

Neuroendocrine effects of cadmium exposure on male reproductive functions

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1. ABSTRACT

In industrialized countries, the use of Cadmium (Cd) produces a form of anthropogenic pollution. Hence, exposure by human populations is becoming a public health problem. With a half-life of up to 40 years, cadmium is now a topic of great interest due to its role as an endocrine disruptor and its effects on male reproduction. Cd's diverse toxic mechanisms are based on its capacity to mimic divalent ions –calcium, zinc, iron– that participate in physiological processes. It alters the mitochondrial function and generates the production of free radicals that can induce apoptosis. In male

reproduction, Cd alters the precise coordination of the hypothalamic-hypophysis-testis axis (HHT), resulting in the loss of testicular functions like steroidogenesis, spermatogenesis and the onset of puberty, sexual maturity, sexual behavior and fertility. Exposure to Cd may even cause changes in the immune system that are associated with the reproductive system. This review analyses the state of the question regarding Cd's cellular and physiological mechanisms and the effects of this heavy metal on the neuroendocrine regulation of male reproduction.

2. INTRODUCTION

In industrialized countries, Cd contamination has become a serious problem for exposed populations due to the severity of its effects, which can cause critical environmental problems and health damage by accumulating in various organs. In 2019, the Metropolitan Area of Mexico City experienced an alarming atmospheric contingency when the concentrations of suspended particles measuring 2.5 micrometers or less (PM 2.5) reached historic levels, as did several other contaminants, such as sulfur dioxide (SO₂), carbon monoxide (CO), nitrogen dioxide (NO₂) and ozone (O₃). If we add pollutants like heavy metals –mercury (Hg), arsenic (As), lead (Pb), cadmium (Cd) and chrome (Cr)– then the outlook for Mexico and other industrialized countries is bleak, indeed. Unfortunately, studies conducted to date on these problems are sorely insufficient (1). The effects of these metal contaminants, especially Cd, on the reproductive health of male and female mammals have been explored to some extent, so we know that they can impact the entire reproductive process (2, 3). However, they do not affect different tissues and systems in precisely the same way because these vary in their degrees of susceptibility (4, 5).

Human activities have increased Cd concentrations in the environment by, for example, using fossil fuels, copper alloys, and coverings to protect iron and steel from corrosion, but this heavy metal is also an ingredient in paints used with ceramics and plastics, and is utilized to manufacture nickel-cadmium hydroxide batteries (Ni-Cd) for the automobile industry. In agriculture, Cd is an element in phosphorus-based fertilizers and pesticides, but is also found in leachates from garbage dumps, run-off from cultivated fields, and mining residues; thus, it is easily incorporated into animal feed and foods for human consumption, mainly in the form of cadmium oxide (CdO), cadmium chloride (CdCl₂) and cadmium sulfate (CdS) (6). Because human and animal populations are exposed to water and foods contaminated with Cd, and to industrial emissions and cigarette smoke that contain it, this metal is considered one of the principal toxic agents present in workplaces and the environment (7). According to the WHO's 2010 report, the maximum allowed

concentration of Cd in the air is ~0.04 µg/m³, while for water it is <1 µg/L, so tolerable concentrations for monthly human ingestion of Cd must be below 25 µg/kg of body weight. However, because this metal is found in foods like mollusks, fish, the viscera of grazing animals, rice and other crops, especially grasses that are consumed in high quantities in human diets, it is relatively easy for people to unknowingly exceed allowable limits. For this reason, authorities have determined that a person can ingest an average of only ~30 µg Cd/day through alimentation (8, 9).

In contrast, in uncontaminated areas where Cd exposure through water and food is low, the most important pathway that allows Cd to enter organisms is by inhaling cigarette smoke. Although environmental contamination is a minor source of exposure, levels of 2-15 ng/m³ can be found in urban areas. Some reports from Mexico City indicate values as high as 35-40 ng/m³, making it one of the metropolises with the highest levels in the world. We can state, therefore, that one of the main exposure routes affecting Mexicans is inhalation, generally in the form of cigarette smoke, which represents approximately 25% (range: 5-50%), while the oral pathway of ingesting contaminated water and foods (viscera, seafood) is estimated to contribute only around 5% (range, 1-10%). It is important to note that one cigarette may contain 1-2.8 µg of Cd, so smokers are more exposed to this contaminant, though most are unaware of its toxic effects (10, 11). We know that Cd accumulates mainly in the kidneys, liver, testicles, ovaries, placenta, lungs and brain, and that its half-life in organisms is very long: from 20 to 40 years in humans. Reports suggest a half-life of 6-38 years in the kidneys, 4-19 years in the liver, and 75-128 days in blood, but the corporal accumulation of this particular metal is dependent on the duration and dosage of exposure, because it can mimic ions that are physiologically necessary and so interfere with biological processes in the organism that allow it to prolong the time required for excretion (9). Cd, therefore, is one of the most widely-distributed environmental contaminants, plus it has a long half-life and the ability to mimic molecules involved in vital biological activities. All this means that it exerts significant effects on human reproduction (12, 13). However, numerous aspects of the effects of Cd

contamination on the neuroendocrine regulation of male sexual maturation and reproduction are not fully understood, so improving our comprehension of the cellular and physiological mechanisms involved will make it possible to implement measures that can reverse Cd's toxic effects.

3. CELLULAR MECHANISMS OF Cd TOXICITY

Cd is a heavy metal that can generate a broad range of toxic effects at both the systemic and cellular levels. Among the former, we can mention nephrotoxicity, carcinogenesis, teratogenesis and damage to the endocrine and reproductive systems. At the cellular level, Cd can damage the structure of DNA and proteins, interfere with several mechanisms of DNA repair, cause genic instability, modify cell proliferation and differentiation, and even activate cell death by apoptosis. Indirectly, Cd induces the generation of reactive oxygen species (ROS), damages DNA, and alters gene expression and signal transduction (14, 15). Any attempt to comprehend the cytotoxicity of Cd mechanisms must begin by clarifying how this metal is transported into the interior of the cells, and then distributed to the intracellular compartments where it accumulates. Therefore, this review analyses the principle entrance mechanisms of Cd into the reproductive cells and then describes its toxic effects and their consequences for the various structures that constitute the male reproductive system.

Cd is a transition element that belongs to Group 12 of the periodic table. Its most frequent oxidation number is +2 (with two positive charges = Cd^{+2} or Cd^{II}). This allows it to mimic and displace divalent cations that have physiological activity, including calcium (Ca^{2+}), iron (Fe^{2+}), magnesium (Mg^{2+}), manganese (Mn^{2+}), zinc (Zn^{2+}) and selenium (Se), in many biochemical and cellular processes. As a result, it can bind to the proteins that transport these metals and form compounds with certain biomolecules that contain negative charges, including amino acids and proteins, nucleic acids, and the sulfhydryl or thiol groups (-SH), among others (16). Thanks to its physicochemical properties, Cd can also interact with membrane transporters involved in carrying these cations to the cells through

a process of ionic mimicking (17, 18). When exposure to Cd is oral, the metal can pass through the intestine by diverse physiological mechanisms common to several species, because it utilizes the same absorption mechanism as Ca, Zn or Fe, as has been widely studied among mammals (19, 20). The mechanism that transports Cd requires proteins from the family of the Ca voltage-gated channel (VGCC) or Fe transporters (DMT family; divalent metal transporters). The DMT-1 protein is the one with the greatest affinity for Cd, but this metal can also utilize Zn transporters (ZIP family of metal transporters; Zrp, Irt-related proteins), especially the ZIP-8 protein, which is present in the gastrointestinal tract (21), and the Ca 1 transporter (CaT-1; also known as TRPV, for its initials in English: transient receptor potential cation channel, subfamily V), which is expressed in highest quantities in the intestines of mammals (22).

The mimicking that Cd shares with divalent metals is what allows it to interrupt certain processes, including the male reproductive function, which is one of the most severely affected since Cd exposure has often been associated with infertility (23). Specifically, Cd penetrates the spermatozooids of mammals through voltage-dependent Ca channels called CatSper. Research shows that Cd sensitivity and permeability depend on the precise composition of the channel, especially the presence of the absolute number of residues of the aspartate amino acid which, at physiological pH, are negatively-charged (24). Once absorbed, Cd binds to albumin and is carried through the bloodstream. It first reaches the liver (25), where it induces synthesis of metallothioneins (MT), a class of proteins rich in cysteines that bind to metals. The albumin and MT together protect against Cd toxicity by limiting its availability in the cells and tissues and then eliminating it in the urine. Cd's ability to induce hepatic and renal lesions intensifies its toxic effects and allows it to accumulate for years, so that its reproductive toxicity may not appear until many years later (26). One of the most comprehensively studied effects of Cd, and the one that affects mainly reproductive tissues, is apoptosis. Studies have demonstrated that this metal can induce oxidative stress and the formation of ROS, while indirectly it can inhibit the activity of antioxidant enzymes and proteins that present Zn finger motifs (27, 28). Cd can

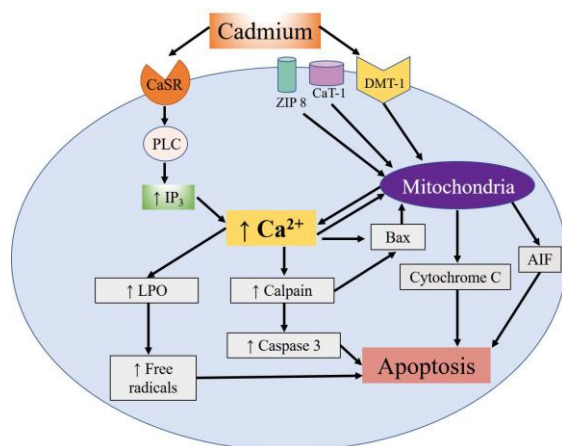


Figure 1. Summary of the apoptotic effects of cellular Cd reported in different cell types. Cd's entry mechanisms are the DMT-1, Zip-8 and CaT-1 ion transporters. Once inside, Cd exerts diverse effects on the mitochondria, including the release of the AIF apoptosis-inducing factor and cytochrome C. In addition, Cd can bond to the Ca-sensitive receptor (CaSR), which activates the PLC-IP3 pathway and leads to an increase in intracellular Ca concentrations that, in turn, promotes activation of apoptotic calpains and caspases. This increase of Ca also fosters activation of the pro-apoptotic Bax protein and release of the mitochondrial cytochrome C and AIF proteins. Moreover, the increase of Ca induces the lipoperoxidation (LPO) of lipids and mitochondrial proteins, leading to the formation of free radicals, and promoting apoptosis.

also reach the mitochondria and damage them by interacting with the –SH groups of the cysteines present in the proteins of the internal mitochondrial membrane, causing it to lose its high impermeability and so dissipate its potential – which is necessary for adenosine triphosphate (ATP) synthesis– by uncoupling the processes of electron transport and oxidative phosphorylation. At the same time, Cd can bind to the hemoproteins that form part of the respiratory chain, causing increased ROS production, which generates mitochondrial lipid peroxidation. Studies have also shown that Cd induces activation of the intrinsic pathway of apoptosis, characterized by the release of mitochondrial cytochrome C and the apoptosis inducing factor (AIF), which can stimulate apoptosis at the nuclear level independently of the caspases. Moreover, Cd can bond to the Ca-sensitive receptor (CaSR) that induces activation of C phospholipase (PLC). This, in turn, increases concentrations of intracellular Ca mediated by the increase of the second inositol triphosphate messenger (IP₃), which promotes the translocation

of the calpains (Ca-dependent cysteine proteases) towards the cytoplasmic membrane, thus inducing autolysis and activation of the pro-apoptotic protein Bax. This protein allows the release of C cytochrome and the induction of initiator and effector caspases that convert mitochondrial apoptosis into an irreversible process (Figure 1). In addition to caspases, the substrates of the calpains contain membrane and cytoskeleton proteins, transcription factors and oncogenes associated with toxic effects that alter the process which allows spermatozooids to bond to ovules (29).

As mentioned above, Cd is directly related to the induction of mitochondrial apoptosis, but it can also inhibit, through indirect mechanisms, anti-apoptotic systems, including such antioxidant enzymes as catalase (CAT), glutathione peroxidase (GPX), and superoxide dismutase (SOD). This results in the accumulation of ROS (30). Exposure to Cd is further associated with reductions of both glutathione (GSH) content and the activities of GPX, CAT and SOD, effects that can be analyzed as functions of the mechanisms of protein synthesis from the nucleus and in the cytoplasm because Cd can increase both the expression of transcription factors and the stability of transduction initiation factors located in eukaryotes, specifically eIF4AE (31). Although Cd's principle target is the mitochondria, other cytotoxic effects occur because it interacts with receptors in the plasmatic membrane thanks to its ability to form multiple unions that include covalent and ionic bonding to the sulfur, oxygen and phosphorus atoms present in some amino acid residues. This explains Cd's ability to modify signal-transducing pathways. Another widely-studied effect is the extracellular entrance of Ca through ionic channels that, as mentioned previously, increases ROS, Bax and AIF concentrations and, consequently, induces apoptosis. Contradictory findings, however, have shown that at sub-lethal concentrations Cd can promote cellular adaptation and survival by activating the extracellular signal-regulated kinase ½ (ERK1/2), phosphatidylinositol 3-kinase (PI3K)/protein kinase B (Akt-PKB) signaling pathway and regulating transcription factors like redox effector factor 1-nuclear factor E2-related factor 2 (Ref1-Nrf2), nuclear transcription factor-kappa B (NF-κB), int/Wingless protein (Wnt), B-cell

lymphoma 2 (Bcl-2) and activator protein-1 (AP-1) (29). Other cellular processes that Cd can alter include adhesion and migration, since it can regulate the structure and function of the cytoskeleton and modify the concentrations and localization of E-cadherin, N-cadherin and β -catenin proteins, all of which are important components of tissue integrity (32). Because the cadherins are Ca-dependent proteins, Cd can mimic them, modify their structure and function, and lead to the loss of intercellular adherence, possibly activating uncontrolled processes of proliferation and cancer in both reproductive and non-reproductive tissues (33).

In summary, Cd is a highly toxic metal that affects various tissues, including those of the reproductive tract. Its principle cytotoxic effects are due to its ability to mimic divalent cations (Ca, Fe, Mn, Mg, Zn, Se) that perform physiological functions. However, this section has presented only a few of Cd's many effects usually associated with reproductive processes. The following sections will, therefore, explain Cd's effects on specific processes and tissues.

4. EFFECTS OF Cd ON THE FUNCTION OF THE HYPOTHALAMIC-HYPOPHYSIS-TESTIS AXIS (HHT)

Male reproduction depends on the precise coordination of the HHT axis, which governs correct testis function and the subsequent production of steroid hormones and spermatogenesis. Male reproductive capacity results from the activation of the HHT axis during puberty, which begins with the pulsatile secretion of the gonadotrophin-releasing hormone (GnRH) by the hypothalamus to stimulate the gonadotrophs in the hypophysis to biosynthesize and secrete hypophysis gonadotrophins and the luteinizing (LH) and follicle-stimulating hormones (FSH). These, in turn, sustain intragonadal testosterone (T) synthesis and spermatozoid production. This function is maintained by a negative feedback mechanism controlled by the increase in the levels of T, which induces the reduction of both the hypothalamic secretion of GnRH in the hypothalamus-hypophyseal portal circulation and the release of both gonadotrophins into the bloodstream by the hypophysis. The hypophysis decreases the

production and release of LH through a negative feedback mechanism that also reduces GnRH and LH and, therefore, lowers plasma levels of T (34). Multiple exogenous factors –including environmental contaminants– can alter the function of the HHT. Long-term exposure to Cd will cause toxic effects due to its accumulation over time in a variety of tissues, including the hypothalamus, hypophysis and testicles (35).

4.1. Synthesis of hypothalamic and hypophysis hormones

The hypothalamus is located at the base of the brain, beneath the thalamus and third ventricle, but above the optic chiasma. It is an extremely complex cerebral structure that forms a functional unity with the hypophysis gland to coordinate such functions of the organism as somatic growth, maturation and gonadal function, among others (36). Its neurosecretory cells synthesize and secrete the GnRH neurohormone, which is responsible for stimulating the HHT axis (37, 38). GnRH is a decapeptide secreted by approximately 1000 neurons located in the hypothalamus (arcuate nucleus and middle eminence). It is responsible for stimulating the secretion of gonadotrophins (FSH and LH) by bonding to the membrane receptors in the gonadotrophs of the hypophysis. The neurovascular linkage between the hypothalamus and hypophysis is the hypophysis stalk, which contains the hypothalamic-hypophyseal portal system. The hypophysis gland is located immediately below the hypothalamus, resting on a depression in the base of the cranium called the *sella turcica*. It is divided into the anterior and posterior hypophysis (39). The FSH and LH secreted by the anterior hypophysis belong to a family of dimeric glycoprotein hormones that share certain structural characteristics (40). Each hormone consists of two sub-units: subunit- α and subunit- β , which are bonded non-covalently. Subunit- α is common to all glycoprotein hormones, but each subunit- β presents a specific sequence of amino acids –111 for FSH and 121 for LH– which gives each one its biological specificity (41). Both FSH and LH act through the classic mechanisms of the protein hormone receptor, which involves transmembrane receptors associated with the G proteins located in organs like the testicle (42).

Several studies have shown that Cd at different concentrations can alter the neuroendocrine function of the hypothalamic-hypophysis axis and so affect the release of various hormones (43-46). While few studies have analyzed Cd's effects on the hypothalamic secretion of GnRH, some reports indicate that Cd exposure increases the expression of ARNm for GnRH 1 and GnRH 2 in salmon, and of GnRH 2 in *Micropterus salmoides* (47, 48), while chronic administration (4 weeks) of 5 mg/kg of CdCl₂ in male rats decreased plasma GnRH levels (49).

Regarding Cd's effect on concentrations of hypophysis hormones, studies in humans show contradictory results. Research on males exposed to Cd in the workplace has found that it reduces FSH secretion after only short exposure periods (50). Other working groups, however, report positive correlations between plasma Cd values and serum LH concentrations, but not with FSH (51). But Cd also reduces concentrations of intrahypophyseal LH (52). Studies in men who suffer from azoospermia or oligospermia associated with infertility have found positive correlations between serum and seminal Cd concentrations and concentrations of FSH (53), but other work failed to find correlations between Cd levels and concentrations of FSH or LH (54-56). Chronic exposure to 50 ppm of Cd introduced into rats' drinking water reduced serum LH concentrations, but increased FSH (57), while chronic intraperitoneal administration of 1 or 5 mg/kg of CdCl₂ decreased concentrations of both gonadotrophins (49, 58). However, the administration of high doses (4 or 6 mg/kg) in an acute or sub-chronic scheme reduced FSH and LH concentrations (59). It is well-known that Cd produces apoptosis in cells of the anterior hypophysis (60) through the mechanisms described above, and that it modifies total lipid content (44). These effects could, therefore, affect the functions performed by the testicle.

4.2. Cd, neurotransmitters and GnRH

Neurosecretion of GnRH in the hypothalamus is regulated by multiple factors, including neurotransmitters, since it receives information from neurons that produce noradrenaline, dopamine, serotonin (5HT), γ -aminobutyric acid (GABA) and glutamate. There are

reports that glutamate and noradrenaline stimulate the HHT axis, while GABA, dopamine and serotonin seem to inhibit it (38, 61-63). Glutamate is an important neurotransmitter that stimulates the GnRH neurons, which are innervated directly by glutamatergic neurons. We also know that the GnRH neurons express receptors for α -amino-3-hydroxy-5-methyl-4-isoxazole propionic acid (AMPA) and kainate and, though to a lesser extent, N-methyl-D-aspartate receptors (NMDA) (64). Hence, it is considered an activator in the pulsatile release of GnRH, since some reports suggest that stimulation of the NMDA receptors leads to early-onset puberty in rats and monkeys. The increases in glutamate synthesis in the hypothalamus initiate puberty in those species (37, 65, 66). In adult rats, stimulating the NMDA receptors triggers the release of LH via GnRH (67).

GABA, meanwhile, is the neurotransmitter that inhibits mature neurons, though it can also perform inhibiting or stimulating functions on GnRH secretion, depending on changes in its receptors during the development of different individuals (68, 69). GnRH neurons express the GABA_A (70, 71) and GABA_B (72) receptors. In rodents, administering bicuculline (a GABA_A antagonist) and phaclofen (a GABA_B antagonist) reduces the release of GnRH in young animals, but increases it in adults (73).

Regarding noradrenaline, we know that this neurotransmitter contributes to facilitating an increase in the release of GnRH/LH. Also, activation of the α 1-adrenergic receptor facilitates the release of noradrenaline, but not α 2 or β , while infusions of the α 1-adrenergic blocker prazosin into the middle eminence suppress GnRH release (74).

With respect to dopamine, the precise role that it plays in initiating puberty has not yet been established completely, though research has demonstrated that hypothalamic concentrations of dopamine increase before and around puberty onset (75).

Turning to 5HT, we find that it has an inhibitory effect on GnRH synthesis, and there are reports that treatment with 5HT or agonists of the serotonin 1A receptor (5HT1A) and serotonin 2

receptor (5HT₂) receptors in male rats reduces the neurons that contain ARNm and codify for GnRH (61).

Exposure to Cd modifies the amounts of neurotransmitters, as has been reported in adult male rats under chronic exposure to high doses of CdCl₂ (25 mg/kg) in drinking water, as they suffered a reduction in 5HT, dopamine and noradrenaline concentrations in the hypothalamus (43, 76). We also know that acute treatment with this same dose of CdCl₂ decreases GABA concentrations in the striate of rats (77), and that chronic treatment in drinking water reduces glutamate concentrations, though without affecting GABA in the hypothalamus (78). These modifications of neurotransmitters could affect GnRH secretion.

4.3. Effects of Cd on the morphophysiology of the male reproductive tract

Testicle development is controlled by a hormonal balance in a microenvironment of somatic and germinal cells that involves cell-to-cell interactions and hormonal signaling by the FSH and LH secreted by the hypophysis (79). It is well-known that while FSH is important for controlling the proliferation of Sertoli cells and inducing spermatogenesis (39, 80, 81), LH is required for steroidogenesis because it stimulates the membrane receptor in the Leydig cells which, in turn, stimulates the enzymatic conversion of cholesterol into T (39). T synthesis and spermatozoid production are the testicle's principle functions, and both are affected by exposure to Cd. The decrease of androgens can alter not only the morphology and functioning of the testicle but also the morphophysiology of the epididymis and accessory glands and reproductive parameters, including fertility.

4.3.1. The testicle

The testicle is the organ that generates gametes; that is, the male sexual cells called spermatozoids. But it is also capable of synthesizing T; hence, it is considered the primary male sex gland. Testicles are found in pairs, are oval-shaped and covered by a layer of connective tissue called the tunica albuginea that invaginates inside the testicular body to form the testicular mediastinum and septa.

The septa separate the testicle into lobes that contain the seminiferous tubules, the site where spermatogenesis—or spermatozoid synthesis—takes place. Spermatogenesis refers to the process of spermatozoid production and their subsequent differentiation from round diploid cells to haploid cells with the characteristic form of spermatozoids. This process is divided into three main phases: 1) mitosis; 2) meiosis; and 3) differentiation. The first phase is also called spermatocytogenesis. During this process, type A spermatogonia divide repeatedly by mitosis to produce other spermatogonia of the same type, which serves to maintain a cellular reserve. These type A spermatogonia then give rise to the type B spermatogonia, which are also divided by mitosis to produce more type B spermatogonia. Studies speak of the “maturation” of the type B spermatogonia because they increase in size and will be transformed into primary spermatocytes. Upon reaching this state, they migrate to the adluminal compartment of the seminiferous tubule, where they will undergo the first meiotic division. Secondary spermatocytes are obtained as a result of this first meiotic division, while spermatids result from the second meiotic division. The phase of differentiation, or spermiogenesis, occurs when the round spermatids are differentiated into spermatozoids. Sertoli cells are of great importance in this, and all, stages of spermatogenesis. They are located in the seminiferous tubules where they provide physical support for germinal cells and maintain the tubular environment to allow the differentiation of spermatocytes into spermatozoids (82).

Although Cd can damage several organs, the testicles are the main targets of Cd toxicity (83, 84), since this metal can migrate to, and cause alterations in, these organs (85) because they are particularly susceptible to the effects of oxidative stress due to their high content of polyunsaturated fatty acids (86). Figure 2 summarizes the principle affectations of the testicle caused by Cd exposure, especially the reduction of testicular size and mass (87-89). Cd exposure reduces the diameter and length of the seminiferous tubules (88, 90, 91). This is explained by their high content of germinal cells that die by apoptosis (89, 92-102), which in some cases is observed as vacuolization of the epithelium, or even the complete loss of cell layers from the

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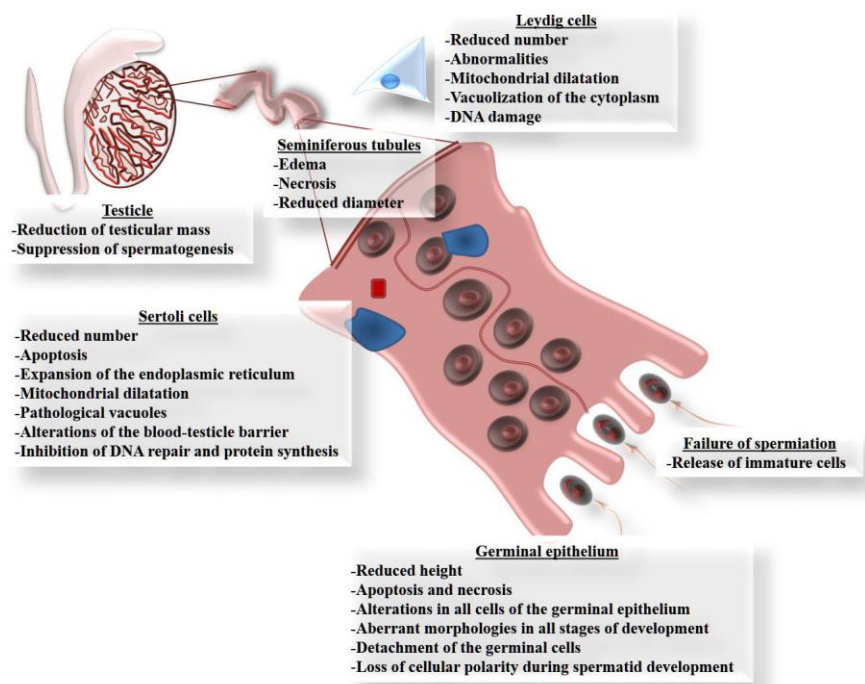


Figure 2. Principle damage that cadmium causes in the testes. All these forms of damage can be explained by the involvement of reactive oxygen species (ROS), because exposure to Cd increases oxidative stress in the testicle, determined as lipoperoxidation.

germinal epithelium (88, 99). This causes interstitial edema and hemorrhaging, together with a marked reduction in the height of the epithelium and its calcification (90, 91, 103). In this regard, alterations have been seen in all cells of the germinal epithelium, including unusual morphologies in all cell types and decreases in the number of spermatogonia and spermatocytes (90, 91, 104-106). But Cd also induces poor orientation and the loss of cellular polarity in the development of spermatids in adult rats (107); a finding that may be associated with failures in spermiation (107) caused by the release of immature cells (104-106).

Sertoli cells play a crucial role in the process of spermatogenesis by participating from the differentiation stage of the germinal cells through to their maturation as spermatozooids. Thus, any changes that these cells suffer due to Cd exposure will affect this process directly. There are reports that, like Leydig cells, Sertoli cells are highly vulnerable to damage by Cd (105, 108), which can even inhibit their proliferation (109) and decrease the number of

cells available (90). This can occur because Cd has been identified as an apoptosis-inducer (110). Other reports on Sertoli cells indicate that Cd causes an expansion of the endoplasmic reticulum, mitochondrial dilatation, vacuolization and DNA damage in birds and mammals like rodents and hogs (100,106,109,111). Another well-known effect of Cd is that it interacts with extracellular Ca binding domains present in E-cadherins (103), which are the principle molecules of cellular adhesion like those found between Sertoli cells as part of the blood-testis barrier (BTB), and between Sertoli cells and germinal cells. The union of E-cadherin with Cd interrupts this cell-cell binding (100). Also, Sertoli cells from human cells, after forming an epithelium, were used as a model to determine the effects of Cd on BHT. In this study, Sertoli cells were sensitive to Cd toxicity, as well as changes in localization in cell adhesion proteins caused by alterations in the cytoskeleton, especially the depolarization of the F-actin filaments caused by the delocalization of the proteins: Eps8 (substrate of the epidermal growth factor 8 receptor pathway) and the branched actin

polymerization protein. Furthermore, Cd prevented endocytic vesicle trafficking and the delocalization of c-Src and annexin II proteins (actin regulatory proteins) (112). In addition, administering Cd can reduce spermatogenesis by affecting the permeability of the BTB (107, 113), which is regulated by certain tight bonds: occludin, N-cadherin and vimentin (100). Leydig cells can also be affected in males exposed to Cd, for studies show abnormalities and reductions in the number of cells in those individuals (87, 90). These affectations include dilatation of the mitochondria, vacuolization in the cytoplasm and DNA damage (114, 115).

Additionally, to these effects Cd reduces the effectiveness of the antioxidant defense system by decreasing the enzymatic activity of SOD, CAT and GPX, as well as that of GSH. This means that levels of oxidative stress in rats' testicles will rise due to alterations of the antioxidant system (116). For this reason, when organisms have been exposed to Cd, Zn can be utilized to reduce this metal's effects by competing with Cd. However, many enzymes require Zn to be activated (117), so it has been associated with numerous enzymes in the body that can prevent damage by activating the antioxidant system (118-121).

4.3.2. The epididymis

When the spermatozooids leave the testicle, they cannot yet fertilize the oocyte. They must pass, first, through the epididymis to acquire this ability. This stage of the process is called epididymal sperm maturation. The epididymis is an elongated, highly-contoured tube, whose length varies by species. It may measure 1 m, as in the mouse, 3 m in rats, between 5 and 7 m in humans, and up to 70 or 80 m in horses (122). It is attached to the testicles by the efferent conducts and contains three main anatomical regions –caput, corpus and cauda– though additional regions can be recognized histologically, so the number of regions depends on both species and the form of the initial segment (123). Observations of most species show that epididymal sperm maturation is achieved before the spermatozooids reach the cauda, where they are stored until ejaculation (82). However, studies of males exposed to Cd reveal that the weight of the epididymis decreases (124) and that the highest

accumulations of Cd occur in the corpus. This suggests that Cd can affect epididymal sperm maturation, the functionality of the spermatozooids and, therefore, the reproductive success of males exposed to this metal (125).

The epididymal tubule consists of a pseudostratified epithelium where we find principle cells, basal cells, clear cells, narrow cells, halo cells and apical cells. However, the principle and basal cells are present throughout the epididymal tubule, while the clear, narrow and apical cells appear only in certain segments of the epididymis (124). The disposition and localization of the cells in the epithelium along the epididymal tubule constitute microenvironments by secreting and absorbing ions, sugars and proteins, etc., that, upon coming into contact with the spermatozooids, undergo biochemical, morphological and physiological modifications that gradually lead them to acquire – during their transit through this organ– their ability to move and become capacitated to interact with the oocyte; that is, the capacity to achieve fertilization (126).

There are reports that the epididymis requires the participation of androgens to maintain its epithelium in good condition (127) and to perform and control its functions. We know that the androgen receptor (AR) is found in great quantities in the caput and corpus, though in lower amounts in the cauda (123, 128). While the epithelium of the epididymis is androgen-dependent, estrogens also participate in maintaining its morphophysiology (129) by secreting T, which is necessary for the correct expression and secretion of the proteins, glycoproteins, glycolipids and phospholipids required for the spermatozooids to mature and survive (123, 130, 131). This becomes especially important when we consider that animals exposed to Cd suffer reductions in their T content (132, 133), which decreases the size and weight of the epididymis (122, 133) as a consequence of histological damage (133). This damage may increase the height of the epithelium in the caput (134). This could be interpreted as indicating a problem with fluid absorption, which depends on a contribution of androgens (135) that, as mentioned above, decreases when Cd is administered. In addition to these problems with morphology and

histology, the epididymis also suffers a reduction in the number of spermatozooids in the lumen of the tubule located in the region of the initial segment; that is, the caput and cauda. This parameter is inversely proportional to the concentration of Cd administered (134, 136).

One of the most important changes that spermatozooids undergo as they pass through the initial segment and caput of the epididymis is hypercompaction of the DNA, which is achieved through the formation of disulfide bridges between cysteine residues that bond to protamines (137). This change is essential for conserving the spermatozoid's genetic material while it transits to the fertilization site (138). However, Cd causes decompaction of the chromatin by eliminating the disulfide bridges, an action that increases the fragmentation of spermatid DNA (139).

Changes in the carbohydrates in the membrane of the spermatozoid constitute other modifications that spermatozooids may undergo while passing through the epididymis, because the adequate distribution of carbohydrates is what gives them the capacity to cross the cervical mucosa, approach the oocyte, and fertilize it (126). This process is dependent on the following enzymes: glycoside hydrolases (hydrolytic enzymes that break down carbohydrate residues) and glycosyltransferases (enzymes that add carbohydrate residues from another donor sugar). Both kinds of enzymes function by modifying the glycoproteins on the spermatozooids' surface during epididymal maturation (140). We have found that administering Cd lowers the concentration of N-acetyl-glucosamine, sialic acid and fucose in the membrane of spermatozooids obtained from the three main regions of the epididymis (122). These changes can be explained by the reduction of androgens caused by exposure to Cd, which may impede the synthesis of such proteins as the glycoside hydrolases and glycosyltransferases, though this remains to be demonstrated.

Other intracellular changes that occur in spermatozooids during their maturation are the modification of Ca and cyclic adenosine monophosphate (cAMP) concentrations, and distinct

patterns of tyrosine phosphorylation, all of which are essential for the adequate development of spermatozoid motility (141-144). For spermatozooids to develop this ability, they must pass through a process of protein activation in the epididymis called tyrosine phosphorylation, which involves a cAMP-dependent tyrosine kinase (145-147). It has been suggested that both non-human animals and humans exposed to Cd present alterations in sperm motility, possibly caused by changes in the process of tyrosine phosphorylation, energy loss, and decreased concentrations of ATP/AMP (148). One *in vitro* study of spermatozooids obtained from the caudal region of the mouse's epididymis (149) found that the percentage of phosphorylated proteins increased under Cd exposure, compared to a control group. This does not necessarily mean that the increase in phosphorylated proteins in tyrosine residues favors the development of sperm motility because this parameter was not analyzed in the study. However, a study by our working group found that Cd also increases the number of phosphorylated proteins, and that phosphorylation increased as the spermatozooids advanced from the caput to the cauda in subjects treated with Cd. This result was distinct from observations of the control subjects, where the pattern of protein phosphorylation was similar between caput and corpus. In contrast, the spermatozooids obtained from the cauda showed a marked reduction (122).

According to these reports on the causes of damage to the morphophysiology of the epididymis in organisms exposed to Cd, it is not a question of oxidative stress, even though numerous studies argue that this is the case for the testicle (150-152). This is affirmed because the activity of the SOD, CAT and GPX enzymes does, indeed, decrease, but does not generate lipoperoxidation (133), as occurs in the testicle. Therefore, the principle factor in the damage that Cd causes in the epididymis is the reduction of androgen concentrations.

4.3.3. Accessory sex glands

The accessory sex glands include the seminal vesicles, prostate, and bulbourethral (or Cowper) glands. The seminal vesicles are joined to the epididymis by efferent ductules and contribute over 60 % of the seminal fluid. They are sack-like

Table 1. Effects of cadmium on parameters of sperm quality

Parameter	Description	Species	Reference
Motility	Reduction	Rats	(135, 151, 158, 161)
		Mice	(164)
	Asthenozoospermia	Humans	(158-163)
Concentration	Reduction	Rats	(123, 154, 161)
		Mice	(164)
	Oligozoospermia Azoospermia	Humans	(158-163)
Morphology	Reduction	Rats	(123)
	Teratozoospermia	Humans	(163, 158, 160, 162)
Live	Reduction	Mice	(164)
		Humans	(158, 160)

structures located on one side of the bladder. The prostate surrounds the ejaculatory ducts at the base of the urethra, just below the bladder. It is nut-shaped and its secretions also form part of the seminal fluid. The bulbourethral (Cowper) glands are located at the base of the penis, where the urethra begins. They secrete an alkaline liquid that lubricates and neutralizes the acidity of the urethra and provide the first fraction of the seminal fluid. The sum of the secretions of these accessory glands constitutes the seminal fluid that, together with the spermatozooids, form the semen (153). High Cd levels in the seminal fluid have been associated with alterations of the sperm parameters in infertile males (154) and a decrease in seminal volume (133). Thus, in addition to the morphophysiological damage that Cd inflicts on the testicles and epididymis, it also reduces the weight of the prostate and seminal vesicles (124, 155), which could be associated with the decreased seminal volume.

Cd also plays a critical role in male infertility by increasing incidences of prostate cancer (156). Cd concentrations in the prostate increase under chronic exposure and, as mentioned above, T synthesis decreases in animals exposed to Cd. Here, it is important to consider that the production of testicular androgens is essential for maintaining prostate tumors since the development of this type of cancer can be delayed –for a time– by castration, estrogen administration, or both (98). The fact that Cd exposure decreases T in organisms might explain the development of prostate cancer in men exposed to

this metal because this condition is an aberrant genic expression that stimulates cellular proliferation or blocks apoptosis (157). Activation of such transcription factors as the metallothionein gene and some proto-oncogenes can increase the proliferation of cells that contain damaged DNA (158) and, perhaps, suppress DNA repair, thus augmenting the population of cells with DNA damage (157). This notion is sustained by the fact that chemically-induced apoptosis can be blocked by Cd and so foster the accumulation of aberrant cells (98).

4.4 Cd's effect on sperm parameters

Most reports on Cd's effects on sperm quality are based on studies of humans exposed to Cd, or experiments that exposed rats and mice to this metal (Table 1). Observations show that individuals exposed to Cd have low sperm concentrations, motility and vitality, accompanied by a higher number of sperm abnormalities (159-163). In addition, infertile patients have Cd in the bloodstream, and studies have determined that as Cd concentrations rise, sperm concentrations and motility decline (162,164). In this regard, when Cd is administered to rats and mice, they also present low sperm concentrations and motility, accompanied by an increase in the incidence of sperm abnormalities (124, 136, 152, 159, 162). Research on another sperm parameter studied in mice exposed to Cd – apart from concentration, motility and vitality, all of which decrease under exposure to Cd– has found marked increases in the number of abnormalities in

the heads of the spermatozooids, associated with a premature acrosomal reaction and an increase in DNA fragmentation (165). Certain studies help us understand the mechanisms through which Cd causes alterations in sperm parameters. Thus, we know that while sperm motility can be affected by various mechanisms, one of the most important ones is Cd, because it is the principle competitor of Ca, an especially important regulator of sperm motility (166, 167). Here, studies with mice have demonstrated that the activity of the CatSper channel is modified by Cd exposure and that this affects sperm motility (136, 168). In addition to this, however, Cd can also reduce levels of sialic acid (the terminal carbohydrate of the glycoproteins) in spermatozooids (169). Sialic acid is important because it enhances sperm motility by preventing “friction”. But Cd also increases ROS concentrations (149) that, according to some reports, can cause oxidative stress that damages the spermatozooids’ membrane and alters tyrosine phosphorylation in the sperm. This further diminishes sperm motility and their capacity to fuse with the oocyte, thus interfering with the fertilization process (170). Cd, however, is not an active metal in the redox system; that is, it does not generate ROS through a pathway similar to the Fenton reaction. Rather, Cd can bond to the sulfhydryl groups of ROS regulators, like GSH, to prevent them from functioning as antioxidants. This allows the ROS to increase –indirectly– and generate greater oxidative stress (171, 172).

5. EFFECTS OF Cd ON PUBERTY AND SEXUAL MATURATION

Pulsatile secretion of GnRH by the hypothalamus is necessary for maintaining male reproduction. However, GnRH and gonadotrophin secretion varies constantly during sexual development. Neuroendocrine stimulation of the HHT reproductive axis begins during fetal development (173) when GnRH neurons are observable, though the connections between these neurons and the portal system of the hypothalamus and hypophysis do not become functional until around week 16 of gestation in humans, thanks to the secretion of GnRH (174). GnRH neurons are functional at birth but remain tonically-repressed during childhood after the cascade of perinatal androgens. Hypothalamic

GnRH secretion increases at the onset of postnatal life and leads to the temporal activation of gonadal steroidogenesis, especially in males, though this remains quiescent because the release of pulsatile GnRH is suppressed until the onset of puberty, when GnRH release is reactivated (175). As puberty progresses, the gonadotropins begin to secrete continuously to stimulate T synthesis, which is what fosters sexual development (176). At that moment, the increase in T levels, together with its conversion into the active metabolite dihydrotestosterone (DHT), triggers the development of the secondary sex characteristics and growth of the reproductive organs, while also increasing the libido by converting T into estradiol (177). In adulthood, pulsatile GnRH continues to stimulate the biosynthesis of LH and FSH that, in turn, maintain the production of intragonadal T and spermatogenesis, as well as the systemic secretion of T (178).

5.1. Puberty

Puberty is a period that comprises a complex series of changes that lead to sexual maturation and the acquisition of reproductive capacity (179). Its onset is controlled by a sophisticated regulatory system in which the brain centers –hypothalamus and hypophysis– govern the peripheral sex glands: the testicles in males. GnRH secretion is of two distinct types: pulsatile and cascade (180), but the latter occurs only in females. The pulsatile mode refers to the episodic release of GnRH through distinct pulsations of GnRH secretion into the circulation portal, while GnRH levels are undetectable during the inter-pulse intervals (181). The precise neuroendocrine detonator of puberty is GnRH which, in turn, is stimulated by kisspeptin (KISS1) (39). During this process, the HHT axis is characterized by a marked increase in the secretion of pulsations of LH that stimulate the testicles to secrete T (39). As puberty advances, the gonadotropins begin to secrete continuously, bringing sexual development to completion (39). KISS1 was identified in 2003 as a 54-amino acid codified peptide and, with its receptor (KISS1R, or GPR54), is now recognized as the key actor in the HHT axis (182). KISS1 is synthesized and secreted by hypothalamic nuclei in the arcuate nucleus and anteroventral periventricular nucleus (183). There

are reports that stimulation of the GnRH neurons by KISS1 activates the hypothalamic-hypophysis-gonad (HHG) axis and secretion of gonadotrophins in the hypophysis (184). We also know that the expression of KISS1 and its receptor increases during puberty in rodents and primates (185, 186). Therefore, administering KISS1 in animals –both males and females– can stimulate the secretion of GnRH and gonadotropins (187).

The rat is a useful animal for studying sexual development, and one widely used to analyze the mechanisms that underlie the process of sexual maturation, which are quite similar across numerous species and so can be extrapolated to other animals, including humans. The mechanisms involved in sexual development include the onset of gonadotropin secretion, the action mechanisms of the sex steroids, and positive and negative feedback circuits (188). In male rats, puberty is considered the stage in which fertility becomes evident, which occurs between days 42 and 49 of postnatal life (188, 189). This phase is associated with behavioral events like genital grooming (GG), which consists of licking the testicles, penis and anogenital region. This behavior intensifies between 46 and 48 days of age (190-192), and its frequency is regulated by LH and T levels (193).

Turning now to the context of male sexual behavior, studies have described that for intromissions to occur, there must be penile reflexes that include erections and dorsiflexion of the penis, described as fast whipping (190, 194). This also requires the formation of a cup in the penis (*i.e.*, the broadening of the glans penis associated with the moment of ejaculation). It is well-known that in prepubescent male rats there is a close relation between circulating T concentrations and the development of penile reflexes (195, 196). Another marker of puberty in males is preputial separation (PS), which occurs in the peripubertal stage, and becomes evident when the prepuce begins to gradually retract from day 41 to day 47 of life (192, 197).

There are reports that peripubertal exposure to Cd affects the functioning of the HHT axis by reducing the weight of the male sex organs –

testicles, epididymis, seminal vesicle, prostate– and decreasing blood T levels in adulthood (132). We also know that prenatal administration of Cd affects some puberty markers in male rats, and studies report that administering high doses of CdCl₂ (10 and 20 mg/Kg) to gestating mothers delays the descent of the testicles in their offspring (198), although other reports have found that maternal exposure to Cd advances this parameter (199). Several other puberty markers, however, remain to be evaluated, including PS and GG, so that exposure to Cd in the perinatal stage could still be found to affect the onset of puberty. Studies in young humans living in contaminated zones of Italy found a delay in the onset of puberty accompanied by a decrease in testicular volume and low T levels (200). However, no one has yet analyzed whether these findings affect the first pulse of GnRH or KISS1 in this stage. In this regard, some studies have found that exposure to Cd and other contaminants increases the expression of KISS1 in the human placenta (201).

6. Cd AS A NEUROENDOCRINE DISRUPTOR

Recently, evidence has accumulated to indicate that certain heavy metals –including Cd– can be considered endocrine disruptors (ED) (202). ED are compounds –natural or synthetic– that interfere with biosynthesis, metabolism, or the action of endogenous hormones. Numerous chemical substances belong to this category, including Cd. Most studies show that ED can imitate the activity of endogenous hormones and so reproduce equivalent effects. The biological effects of these compounds occur through a genomic action mechanism; that is, they act as hormonal agonists for a specific receptor (203). However, there are also clear indications that non-genomic action mechanisms are present and fully capable of altering, or at least affecting, the synthesis, transport and availability of endogenous hormones (203). Also, ED blocks the activity of hormones by competing for receptors or affecting the physiological concentration of a specific one (203). There is disagreement in the scientific community regarding evaluations of the possible risks of exposure to ED. Several researchers believe that this is harmful to health because ED can contribute significantly to the development of breast, ovarian, or

testicle and prostate cancer (204-208). Others, however, sustain that additional studies are needed to adequately analyze whether exposure to ED can produce adverse effects on human health (209-211). The effects of both ED and Cd, however, do include inhibition of the steroidogenic enzymes and activation of estrogen receptors (ER) or AR, as will be described in greater detail below. Cd is included among the ED because it affects the synthesis and/or regulation of various hormones, including LH and FSH (212, 213). Cd also affects the synthesis of progesterone in JC-410 porcine granulosa cells and activates estrogen receptor alpha (ER α) and/or mimics its estrogenic effect in various tissues, such as the uterus and mammary gland, as well as in breast cancer cell lines (214-216).

6.1. Estrogenic-type effects of Cd

In recent years, we have learned that many environmental compounds mimic the physiological activity of estrogens. These include xenoestrogens, phytoestrogens and metalloestrogens (217). Cd is a predominant environmental contaminant that is reported to have effects as a potent metalloestrogen (218) which mimics the activity of the estrogenic hormones (219-222) by bonding to the nuclear-type ERs denominated ER α and estrogen receptor beta (ER β) (216, 220, 223-229), and to the transmembrane estrogen receptor GPR30 (228, 230). ER α and ER β are codified by distinct genes and their expression varies with the type of tissue involved. ER α is expressed predominantly in reproductive organs like the uterus, breast and ovaries, as well as in the liver and central nervous system, while the β form is expressed mainly in other tissues, including bone, the endothelium, the lung and the urogenital tract, but also the ovaries, the central nervous system and the prostate (231, 232). In the absence of the hormone, ER α is sequestered in an inactive compound and repressed by such molecular chaperones as thermal shock proteins. But the union with the hormone induces a change in the conformation of the ligand-binding domain that releases the receptor from the inactive compound and, as a result, eliminates the masking effect of the ligand-binding domain. The receptor is dimerized and later bonds to the response elements of the DNA in a complex process that requires recruiting coactivators

of steroid receptors (SRC-1, SRC-2 and SRC-3) (233). Once bonded, the coactivators recruit the co-integrator p300/CBP (CREB-binding protein). The co-regulator compound then stimulates transcription by remodeling the chromatin through its ability to acetylate histones and interact with the machinery of basal transcription (234).

Using transitory transfection assays to block the union of 17 β estradiol to ER α in a non-competitive manner, studies have demonstrated that Cd activates ER α (219, 220, 225, 227). This suggests that it interacts with the receptor's ligand-binding domain. The amino acids cys381, cys447, glu523, his524 and asp538 have been identified as possible interaction sites of Cd with the ER α 's ligand-binding domain (235). These amino acids, which play a role in the interaction of Cd with ER α , are located in the H4, H8, H11 helices and the interface of the loop with the H12 helix. In this way, Cd's interaction with different amino acids can promote localized folding, as in the case of the zinc finger domain, or the assemblage of different regions of the protein in a domain (235). Finally, the effects of Cd can be blocked by an anti-estrogen, suggesting that this metal's effects are mediated by the genomic ER α pathway and activate the non-genomic ER α pathway through ERK1/2 and Akt (216, 228).

Like E₂, Cd induces cellular proliferation (216, 219, 225) and increases the transcription and expression of estrogen-regulated genes, such as the progesterone receptor (PR) (216, 219, 225). In fact, in studies with rodents Cd begins to mimic the effects of E₂ in target organs (236) after just one intraperitoneal injection of 5 μ g/kg of CdCl₂. In ovariectomized female Sprague-Dawley rats, an increase in the net uterine weight was observed, accompanied by a proliferative response by the endometrium, which promotes the growth and development of the mammary glands and induces hormone-regulated genes. Also, the female descendants of those rats experienced early onset puberty (236).

Various reports indicate that Cd has a potential role in the development of hormone-dependent cancers (218, 219, 221, 222, 230). For example, *in utero* exposure to Cd increases the risk

of developing breast cancer. Parodi *et al.* (237) observed that treating pregnant female rats with Cd altered the development of the mammary gland before the onset of puberty in female offspring by increasing the number of mother/progenitor cells, cell density and ER α expression, all of which increase the risk of developing breast cancer. Finally, *in vitro* tests have demonstrated that Cd has proliferative action in human prostate cells through an estrogen-dependent mechanism that increases ER α and ER β expression independently of the androgens (238).

6.2. Androgenic-type effects of Cd

Other research has shown that prostate tumors can be induced experimentally by oral exposure to Cd (157). While some important epidemiological reports indicate a relation between Cd exposure and rates of prostate cancer, their findings have been refuted (239). Over the past decade, research has shown that Cd has potent androgenic-type activities both *in vivo* and *in vitro* by bonding directly to AR. In transfection assays, Cd activates a chimera that contains the ligand-binding domain of AR (240), allowing it to bond to the ligand-binding domain with a high degree of affinity (240, 241) and to block androgen's union to the receptor. Moreover, it can compete with DHT, the natural ligand of AR (240). The union of Cd with AR modifies the conformation of the receptor (242), and so could change its transcription potential. Scatchard analysis demonstrated that Cd binds to AR with a dissociation constant in an equilibrium of 1.19×10^{-10} M. Work with cell lines has shown changes in AR activity or AR expression when cells are exposed to Cd (240, 243-245). Epidemiological studies evaluating the relation between Cd and AR have analyzed blood, toenails and urine, exposure to Cd and indirect measurements of the function of AR, such as the level of the specific prostate antigen in serum (246-249). To date, however, these approaches have produced only mixed results, while failing to confirm any clear interaction between Cd exposure and AR signaling in the human prostate. Evidence from basic scientific studies suggests that Cd may play a role in prostate cancer by interrupting AR, a hormone-activated transcription factor that is the key driver of the progression of cancer in that

organ (250) although, ironically, AR is necessary for normal prostate growth and development. Finally, research has demonstrated that prostate tumors can be induced experimentally by oral exposure to Cd (251).

6.3. Testosterone synthesis

Testosterone (4-androstenol, 14-ol, 3-one) is a steroid hormone belonging to the family of the androgens, with a structure derived from cyclopentanoperhydrophenanthrene. It is composed of 19 carbon atoms, with methyl groups at carbons 10 and 13, and a hydroxyl group at carbon 17 (252). This hormone is synthesized from cholesterol in the Leydig cells in the testicles (253). The biosynthesis of T is subject to short-term regulation controlled by LH, which binds to specific receptors on the surface of the Leydig cells and stimulates cAMP production. cAMP performs two principle activities in controlling steroidogenesis in the Leydig cells. The first is to acutely stimulate the biosynthesis of T by translocating cholesterol from the external mitochondrial membrane towards the internal one, which generates the synthesis of pregnenolone through the action of the cytochrome enzyme P450_{scc}/CYP11A1 (cholesterol side cleavage enzyme), which is found in the internal mitochondrial membrane. cAMP's second action is to activate A kinase (PKA) that, through the phosphorylation of transcription factors related to the activation of genes that codify for steroidogenic enzymes, phosphorylates proteins involved in transporting cholesterol towards the mitochondria, as is the case of the StAR protein (steroidogenic acute regulatory protein) (254). Pregnenolone is biotransformed into progesterone by the 3 β hydroxysteroid dehydrogenase Δ 4-5 isomerase enzyme (3 β -HSD) (252). Then the progesterone obtained through the Δ 4 pathway is transformed into androstenedione by the enzymatic compound 17 α hydroxylase C17-20 lyase, which is dependent on cytochrome P-450 (P-450_{c17}). The final step in T synthesis is regulated by the microsomal enzyme 17 β -hydroxysteroid dehydrogenase (17 β HSD), which catalyzes the conversion of androstenedione into T. The endogenous production of steroids in the Leydig cells regulates the expression of the enzymes involved in T biosynthesis (253).

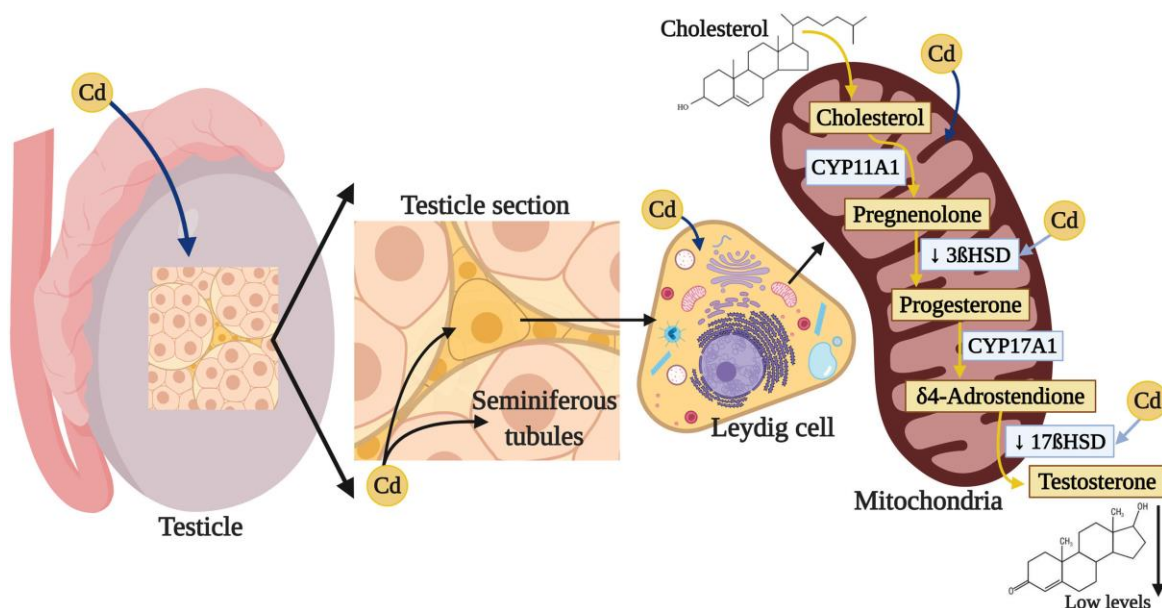


Figure 3. Damage of cadmium in testicular steroidogenesis. Cadmium can affect various enzymes that participate in T synthesis by reducing the expression and activity of 3β-HSD and 17β-HSD, which interfere in steroidogenesis, specially in the Leydig cells, the result of this process is a decrease in T.

Multiple studies show that exposure to Cd can affect various enzymes that participate in T synthesis; for example, chronic exposure to CdCl₂ in adult male rats increased testicular cholesterol (255) but, in contrast, decreased expression of the ARNm of class B type I Scavenger receptor (SR-BI), which facilitates the capture of cholesterol esters (256) from the StAR protein (257, 258), since Cd increases the activation of the cAMP-responsive element-binding protein (CREB) transcription factor (258). In addition, Cd reduces the expression and activity of the 3β-HSD and 17β-HSD enzymes (255, 259), while decreasing the expression ARNm by the CYP11A1, 17α-hydroxylase and 11β-hydroxysteroid dehydrogenase enzymes (11β-HSD), which intervene in steroidogenesis in the Leydig cells (256) (Figure 3). The result of this complex process is a decrease in T synthesis that has been amply reported after exposure to Cd (132).

7. IMPACT OF CADMIUM ON THE IMMUNE AND MALE REPRODUCTIVE SYSTEMS

A considerable amount of information has been amassed indicating that the immune system can have profound effects on the hypothalamic-

hypophysis-testis axis. Figure 4 summarizes the principle affectations of the Cd on the HHT axis, and the effects of it related to the release of cytokines from a variety of activated immune cells that can regulate the secretion of GnRH and LH or have direct actions on Leydig and Sertoli cells (260-263). Cytokines that play important regulatory roles in the normal functioning of the HHT axis include the proinflammatory cytokines tumor necrosis factor-α (TNF-α), interleukin-1β (IL-1 β), and interleukin-6 (IL-6). TNF-α is released from a variety of cell types, including macrophages, and is a marker of systemic and local inflammation in smokers (264). Moreover, a recent study reports that these three cytokines cause a dose-dependent decline in steroidogenesis in the TM3 mouse Leydig cell line (265); whereas TNF-α infusion into hypothalamic fetal cells in culture decreases GnRH secretion (266). Research has also shown that chronic low-dose cadmium exposure in rats can significantly increase the interferon gamma (IFNγ) and interleukin-10 (IL-10) levels, suggesting that Cd may enhance inflammatory responses (267). It is well-known that inflammation is also associated with oxidative stress and can impair male reproductive functions (268-270).

Cadmium exposure and male reproductive functions

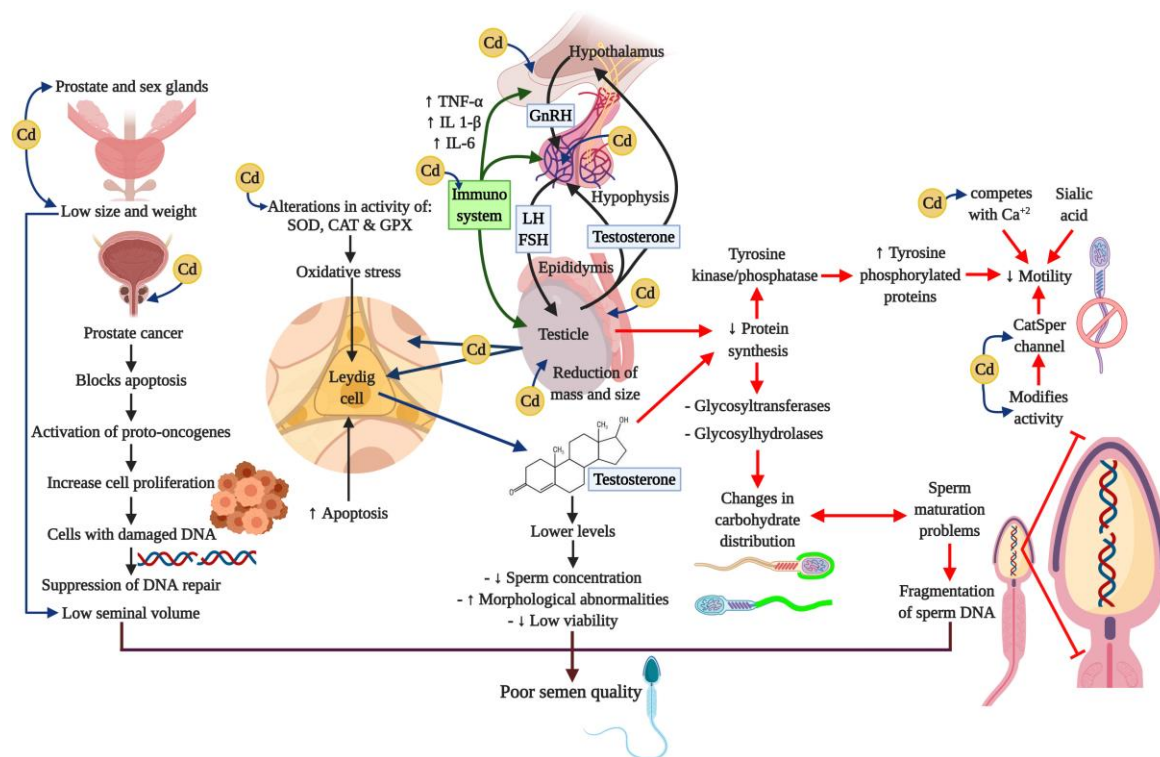


Figure 4. Cadmium and negative impacts on neuroendocrine axis. Cadmium alters the oxidative stress, the signaling pathways, and interrupts neuroendocrine processes. These actions cause serious damages in male reproductive organs such as the prostate, the testis and the epididymis, and prevents processes like steroidogenesis, spermatogenesis, and epididymal maturation. Besides, Cd can induce adverse effects in immune system through the cytokines. All these alterations may compromise the reproductive capacity.

Adverse effects of Cd exposure induce testicular dysfunction, inhibition of spermatogenesis, and male infertility. This phenomenon is attributed to activation of the “inflammasome” and augments the inflammatory response, an intracellular multiprotein complex, which is activated by tissue damage induced by Cd (271). Several studies have reported that Cd exposure increased TNF- α , IL-6 and IL1 β generation in testicular tissue. When adult male mice were injected intraperitoneally with CdCl₂ at a dose of 2 mg/kg body weight per day for seven consecutive days, Cd-treatment increased IL-1 β and TNF- α concentrations in a statistically-significant manner, though this effect was precluded by incubation with melatonin, thus decreasing oxidative damage to the sperm (272). Similar results were reported recently in rat and chicken testes. In the first experiment, male rats received only one injection of CdCl₂ (2 mg/kg, i.p.) on day 9 and TNF- α increased 2-fold; while the administration of a potent anti-inflammatory

compound (Diacerein) inhibited the Cd-induced effect (271). In the latter report, chickens were fed with normal full-fodder with 140 mg/kg of CdCl₂, and their TNF- α , IL-1 β , and IL-6 mRNA levels were significantly higher in the Cd group. In those experiments, levels of an inflammatory factor were precluded with Ganoderma, a medicinal mushroom that has enhanced anti-inflammatory effects (273).

Taken together, these studies suggest that exposure to Cd induces the inflammatory response by activating the “inflammasome” in different cells of the male reproductive system.

8. CONCLUSION

Cd is a toxic agent present in the environment that can have negative impacts on the HHT axis through diverse toxicity mechanisms, including oxidative stress, inhibition of the Ca channels, alterations of the signaling

pathways, and immuno-endocrine interruption. These actions cause the primary male reproductive organ –the testicle– to lose such key functions as steroidogenesis, spermatogenesis, and epididymal maturation. Moreover, Cd can exert adverse effects on the HHT axis through the activation of the “inflammasome” in different cells of the male reproductive system, modifying the onset of puberty, the development of sexual maturity, adult sexual behavior and even fertility; thus compromising the reproductive capacity of individuals so affected.

9. ACKNOWLEDGMENTS

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10. REFERENCES

1. S.A. Covarrubias, J.J. Peña-Cabriales: Contaminación ambiental por metales pesados en México. *Rev Int Contam Ambie* 33, 7-21 (2017). DOI: 10.20937/RICA.2017.33.esp01.01
2. L. Torres-Sánchez, R.A. Vázquez-Salas, A. Vite, M. Galván-Portillo, M.E. Cebrián, A.P. Macías-Jiménez, C. Ríos, S. Montes: Blood cadmium determinants among males over forty living in Mexico City. *Sci Total Environ* 637-638, 686-694 (2018). DOI: 10.1016/j.scitotenv.2018.04.371
3. J. Ramos-Treviño, S. Bassol-Mayagoitia, J.A. Hernández-Ibarra, P. Ruiz-Flores, M.P. Nava-Hernández: Toxic Effect of Cadmium, Lead, and Arsenic on the Sertoli Cell: Mechanisms of Damage Involved. *DNA Cell Biol* 37, 600-608 (2018). DOI: 10.1089/dna.2017.4081
4. J.J. Wirth, R.S. Mijal: Adverse effects of

low level heavy metal exposure on male reproductive function. *Syst Biol Reprod Med* 56(2), 147-167 (2010). DOI: 10.3109/19396360903582216

5. H.X. Geng, L. Wang: Cadmium: Toxic effects on placental and embryonic development. *Environ Toxicol Pharmacol* 67, 102-107 (2019). DOI: 10.1016/j.etap.2019.02.006
6. M.N. Rana, J. Tangpong, M.M. Rahman: Toxicodynamics of Lead, Cadmium, Mercury and Arsenic- induced kidney toxicity and treatment strategy: A mini review. *Toxicol Rep* 5, 704-713 (2018). DOI: 10.1016/j.toxrep.2018.05.012
7. H. Zhang, M. Reynolds: Cadmium exposure in living organisms: A short review. *Sci Total Environ* 678, 761-767 (2019). DOI: 10.1016/j.scitotenv.2019.04.395
8. Y. Zang, B. Devleesschauwer, P.M. Bolger, E. Goodman, H.J. Gibb: Global burden of late-stage chronic kidney disease resulting from dietary exposure to cadmium, 2015. *Environ Res* 169, 72-78 (2019). DOI: 10.1016/j.envres.2018.10.005
9. WHO (World Health Organization): Exposure to cadmium: a major public health concern, Public Health and Environment WHO. 20 Avenue Appia, 1211 Geneva 27, Switzerland, (2010).
10. ATSDR (Agency for Toxic Substances and Disease Registry): Toxicological profile for Cadmium, Atlanta, GA: U.S. Department of Health and Human Services, Public Health Service, (2012).
11. P. Richter, O. Faroon, R.S. Pappas: Cadmium and Cadmium/Zinc Ratios and

- Tobacco-Related Morbidities. *Int J Environ Res Public Health* 14(10), 1-19 (2017).
DOI: 10.3390/ijerph14101154
12. N. Pant, A.K. Banerjee, S. Pandey, N. Mathur, D.K. Saxena, S.P. Srivastava: Correlation of lead and cadmium in human seminal plasma with seminal vesicle and prostatic markers. *Hum Exp Toxicol* 22(3), 125-128 (2003).
DOI: 10.1191/0960327103ht336oa
13. L.L. Zhao, Y.F. Ru, M. Liu, J.N. Tang, J.F. Zheng, B. Wu Y.H. Gu, H.J. Shi: Reproductive effects of cadmium on sperm function and early embryonic development *in vitro*. *PLoS One* 12, 1-12 (2017).
DOI: 10.1371/journal.pone.0186727
14. G. Bertin, D. Averbeck: Cadmium: cellular effects, modifications of biomolecules, modulation of DNA repair and genotoxic consequences (a review). *Biochimie* 88(11), 1549-1559 (2006).
DOI: 10.1016/j.biochi.2006.10.001
15. A. Rani, A. Kumar, A. Lal, M. Pant: Cellular mechanisms of cadmium-induced toxicity: a review. *Int J Environ Health Res* 24(4), 378-399 (2014).
DOI: 10.1080/09603123.2013.835032
16. S. Azam, G.S. Louis, J. Miksovská: Cadmium association with DREAM promotes DREAM interactions with intracellular partners in a similar manner to its physiological ligand, calcium. *Metallomics* 11, 1115-1127 (2019).
DOI: 10.1039/c9mt00059c
17. K.I. Ohba: Transport and Toxicity of Cadmium. *Nihon Eiseigaku Zasshi* 73(3), 269-274 (2018).
DOI: 10.1265/jjh.73.269
18. T.N. Gerasimenko, N.V. Senyavina, N.U. Anisimov, S.A. Tonevitskaya: A Model of Cadmium Uptake and Transport in Caco-2 Cells. *Bull Exp Biol Med* 161(1), 187-192 (2016).
DOI: 10.1007/s10517-016-3373-7
19. J.S. Klinck, C.M. Wood: Gastro-intestinal transport of calcium and cadmium in fresh water and seawater acclimated trout (*Oncorhynchus mykiss*). *Comp Biochem Physiol C Toxicol Pharmacol* 157(2), 236-250 (2013).
DOI: 10.1016/j.cbpc.2012.11.006
20. J. Breton, K. LE Clère, C. Daniel, M. Sauty, L. Nakab, T. Chassat, J. Dewulf, S. Penet, C. Carnoy, P. Thomas, B. Pot, F. Nessler, B. Foligné: Chronic ingestion of cadmium and lead alters the bioavailability of essential and heavy metals, gene expression pathways and genotoxicity in mouse intestine. *Arch Toxicol* 87(10), 1787-1795 (2013).
DOI: 10.1007/s00204-013-1032-6
21. H. Fujishiro, S. Hamano, R. Tanaka, T. Kambe, S. Himeno: Concentration-dependent roles of DMT1 and ZIP14 in cadmium absorption in Caco-2 cells. *J Toxicol Sci* 42(5), 559-567 (2017).
DOI: 10.2131/jts.42.559
22. G. Kovacs, N. Montalbetti, M.C. Franz, S. Graeter, A. Simonin, M.A. Hediger: Human TRPV5 and TRPV6: key players in cadmium and zinc toxicity. *Cell Calcium* 54(4), 276-286 (2013).
DOI: 10.1016/j.ceca.2013.07.003
23. C. de Angelis, M. Galdiero, C. Pivonello, C. Salzano, D. Gianfrilli, P. Piscitelli, A. Lenzi, A. Colao, R. Pivonello: The environment and male reproduction: The effect of cadmium exposure on reproductive function and its implication

- in fertility. *Reprod Toxicol* 73, 105-127 (2017).
DOI: 10.1016/j.reprotox.2017.07.021
24. E. Garza-López, J.C. Chávez, C. Santana-Calvo, I. López-González, T. Nishigaki: Cd(2+) sensitivity and permeability of a low voltage-activated Ca(2+) channel with CatSper-like selectivity filter. *Cell Calcium* 60(1), 41-50 (2016).
DOI: 10.1016/j.ceca.2016.03.011
25. J. Fels, B. Scharner, R. Zarbock, I.P. Zavala Guevara, W.K. Lee, O.C. Barbier, F. Thévenod: Cadmium Complexed with β 2-Microglobulin, Albumin and Lipocalin-2 rather than Metallothionein Cause Megalin: Cubilin Dependent Toxicity of the Renal Proximal Tubule. *Int J Mol Sci* 20(10), 1-15 (2019).
DOI: 10.3390/ijms20102379
26. M.K. Driessnack, A. Jamwal, S. Niyogi: Effects of chronic waterborne cadmium and zinc interactions on tissue-specific metal accumulation and reproduction in fathead minnow (*Pimephales promelas*). *Ecotoxicol Environ Saf* 140, 65-75 (2017).
DOI: 10.1016/j.ecoenv.2017.02.023
27. T. Hamada, A. Tanimoto, Y. Sasaguri: Apoptosis induced by cadmium. *Apoptosis* 2(4), 359-367 (1997).
DOI: 10.1023/a:1026401506914
28. S. Wang, X. Ren, X. Hu, L. Zhou, C. Zhang, M. Zhang: Cadmium-induced apoptosis through reactive oxygen species-mediated mitochondrial oxidative stress and the JNK signaling pathway in TM3 cells, a model of mouse Leydig cells. *Toxicol Appl Pharmacol* 368, 37-48 (2019).
DOI: 10.1016/j.taap.2019.02.012
29. F. Thévenod, W.K. Lee: Cadmium and cellular signaling cascades: interactions between cell death and survival pathways. *Arch Toxicol* 87(10), 1743-1786 (2013).
DOI: 10.1007/s00204-013-1110-9
30. T. Liu, W. He, C. Yan, Y. Qi, Y. Zhang: Roles of reactive oxygen species and mitochondria in cadmium-induced injury of liver cells. *Toxicol Ind Health* 27(3), 249-256 (2011).
DOI: 10.1177/0748233710386408
31. M.D.C. Cupertino, R.D. Novaes, E.C. Santos, D.S.S. Bastos, D.C. Marques Dos Santos, M. do Carmo Queiroz Fialho, S.L.P.D. Matta: Cadmium-induced testicular damage is associated with mineral imbalance, increased antioxidant enzymes activity and protein oxidation in rats. *Life Sci* 175, 23-30 (2017).
DOI: 10.1016/j.lfs.2017.03.007
32. W.C. Prozialeck, P.C. Lamar, S.M. Lynch: Cadmium alters the localization of N-cadherin, E-cadherin, and beta-catenin in the proximal tubule epithelium. *Toxicol Appl Pharmacol* 189(3), 180-195 (2003).
DOI: 10.1016/s0041-008x(03)00130-3
33. Z. Wei, Z. Shan, Z.A. Shaikh: Epithelial-mesenchymal transition in breast epithelial cells treated with cadmium and the role of Snail. *Toxicol Appl Pharmacol* 344, 46-55 (2018).
DOI: 10.1016/j.taap.2018.02.022
34. L.M. Bilezikjian, A.L. Blount, A.M. Leal, C.J. Donaldson, W.H. Fischer, W.W. Vale: Autocrine/paracrine regulation of pituitary function by activin, inhibin and follistatin. *Mol Cell Endocrinol* 225(1-2), 29-36 (2004).
DOI: 10.1016/j.mce.2004.02.010

35. A. Lafuente, N. Márquez, M. Pérez-Lorenzo, D. Pazo, A.I. Esquifino: Cadmium effects on hypothalamic-pituitary-testicular axis in male rats. *Exp Biol Med (Maywood)* 226(6), 605-611 (2001).
DOI: 10.1177/153537020122600615
36. B. Halász. Anatomy of Hypothalamus. In: Encyclopedia of Endocrine Diseases. Eds: I. Huhtaniemi, L. Martini, Academic Press Oxford, UK. Second Edition, 707-715 (2004).
37. C. Teinturier: Mécanismes neuroendocriniens de la maturation pubertaire Neurobiological mechanisms of the onset of puberty. *Gynecol Obstet Fertil* 30(10), 809-813 (2002).
DOI: 10.1016/S1297-9589(02)00450-2
38. E. Terasawa, J.R. Kurian: Neuroendocrine Mechanism of Puberty. In: Handbook of Neuroendocrinology. Eds: G. Fink, D.W. Pfaff, J.E. Levine, Academic Press Oxford, UK. First Edition, 433-484 (2012).
DOI: 10.1016/C2009-0-04284-6
39. P.F. Corradi, R.B. Corradi, L.W. Greene: Physiology of the Hypothalamic Pituitary Gonadal Axis in the Male. *Urol Clin North Am* 43(2), 151-162 (2016).
DOI: 10.1016/j.ucl.2016.01.001
40. D.J. Bernard, J. Fortin, Y. Wang, P. Lamba: Mechanisms of FSH synthesis: what we know, what we don't, and why you should care. *Fertil Steril* 93(8), 2465-2485 (2010).
DOI: 10.1016/j.fertnstert.2010.03.034
41. U.B. Kaiser. Gonadotropin Hormones. In: The Pituitary. Eds: M. Shlomo, ELSEVIER Inc., London, England, 205-260 (2011).
42. M. Simoni, J. Gromoll, E. Nieschlag: The Follicle-Stimulating Hormone Receptor: Biochemistry, Molecular Biology, Physiology, and Pathophysiology. *Endocr Rev* 18(6), 739-773 (1997).
DOI: 10.1210/edrv.18.6.0320
43. A. Lafuente, A. González-Carracedo, A. Romero, A.I. Esquifino: Effect of cadmium on 24-h variations in hypothalamic dopamine and serotonin metabolism in adult male rats. *Exp Brain Res* 149(2), 200-206 (2003).
DOI: 10.1007/s00221-002-1356-6
44. A.M. Calderoni, L. Oliveros, G. Jahn, R. Anton, J. Luco, M.S. Giménez: Alterations in the lipid content of pituitary gland and serum prolactin and growth hormone in cadmium treated rats. *Biometals* 18(3), 213-220 (2005).
DOI: 10.1007/s10534-005-0581-4
45. A. Caride, B. Fernández-Pérez, T. Cabaleiro, M. Tarasco, A.I. Esquifino, A. Lafuente: Cadmium chronotoxicity at pituitary level: effects on plasma ACTH, GH, and TSH daily pattern. *J Physiol Biochem* 66(3):213-220 (2010).
DOI: 10.1007/s13105-010-0027-5
46. Z.H. Li, L. Chen, Y.H. Wu, P. Li, Y.F. Li, Z.H. Ni: Effects of waterborne cadmium on thyroid hormone levels and related gene expression in Chinese rare minnow larvae. *Comp Biochem Physiol C Toxicol Pharmacol* 161:53-57 (2014).
DOI: 10.1016/j.cbpc.2014.02.001
47. A. Vetillard, T. Bailhache: Cadmium: an endocrine disrupter that affects gene expression in the liver and brain of juvenile rainbow trout. *Biol Reprod* 72(1):119-126 (2005).
DOI: 10.1095/biolreprod.104.029520

48. C.J. Martyniuk, B.C. Sanchez, N.J. Szabo, N.D. Denslow, M.S. Sepúlveda: Aquatic contaminants alter genes involved in neurotransmitter synthesis and gonadotropin release in largemouth bass. *Aquat Toxicol* 95(1), 1-9 (2009). DOI: 10.1016/j.aquatox.2009.06.009
49. V.U. Nna, U.Z. Usman, E.O. Ofutet, D.U. Owu: Quercetin exerts preventive, ameliorative and prophylactic effects on cadmium chloride - induced oxidative stress in the uterus and ovaries of female Wistar rats. *Food Chem Toxicol* 102, 143-155 (2017). DOI: 10.1016/j.fct.2017.02.010
50. M. Ciarrocca, A. Capozzella, F. Tomei, G. Tomei, T. Caciari: Exposure to cadmium in male urban and rural workers and effects on FSH, LH and testosterone. *Chemosphere* 90 (7), 2077–2084 (2013). DOI: 10.1016/j.chemosphere.-2012.10.060
51. X. Zeng, T. Lin, Y. Zhou, Q. Kong: Alterations of serum hormone levels in male workers occupationally exposed to cadmium, *J Toxicol Environ Health A* 65(7), 513-521 (2002). DOI: 10.1080/15287390252807975
52. S.A. Ronchetti, G.V. Novack, M.S. Bianchi, M.C. Crocco, B.H. Duvilanski, J.P. Cabilla: *In vivo* xenoestrogenic actions of cadmium and arsenic in anterior pituitary and uterus. *Reproduction* 152(1), 1-10 (2016). DOI: 10.1530/rep-16-0115
53. O. Akinloye, A.O. Arowojolu, O.B. Shittu, J.I. Anetor: Cadmium toxicity: a possible cause of male infertility in Nigeria. *Reprod Biol* 6(1), 17-30 (2006).
54. J. Mendiola, J.M. Moreno, M. Roca, N. Vergara-Juarez, M.J. Martinez-Garcia, A. Garcia-Sanchez, B. Elvira-Rendueles, S. Moreno-Grau, J.J. López-Espin, J. Ten, R. Bernabeu, A.M. Torres-Cantero: Relationships between heavy metal concentrations in three different body fluids and male reproductive. *Environ Health* 10(6), 1-7 (2011). DOI: 10.1186/1476-069x-10-6
55. S. Benoff, I.R. Hurley, M. Barcia, F.S. Mandel, G.W. Cooper, A. Hershlag: A potential role for cadmium in the etiology of varicocele-associated infertility. *Fertil Steril* 67(2), 336-347 (1997). DOI: 10.1016/s0015-0282(97)81921-8
56. X. Zeng, T. Jin, J.P. Buchet, X. Jiang, Q. Kong, T. Ye, A. Bernard, G.F. Nordberg: Impact of cadmium exposure on male sex hormones: a population-based study in China. *Environ Res* 96(3), 338-344 (2004). DOI: 10.1016/j.envres.2004.02.004
57. A. Lafuente, A. Gonzalez-Carracedo, A. Romero, P. Cano, A.I. Esquifino: Cadmium exposure differentially modifies the circadian patterns of norepinephrine at the median eminence and plasma LH, FSH and testosterone levels. *Toxicol Lett* 146(2), 175-182 (2004). DOI: 10.1016/j.toxlet.2003.10.004
58. M.M. Ahmed, S.A. El-Shazly, M.E. Alkafafy, A.A. Mohamed, A.A. Mousa: Protective potential of royal jelly against cadmium-induced infertility in male rats. *Andrologia* 50(5), 1-12 (2018). DOI: 10.1111/and.12996
59. A. Lafuente, N. Márquez, S. Piquero, A.I. Esquifino: Cadmium affects the episodic luteinizing hormone secretion in male rats: possible age-dependent effects. *Toxicol Lett* 104(1-2), 27-33 (1999).

- DOI: 10.1016/s0378-4274(98)00349-x
60. A.H. Poliandri, J.P. Cabilla, M.O. Velardez, C.C. Bodo, B.H. Duvilanski: Cadmium induces apoptosis in anterior pituitary cells that can be reversed by treatment with antioxidants. *Toxicol Appl Pharmacol* 190(1), 17-24 (2003). DOI: 10.1016/s0041-008x(03)00191-1
 61. S. Li, G. Pelletier: Involvement of serotonin in the regulation of GnRH gene expression in the male rat brain. *Neuropeptides* 29(1), 21-25 (1995). DOI: 10.1016/0143-4179(95)90052-7
 62. X. Liu, A.E. Herbison: Dopamine regulation of gonadotropin-releasing hormone neuron excitability in male and female mice. *Endocrinology* 154(1), 340-350 (2013). DOI: 10.1210/en.2012-1602
 63. A. Ezzat, A. Pereira, I.J. Clarke: Kisspeptin is a component of the pulse generator for GnRH secretion in female sheep but not the pulse generator. *Endocrinology* 156(5), 1828-1837 (2015). DOI: 10.1210/en.2014-1756
 64. K.J. Iremonger, S. Constantin, X. Liu, A.E. Herbison: Glutamate regulation of GnRH neuron excitability. *Brain Res* 1364, 35-43 (2010). DOI: 10.1016/j.brainres.2010.08.071
 65. T.J. Cicero, E.R. Meyer, B.D. Bell: Characterization and possible opioid modulation of N-methyl-D-aspartic acid induced increases in serum luteinizing hormone levels in the developing male rat. *Life Sci* 42(18), 1725-1732 (1988). DOI: 10.1016/0024-3205(88)90038-0
 66. R. Medhamurthy, V.L. Gay, T.M. Plant: Repetitive injections of L-glutamic acid, in contrast to those of N-methyl-D,L-aspartic acid, fail to elicit sustained hypothalamic GnRH release in the prepubertal male rhesus monkey (*Macaca mulatta*). *Neuroendocrinology* 55(6), 660-66 (1992). DOI: 10.1159/000126186
 67. J.J. Bonavera, R.S. Swerdloff, A.P. Sinha Hakim, Y.H. Lue, C. Wang: Aging results in attenuated gonadotropin releasing hormone-luteinizing hormone axis responsiveness to glutamate receptor agonist N-methyl-D-aspartate. *J Neuroendocrinol* 10(2), 93-99 (1998). DOI: 10.1046/j.1365-2826.1998.00177.x
 68. S. Tzanoulinou, O. Riccio, M.W. de Boer, C. Sandi: Peripubertal stress-induced behavioral changes are associated with altered expression of genes involved in excitation and inhibition in the amygdala. *Transl Psychiatry* 4(7), 1-9 (2014). DOI: 10.1038/tp.2014.54
 69. M. Watanabe, A. Fukuda, J. Nabekura: The role of GABA in the regulation of GnRH neurons. *Front Neurosci* 8, 1-9 (2014). DOI: 10.3389/fnins.2014.00387
 70. J.A. Sim, M.J. Skynner, J.R. Pape, A.E. Herbison: Late postnatal reorganization of GABA(A) receptor signalling in native GnRH neurons. *Eur J Neurosci* 12(10), 3497-3504 (2000). DOI: 10.1046/j.1460-9568.2000.00261.x
 71. J.L. Temple, S. Wray: Developmental changes in GABA receptor subunit composition within the gonadotrophin-releasing hormone-1 neuronal system. *J Neuroendocrinol* 17(9), 591-599 (2005). DOI: 10.1111/j.1365-2826.2005.01348.x
 72. C. Zhang, M.A. Bosch, O.K. Rønnekleiv,

- M.J. Kelly: Gamma-aminobutyric acid B receptor mediated inhibition of gonadotropin-releasing hormone neurons is suppressed by kisspeptin-G protein-coupled receptor 54 signaling. *Endocrinology* 150(5), 2388–2394 (2009).
DOI: 10.1210%2Fen.2008-1313
73. C. Feleder, H. Jarry, S. Leonhardt, J.A. Moguilevsky, W. Wuttke: Evidence to suggest that gonadotropin-releasing hormone inhibits its own secretion by affecting hypothalamic amino acid neurotransmitter release. *Neuroendocrinology* 64(4), 298-304 (1996).
DOI: 10.1159/000127132
74. M. Gearing, E. Terasawa: The alpha-1-adrenergic neuronal system is involved in the pulsatile release of luteinizing hormone-releasing hormone in the ovariectomized female rhesus monkey. *Neuroendocrinology* 53(4), 373-381 (1991).
DOI: 10.1159/000125744
75. A. Krieger, W. Wuttke: Ontogeny of tyrosine hydroxylase and dopamine- β -hydroxylase activity in discrete limbic and hypothalamic structures of female rats. *Brain Res* 193(7), 181-188 (1980).
DOI: 10.1016/0006-8993(80)90955-5
76. A. Romero, A. Caride, N. Pereiro, A. Lafuente: Modulatory effects of melatonin on cadmium-induced changes in biogenic amines in rat hypothalamus. *Neurotox Res* 20(3), 240-249 (2011).
DOI: 10.1007/s12640-010-9237-4
77. B. Fernández-Pérez, A. Caride, T. Cabaleiro, A. Lafuente: Cadmium effects on 24h changes in glutamate, aspartate, glutamine, GABA and taurine content of rat striatum. *J Trace Elem Med Biol* 24(3), 212-218 (2010).
DOI: 10.1016/j.jtemb.2010.01.005
78. A. Lafuente, A.I. Esquifino: Possible role of glutamate, aspartate, glutamine, GABA or taurine on cadmium toxicity on the hypothalamic pituitary axis activity in adult male rats. *Biometals* 15(2), 183-187 (2002).
DOI: 10.1023/A:1015255406461
79. R.O. Greep, H.C. Fevold: The spermatogenic and secretory function of the gonads of hypophysectomised adult rats treated with pituitary FSH and LH. *Endocrinology* 21, 611-618 (1937).
DOI: 10.1210/endo-21-5-611
80. T.R. Kumar, Y. Wang, N. Lu, M.M. Matzuk: Follicle stimulating hormone is required for ovarian follicle maturation but not male fertility. *Nat Genet* 15(2), 201-204 (1997).
DOI: 10.1038/ng0297-201
81. M.B. Abel, A.N. Wootton, V. Wilkins, I. Huhtaniemi, P.G. Knight, H.M. Charlton: The effect of a null mutation in the follicle-stimulating hormone receptor gene on mouse reproduction. *Endocrinology* 141(5), 1795-1803 (2000).
DOI: 10.1210/endo.141.5.7456
82. E. Arenas-Ríos, M.A. León-Galván, A. Rosado-García. Espermatogénesis, maduración y almacenamiento epididimario de espermatozoides en mamíferos. In: Avances en la Biología de la Reproducción. Eds: S. Retana-Márquez. Ciudad de México. UAM-SEP, 12-42 (2012).
83. A. Stajn, R.V. Zikić, B. Ognjanović, Z.S. Saicić, S.Z. Pavlović, M.M. Kostić, V.M. Petrović: Effect of cadmium and selenium on the antioxidant defense system in rat

- kidneys. *Comp Biochem Physiol C Pharmacol Toxicol Endocrinol* 117(2), 167-172 (1997).
DOI: 10.1016/s0742-8413(97)00063-7
84. N. Khansakorn, W. Wongwit, P. Tharnpoophasiam, B. Hengprasith, L. Suwannathon, S. Chanprasertyothin, T. Sura, S. Kaojarern, P. Sritara, J. Sirivarasai: Genetic Variations of Glutathione S-Transferase Influence on Blood Cadmium Concentration. *J Toxicol* 2012, 1-6 (2012).
DOI: 10.1155%2F2012%2F356126
85. S. Jahan, A. Zahra, U. Irum, N. Iftikhar, H. Ullah: Protective effects of different antioxidants against cadmium induced oxidative damage in rat testis and prostate tissues. *Syst Biol Reprod Med* 60(4), 199-205 (2014).
DOI: 10.3109/19396368.2014.912363
86. A. Kheradmand, O. Dezfoulan, M. Alirezaei, B. Hadian: Ghrelin is a suppressor of testicular damage following experimentally induced cryptorchidism in the rat. *J Pediatr Surg* 49(4), 593-598 (2014).
DOI: 10.1016/j.jpedsurg.2013.10.003
87. R.J. Niewenhuis: Effects of cadmium upon regenerated testicular vessels in the rat. *Biol Reprod* 23(1), 171-179 (1980).
DOI: 10.1095/biolreprod23.1.171
88. F. De Sousa Predes, M.A. Diamante, H. Dolder: Testis response to low doses of cadmium in Wistar rats. *Int J Exp Pathol* 91(2), 125-131 (2010).
DOI: 10.1111/j.1365-2613.2009.00692.x
89. N. Babaknejad, S. Bahrami1, A.A. Moshtaghi1, H. Nayeri1, P. Rajabi, F.G. Iranpour: Cadmium Testicular Toxicity in Male Wistar Rats: Protective Roles of Zinc and Magnesium. *Biol Trace Elem Res* 185(1), 106-115 (2018).
DOI: 10.1007/s12011-017-1218-5
90. R. Mahmoudi, A. Azizi, S. Abedini, V. Hemayatkah Jahromi, H. Abidi, M. Jafari Barmak: Green tea improves rat sperm quality and reduced cadmium chloride damage effect in spermatogenesis cycle. *J Med Life* 11(4), 371-380 (2018).
DOI: 10.25122%2Fjml-2018-0005
91. S.O. Abarikwu, A.F.S. Wokoma, C.J. Mgbudom-Okah, S.I. Omeodu, R. Ohanador: Effect of Fe and Cd Co-Exposure on Testicular Steroid Metabolism, Morphometry, and Spermatogenesis in Mice. *Biol Trace Elem Res* 190(1), 109-123 (2018).
DOI: 10.1007/s12011-018-1536-2
92. J. Skolarczyk, M. Budzyński, J. Pekar, T. Małecka-Massalska, K. Skórzyńska-Dzidusko: The impact of cadmium on male infertility. *J Elem* 23(1), 35-44 (2018).
DOI: 10.5601/jelem.2017.22.2.1320
93. C. Xu, J.E. Johnson, P.K. Singh, M.M. Jones, H. Yan, C.E. Carter: *In vivo* studies of cadmium induced apoptosis in testicular tissue of the rat and its modulation by a chelating agent. *Toxicology* 107(1), 1-8 (1996).
DOI: 10.1016/0300-483x(95)03195-I
94. H. Yan, C.E. Carter, C. Xu, P.K. Singh, M.M. Jones, J.E. Johnson, M.S. Dietrich: Cadmium-induced apoptosis in the urogenital organs of the male rat and its suppression by chelation. *J Toxicol Environ Health* 52(2), 149-168 (1997).
DOI: 10.1080/00984109708984058
95. T. Zhou, G. Zhou, W. Song, N. Eguchi,

- W. Lu, E. Lundin, T. Jin, G. Nordberg: Cadmium-induced apoptosis and changes in expression of p53, c-jun and MT-I genes in testes and ventral prostate of rats. *Toxicology* 142(1), 1-13 (1999). DOI: 10.1016/s0300-483x(99)00115-8
96. J. Li, J. Yi, C. Wang, P. Xu: Effect of cadmium on apoptosis of spermatogenic cells of rat testis and the protection effect of zinc against it. *Wei Sheng Yan Jiu* 29(3), 135-137 (2000).
97. R. Toman, P. Massányi, V. Uhrin: Changes in the testis and epididymis of rabbits after an intraperitoneal and peroral administration of cadmium. *Trace Elem Med* 19(3), 114-117 (2002).
98. R.A. Goyer, J. Liu, M.P. Waalkes: Cadmium and cancer of prostate and testis. *Biometals* 17(5), 555-558 (2004). DOI: 10.1023/b:biom.0000045-738.59708.20
99. A.W. Obianime, I.I. Roberts: Antioxidants, cadmium-induced toxicity, serum biochemical and the histological abnormalities of the kidney and testes of the male Wistar rats. *Niger J Physiol Sci* 24(2), 177-185 (2009). DOI: 10.4314/njps.v24i2.52910
100. S. Bekheet: Cadmium chloride rapidly alters both BTB tight junction proteins and germ cells in young rat testes. *Egypt Acad J Biolog Sci* 2(1), 59-74 (2010). DOI: 10.21608/eajbsz.2010.15916
101. L.M. Herranz, F. Teba, R. Martín, I. Ingelmo, V.V. Gómez, J. Codesal, J.M. Pozuelo, B. Oltra, E. Serna, L. Santamaría: Quantitative Changes in Rat Seminiferous Epithelium After Chronic Administration of Low Doses of Cadmium and Zinc: A Stereological Study. *The Open Androl J* 2, 27-36 (2010). DOI: 10.2174/1876827X01002010027
102. A. Hirako, Y. Takeoka, T. Hayashi, T. Takeuchi, S. Furukawa, A. Sugiyama: Effects of cadmium exposure on Iberian ribbed newt (*Pleurodeles waltl*) testes. *J Toxicol Pathol* 30(4), 345-350 (2017). DOI: 10.1293%2Ftox.2017-0032
103. D.D. Mruk, C.Y. Cheng: Environmental contaminants: Is male reproductive health at risk? *Spermatogenesis* 1(4), 1-8 (2011). DOI: 10.4161/spmg.1.4.18328
104. T. Aoyagi, H. Ishikawa, K. Miyaji, K. Hayakawa, M. Hata: Cadmium-induced testicular damage in a rat model of subchronic intoxication. *Reprod Med Biol* 1(2), 59-63 (2002). DOI: 10.1046%2Fj.1445-5781.2002.00010.x
105. T. Zhou, X. Jia, R.E. Chapin, R.R. Maronpot, M.W. Harris, J. Liu, M.P. Waalkes, E.M. Eddy: Cadmium at a non-toxic dose alters gene expression in mouse testes. *Toxicol Lett* 154(3), 191-200 (2004). DOI: 10.1016/j.toxlet.2004.07.015
106. E. Maretová, M. Maretta, J. Legáth: Toxic effects of cadmium on testis of birds and mammals: Toxic effects of cadmium on testis of birds and mammals: a review. *Anim Reprod Sci* 155:1-10 (2015). DOI: 10.1016/j.anireprosci.2015.01.007
107. K.W. Hew, W.A. Ericson, M.J. Welsh: A single low cadmium dose causes failure of spermiation in the rat. *Toxicol Appl Pharmacol* 121(1), 15-21 (1993). DOI: 10.1006/taap.1993.1123

108. S.R. Clough, M.J. Welsh, A.H. Payne, C.D. Brown, M.J. Brabec: Primary rat Sertoli and interstitial cells exhibit a differential response to cadmium. *Cell Biol Toxicol* 6(1), 63-79 (1990). DOI: 10.1007/bf00135027
109. M. Zhang, Z. He, L. Wen, J. Wu, L. Yuan, Y. Lu, C. Guo, L. Zhu, S. Deng, H. Yuan: Cadmium suppresses the proliferation of piglet Sertoli cells and causes their DNA damage, cell apoptosis and aberrant ultrastructure. *Reprod Biol Endocrinol* 8:97, 1-12 (2010). DOI: 10.1186/1477-7827-8-97
110. X. Yu, S. Hong, E.M. Faustman: Cadmium-induced Activation of Stress Signaling Pathways, Disruption of Ubiquitin-dependent Protein Degradation and Apoptosis in Primary Rat Sertoli Cell-Gonocyte Cocultures. *Toxicol Sci* 104(2), 385-396 (2008). DOI: 10.1093/toxsci/kfn087
111. P. Bizarro, S. Acevedo, G. Niño-Cabrera, P. Mussali-Galante, F. Pasos, M.R. Avila-Costa, T.I. Fortoul: Ultrastructural modifications in the mitochondrion of mouse Sertoli cells after inhalation of lead, cadmium or lead-cadmium mixture. *Reprod Toxicol* 17(5), 561-566 (2003). DOI: 10.1016/s0890-6238(03)00096-0
112. X. Xiao, D.D. Mruk, E.I. Tang, C.K. Wong, W.M. Lee, C.M. John, P.J. Turek, B. Silvestrini, C.Y. Cheng: Environmental toxicants perturb human Sertoli cell adhesive function via changes in F-actin organization mediated by actin regulatory proteins. *Hum Reprod* 29(6), 1279-1291 (2014). DOI: 10.1093/humrep/deu011
113. N.P. Chung, C.Y. Cheng: Is cadmium chloride-induced inter-sertoli tight junction permeability barrier disruption a suitable *in vitro* model to study the events of junction disassembly during spermatogenesis in the rat testis? *Endocrinology* 142(5), 1878-1888 (2001). DOI: 10.1210/endo.142.5.8145
114. J.L. Li, R. Gao, S. Li, J.T. Wang, Z.X. Tang, S.W. Xu: Testicular toxicity induced by dietary cadmium in cocks and ameliorative effect by selenium. *Biometals* 23(4), 695-705 (2010). DOI: 10.1007/s10534-010-9334-0
115. Q. Liu, J.H. Gu, Y. Yuan, X.Z. Liu, H.D. Wang, Z.P. Liu, Z.Y. Wang, J.C. Bian: Effect of cadmium on rat Leydig cell testosterone production and DNA integrity *in vitro*. *Biomed Environ Sci* 26(9), 769-773 (2013). DOI: 10.3967/0895-3988.2013.09.009
116. B. Rajendar, K. Bharavi, G.S. Rao, P.V. Kishore, P.R. Kumar, C.S. Kumar, D.S. Kumar: Protective effect of alpha-tocopherol on biochemical and histological alterations induced by cadmium in rat testes. *Indian J Physiol Pharmacol* 55(3), 213-220 (2011).
117. C.J. Frederickson: Neurobiology of zinc and zinc-containing neurons. *Int Rev Neurobiol* 31, 145-238 (1989). DOI: 10.1016/s0074-7742(08)60279-2
118. S.R. Powell: Antioxidant properties of zinc. *J Nutr* 130(5), 1447s-1454s (2000). DOI: 10.1093/jn/130.5.1447s
119. A. Ozturk, A.K. Baltaci, R. Mogulkoc, E. Oztekin, A. Sivrikaya, E. Kurtoglu, A. Kul: Effects of zinc deficiency and supplementation on malondialdehyde and glutathione levels in blood and tissues of rats performing swimming exercise. *Biol Trace Elem Res* 94(2),

- 157-166 (2003).
DOI: 10.1385/bter:94:2:157
120. G. Ozdemir, F. Inanc: Zinc may protect remote ocular injury caused by intestinal ischemia reperfusion in rats. *Tohoku J Exp Med* 206(3), 247-251 (2005).
DOI: 10.1620/tjem.206.247
121. S. Amara, H. Abdelmelek, C. Garrel, P. Guiraud, T. Douki, J.L. Ravanat, A. Favier, M. Sakly, K. Ben Rhouma: Preventive effect of zinc against cadmium-induced oxidative stress in the rat testis. *J Reprod Dev* 54(2), 129-134 (2008).
DOI: 10.1262/jrd.18110
122. J. Hernández Rodríguez, S. Montes López, I. Arrieta Cruz, E. Arenas Ríos, I. Jiménez Morales, M. Arteaga Silva. Efecto de la exposición al cadmio sobre los carbohidratos de la membrana plasmática y la fosforilación de proteínas de espermatozoides epididimarios de la rata Wistar. In: *El espermatozoide: una mirada desde México*. Eds: E. Arenas Ríos, G. Fuentes Mascorro, México. First Edition, 193-230 (2018).
123. B. Robaire, B.T. Hinton. The Epididymis. In: *Knobil and Neill's Physiology of Reproduction*. Eds: T.M. Plant, A.J. Zeleznik. USA: Elsevier. Fourth Edition, 619-677 (2015).
DOI: 10.1016/B978-0-12-397175-3.00017-X
124. R.A. Rahman Elgawish, M. Elshabrawy Ghanem: Effect of Long Term Cadmium Chloride Exposure on Testicular Functions in Male Albino Rats. *Am J Anim Vet Sci* 9(4), 182-188 (2014).
DOI: 10.3844/ajavsp.2014.182.188
125. J. Hernández-Rodríguez, E. Arenas-Ríos, I. Jiménez-Morales, E. Cortés-Barberena, S. Montes, R.M. Vigueras-Villaseñor, M. Arteaga-Silva: Cadmium postnatal administration affects the presence and distribution of carbohydrates of the sperm membrane during its maturation on the epididymis. Manuscript in preparation, 2020.
126. J. Hernández-Rodríguez, C. Togno-Peirce, P. López de Jesús, S.G. Pérez-Aguirre, E. Arenas-Ríos, R.M. Vigueras-Villaseñor, S. Montes-López, H. Bonilla-Jaime, P. Damián-Matzumura, M. Arteaga-Silva: Efecto del cadmio en la maduración espermática epididimaria. *RelbCi* 3(3), 11-21 (2016).
127. B. Robaire. H. Scheer, C. Hachey. Regulation of epididymal steroid metabolizing enzymes. In: *Bioregulators of Reproduction*. Eds: G. Jagiello, H.J. Vogel, New York. Academic Press. 487-498 (1981).
128. B. Robaire, M. Hamzeh: Androgen action in the epididymis. *J Androl* 32(6), 592-599 (2011).
DOI: 10.2164/jandrol.111.014266
129. K.F. Arrotéia, P. Vianna-García, M. Ferreira-Barbieri, M. Lopes-Justino, L.A. Violin-Pereira. The Epididymis: Embryology, Structure, Function and Its Role in Fertilization and Infertility. In: *Embryology - Updates and Highlights on Classic Topics*. Eds: L. Violin-Pereira, InTech, Rijeka, Croatia, 41-66 (2012).
DOI: 10.5772/35847
130. L. O'Hara, M. Welsh, P.T. Saunders, L.B. Smith: Androgen receptor expression in the caput epididymal epithelium is essential for development of the initial segment and epididymal spermatozoa transit. *Endocrinology* 152(2), 718-729

- (2011).
DOI: 10.1210/en.2010-0928
131. S. Kerkhofs, V. Dubois, K. De Gendt, C. Helsen, L. Clinckemalie, L. Spans, F. Schuit, S. Boonen, D. Vanderschueren, P.T. Saunders, G. Verhoeven, F. Claessens: A role for selective androgen response elements in the development of the epididymis and the androgen control of the 5 α reductase II gene. *FASEB J* 26(10), 4360-4372 (2012).
DOI: 10.1096/fj.11-202283
 132. Y.L. Ji, H. Wang, P. Liu, Q. Wang, X.F. Zhao, X.H. Meng, T. Yu, H. Zhang, C. Zhang, Y. Zhang, D.X. Xu: Pubertal cadmium exposure impairs testicular development and spermatogenesis via disrupting testicular testosterone synthesis in adult mice. *Reprod Toxicol* 29(2), 176-183 (2010).
DOI: 10.1016/j.reprotox.2009.10.014
 133. S.O. Abarikwu, S. Oruitemeka, I.A. Uwadileke S.I. Omeodu, N.F. Okoye, C.J. Mgbudom-Okah, R. Ohanador: Oral administration of cadmium depletes intratesticular and epididymal iron levels and inhibits lipid peroxidation in the testis and epididymis of adult rats. *J Trace Elem Med Biol* 48, 213-223 (2018).
DOI: 10.1016/j.jtemb.2018.04.011
 134. S.P. Ribeiro: Efeitos do cádmio, chumbo e zinco em epidídimo de ratos Wistar. Unpublished Masters Thesis. Federal University of Viçosa, Brazil, 1-67 (2013).
 135. A.C. Kohane: Contribución al estudio del proceso de maduración de los espermatozoides en el epididimo. Unpublished PhD thesis. Faculty of Exact and Natural Sciences. Buenos Aires' University. 1-240 (1980).
 136. S. Benoff, K. Auborn, J.L. Marmar, I.R. Hurley: Link between low-dose environmentally relevant cadmium exposures and asthenozoospermia in a rat model. *Fertil Steril* 89, 73-79 (2008).
DOI: 10.1016/j.fertnstert.2007.12.035
 137. R. Oliva: Protamines and male infertility. *Hum Reprod Update* 12(4), 417-435 (2006).
DOI: 10.1093/humupd/dml009
 138. G.A. Quintero-Vásquez, R.M. Bermúdez-Cruz, J. Castillo-Cadena: Infertilidad masculina y fragmentación del ADN espermático: Un problema actual. *TIP Rev Esp Cienc Quím Biol* 18(2), 144-151 (2015).
DOI: 10.1016/j.recqb.2015.09.006
 139. Y. Méndez, F. Báez, P. Villamediana: Efecto de la exposición *in vitro* de espermatozoides humanos a cadmio (CdCl₂). *Perinatol Reprod Hum* 25(4), 198-204 (2011).
 140. D.R. Tulsiani: Glycan-modifying enzymes in luminal fluid of the mammalian epididymis: an overview of their potential role in sperm maturation. *Mol Cell Endocrinol* 250(1-2), 58-65 (2006).
DOI: 10.1016/j.mce.2005.12.025
 141. D.R. White, R.J. Aitken: Influence of epididymal maturation on cyclic AMP levels in hamster spermatozoa. *Int J Androl* 12(1), 29-43 (1989).
DOI: 10.1111/j.1365-2605.1989.tb01283.x
 142. C.H. Yeung, G.F. Weinbauer, T.G. Cooper: Responses of monkey epididymal sperm of different maturational status to second messengers mediating protein tyrosine phosphorylation, acrosome reaction, and

- motility. *Mol Reprod Dev* 54(2), 194-202 (1999).
DOI: 10.1002/(sici)1098-2795(199910)54:2%3C194::aid-mrd12%3E3.0.co;2-c
143. S. Vijayaraghavan, J. Mohan, H. Gray, B. Khatra, D.W. Carr: A Role for Phosphorylation of Glycogen Synthase Kinase-3 α in Bovine Sperm Motility Regulation. *Biol Reprod* 62(6), 1647-1654 (2000).
DOI: 10.1095/biolreprod62.6.1647
144. A. Contri, A. Gloria, D. Robbe, I. De Amicis, A. Carluccio: Characteristics of donkey spermatozoa along the length of the epididymis. *Theriogenology* 77(1), 166-173 (2012).
DOI: 10.1016/j.theriogenology.-2011.07.031
145. S. Tardif, C. Dube, S. Chevalier, J.L. Bailey: Capacitation is associated with tyrosine phosphorylation and tyrosine kinase-like activity of pig sperm proteins. *Biol Reprod* 65(3), 784-792 (2001).
DOI: 10.1095/biolreprod65.3.784
146. F. Urner, D. Sakkas: Protein phosphorylation in mammalian spermatozoa. *Reproduction* 125(1), 17-26 (2003).
DOI: 10.1530/rep.0.1250017
147. R.K. Naz, P.B. Rajesh: Role of tyrosine phosphorylation in sperm capacitation / acrosome reaction. *Reprod Biol Endocrinol* 2:75, 1-12 (2004).
DOI: 10.1186%2F1477-7827-2-75
148. R. Da Costa, D. Botana, S. Piñero, F. Proverbio, R. Marín: Cadmium inhibits motility, activities of plasma membrane Ca(2+)-ATPase and axonemal dynein-ATPase of human spermatozoa. *Andrologia* 48(4), 464-469 (2016).
DOI: 10.1111/and.12466
149. L. Wang, Y. Li, J. Fu, L. Zhen, N. Zhao, Q. Yang, S. Li, X. Li: Cadmium inhibits mouse sperm motility through inducing tyrosine phosphorylation in a specific subset of proteins. *Reprod Toxicol* 63, 96-106 (2016).
DOI: 10.1016/j.reprotox.2016.05.018
150. N. Sugawara, C. Sugawara: Selenium protection against testicular lipid peroxidation from cadmium. *J Appl Biochem* 6(4), 199-204 (1984).
151. T. Koizumi, Z.G. Li: Role of oxidative stress in single-dose, cadmium-induced testicular cancer. *J Toxicol Environ Health* 37(1), 25-36 (1992).
DOI: 10.1080/15287399209531654
152. O.A. Adaramoye, O.O. Akanni: Protective effects of *Artocarpus altilis* (Moraceae) on cadmium-induced changes in sperm characteristics and testicular oxidative damage in rats. *Andrologia* 48(2), 152-163 (2016).
DOI: 10.1111/and.12426
153. M. Jodar, A. Soler-Ventura, R. Oliva: Semen proteomics and male infertility. *J Proteomics* 162, 125-134 (2017).
DOI: 10.1016/j.jprot.2016.08.018
154. S. Benoff, I.R. Hurley, M. Barcia, F.S. Mandel, G.W. Cooper, A. Herschlag: A potential role for cadmium in the etiology of varicocele-associated infertility. *Fertil Steril* 67(2), 336-347 (1997).
DOI: 10.1016/s0015-0282(97)81921-8
155. J.W. Laskey, G.L. Rehnberg, S.C. Laws, J.F. Hein: Reproductive effects of low acute doses of cadmium chloride in adult male rats. *Toxicol Appl Pharmacol* 73(2),

- 250-255 (1984).
DOI: 10.1016/0041-008x(84)90330-2
156. M.D. Kipling, J.A.H. Waterhouse: Cadmium and prostatic carcinoma. *Lancet* 289(7482), 730-731 (1967).
DOI: 10.1016/S0140-6736(67)92222-2
157. M.P. Waalkes: Cadmium carcinogenesis in review. *J Inorg Biochem* 79(1-4), 241-244 (2000).
DOI: 10.1016/s0162-0134(00)00009-x
158. C.D. Klaassen, J. Liu, S. Choudhuri: Metallothionein: an intracellular protein to protect against cadmium toxicity. *Annu Rev Pharmacol Toxicol* 39, 267-294 (1999).
DOI: 10.1146/annurev.pharmtox.39.1.267
159. L.C. Xu, S.Y. Wang, X.F. Yang, X.R. Wang: Effects of cadmium on rat sperm motility evaluated with computer assisted sperm analysis. *Biomed Environ Sci* 14(4), 312-317 (2001).
160. N. Pant, G. Upadhyay, S. Pandey, N. Mathur, D.K. Saxena, S.P. Srivastava: Lead and cadmium concentration in the seminal plasma of men in the general population: correlation with sperm quality. *Reprod Toxicol* 17(4), 447-450 (2003).
DOI: 10.1016/s0890-6238(03)00036-4
161. O. Akinloye, A.O. Arowojolu, O.B. Shittu, J.I. Anetor: Cadmium toxicity: a possible cause of male infertility in Nigeria. *Reprod Biol* 6(1), 17-30 (2006).
162. S. Benoff, R. Hauser, J.L. Marmar, I.R. Hurley, B. Napolitano, G.M. Centola: Cadmium concentrations in blood and seminal plasma: correlations with sperm number and motility in three male populations (infertility patients, artificial insemination donors, and unselected volunteers). *Mol Med* 15(7-8), 248-262 (2009).
DOI: 10.2119/molmed.2008.00104
163. W. Guzikowski, M.I. Szykowska, H. Motak-Pochrzest, A. Pawlaczyk, S. Sypniewski: Trace elements in seminal plasma of men from infertile couples. *Arch Med Sci* 11(3), 591-598 (2015).
DOI: 10.5114%2Faoms.2015.52363
164. S.E. Chia, C.N. Ong, S.T. Lee, F.H. Tsakok: Blood concentrations of lead, cadmium, mercury, zinc, and copper and human semen parameters. *Arch Androl* 29(2), 177-183 (1992).
DOI: 10.3109/01485019208987722
165. H. Oliveira, M. Spanò, C. Santos, M.L. Pereira: Adverse effects of cadmium exposure on mouse sperm. *Reprod Toxicol* 28(4), 550-555 (2009).
DOI: 10.1016/j.reprotox.2009.08.001
166. D. Beyersmann, S. Hechtenberg: Cadmium, gene regulation, and cellular signalling in mammalian cells. *Toxicol Appl Pharmacol* 144(2), 247-261 (1997).
DOI: 10.1006/taap.1997.8125
167. A. Martelli, E. Rousselet, C. Dycke, A. Bouron, J.M. Moulis: Cadmium toxicity in animal cells by interference with essential metals. *Biochimie* 88(11), 1807-1814 (2006).
DOI: 10.1016/j.biochi.2006.05.013
168. H.F. Wang, M. Chang, T.T. Peng, Y. Yang, N. Li, T. Luo, Y.M. Cheng, M.Z. Zhou, X.H. Zeng, L.P. Zheng: Exposure to Cadmium Impairs Sperm Functions by Reducing CatSper in Mice. *Cell Physiol Biochem* 42(1), 44-54 (2017).
DOI: 10.1159/000477113

169. S.S. Riar, B.S. Setty, A.B. Kar: Studies on the physiology and biochemistry of mammalian epididymis: biochemical composition of epididymis. A comparative study. *Fertil Steril* 24(5):353-363 (1973).
DOI: 10.1016/S0015-0282(16)39673-X
170. R.J. Aitken, M. Paterson, H. Fisher, D.W. Buckingham, M. van Duin: Redox regulation of tyrosine phosphorylation in human spermatozoa and its role in the control of human sperm function. *J of Cell Sci* 108, 2017-2025 (1995).
171. S.J. Stohs, D. Bagchi: Oxidative mechanisms in the toxicity of metal ions. *Free Radic Biol Med* 18(2), 321-336 (1995).
DOI: 10.1016/0891-5849(94)00159-h
172. M. Valko, H. Morris, M.T. Cronin: Metals, toxicity and oxidative stress. *Curr Med Chem* 12(10), 1161-1208 (2005).
DOI: 10.2174/0929867053764635
173. S. Takagi, T. Yoshida, K. Tsubata, H. Ozaki, T.K. Fujii, Y. Nomura, M. Sawada: Sex differences in fetal gonadotropins and androgens. *J Steroid Biochem* 8, 609-620 (1977).
DOI: 10.1016/0022-4731(77)90270-9
174. S.K. Kota, K. Gayatri, S. Jammula, L.K. Meher: Fetal endocrinology. *Indian J Endocrinol Metab* 17(4), 568-579 (2013).
DOI: 10.4103%2F2230-8210.113722
175. A. Kaprara, I.T. Huhtaniemi: The hypothalamus-pituitary-gonad axis: Tales of mice and men. *Metabolism* 86, 3-17 (2018).
DOI: 10.1016/j.metabol.2017.11.018
176. A.P. Abreu, U.B. Kaiser: Pubertal development and regulation. *Lancet Diabetes Endocrinol* 4(3), 254-264 (2016).
DOI: 10.1016/s2213-8587(15)00418-0
177. P. Celec, D. Ostatníková, J. Hodosy: On the effects of testosterone on brain behavioral functions. *Front Neurosci* 17, 9-12 (2015).
DOI: 10.3389%2Ffnins.2015.00012
178. F. Hayes, A. Dwyer, N. Pitteloud: Hypogonadotropic Hypogonadism (HH) and Gonadotropin Therapy. In: Endotext. Eds: K.R. Feingold, B. Anawalt, A. Boyce, G. Chrousos, K. Dungan, A. Grossman, J.M. Hershman, G. Kaltsas, C. Koch, P. Kopp, M. Korbonits, R. McLachlan, J.E. Morley, M. New, L. Perreault, J. Purnell, R. Rebar, F. Singer, D.L. Trence, A. Vinik,, D.P. Wilson. South Dartmouth (MA): MDText.com, Inc.; 2000-2013.
179. L. Dunkel, R. Quinton: Transition in endocrinology: induction of puberty. *Eur J Endocrinol* 170(6), R229-R239 (2014).
DOI: 10.1530/eje-13-0894
180. k. Maeda, S. Ohkura, Y. Uenoyama, Y. Wakabayashi, Y. Oka, H. Tsukamura, H. Okamura: Neurobiological mechanisms underlying GnRH pulse generation by the hypothalamus. *Brain Res* 1364, 103-115 (2010).
DOI: 10.1016/j.brainres.2010.10.026
181. J.L. Antunes, P.W. Carmel, E.M. Housepian, M. Ferin: Luteinizing hormone-releasing hormone in human pituitary blood. *J Neurosurg* 49(3), 382-386 (1978).
DOI: 10.3171/jns.1978.49.3.0382
182. C. Sonigo, N. Binart: Overview of the impact of kisspeptin on reproductive function. *Ann Endocrinol (Paris)* 73(5), 448-458 (2012).

- DOI: 10.1016/j.ando.2012.07.680
183. J.T. Smith, M.J. Cunningham, E.F. Rissman, D.K. Clifton, R.A. Steiner: Regulation of Kiss1 gene expression in the brain of the female mouse. *Endocrinology* 146(9), 3686-3692 (2005). DOI: 10.1210/en.2005-0488
184. N. Richard, G. Galmiche, S. Corvaisier, A. Caraty, M.L. Kottler: KiSS-1 and GPR54 genes are co-expressed in rat gonadotrophs and differentially regulated *in vivo* by oestradiol and gonadotrophin-releasing hormone. *J Neuroendocrinol* 20(3), 381-393 (2008). DOI: 10.1111/j.1365-2826.2008.01653.x
185. V.M. Navarro, J.M. Castellano, R. Fernández-Fernández, M.L. Barreiro, J. Roa, J.E. Sanchez-Criado, C. Dieguez, L. Pinilla, M. Tena-Sempere: Developmental and hormonally regulated messenger ribonucleic acid expression of KiSS-1 and its putative receptor, GPR54, in rat hypothalamus and potent luteinizing hormone-releasing activity of KiSS-1 peptide. *Endocrinology* 145(10), 4565-4574 (2004). DOI: 10.1210/en.2004-0413
186. M. Shahab, C. Mastronardi, S.B. Seminara, W.F. Crowley, S.R. Ojeda, T.M. Plant: Increased hypothalamic GPR54 signaling: a potential mechanism for initiation of puberty in primates. *Proc Natl Acad Sci U S A* 102(6), 2129-2134 (2005). DOI: 10.1073/pnas.0409822102
187. J. Roa, E. Aguilar, C. Dieguez, L. Pinilla, M. Tena-Sempere: New frontiers in kisspeptin/GPR54 physiology as fundamental gatekeepers of reproductive function. *Front Neuroendocrinol* 29(1), 48-69 (2008)
- DOI: 10.1016/j.yfrne.2007.07.002
188. S. Ojeda, M.K. Skinner. Puberty in the rat. In: Knobil and Neill's Physiology of Reproduction. Eds: E.M. Knobil, J.D. Nelly, Raven Press Ltd, New York, 2061-2126 (2006). DOI: 10.1016/B978-012515400-0/50043-9
189. M. Arteaga-Silva, R.M. Vigueras-Villaseñor, S. Retana-Márquez, M. Hernández-González, H. Bonilla-Jaime, X. Guzmán-García, J.L. Contreras-Montiel: Testosterone Levels and Development of the Penile Spines and Testicular Tissue during the Postnatal Growth in Wistar Rats. *ASM* 3(3), 1-9 (2013). DOI: 10.4236/asm.2013.33A001
190. B.D. Sachs, G.B. Glater, J.K. O'Hanlon: Morphology of the erect glans penis in rats under various gonadal hormone conditions. *Anat Rec* 210(1), 45-52 (1984). DOI: 10.1002/ar.1092100108
191. C.L. Moore: A hormonal basis for sex differences in the self-grooming of rats. *Horm Behav* 20(2), 155-165 (1986). DOI: 10.1016/0018-506x(86)90014-0
192. M. Hernández-González: Prepubertal genital grooming and penile erections in relation to sexual behavior of rats. *Physiol Behav* 71(1-2), 51-56 (2000). DOI: 10.1016/s0031-9384(00)00320-6
193. J.K. O'Hanlon, B.D. Sachs: Fertility of mating in rats (*Rattus norvegicus*): contributions of androgen-dependent morphology and actions of the penis. *J Comp Psychol* 100(2), 178-187 (1986).
194. B.D. Sachs, L.D. Garinello: Interaction

- between penile reflexes and copulation in male rats. *J Comp Physiol Psychol* 92(4), 759-767 (1978).
DOI: 10.1037/h0077498
195. B.L. Hart: Testosterone regulation of sexual reflexes in spinal male rats. *Science* 155(3767), 1283-1284 (1967).
DOI: 10.1126/science.155.3767.1283
196. J.M. Davidson, M.L. Stefanick, B.D. Sachs, E.R. Smith: Role of androgen in sexual reflexes of the male rat. *Physiol Behav* 21(2), 141-146 (1978).
DOI: 10.1016/0031-9384(78)90033-1
197. C.C. Korenbrot, I.T. Huhtaniemi, R.I. Weiner: Preputial separation as an external sign of pubertal development in the male rat. *Biol Reprod* 17(2), 298-303 (1977).
DOI: 10.1095/biolreprod17.2.298
198. R. Couto-Moraes, L.F. Felicio, M.M. Bernardi: Postpartum testosterone administration does not reverse the effects of perinatal exposure to cadmium on rat offspring development. *J Appl Toxicol* 30(3), 233-241 (2010).
DOI: 10.1002/jat.1489
199. F. Salvatori, C.B. Talassi, S.A. Salzgeber, H.S. Spinosa, M.M. Bernardi: Embryotoxic and long-term effects of cadmium exposure during embryogenesis in rats. *Neurotoxicol Teratol* 26(5), 673-680 (2004).
DOI: 10.1016/j.ntt.2004.05.001
200. M. Interdonato, G. Pizzino, A. Bitto, F. Galfo, N. Irrera, A. Mecchio, G. Pallio, V. Ramistella, F. De Luca, A. Santamaria, L. Minutoli, H. Marini, F. Squadrito, D. Altavilla: Cadmium delays puberty onset and testis growth in adolescents. *Clin Endocrinol (Oxf)* 83(3), 357-362 (2015).
DOI: 10.1111/cen.12704
201. X. Xu, Y.M. Chiung, F. Lu, S. Qiu, M. Ji, X. Huo: Associations of cadmium, bisphenol A and polychlorinated biphenyl co-exposure in utero with placental gene expression and neonatal outcomes. *Reprod Toxicol* 52, 62-70 (2015).
DOI: 10.1016/j.reprotox.2015.02.004
202. P.J. Chedrese, M. Piasek, M.C. Henson: Cadmium as an Endocrine Disruptor in the Reproductive System. *Immunol Endocr Metab Agents Med Chem* 6(1), 27-35 (2006).
DOI: 10.2174/187152206775528941
203. R.H. Waring, R.M. Harris: Endocrine disrupters: A human risk? *Mol Cell Endocrinol* 244(1-2), 2-9 (2005).
DOI: 10.1016/j.mce.2005.02.007
204. A. Donna, P. Crosignani, F. Robutti, P.G. Betta, R. Bocca, N. Mariani, F. Ferrario, R. Fissi, F. Berrino: Triazine herbicides and ovarian epithelial neoplasms. *Scand J Work Environ Health* 15(1), 47-53 (1989).
DOI: 10.5271/sjweh.1882
205. J.E. Keller-Byrne, S.A. Khuder, E.A. Schaub: Meta-analyses of prostate cancer and farming. *Am J Ind Med* 31(5), 580-586 (1997).
DOI: 10.1002/(sici)1097-0274(199705)31:5%3C580::aid-ajim13%3E3.0.co;2-v
206. K.J. Aronson, A.B. Miller, C.G. Woolcott, E.E. Sterns, D.R. McCready, L.A. Lickley, E.B. Fish, G.Y. Hiraki, C. Holloway, T. Ross, W.M. Hanna, S.K. SenGupta, J.P. Weber: Breast adipose tissue concentrations of polychlorinated biphenyls and other organochlorines and breast cancer risk. *Cancer Epidemiol*

- Biomarkers Prev* 9(1), 55-63 (2000).
DOI: 10.1002/jat.899
207. H.K. Weir, L.D. Marrett, N. Kreiger, G.A. Darlington, L. Sugar: Pre-natal and perinatal exposures and risk of testicular germ-cell cancer. *Int J Cancer* 87(3), 438-443 (2000).
DOI: 10.1002/1097-0215(20000801)87:3%3C438::aid-ijc20%3E3.0.co;2-1
 208. P.R. Band, N.D. Le, R. Fang, M. Deschamps: Carcinogenic and endocrine disrupting effects of cigarette smoke and risk of breast cancer. *Lancet* 360(9339), 1044-1049 (2002).
DOI: 10.1016/S0140-6736(02)11140-8
 209. T. Colborn, D. Dumanoski, J. Myers. *Our Stolen Future: Are We Threatening Our Fertility, Intelligence, and Survival?--A Scientific Detective Story*. Penguin Books, New York, 336 (1996).
 210. B.N. Ames, L.S. Gold: Paracelsus to parascience: the environmental cancer distraction. *Mutat Res* 447(1), 3-13 (2000).
DOI: 10.1016/S0027-5107(99)00194-3
 211. S.H. Safe: Endocrine disruptors and human health--is there a problem? An update. *Environ Health Perspect* 108(6), 487-493 (2000).
DOI: 10.1289/ehp.00108487
 212. M.C. Henson, P.J. Chedrese: Endocrine disruption by cadmium, a common environmental toxicant with paradoxical effects on reproduction. *Exp Biol Med (Maywood)* 229(5), 383-392 (2004).
DOI: 10.1177/153537020422900506
 213. P.D. Darbre: Underarm cosmetics and breast cancer. *J Appl Toxicol* 23(2), 89-95 (2003).
 214. A.D. Smida, X.P. Valderrama, M.C. Agostini, M.A. Furlan, J. Chedrese: Cadmium stimulates transcription of the cytochrome p450 side chain cleavage gene in genetically modified stable porcine granulosa cells. *Biol Reprod* 70(1), 25-31 (2004).
DOI: 10.1095/biolreprod.103.019000
 215. M.D. Johnson, N. Kenney, A. Stoica, L. Hilakivi-Clarke, B. Singh, G. Chepko, R. Clarke, P.F. Sholler, A.A. Lirio, C. Foss, R. Reiter, B. Trock, S. Paik, M.B. Martin: Cadmium mimics the *in vivo* effects of estrogen in the uterus and mammary gland. *Nat Med* 9(8), 1081-1084 (2003).
DOI: 10.1038/nm902
 216. M. Brama, L. Gnessi, S. Basciani, N. Cerulli, L. Politi, G. Spera, S. Mariani, S. Cherubini, A. Scotto d'Abusco, R. Scandurra, S. Migliaccio: Cadmium induces mitogenic signaling in breast cancer cell by an ERalpha-dependent mechanism. *Mol Cell Endocrinol* 264(1-2), 102-108 (2007).
DOI: 10.1016/j.mce.2006.10.013
 217. P.D. Darbre: Metalloestrogens: an emerging class of inorganic xenoestrogens with potential to add to the oestrogenic burden of the human breast. *J Appl Toxicol* 26(3), 191-197 (2006).
DOI: 10.1002/jat.1135
 218. C. Byrne, S.D. Divekar, G.B. Storch, D.A. Parodi, M.B. Martin: Cadmium--a metalloestrogen? *Toxicol Appl Pharmacol* 238(3), 266-271 (2009).
DOI: 10.1016/j.taap.2009.03.025
 219. P. Garcia-Morales, M. Saceda, N. Kenney, N. Kim, D.S. Salomon, M.M. Gottardis, H.B. Solomon, P.F. Sholler,

- V.C. Jordan, M.B. Martin: Effect of cadmium on estrogen receptor levels and estrogen-induced responses in human breast cancer cells. *J Biol Chem* 269(24), 16896-16901 (1994)
220. A. Stoica, B.S. Katzenellenbogen, M.B. Martin: Activation of estrogen receptor- α by the heavy metal cadmium. *Mol Endocrinol* 14(4), 545-553 (2000). DOI: 10.1210/mend.14.4.0441
221. M.B. Martin, R. Reiter, T. Pham, Y.R. Avellanet, J. Camara, M. Lahm, E. Pentecost, K. Pratap, B.A. Gilmore, S. Divekar, R.S. Dagata, J.L. Bull, A. Stoica: Estrogen-like activity of metals in MCF-7 breast cancer cells. *Endocrinology* 144(6), 2425-2436 (2003). DOI: 10.1210/en.2002-221054
222. N.B. Aquino, M.B. Sevigny, J. Sabangan, M.C. Louie: Role of Cadmium and Nickel in Estrogen Receptor Signaling and Breast Cancer: Metalloestrogens or Not? *J Environ Sci Health C Environ Carcinog Ecotoxicol Rev* 30(3), 189-224 (2012). DOI: 10.1080%2F10590501.-2012.705159
223. G.G. Kuiper, E. Enmark, M. Peltö-Huikko, S. Nilsson, J.A. Gustafsson: Cloning of a novel receptor expressed in rat prostate and ovary. *Proc Natl Acad Sci USA* 93(12), 5925-5930 (1996). DOI: 10.1073%2Fpnas.93.12.5925
224. S.Y. Choe, S.J. Kim, H.G. Kim, J.H. Lee, Y. Choi, H. Lee, Y. Kim: Evaluation of estrogenicity of major heavy metals. *Sci Total Environ* 312(1-3), 15-21 (2003). DOI: 10.1016/S0048-9697(03)00190-6
225. C. Martinez-Campa, C. Alonso-Gonzalez, M.D. Mediavilla, S. Cos, A. Gonzalez, S. Ramos, E.J. Sanchez-Barcelo: Melatonin inhibits both ER α activation and breast cancer cell proliferation induced by a metalloestrogen, cadmium. *J Pineal Res* 40(4), 291-296 (2006). DOI: 10.1111/j.1600-079x.2006.00315.x
226. C.L. Siewit, B. Gengler, E. Vegas, R. Puckett, M.C. Louie: Cadmium promotes breast cancer cell proliferation by potentiating the interaction between ER α and c-Jun. *Mol Endocrinol* 24(5), 981-992 (2010). DOI: 10.1210%2Fme.2009-0410
227. V.S. Wilson, K. Bobseine, L.E. Gray: Development and characterization of a cell line that stably expresses an estrogen-responsive luciferase reporter for the detection of estrogen receptor agonist and antagonists. *Toxicol Sci* 81(1), 69-77 (2004). DOI: 10.1093/toxsci/kfh180
228. Z. Liu, X. Yu, Z.A. Shaikh: Rapid activation of ERK1/2 and AKT in human breast cancer cells by cadmium. *Toxicol Appl Pharmacol* 228(3), 286-294 (2008). DOI: 10.1016/j.taap.2007.12.017
229. Y. Zang, S. Odwin-Dacosta, J.D. Yager: Effects of cadmium on estrogen receptor mediated signaling and estrogen induced DNA synthesis in T47D human breast cancer cells. *Toxicol Lett* 184(2), 134-138 (2009). DOI: 10.1016%2Fj.toxlet.2008.10.032
230. M.O. Huff, S.L. Todd, A.L. Smith, J.T. Elpers, A.P. Smith, R.D. Murphy, A.S. Bleser-Shartz, J.E. Hoerter, B.N. Radde, C.M. Klinge: Arsenite and Cadmium Activate MAPK/ERK via Membrane Estrogen Receptors and G-Protein Coupled Estrogen Receptor Signaling in Human Lung

- Adenocarcinoma Cells. *Toxicol Sci* 152(1), 62-71 (2016).
DOI: 10.1093/toxsci/kfw064
231. S. Nilsson, S. Mäkelä, E. Treuter, M. Tujague, J. Thomsen, G. Andersson, E. Enmark, K. Pettersson, M. Warner, J.A. Gustafsson: Mechanisms of estrogen action. *Physiol Rev* 81(4), 1535-1565 (2001).
DOI: 10.1152/physrev.2001.81.4.1535
232. W.F. Anderson, N. Chatterjee, W.B. Ershler, O.W. Brawley: Estrogen receptor breast cancer phenotypes in the Surveillance, Epidemiology, and End Results database. *Breast Cancer Res Treat* 6(1), 27-36 (2002).
DOI: 10.1023/a:1020299707510
233. S.J. Han, F.J. DeMayo, J. Xu, S.Y. Tsai, M.J. Tsai, B.W. O'Malley: Steroid receptor coactivator (SRC)-1 and SRC-3 differentially modulate tissue-specific activation functions of the progesterone receptor. *Mol Endocrinol* 20(1), 45-55 (2006).
DOI: 10.1210/me.2005-0310
234. H. Lu, R.P. Fisher, P. Bailey, A.J. Levine: The CDK7-cycH-p36 complex of transcription factor IIH phosphorylates p53, enhancing its sequence-specific DNA binding activity *in vitro*. *Mol Cell Biol* 17(10), 5923-5934 (1997).
DOI: 10.1128/mcb.17.10.5923
235. L. Rulisek, J. Vondrasek: Coordination geometries of selected transition metal ions (Co²⁺, Ni²⁺, Cu²⁺, Zn²⁺, Cd²⁺, and Hg²⁺) in metalloproteins. *J Inorg Biochem* 71(3-4), 115-127 (1998).
DOI: 10.1016/s0162-0134(98)10042-9
236. M.D. Johnson, N. Kenney, A. Stoica, L. Hillakivi-Clarke, B. Singh, G. Chepko, R. Clarke, P.F. Sholler. A.A. Lirio, C. Foss, R. Reiter, B. Trock, S. Paik, M.B. Martin: Cadmium mimics the *in vivo* effects of estrogen in the uterus and mammary gland. *Nat Med* 9(8), 1081-1084 (2003).
DOI: 10.1038/nm902
237. D.A. Parodi, M. Greenfield, C. Evans, A. Chichura, A. Alpaugh, J. Williams, K.C. Cyrus, M.B. Martin: Alteration of Mammary Gland Development and Gene Expression by In Utero Exposure to Cadmium. *Int J Mol Sci* 18(9), 1939 (2017).
DOI: 10.3390/ijms18091939
238. L. Benbrahim-Tallaa, J. Liu, M.M. Webber, M.P. Waalkes: Estrogen signaling and disruption of androgen metabolism in acquired androgen-independence during cadmium carcinogenesis in human prostate epithelial cells. *Prostate* 67(2), 135-145 (2007).
DOI: 10.1002/pros.20479
239. M.É. Parent, J. Siemiatycki: Occupation and Prostate Cancer. *Epidemiol Rev* 23(1), 138-143 (2001).
DOI: 10.1093/oxfordjournals.-epirev.a000779
240. M.B. Martin, H.J. Voeller, E.P. Gelmann, J. Lu, E.G. Stoica, E.J. Hebert, R. Reter, B. Singh, M. Danielsen, E. Pentecost, A. Stoica: Role of cadmium in the regulation of AR gene expression and activity. *Endocrinol* 143(1), 263-275 (2002).
DOI: 10.1210/endo.143.1.8581
241. M.P. Donovan, L.G. Schein, J.A. Thomas: Inhibition of androgen-receptor interaction in mouse prostate gland cytosol by divalent metal ions. *Mol Pharmacol* 17(2), 156-162 (1980).

242. E.M. Wilson: Interconversion of androgen receptor forms by divalent cations and 8 S androgen receptor-promoting factor. Effects of Zn²⁺, Cd²⁺, Ca²⁺, and Mg²⁺. *J Biol Chem* 260(15), 8683–8689 (1985).
243. J. Ye, S. Wang, M. Barger, V. Castranova, X. Shi: Activation of androgen response element by cadmium: a potential mechanism for a carcinogenic effect of cadmium in the prostate. *J Environ Pathol Toxicol Oncol* 19(3), 275-280 (2000).
244. L.M. Lacorte, F.K. Delella, E.M. Porto Amorim, L.A. Justulin, A.F. Godinho, A.A. Almeida, P.F. Felipe Pinheiro, R.L. Amorim, S.L. Felisbino: Early changes induced by short-term low-dose cadmium exposure in rat ventral and dorsolateral prostates. *Microsc Res Tech* 74(11), 988-997 (2011).
DOI: 10.1002/jemt.20985
245. R. Wu, Y. Cui, X. Yuan, H. Yuan, Y. Wang, J. He, J. Zhao, S. Peng: SUMO-specific protease 1 modulates cadmium-augmented transcriptional activity of androgen receptor (AR) by reversing AR SUMOylation. *Toxicol Lett* 229(2), 405-413 (2014).
DOI: 10.1016/j.toxlet.2014.07.003
246. M.A. Gray, J.A. Centeno, D.P. Slaney, J.W. Ejnik, T. Todorov, J.N. Nacey: Environmental exposure to trace elements and prostate cancer in three New Zealand ethnic groups. *Int J Environ Res Public Health* 2(3–4), 374-384 (2005).
DOI: 10.3390/ijerph2005030001
247. E. van Wijngaarden, E.A. Singer, G.S. Palapattu: Prostate-specific antigen levels in relation to cadmium exposure and zinc intake: results from the 2001-2002 National Health and Nutrition Examination Survey. *Prostate* 68(2), 122-128 (2008).
DOI: 10.1002/pros.20668
248. C.C. Wu, Y.S. Pu, H.C. Wu, C.Y. Yang, Y.C. Chen: Reversed association between levels of prostate specific antigen and levels of blood cadmium and urinary cadmium. *Chemosphere* 83(8), 1188–1191 (2011).
DOI: 10.1016/j.chemosphere.-2010.12.085
249. A. Andreucci, E. Mocevic, B.A. Jonsson, A. Giwercman, Y.L. Giwercman, G. Toft, T. Lundh, D. Bizzaro, I.O. Specht, J.P. Bonde: Cadmium may impair prostate function as measured by prostate specific antigen in semen: a cross-sectional study among European and Inuit men. *Reprod Toxicol* 53, 33-38 (2015).
DOI: 10.1016/j.reprotox.2015.01.010
250. E.A. Platz, E. Giovannucci: The epidemiology of sex steroid hormones and their signaling and metabolic pathways in the etiology of prostate cancer. *J Steroid Biochem Mol Biol* 92(4), 237-253 (2004).
DOI: 10.1016/j.jsbmb.2004.10.002
251. M.P. Waalkes, S. Rehm, M.G. Cherian: Repeated cadmium exposures enhance the malignant progression of ensuing tumors in rats. *Toxicol Sci* 54(1), 110-120 (2000).
DOI: 10.1093/toxsci/54.1.110
252. J.I. Borráz-León, J. Herrera-Pérez, A.L. Cerda-Molina, L. Martínez-Mota: Testosterona y salud mental: una revisión. *Psiquiatría Biológica* 22(2), 44-49 (2015).
DOI: 10.1016/j.psiq.2015.10.005
253. I. Huhtaniemi, L.J. Pelliniemi: Fetal

- Leydig cells: cellular origin, morphology, life span, and special functional features. *Proc Soc Exp Biol Med* 201(2), 125-140 (1992).
DOI: 10.3181/00379727-201-43493
254. W.L. Miller, R.J. Auchus: The molecular biology, biochemistry, and physiology of human steroidogenesis and its disorders. *Endocr Rev* 32(1), 81-151 (2011).
DOI: 10.1210/er.2010-0013
 255. G.A. Ujah, V.U. Nna, M.I. Agah, L.O. Omue, C.B. Leku, E.E. Osim: Effect of quercetin on cadmium chloride-induced impairments in sexual behaviour and steroidogenesis in male Wistar rats. *Andrologia* 50(2), 1-9 (2018).
DOI: 10.1111/and.12866
 256. X. Wu, X. Guo, H. Wang, S. Zhou, L. Li, X. Chen, G. Wang, J. Liu, H.S. Ge, R.S. Ge: A brief exposure to cadmium impairs Leydig cell regeneration in the adult rat testis. *Sci Rep* 7(1), 6337 (2017).
DOI: 10.1038/s41598-017-06870-0
 257. D. Gunnarsson, M. Svensson, G. Selstam, G. Nordberg. Pronounced induction of testicular PGF (2 alpha) and suppression of testosterone by cadmium-prevention by zinc. *Toxicology* 200, 49-58 (2004).
DOI: 10.1016/j.tox.2004.03.003
 258. S.Y. Park, C. Gomes, S.D. Oh, J. Soh: Cadmium up-regulates transcription of the steroidogenic acute regulatory protein (StAR) gene through phosphorylated CREB rather than SF-1 in K28 cells. *J Toxicol Sci* 40(2), 151-161 (2015).
DOI: 10.2131/jts.40.151
 259. M.R. Farag, R.M.A. El-Aziz, H.A. Ali, S.A. Ahmed: Evaluating the ameliorative efficacy of *Spirulina platensis* on spermatogenesis and steroidogenesis in cadmium-intoxicated rats. *Environ Sci Pollut Res Int* 23(3), 2454-2466 (2016).
DOI: 10.1007/s11356-015-5314-9
 260. D.B. Hales, T. Diemer, K.H. Hales: Role of cytokines in testicular function. *Endocrine* 10(3), 201-217 (1999).
DOI: 10.1007/bf02738619
 261. S.R. Bornstein, H. Rutkowski, I. Vrezas: Cytokines and Steroidogenesis. *Mol Cell Endocrinol* 215(1-2), 135-141 (2004).
DOI: 10.1016/j.mce.2003.11.022
 262. K. Tremellen, N. McPhee, K. Pearce, S. Benson, M. Schedlowski, H. Engler: Endotoxin-initiated inflammation reduces testosterone production in men of reproductive age. *Am J Physiol Endocrinol Metab* 314(3), E206-E213 (2018).
DOI: 10.1152/ajpendo.00279.2017
 263. C. Togno-Peirce, O. Limón-Morales, S. Montes-López, J. Rojas-Castañeda, D. Márquez-Aguiluz, H. Bonilla-Jaime, M. Arteaga-Silva: Pleiotropic Effects of Cadmium Toxicity on the Neuroendocrine-Immune Network. *Advances in Neuroimmune Biology* 7(2), 1-13 (2018).
DOI: 10.3233/nib-180138
 264. J.M. Díez-Pina, M.J. Fernández-Aceñero, M.J. Llorente-Alonso, S. Díaz-Lobato, S. Mayoralas, A. Florez: Tumor necrosis factor alpha as a marker of systemic and local inflammation in "healthy" smokers. *Int J Gen Med* 2, 9-14 (2009).
DOI: 10.2147/ijgm.s4723
 265. K. Leisegang, R. Henkel: The *in vitro* modulation of steroidogenesis by inflammatory cytokines and insulin in TM3 Leydig cells. *Reprod Biol Endocrinol*

- 16(1), 26 (2018).
DOI: 10.1186/s12958-018-0341-2
266. E. Sarchielli, P. Comeglio, R. Squecco, L. Ballerini, T. Mello, G. Guarnieri, E. Idrizaj, B. Mazzanti, L. Vignozzi, P. Gallina, M. Maggi, G.B. Vannelli, A. Morelli: Tumor Necrosis Factor- α Impairs Kisspeptin Signaling in Human Gonadotropin-Releasing Hormone Primary Neurons. *J Clin Endocrinol Metab* 102(1), 46-56 (2017).
DOI: 10.1210/jc.2016-2115
 267. A.E. Turley, J.W. Zagorski, R.C. Kennedy, R.A. Freeborn, J.K. Bursley, J.R. Edwards, C.E. Rockwell: Chronic low-level cadmium exposure in rats affects cytokine production by activated T cells. *Toxicol Res (Camb)* 8(2), 227-237 (2019).
DOI: 10.1039/c2Fc8tx00194d
 268. A. Azenabor, A.O. Ekun, O. Akinloye: Impact of Inflammation on Male Reproductive Tract. *J Reprod Infertil* 16(3), 123-129 (2015).
 269. J. Veldhuis, R. Yang, F. Roelfsema, P. Takahashi: Proinflammatory Cytokine Infusion Attenuates LH's Feedforward on Testosterone Secretion: Modulation by Age. *J Clin Endocrinol Metab* 101(2), 539-549 (2016).
DOI: 10.1210/jc.2015-3611
 270. K. Tremellen, N. McPhee, K. Pearce, S. Benson, M. Schedlowski, H. Engler: Endotoxin-initiated inflammation reduces testosterone production in men of reproductive age. *Am J Physiol Endocrinol Metab* 314(3), E206-E213 (2018).
DOI: 10.1152/ajpendo.00279.2017
 271. A.A. Fouad, A.M. Abdel-Aziz, A.A.H. Hamouda: Diacerein Downregulates NLRP3/Caspase-1/IL-1 β and IL-6/STAT3 Pathways of Inflammation and Apoptosis in a Rat Model of Cadmium Testicular Toxicity. *Biol Trace Elem Res* (2019).
DOI: 10.1007/s12011-019-01865-6
 272. R. Li, X. Luo, L. Li, Q. Peng, Y. Yang, L. Zhao, M. Ma, Z. Hou: The Protective Effects of Melatonin Against Oxidative Stress and Inflammation Induced by Acute Cadmium Exposure in Mice Testis. *Biol Trace Elem Res* 170(1), 152-164 (2016).
DOI: 10.1007/s12011-015-0449-6
 273. H. Wang, R. Zhang, Y. Song, T. Li, M. Ge: Protective Effect of Ganoderma Triterpenoids on Cadmium-Induced Testicular Toxicity in Chickens. *Biol Trace Elem Res* 187(1), 281-290 (2019).
DOI: 10.1007/s12011-018-1364-4

Abbreviations: Abbreviations: HHT: hypothalamic-hypophysis-testis; Cd: cadmium; PM 2.5: particles measuring 2.5 micrometers; SO₂: sulfur dioxide, CO: carbon monoxide, NO₂: nitrogen dioxide, O₃: ozone, Hg: mercury, As: arsenic, Pb: lead, Cr: chrome, Ni-Cd: nickel-cadmium, CdO: cadmium oxide, CdCl₂: cadmium chloride, CdS: cadmium sulfate, WHO: World Health Organization, ROS: oxygen species, Cd²⁺: positive charges. Cd⁰: positive charges; Ca²⁺: calcium, Fe²⁺: iron, Mg²⁺: magnesium, Mn²⁺: manganese, Zn²⁺: zinc, Se: selenium, -SH: sulfhydryl or thiol group, VGCC: Ca voltage-gated channel, DMT: family; divalent metal transporters, ZIP: family of metal transporters, CaT 1: transporter CaT-1, TRPV: transient receptor potential cation channel, subfamily V, CatSper: voltage-dependent Ca channels, MT: metallothioneins, ATP: adenosine triphosphate, AIF: apoptosis-inducing factor, CaSR: Ca-sensitive receptor, PLC: C phospholipase, IP₃: inositol triphosphate, Bax: pro-apoptotic protein, CAT:

catalase, GPX: glutathione peroxidase, SOD: superoxide dismutase, GSH: glutathione, eIF4AE: transduction initiation factors located in eukaryotes, ERK1/2: extracellular signal-regulated kinase $\frac{1}{2}$, PI3K: phosphatidylinositol 3-kinase, Akt-PKB: protein kinase B, Ref1: redox effector factor 1, Nrf2: nuclear factor E2-related factor 2, NF- κ B: nuclear transcription factor-kappa B, Wnt: int/Wingless protein, Bcl-2: B-cell lymphoma 2, AP-1: activator protein-1, GnRH: gonadotrophin-releasing hormone, LH: luteinizing hormone, FSH: follicle-stimulating hormone, T: testosterone, 5HT: serotonin, GABA: γ -aminobutyric acid, AMPA: α -amino-3-hydroxy-5-methyl-4-isoxazole propionic acid, NMDA: N-methyl-D-aspartate, 5HT1A:serotonin 1A receptor, 5HT2: serotonin 2 receptor, BTB: blood-testicle barrier, Eps8: substrate of the epidermal growth factor 8 receptor pathway, AR: androgen receptor, cAMP: cyclic adenosine monophosphate, DHT: dihydrotestosterone, KISS: kisspeptin, KISS1R: kisspeptin receptor, GPR54: kisspeptin receptor, HHG: hypothalamic-hypophysis-gonad, GG: genital grooming, PS: preputial separation, ED: endocrine disruptors, ER: estrogen receptor, ER α : estrogen receptor alpha, ER β : estrogen receptor beta, GPR30: transmembrane estrogen receptor, SRC-1: steroid receptor coactivator-1, CBP: CREB-binding protein, P450scc: cytochrome enzyme, CYP11A: cholesterol side cleavage enzyme, PKA: kinase A, StAR: steroidogenic acute regulatory protein, 3 β -HSD: 3 β hydroxysteroid dehydrogenase Δ 4-5 isomerase enzyme, P-450c17: 17 α hydroxylase C17-20 lyase, 17 β HSD: 17 β -hydroxysteroid dehydrogenase enzyme, SR-BI: B type I Scavenger receptor, CREB: cAMP Response Element-Binding Protein, TNF- α : tumor necrosis factor-alpha, IL-1 β : interleukin-1beta, IL-6: interleukin-6, TM3: mouse Leydig cell line, IFN γ : interferon gamma, IL-10: interleukin-10.

Key Words: Cadmium, Male Reproduction, Fertility, Testicle, Epididymis, Prostate, Semen

quality, Immune system, Review

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