complications (n = 191) and chorioamnionitis (n = 4). Additionally, 20 outlier ALP values, all above 1000, were excluded. Accordingly, 5285 women, with 6610 ALP measurements were included.

Demographic, clinical and obstetrical characteristics of the study group, are presented in Table 1, and compared to the population without an available ALP, meeting the same in-

Volume 48, Number 6, 2021 1395

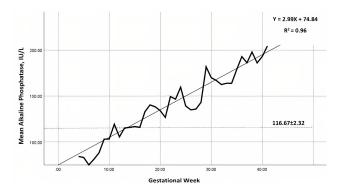


Fig. 1. Nomogram of mean alkaline phosphatase (IU/L) levels, throughout gestation (———), per gestational week. Linear regression line is presented (———), as well as the reference (- - -) for non-pregnant women in the study population ($116.7 \pm 2.3 \text{ IU/L}$).

clusion and exclusion criteria. Although there were some statistically significant differences between the two populations in gravidity (2.7 \pm 1.8 vs. 2.7 \pm 1.7, p = 0.025), parity (2.2 \pm 1.3 vs. 2.3 \pm 1.3, p < 0.001), nulliparity (36.1% vs. 31.8%, p < 0.0001), birthweight (3213 \pm 313 vs. 3225 \pm 309 grams, p = 0.015) and mode of delivery; they were not clinically relevant.

The distribution of ALP measurements throughout gestation is presented in Table 2, as a nomogram with mean, standard deviation, minimum and maximum range, as well as 2.5th–97.5th percentiles per gestational week. The means and standard errors according to possible modifiers, are presented in Table 3. The mean ALP distribution is graphically presented in Fig. 1, per gestational week and the median, interquartile range and minimum-maximum values, in Fig. 2, per trimester.

The association of possible physiological modifiers to ALP levels are presented in Table 3. ALP is significantly elevated during pregnancy compared to a non-pregnant state (132.2 \pm 3.0 vs. 116.7 \pm 2.3 IU/L, p < 0.001), and during the third and second trimesters compared to the first trimester (166.4 \pm 1.66, 123.7 \pm 2.4 and 95.7 \pm 1.6 IU/L, p < 0.001). ALP levels remained unaffected by the modifiers we explored: neonatal sex, neonatal birthweight and maternal age.

4. Discussion

We constructed an ALP nomogram among healthy women during the course of an uncomplicated pregnancy. Our main results indicate that ALP gradually increases during pregnancy, starting from early gestation and that other than gestational week, physiological parameters of maternal age, fetal sex and neonatal birthweight, did not affect the levels.

In accordance with our results, previous studies reported similar trends for both total and heat stable ALP, throughout pregnancy [21–27]. In a longitudinal study among 52 women, Larsson *et al.* [24] evaluated 25 common biochemical indices, including ALP. Similar to our results, they demonstrated rising ALP levels through gestation. However, their cohort was smaller and less homogenous, included

 $Table \ 3. \ Possible \ modifiers \ affecting \ alkaline \ phosphatase$

levels.					
Modifiers	n	ALP (U/L)	<i>p</i> -value		
Pregnancy status [†]					
Pregnant	2014	$\textbf{132.19} \pm \textbf{3.01}$	<0.001		
Non-pregnant	2014	116.67 ± 2.32	< 0.001		
Pregnancy trimester					
First	2540	95.69 ± 1.64			
Second	1596	123.74 ± 2.40	< 0.001		
Third	2474	166.39 ± 1.66			
Neonatal sex					
Males	2684	136.03 ± 2.45	0.510		
Female	2620	133.87 ± 2.16	0.510		
Neonatal birthweight [‡]					
>2500	22	161.84 ± 28.03	0.707		
<2500	2452	166.42 ± 1.66	0.797		
Maternal age					
<25	320	145.79 ± 6.56			
25130	3178	$\textbf{135.59} \pm \textbf{2.05}$	0.130		
>35	1795	131.67 ± 2.99			
Parity					
Nulliparous	3393	136.40 ± 2.21	0.211		
Multiparous	1911	132.41 ± 2.30	0.211		

Data presented as mean \pm standard error.

preterm deliveries and small for gestational age newborns, as well as mothers who were smokers and used medication. Abbassi-Ghanavati *et al.* [25] established a normal reference range during pregnancy for multiple blood components, derived from a comprehensive literature review. Specifically, for ALP, they reported the upper and lower limits during a non-pregnant state and per trimester, based on 5 prior publications [22–24, 27, 33], again demonstrating gestational rise of ALP. Selected comparisons of ALP levels between the available results of two studies [24, 25] and our own, are presented in Table 4 (Ref. [23, 24]).

Table 4. Selected comparisons, according to data availability between the current study and published nomograms

(2.5–97.5th percentile) for ALP (IU/L).

(2.5-77.5th percentile) for AET (107E).						
Timing of measurement	Current study	Larsson [23]	Abbassi-Ghanavati [24]			
7–17 weeks	33.0-334.5	34.8-79.8	-			
17-24 weeks	39.0-374.3	39.0-105	-			
24-28 weeks	47.2-388.9	46.2-115.2	-			
28-31 weeks	57.3-376.5	52.8-118.8	-			
31-34 weeks	65.0-353.7	66.6-177.6	-			
34-38 weeks	80.0-360.4	87.6-228.6	-			
First trimester	33.0-315.3	-	17-88			
Second trimester	36.0-366.6	-	25–126			
Third trimester	58.0-368.0	-	38-229			

1396 Volume 48, Number 6, 2021

[†]For a subgroup of women with an ALP measurement also available in a non-pregnant state.

[‡]ALP levels considered only for the third trimester.

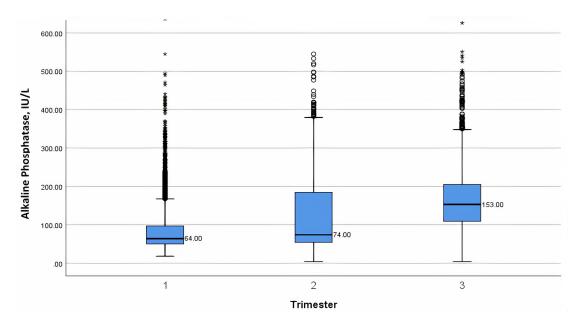


Fig. 2. Box-plot nomogram of median alkaline phosphatase (IU/L) levels throughout pregnancy, per trimester week - with 25th and 75th interquartile ranges, as well as minimum and maximum values.

Okesina *et al.* [21] in a longitudinal study of 67 pregnant women, determined the rise in total ALP and demonstrated that both placental and bone isoenzymes contribute to this increase. Placental ALP appears in serum as early as 13 weeks and becomes a significant contributor beyond 16 weeks, whereas bone ALP is a significant contributor at 32 and 38 weeks.

This is the first study to demonstrate that other than gestational age, possible modifiers of fetal sex, birthweight (as a surrogate for the fetal-placental mass) and maternal age did not affect ALP. This observation contradicts a small-scale study by Gol *et al.* [26], who compared total and placental ALP, according to fetal sex, among 30 women. Unlike our results they concluded that fetal sex affects ALP levels, across all gestational ages, with ALP significantly higher in mothers carrying a female versus a male fetus.

Establishing a normal reference range is of paramount importance, especially during pregnancy. Maternal adaptation to pregnancy induces profound physiological changes, in all organ systems, resulting in substantial changes in multiple blood analytes. An appropriate, robust, gestational nomogram is important to truly identify normal and pathological deviations. As there are only limited nomograms for pregnancy, and those for ALP are minimal, we constructed a large-scale, week-specific nomogram. Our planned research will use this nomogram to evaluate the association between abnormal ALP and pregnancy complications, all of which were not included in the current study.

We speculate that ALP is released following placental damage. Accordingly, higher than normal ALP may be associated with adverse pregnancy outcomes. This pathogenesis was occasionally supported by compatible histological findings associated with elevated ALP and pregnancy complications, such as placental infarcts [10, 11], abnormal syncytial and villus formation [12], chronic histiocytic intervillositis [34], and uteroplacental vascular disease [35]. Alternatively, others have suggested a different pathway through which placental ALP rises insufficiently in pregnancies complicated with fetal growth restriction and preeclampsia [36, 37], possibly reflecting decreased placental mass or malfunction, and relative ALP deficiency. Interestingly, there is a possible role in this mechanism to insufficient plasma level of the ectopic mineralization inhibitor inorganic pyrophosphate, which is a substrate of alkaline phosphatase [19].

The current study is not without limitations, mainly due to its retrospective design and the lack of some data (body mass index, mode of conception and smoking status), which may theoretically impact ALP levels. Additionally, we analyzed total ALP only and not the direct placental fraction. However, previous studies have demonstrated that placental isoenzymes have a half-life up to 7 days [38], which is the longest of all isoenzymes, and as such, have the most impact on the total ALP [5, 21]. As we included only healthy women, without any chronic or gestational morbidity, we safely assume that the rising levels of ALP represent the placental component and not a pathological source, from either maternal or fetal origin. Comparing the included and excluded population, i.e., those without an ALP measurement, demonstrated that the excluded population was similar to those included, confirming that there was no selection bias.

5. Conclusions

In conclusion, normal ALP levels associated with pregnancy are higher than in the non-pregnant state, gradually increasing from the end of the first trimester. This should be accounted for when evaluating lab results obtained for preg-

Volume 48, Number 6, 2021 1397

nant woman. In future studies, this nomogram can serve as the basis for comparison between a normal population and those with various pregnancy complications, to determine how ALP is affected during pathological events, the corresponding placental histopathology and whether ALP has a predictive role for adverse pregnancy outcomes.

Author contributions

NA—Conceptualization; Data curation; Investigation; Methodology; Validation; Supervision; Visualization; Writing - review & editing. Final approval. KNS—Investigation; Methodology; Validation; and data analysis and interpretation. LS, OSA, AB, EK—Data curation, Review and editing, Final approval. EH—Conceptualization; Data curation; Investigation; Methodology; Project administration; Supervision; Validation; Visualization; Roles/Writing - original draft; Writing - review and editing, Final approval. All authors agreed to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Ethics approval and consent to participate

The study was approved by the Institutional Review Board of Rabin Medical Center (Approval No. 0121-13-RMC). Informed consent was waived due the retrospective design of the study.

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Conflict of interest

The authors declare no conflict of interest.

References

- [1] Buchet R, Millán JL, Magne D. Multisystemic Functions of Alkaline Phosphatases. Methods in Molecular Biology. 2013; 263: 27–51.
- [2] Quigley GJ, Richards RT, Shier KJ. Heat-stabile alkaline phosphatase. a parameter of placental function. American Journal of Obstetrics and Gynecology. 1970; 106: 340–351.
- [3] Le Du MH, Stigbrand T, Taussig MJ, Menez A, Stura EA. Crystal structure of alkaline phosphatase from human placenta at 1.8 A resolution. Implication for a substrate specificity. Journal of Biological Chemistry. 2001; 276: 9158–9165.
- [4] Moss DW. Physicochemical and pathophysiological factors in the release of membrane-bound alkaline phosphatase from cells. Clinica Chimica Acta. 1997; 257: 133–140.
- [5] Valenzuela GJ, Munson LA, Tarbaux NM, Farley JR. Time-dependent changes in bone, placental, intestinal, and hepatic alkaline phosphatase activities in serum during human pregnancy. Clinical Chemistry. 1987; 33: 1801–1806.
- [6] Leroux M, Perry WF. Serum heat-stable alkaline phosphatase in

- pregnancy. American Journal of Obstetrics and Gynecology. 1990; 108: 235–239.
- [7] Heazell AEP, Judge JK, Bhatti NR. A case of isolated peripartum elevation of alkaline phosphatase in pregnancy complicated by gestational diabetes. The Journal of Maternal-Fetal & Neonatal Medicine. 2006; 19: 311–313.
- [8] Wojcicka-Bentyn J, Czajkowski K, Sienko J, Grymowicz M, Bros M. Extremely elevated activity of serum alkaline phosphatase in gestational diabetes: a case report. American Journal of Obstetrics and Gynecology. 2004; 190: 566–567.
- [9] Lozo S, Atabeygi A, Healey M. Extreme Elevation of Alkaline Phosphatase in a Pregnancy Complicated by Gestational Diabetes and Infant with Neonatal Alloimmune Thrombocytopenia. Case Reports in Obstetrics and Gynecology. 2016; 2016: 4896487.
- [10] Bashiri A, Katz O, Maor E, Sheiner E, Pack I, Mazor M. Positive placental staining for alkaline phosphatase corresponding with extreme elevation of serum alkaline phosphatase during pregnancy. Archives of Gynecology and Obstetrics. 2007; 275: 211–214.
- [11] Ranganath L, Taylor W, John L, Alfirevic Z. Biochemical diagnosis of placental infarction/damage: acutely rising alkaline phosphatase. Annals of Clinical Biochemistry. 2008; 45: 335–338.
- [12] Boronkai A, Than NG, Magenheim R, Bellyei S, Szigeti A, Deres P, *et al.* Extremely high maternal alkaline phosphatase serum concentration with syncytiotrophoblastic origin. Journal of Clinical Pathology. 2005; 58: 72–76.
- [13] Vongthavaravat V, Nurnberger MM, Balodimos N, Blanchette H, Koff RS. Isolated elevation of serum alkaline phosphatase level in an uncomplicated pregnancy: a case report. American Journal of Obstetrics and Gynecology. 2000; 183: 505–506.
- [14] Moawad AH, Goldenberg RL, Mercer B, Meis PJ, Iams JD, Das A, et al. The Preterm Prediction Study: the value of serum alkaline phosphatase, alpha-fetoprotein, plasma corticotropin-releasing hormone, and other serum markers for the prediction of spontaneous preterm birth. American Journal of Obstetrics and Gynecology. 2002; 186: 990–996.
- [15] Meyer RE, Thompson SJ, Addy CL, Garrison CZ, Best RG. Maternal serum placental alkaline phosphatase level and risk for preterm delivery. American Journal of Obstetrics and Gynecology. 1995; 173: 181–186.
- [16] Goldenberg RL, Tamura T, Dubard M, Johnston KE, Copper RL, Neggers Y. Plasma Alkaline Phosphatase and Pregnancy Outcome. Journal of Maternal-Fetal and Neonatal Medicine. 1997; 6: 140-145.
- [17] BEST RG, MEYER RE, SHIPLEY CF. Maternal Serum Placental Alkaline Phosphatase as a Marker for Low Birth Weight. Southern Medical Journal. 1991; 84: 740–742.
- [18] Brock DJ, Barron L. Measurement of placental alkaline phosphatase in maternal plasma as an indicator of subsequent low birthweight outcome. British Journal of Obstetrics and Gynaecology. 1988; 95: 79–83.
- [19] Lee J, Lee Y. The Relationship between Serum Alkaline Phosphatase and Arterial Stiffness in Korean Adults. Journal of Atherosclerosis and Thrombosis. 2019; 26: 1084–1091.
- [20] Veiga-Lopez A, Sethuraman V, Navasiolava N, Makela B, Olomu I, Long R, et al. Plasma Inorganic Pyrophosphate Deficiency Links Multiparity to Cardiovascular Disease Risk. Frontiers in Cell and Developmental Biology. 2020; 8: 573727.
- [21] Okesina AB, Donaldson D, Lascelles PT, Morris P. Effect of gestational age on levels of serum alkaline phosphatase isoenzymes in healthy pregnant women. International Journal of Gynaecology and Obstetrics. 1995; 48: 25–29.
- [22] van Buul EJ, Steegers EA, Jongsma HW, Eskes TK, Thomas CM, Hein PR. Haematological and biochemical profile of uncomplicated pregnancy in nulliparous women; a longitudinal study. The Netherlands Journal of Medicine. 1995; 46: 73–85.
- [23] Bacq Y, Zarka O, Bréchot JF, Mariotte N, Vol S, Tichet J, et al. Liver function tests in normal pregnancy: a prospective study of 103 pregnant women and 103 matched controls. Hepatology. 1996; 23: 1030–1034.

1398 Volume 48, Number 6, 2021

- [24] Larsson A, Palm M, Hansson L, Axelsson O. Reference values for clinical chemistry tests during normal pregnancy. BJOG: An International Journal of Obstetrics and Gynaecology. 2008; 115: 874– 881.
- [25] Abbassi-Ghanavati M, Greer LG, Cunningham FG. Pregnancy and laboratory studies: a reference table for clinicians. Obstetrics & Gynecology. 2009; 114: 1326–1331.
- [26] Gol M, Sisman AR, Guclu S, Altunyurt S, Onvural B, Demir N. Fetal gender affects maternal serum total and placental alkaline phosphatase levels during pregnancy. European Journal of Obstetrics, Gynecology, and Reproductive Biology. 2006; 128: 253–256.
- [27] Ardawi M, Nasrat H, BA'Aqueel H. Calcium-regulating hormones and parathyroid hormone-related peptide in normal human pregnancy and postpartum: a longitudinal study. European Journal of Endocrinology. 1997; 137: 402–409.
- [28] ACOG practice bulletin No. 101: Ultrasonography in pregnancy. Obstetrics & Gynecology. 2009; 113: 451–461.
- [29] ACOG Committee on Obstetric Practice. Practice bulletin #33: diagnosis and management of preeclampsia and eclampsia. Obstetrics & Gynecology. 2002; 99: 159–167.
- [30] Carpenter MW, Coustan DR. Criteria for screening tests for gestational diabetes. American Journal of Obstetrics and Gynecology. 1982; 144: 768–773.
- [31] Dollberg S, Haklai Z, Mimouni FB, Gorfein I, Gordon E. Birth weight standards in the live-born population in Israel. the Israel Medical Association Journal. 2005; 7: 311–314.
- [32] Tietz NW, Rinker AD, Shaw LM. IFCC methods for the measure-

- ment of catalytic concentration of enzymes Part 5. IFCC method for alkaline phosphatase (orthophosphoric-monoester phosphohydrolase, alkaline optimum, EC 3.1.3.1). Journal of Clinical Chemistry and Clinical Biochemistry. 1983; 21: 731–748.
- [33] Gillian L. Handbook of Diagnostic Biochemistry and Hematology in Normal Pregnancy. CRC press: Boca Raton. 1993. Available at: https://lib.ugent.be/en/catalog/rug01:000308238 (Accessed: 28 June 2021).
- [34] Marchaudon V, Devisme L, Petit S, Ansart-Franquet H, Vaast P, Subtil D. Chronic histiocytic intervillositis of unknown etiology: clinical features in a consecutive series of 69 cases. Placenta. 2011; 32: 140–145.
- [35] Davis CJ, Booth J, Summerfield J, Lazda EJ, Regan L. Grossly elevated placental derived alkaline phosphatase in pregnancy as a marker for uteroplacental vascular disease. Journal of Obstetrics and Gynaecology. 1999; 19: 533–534.
- [36] Holmgren PA, Stigbrand T, Damber MG, von Schoultz B. Serum levels of placental alkaline phosphatase in high-risk pregnancies. Obstetrics and Gynecology. 1979; 54: 631–634.
- [37] Adeniyi FA, Olatunbosun DA. Origins and significance of the increased plasma alkaline phosphatase during normal pregnancy and pre-eclampsia. British Journal of Obstetrics and Gynaecology. 1984; 91: 857–862.
- [38] Clubb JS, Neale FC, Posen S. The behavior of infused human placental alkaline phosphatase in human subjects. Journal of Laboratory and Clinical Medicine. 1965; 66: 493–507.

Volume 48, Number 6, 2021 1399