

Review

Antioxidant Cardioprotection in Acute Myocardial Infarction: From Mechanisms to Therapeutic Strategies

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Abstract

Acute myocardial infarction (AMI) is one of the main causes of mortality worldwide. Currently, the most effective treatment is percutaneous coronary angioplasty (PCA). However, paradoxically, the restoration of blood flow induces myocardial reperfusion injury (MRI), contributing up to 50% of the final infarct size. Oxidative stress, characterized by a burst of reactive oxygen species (ROS) following reperfusion, plays a fundamental role in its pathophysiology, causing inflammation, endothelial dysfunction, and cell death mainly through autophagy, apoptosis, ferroptosis, necroptosis, and pyroptosis. To mitigate these injury mechanisms, numerous antioxidant strategies have been evaluated using both *in vitro* and *in vivo* models with promising results, but limited benefit when tested in humans. Several antioxidants have biological properties that counteract ROS-induced damage by acting as ROS scavengers, metal chelators, and antioxidant enzyme enhancers. In this review, we focus on the mechanisms by which oxidative stress induces cell death after AMI and highlight the most promising therapeutic antioxidant agents that could provide comprehensive protection against MRI. A multitarget cardioprotective strategy, combining interventions with strong preclinical evidence, could provide a more effective approach for reducing MRI. Our study aims to bridge the gap between basic and clinical research and explore the potential clinical applications of antioxidants.

Keywords: acute myocardial infarction; oxidative stress; antioxidants; pharmacological cardioprotection

1. Introduction

Acute myocardial infarction (AMI) is a leading cause of mortality worldwide, causing 9 million deaths globally each year and more than 10% of the annual loss of disability-adjusted life-years [1]. Rapid and effective reperfusion therapy, primarily through percutaneous coronary intervention (PCI), has improved the treatment of AMI and improved survival rates [2]. However, paradoxically, following the restoration of blood flow, myocardial reperfusion injury (MRI) ensues, accounting for up to 50% of the final infarct size [3].

The pathophysiology of MRI is complex and multifactorial, involving oxidative stress, calcium overload, mitochondrial dysfunction, and inflammation, among other cellular mechanisms, that ultimately lead to programmed cell death pathways such as autophagy, apoptosis, necroptosis, ferroptosis, and pyroptosis [4].

Despite extensive research efforts, there are currently no effective therapies that can completely reduce MRI. Numerous pharmacological and mechanical interventions have been explored, including ischemic conditioning strategies and antioxidant therapies, but their translation into clinical practice has been largely unsuccessful [5]. One of the critical challenges in developing cardioprotective strategies is the multifactorial nature of MRI, which may require a multitarget approach to address the involvement of various mechanisms of injury [6].

In this review, we explore the current knowledge of MRI, focusing on the potential of combined antioxidant therapies to provide enhanced cardioprotection. We will also discuss the underlying mechanisms of MRI and programmed cell death, recent advances in therapeutic strategies, and the current challenges in translating preclinical findings into effective clinical treatments. Finally, we will

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present a novel multitherapy approach based on antioxidant cardioprotection.

2. The Challenge of Ischemia-Reperfusion Injury

2.1 Myocardial Ischemia

Ischemia initiates a cascade of cellular events characterized by hypoxia, which arrests oxidative phosphorylation, leading to mitochondrial membrane depolarization, adenosine triphosphate (ATP) depletion, and inhibition of myocardial contractile function [3]. A burst of reactive oxygen species (ROS) is generated early upon reperfusion. When the rate of ROS production exceeds the antioxidant potential of the heart, the increased concentration of these small reactive molecules, including oxygen free radicals such as superoxide and hydroxyl radicals, as well as non-radical species such as hydrogen peroxide, will exacerbate tissue damage [7].

As the duration of ischemia increases, maladaptive processes emerge, notably the accumulation of ROS from sources such as nicotinamide adenine dinucleotide phosphate (NADPH) oxidase (NOX) and mitochondrial reverse electron transport (RET) at complex I, triggered by the accumulation of succinate [8]. ROS production exacerbates mitochondrial dysfunction, leading to mitochondrial swelling and rupture, primarily through the opening of the mitochondrial permeability transition pore (mPTP). This event releases cytochrome-c into the cytosol, initiating apoptotic signaling [9].

The ischemic-induced proton accumulation within the cell further reduces intracellular pH, activating the Na⁺/H⁺ exchanger (NHE), which, combined with inhibition of the Na⁺/K⁺ ATPase due to lack of ATP, leads to excessive intracellular sodium. This sodium overload reverses the action of the Na⁺/Ca²⁺ exchanger, resulting in an accumulation of intracellular calcium, which exacerbates mitochondrial dysfunction and promotes further ROS generation. The resulting calcium overload and mitochondrial and DNA damage trigger programmed cell death pathways [10]. Once cytosolic membranes rupture, cellular contents are released into the extracellular space, acting as damage-associated molecular patterns (DAMPs) that can propagate cellular injury across neighboring cells [3].

2.2 Myocardial Reperfusion Injury

When blood flow is suddenly restored to the ischemic myocardium, the heart undergoes a complex series of damaging processes collectively called MRI. This phenomenon occurs due to several interconnected mechanisms, including oxidative stress, calcium overload, pH changes, and inflammation, all of which exacerbate the original ischemic damage [11].

Excessive ROS generation is a key driver of reperfusion injury, promoting inflammation, endothelial dysfunction, and programmed cell death through lipid peroxida-

tion and protein damage, ultimately leading to greater infarct size and impaired cardiac function. Mitochondria are the main source of intracellular ROS, mainly through the mitochondrial electron transport chain. RET at complex I plays a significant role in ROS generation, especially in the early stages of reperfusion [12]. In addition to mitochondria, NOX is activated during reperfusion and contributes to ROS production [13]. Moreover, the activity of xanthine oxidase (XO), uncoupled nitric oxide synthase (NOS), and activation of NOX in infiltrating leukocytes, such as neutrophils and macrophages, further exacerbate ROS accumulation and tissue damage [14]. Iron dysregulation also plays a critical role, primarily through increased intracellular free iron during reperfusion, which participates in the Fenton reaction. This reaction involves ferrous iron (Fe²⁺) reacting with hydrogen peroxide (H₂O₂), generating hydroxyl radicals (•OH) that exacerbate oxidative damage [15] (Fig. 1).

Oxidative stress is not solely driven by oxygen radicals; it also involves reduced bioavailability of nitric oxide (NO), a molecule with cardioprotective roles, including inhibition of neutrophil accumulation, superoxide scavenging, and enhancement of coronary blood flow through vasodilation [16]. However, NO's therapeutic potential is controversial, as it can form peroxynitrite (ONOO-), a reactive oxidant from NO and superoxide that oxidizes protein and non-protein thiols [17] and increases mitochondrial protein tyrosine nitration [18]. NO may either mitigate oxidative stress and prevent cell death [19] or, at higher concentrations, contribute to cellular damage [20].

In addition to oxidative stress, reperfusion leads to a phenomenon known as the calcium paradox. Normally, calcium is necessary for oxidative phosphorylation in the mitochondria. Ischemia and ATP depletion lead to intracellular calcium accumulation in cardiomyocytes and mitochondria primarily through the voltage-dependent anion channel (VDAC) and the mitochondrial calcium uniporter complex [21]. During reperfusion, increased ROS levels and mitogen-activated protein kinase (MAPK) activation [22] contribute to a feedback loop through phosphorylation of the sodium-hydrogen exchanger (NHE), raising intracellular sodium and, consequently, intracellular and mitochondrial calcium [10]. This calcium overload triggers a cascade of events, including mitochondrial dysfunction, cardiomyocyte hypercontracture, and the opening of the mPTP with the release of cytochrome-c, leading to further cell death. Although experimental evidence suggests that blocking this mechanism may protect against reperfusion injury [23,24], it has yet to be successfully translated to clinical trials [25,26].

Metabolic alterations add another layer of complexity to MRI. During ischemia, the myocardium switches from fatty acid metabolism to glucose metabolism due to limited oxygen availability [27], contributing to acidosis. After reperfusion, mPTP opens in the mitochondria and generates depolarization its membrane, which contributes to



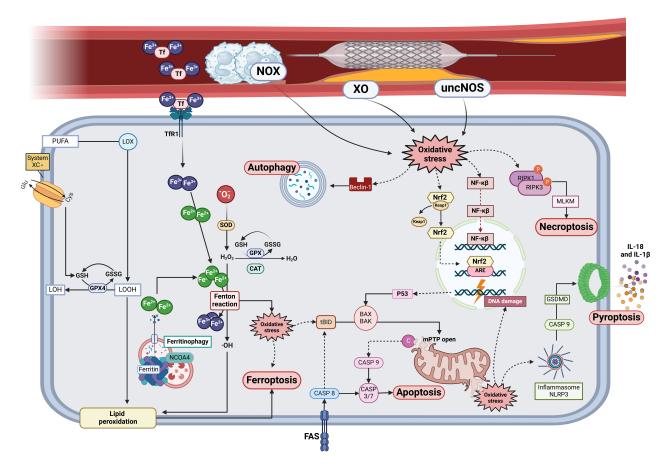


Fig. 1. Molecular mechanisms of generation of reactive oxygen species (ROS) and programmed cell death in myocardial reperfusion injury after percutaneous coronary angioplasty. ARE, antioxidant response element; BAX, Bcl-2 associated X protein; BAK, Bcl-2 homologous antagonist/killer; CAT, catalase; CASP, Caspase; Fe³⁺, ferric iron; Fe²⁺, ferrous iron; GPX, glutathione peroxidase; GSH, reduced glutathione; GSSG, oxidized glutathione; GSDMD, gasdermin D; GST, glutathione transferase; HO-1, heme oxygenase-1; IκB, NF-κB inhibitor protein; KEAP1, Kelch-like ECH-associated protein 1; LOX, lipoxygenases; LOH, lipid alcohol; LOOH, lipidhydroperoxides; MLKL, mixed lineage kinase domain-like; NCOA4, nuclear receptor coactivator 4; NF-κB, nuclear factor kappalight-chain-enhancer of activated B cells; NLRP3, NLR family pyrin domain containing 3; Nrf2, nuclear factor-erythroid 2-related factor 2; NOX, nicotinamide adenine dinucleotide phosphate oxidase; PUFA, polyunsaturated fatty acid; RIPK1, receptor interacting serine/threonine kinase 1; RIPK3, receptor interacting serine/threonine kinase 3; SOD, superoxide dismutase; Tf, transferrin; TfR1, transferrin receptor 1; tBID, truncated BCL homology domain 3 interacting domain; uncNOS, uncouple Nitric oxide synthase; Xc⁻ System, Cysteine/Glutamate Antiporter System; XO, xanthine oxidase. Created with BioRender.com.

the uncoupling of oxidative phosphorylation, aggravating ATP depletion [28]. This insight has led to therapeutic approaches such as glucose-insulin-potassium (GIK) therapy, which will be discussed later.

Inflammation also plays a critical role in reperfusion injury. Within hours of reperfusion, neutrophils are recruited to the damaged myocardium, adhering to the endothelium, releasing ROS, and secreting degradative enzymes. This inflammatory response exacerbates tissue damage by clogging capillaries and increases oxidative stress [29]. Therefore, the goal would be to diminish the initial proinflammatory phase. While experimental interventions aimed at reducing neutrophil activity, such as using leukocyte-depleted blood, blocking cell adhesion molecules, or inhibiting complement, have shown potential

[30–32], these strategies have not been consistently translated into clinical success [33,34].

2.3 Second Messenger Pathways in Myocardial Reperfusion Injury

Normally, 2 to 5 percent of the oxygen consumed by mitochondria is converted to superoxide and neutralized by the cell's antioxidant machinery, which involves catalase, glutathione peroxidase, and superoxide dismutase, among others [35]. In MRI, excessive ROS production also activates multiple second messenger pathways, besides all the previously described mechanisms of cell injury, which have also been the target of direct pharmacological interventions or are aimed to be regulated if oxidative stress is controlled.



MAPK is a group of enzymes that includes extracellular regulated kinases (ERKs), p38 kinases, and c-Jun N-terminal kinases (JNKs). The two latter are activated by oxidative stress and are pro-apoptotic [36]. The phosphoinositide-3 kinase/protein kinase B (PI3K/Akt) pathway is upregulated after MRI [37,38]. Downstream effectors of Akt include the endothelial Nitric Oxide Synthase (eNOS), mammalian target of Rapamycin (mTOR), glycogen synthase kinase 3β (GSK 3β), and forkhead box subfamily O (FOXO), which also regulates survival, apoptotic and autophagy signaling [39,40].

The Janus kinase/signal transducer and activator of transcription (JAK/STAT) pathway is a signaling mechanism involving tyrosine kinases associated with the intracellular domains of membrane-bound receptors. This pathway facilitates communication between cell surface receptors and the nucleus, which regulates cell survival and apoptosis. Specifically, STAT1 promotes apoptosis, while STAT3 exerts an anti-apoptotic effect [41].

3. Programmed Cell Death in Myocardial Ischemia-Reperfusion Injury

The loss of cardiomyocytes is the direct consequence of cell death. Clinically, this translates into a greater infarct size, a significant determinant in the prognosis of these patients [2]. Understanding the mechanisms of cell death and their interactions is crucial for developing targeted therapies aimed at mitigating damage. Such insights could help design synergistic treatments to improve outcomes in patients with AMI. The connection between the oxidative stress generated by ischemia/reperfusion (IR) and programmed cell death is outlined in Fig. 1.

3.1 Autophagy

Autophagy is a component of cellular homeostasis responsible for degrading and recycling damaged proteins and cytoplasmic organelles. In response to ischemic stress, autophagy is activated mainly by inhibiting mTOR [42]. This process degrades macromolecules and organelles by transporting them to lysosomes in double-membrane vesicles termed autophagosomes. Autophagy is regulated by complexes involving beclin-1 and autophagy-related genes (ATGs), ensuring that damaged proteins and organelles are compartmentalized and removed before they trigger pro-apoptotic pathways [43]. However, autophagy can be double-edged. Excessive autophagy can cause autodigestion and cell death, while its inhibition could prevent the removal of damaged organelles, pushing cells towards apoptosis or necroptosis.

Beclin-1 is essential for autophagosome formation. Discordantly, its deletion ameliorates cardiac dysfunction resulting from severe pressure overload. Its overexpression leads to increased autophagy and an exacerbation of cardiac dysfunction [44], and increased apoptosis due to decreased

beclin-1 leads to a reduction in infarct size [45]. In contrast, deletion of ATG5 in adult cardiomyocytes caused heart failure under baseline and pressure overload conditions. These opposing results highlight the complex role of autophagy in cardiac pathology [46].

3.2 Apoptosis

Apoptosis is a regulated form of cell death characterized by cell shrinkage, chromatin condensation, and plasma membrane blebbing without rupture [47]. This pathway is initiated through two main mechanisms: the extrinsic pathway, which involves the activation of cell surface receptors (e.g., Tumor necrosis factor receptor 1 [TNFR1], Fas receptor, TNF-related apoptosis-inducing ligand [TRAIL]) leading to caspase-8 activation, and the intrinsic pathway, which is triggered by mitochondrial signals, such as cytochrome-c release and the formation of the apoptosome complex (Apoptotic protease activating factor 1, caspase-9) that ultimately activates caspase-3. Besides ROS production and mitochondrial calcium overload, the B-cell lymphoma 2 (Bcl-2) protein family proteins contribute to pore formation and mitochondrial outer membrane permeabilization allowing the release of cytochrome-c. Bcl-2 associated X protein (BAX)/Bcl-2 homologous antagonist/killer (BAK) promote apoptosis through mitochondrial membrane permeabilization, while survival pathways upregulate anti-apoptotic members such as Bcl-2. Also, VDAC increases mitochondrial matrix calcium concentration, contributing to ROS generation [48].

Apoptosis in myocardial reperfusion injury is notable for its non-inflammatory nature, as macrophages clear apoptotic cells without inducing an inflammatory response. However, excessive apoptosis can interfere with cell survival mechanisms, including autophagy, and may lead to unnecessary cell death. By reducing apoptosis, broad caspase inhibitors have been shown to limit MRI settings, supporting the therapeutic potential of targeting apoptotic pathways to mitigate damage [49].

Another key apoptosis regulator in cardiomyocytes is the apoptosis repressor with caspase recruitment domain (ARC). ARC inhibits apoptosis by interacting with both death receptors and mitochondrial pathways. ARC also interacts with p53, preventing its pro-apoptotic transcriptional activities [50]. Overexpression of ARC in cardiomyocytes reduces infarct size in animal models, while global ARC deletion exacerbates myocardial damage [51]. Furthermore, caspase inhibitors have shown variable success in reducing infarct size and cardiac dysfunction post-AMI [52]. Through caspase inhibition, the preservation of contractile proteins such as troponin T may mitigate some of the functional losses associated with reperfusion injury.

3.3 Necroptosis

Programmed necrosis, also called necroptosis, leads to cell swelling, formation of a necrosome, and plasma



membrane rupture, releasing cellular content and promoting an inflammatory response. The process is triggered through death receptors such as TNFR1 and Fas, activating receptor-interacting serine/threonine-protein kinase (RIPK) 1 and RIPK3. When caspase 8 activity is suppressed, RIP1 and RIP3 interact, forming the necrosome complex with mixed lineage kinase domain-like (MLKL), which subsequently oligomerizes and translocates to the plasma membrane, causing rupture [53].

Additionally, RIPK3 can phosphorylate Ca²⁺/calmodulin-dependent protein kinase (CaMKII) and Phosphoglycerate mutase, contributing to mPTP opening and mitochondrial dysfunction, which enhances ROS production and cell death. Also, extracellular RIPK3 has a role as a DAMP, binding to the receptor for advanced glycation end-products (RAGE), activating CaMKII and exacerbating myocardial injury [54]. Notably, it has been shown that necrostatin-1, an inhibitor of RIPK1, reduces IR injury (IRI), suggesting potential for therapeutic intervention [55].

3.4 Ferroptosis

Iron dysregulation has emerged as a key player in MRI and the pathophysiology of other cardiovascular diseases such as heart failure, cardiac hypertrophy, diabetic cardiomyopathy, and septic heart injury, positioning iron as a substrate for cardiovascular disease [56–58].

Ferroptosis, a form of regulated cell death driven by iron and ROS, is increasingly recognized as a critical mechanism in MRI and occurs when imbalances in iron and ROS lead to lipid peroxidation, mitochondrial damage, and overall organelle dysfunction [59].

Transferrin receptor 1 (TfR1) is a membrane protein that facilitates iron transfer from the extracellular environment into cells, thereby contributing to the intracellular iron pool necessary for ferroptosis. It regulates iron uptake and ensures iron is stored in a non-toxic form within ferritin [60]. Notably, TfR1 is highly expressed in the ischemic myocardium, leading to an influx of iron ions into cardiomyocytes during this phase [61].

Ferritin is a cellular iron storage protein that regulates iron efflux [62]. Its heavy subunit (FTH) has ferroxidase activity that converts Fe²⁺ to Fe³⁺ for storage inside the shell, being the main iron storage protein [63]. Ferritinophagy is an autophagy process that degrades ferritin [64]. This process is mediated by the nuclear receptor coactivator 4 (NCOA4), an autophagy cargo receptor that binds FTH1 and is delivered into the autolysosome for degradation, releasing free iron [65]. Notably, levels of FTH1 are decreased in myocardial tissues that undergo IR [66]. Subsequently, it was shown that glutaminolysis inhibition attenuated myocardial IRI by blocking ferroptosis [67].

The inhibition of the cystine/glutamate antiporter (system Xc⁻) depletes intracellular reduced glutathione (GSH), leading to the inactivation of glutathione peroxidase 4

(GPX4), which, under normal conditions, plays a key role in neutralizing lipid peroxides [68]. Down-regulation of GPX4, oxidizing GSH, and generating ferric iron from the Fenton reaction lead to lipid peroxidation and ferroptotic mitochondrial injury. Ferroptotic ROS generation contributes to myocardial cell death, and interestingly, iron chelators improve myocardial survival [69]. The substrates for peroxidation are phospholipids with polyunsaturated fatty acids (PL-PUFAs) [70]. ROS that originate from the Fenton reaction are catalyzed by iron-dependent lipoxygenases (LOXs). The obtained PL-PUFAs are then oxidized into lipid hydroperoxides (PL-PUFAs-OOH), which trigger ferroptosis [71].

Ferroptosis is primarily initiated during the reperfusion phase rather than during ischemia [72]. This observation underscores the importance of post-ischemic iron homeostasis, as clinical data indicate that iron overload is an independent risk factor for adverse left ventricular remodeling following reperfusion [73,74].

Although ferroptosis is closely related to other types of programmed cell death, it is distinct from other forms of cell death, such as apoptosis, as it primarily involves oxidative membrane damage without significant nuclear involvement. Its regulation is influenced by key enzymes such as acyl-CoA synthetase long-chain family member 4 (ACSL4), which promotes the incorporation of PUFA into phospholipids, making membranes more susceptible to peroxidation [75]. Indeed, ACSL4 overexpression transfection blocked cardiomyocyte protective effects by an antioxidant substance [76]. Other molecules are the ferroptosis suppressor protein 1 (FSP1), which works independently of GPX4 and helps reduce lipid peroxides, providing an alternative mechanism of ferroptosis resistance [77] and the Coenzyme Q10 (CoQ10), which is another antioxidant involved in preventing lipid peroxidation. FSP1 uses CoQ10 to counteract oxidative damage [78]. A specific system, Xc-SLC7A11, is tightly involved in ferroptosis regulation, and modulates different pathways inhibiting ferroptosis [79]. Nuclear factor-erythroid 2-related factor 2 (Nrf2) has been widely investigated as a negative ferroptosis regulator, as it inhibits ROS production and reduces intracellular iron uptake. A previous study identified GPX4 and SLC7A11 as transcriptional downstream targets of Nrf2 [80].

So far, there is some monotherapy in human studies of several compounds focused on targeting ferroptosis in MRI *in vitro* and *in vivo*, in which different mechanisms have been proposed as potential pathways and strategies to mitigate the MRI. While most compounds are studied in the form of monotherapy, some that act on different pathways such as apoptosis and ferroptosis—referred to by some authors as the "strategy of killing two birds with one stone"—greatly enhance the therapeutic effect on MRI without the need for additional pharmaceutical excipients, offering strong potential for clinical application [61]. Iron



chelators like Deferoxamine and Ferrostatine-1 have been effective in reducing cardiac dysfunction, cell death, and infarct size [15,67].

3.5 Pyroptosis

Pyroptosis is a regulated form of cell death characterized by the permeabilization of the plasma membrane and the release of inflammatory cytokines into the extracellular space, exacerbating inflammation and tissue damage. This process is mediated by gasdermin D (GSDMD), a protein that in its inactive form is auto inhibited. Upon activation by caspases, such as caspase-1 or caspase-11, GSDMD undergoes proteolytic cleavage, which then embeds into the cell membrane to form pores. These pores lead to cell swelling, membrane rupture, and the release of inflammatory signals [81].

The NLR family pyrin domain containing 3 (NLRP3) inflammasome is a key activator of pyroptosis, mainly through the activation of caspase-1, which not only cleaves GSDMD but also processes the proinflammatory cytokines interleukin-1 β (IL-1 β) and interleukin-18 (IL-18) for release. The release of IL-1 β and IL-18 drives inflammation, contributing to tissue damage in IRI. Evidence shows that in myocardial infarction, particularly following PCI in patients with AMI, serum levels of both GSDMD and IL-18 increase, highlighting the role of pyroptosis in cardiac damage [82,83].

4. Therapies for Myocardial Ischemia-Reperfusion Injury Against Oxidative Stress

Preclinical studies have shown numerous promising cardioprotective agents, some of which will be mentioned in the following sections. In particular, oxidative stress plays a central role in MRI, prompting extensive research into natural and synthetic antioxidant compounds. While many antioxidant agents have shown promising results in preclinical models, few have progressed to clinical trials. This section highlights the results from clinical studies evaluating that have assessed antioxidant agents that have been tested in humans. Rather than providing an exhaustive list, we have focused on the largest trials and recent metaanalyses that have assessed the potential cardioprotective effects of these agents (Table 1, Ref. [84-138]). Additionally, we provide a brief overview of the mechanisms of action of these antioxidants, along with the challenges and limitations that have hindered their translation into routine clinical practice. This perspective aims to offer a clearer understanding of both their therapeutic potential and the barriers to their widespread adoption.

4.1 Melatonin

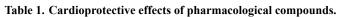
Melatonin is a lipophilic hormone predominantly produced predominantly by the pineal gland in most living organisms, primarily responsible for regulating circadian rhythms, particularly the sleep-wake cycle. Clinically, melatonin has been employed for decades to treat conditions related to altered biological rhythms and sleep disorders [84]. Over time, additional therapeutic properties of melatonin have been identified, including its potent anti-inflammatory and antioxidant effects, leading to its investigation in various conditions [84].

More recently, the potential of melatonin to mitigate IRI has attracted increasing attention. Its antioxidant and cardioprotective effects are mainly attributed to its ability to directly scavenge free radicals, an action also shared by several of its metabolites [85]. Furthermore, melatonin enhances the activity of endogenous antioxidant enzymes by upregulating the transcription factor Nrf2 and sirtuin 1 (SIRT1) [86,139]. SIRT1, a protein deacetylase, is stimulated by melatonin to enhance the stability and nuclear translocation of Nrf2, boosting the production of antioxidant enzymes and suppressing pro-inflammatory signaling pathways, such as nuclear factor kappa-light-chainenhancer of activated B cells (NF- κ B), p53 [140] and inhibiting the NLRP3 inflammasome [141], while promoting mitochondrial function and biogenesis [142], thereby preserving energy production in cardiac cells, which protect the heart from myocardial IRI.

It also exerts anti-apoptotic effects by activating the JAK2/STAT3 signaling pathway [87]. Additional proposed mechanisms include inhibiting autophagy and suppressing quinone oxidoreductase two activity, although these remain debated within the scientific community [84,87].

These properties have provided the foundation for randomized controlled trials (RCTs) evaluating the efficacy of melatonin in preventing IRI in patients with AMI with mixed results. In patients with ST-segment elevation myocardial infarction (STEMI) melatonin administration during reperfusion did not significantly improve the myocardial salvage index or reduce cardiac biomarkers (hs-cTnT, creatine kinase (CK)-MB) and oxidative stress markers compared to placebo [88]. However, a post-hoc analysis from the Melatonin Adjunct in Acute Myocardial Infarction Treated with Angioplasty trial suggested a time-dependent effect, with early melatonin administration (within 136 minutes of symptom onset) associated with smaller infarct size, whereas delayed administration (249 minutes) increased infarct size [89]. In coronary artery bypass graft (CABG) surgery patients, melatonin demonstrated dosedependent benefits, including improved left ventricular ejection fraction (LVEF), reduced heart rate, cardiac injury biomarkers, inflammatory markers, and apoptosis, with higher doses (20 mg) yielding stronger effects [90]. Another CABG trial reported reduced postoperative CK-MB levels and shorter intensive care unit (ICU) stays with melatonin supplementation, though without significant changes in other biomarkers [91].





Compound	Pathway/Mechanism involved	Results in humans
Melatonin	Free radical scavenger [85].	Lowered CK-MB levels on postoperative days 2 and 3 after CABG surgery, and shortened ICU stay, without significant changes
		in other inflammatory markers [88].
	Nrf2/ARE [86].	Improved LVEF and reduced HR. Reduced levels of cTnI, IL-1 β , iNOS, and caspase-3, with greater effects in the high-dose group
		[89].
	SIRT1 [84].	Early administration significantly reduced infarct size, while late administration was associated with a larger infarct size [90].
	Decreases apoptosis (JAK2/STAT3) [87].	Did not improve the myocardial salvage index or levels of troponin T, CK-MB, or oxidative stress biomarkers compared to placebo [91].
N-acetylcysteine	ROS scavenger and precursor for glutathione biosynthesis [116].	Decreased plasma MDA levels, LVEF was higher, there was no difference in reduction of infarct size according to CK-MB levels [93].
		Reduced infarct size in patients with STEMI undergoing PCI [92].
		Reduced early cardiac remodeling by diminishing levels of MMP-2 and MMP-9, reduced hospital stays and reduced the incidence of MACE [94].
Allopurinol	Inhibits XO. Free radical scavenger [95].	Treatment improves the TIMI flow. No improvement in cardiovascular events, complications, troponin levels and ECG ST-
		elevation regression [96].
		Reduces urinary isoprostanes levels. Higher recovery of LVEF [96].
		$More\ effective\ ST-elevation\ recovery\ and\ lower\ cTnI, CK\ and\ CK-MB\ peak\ values.\ Lower\ incidence\ of\ MACE, with\ no\ significant$
		differences in LVEF [97].
Edaravone	Free radical scavenger [98].	Reduced incidence of ventricular tachyarrhythmias [101].
	Decreases apoptosis (JAK2/STAT3) [99].	Reduction of reperfusion arrhythmias, myocardial stunning, and lethal reperfusion injury [100].
		Improved left ventricular systolic function immediately after reperfusion.
		Higher LVEF and fewer heart failure-related rehospitalizations at 12 months [100].
Sodium thiosulphate	Inhibits mitochondrial complex IV, increases SOD activity [102].	No clinical benefit in reducing reperfusion injury in a sample with low risk for large infarction [104].
NT: 111	Activates Nrf2 [103].	
Nicorandil	Stimulates mitochondrial ATP-sensitive potassium channels [105].	Reduced the incidence of the no-reflow phenomenon and MACEs, A combination of intracoronary and intravenous administration further reduced the incidence of MACEs [107].
	Decreases pyroptosis (TLR4/MyD88/NF-κB/NLRP3)	No significant difference in infarct size, cardiac function, or MACEs between the nicorandil and placebo groups over 6 months
	[106].	[108].
Quercetin	SIRT1 [109].	Lower CK-MB AUC, suggesting a smaller infarct size. Reduced the incidence of reperfusion-induced intramyocardial hemor-
		rhage. No significant differences were noted in LVEF or LV remodeling indicators [110].
Ascorbic Acid	Free radical scavenger. Nrf2/ARE [117].	Meta-analysis of seven controlled trials (872 patients) examining peak cTn and CK-MB levels post-procedure and oxidative stress
		biomarkers. AA reduced peak cTn levels by 43% and peak CK-MB levels by 14% and decreased oxidative stress biomarkers [120].
	Inhibits NADPH oxidase synthesis [118].	Randomized study with 532 patients receiving either a 3-g AA infusion or normal saline before PCI. The incidence of myocardial
		injury was significantly lower in the AA group. AA use was an independent predictor of reduced myocardial injury [121].
	Prevents the oxidation of BH4 [119].	

Table 1. Continued.

Compound	Pathway/Mechanism involved	Results in humans	
Statins	Nrf2/ARE [115].	Early high-dose rosuvastatin therapy did not improve myocardial perfusion or reduce infarct volume compared to convention	
		low-dose therapy in STEMI patients undergoing primary PCI [116].	
	Decreases the activity of NADPH oxidase [114].	High-dose atorvastatin pretreatment followed by continued treatment for 5 days did not reduce infarct size in STEMI patients undergoing primary PCI [122].	
	Reduces the expression of LOX-1 [111–113].		
Dexmedetomidine	AMPK/GSK-3 β /Nrf2 axis [123].	Decrease in cTnI and TNF- α levels, and increased concentrations of IL-6 and IL-8 after 24 hours of cardiopulmonary bypass. No significant changes in MDA content or SOD activity. Lower proportion of anemia cases [124].	
Dapagliflozin	MAPK signaling inhibition [125].	The primary hierarchical composite outcome showed significantly more wins for DAPA compared to placebo. No significant difference in the composite of cardiovascular death or hospitalization for heart failure.	
		No significant reduction in cardiovascular death or hospitalization for heart failure compared to placebo [126].	
Empagliflozin	AMPK/Nrf2 [127].	No significant difference of either a first hospitalization for heart failure or death from any cause compared to placebo [128].	
Puerarin	AMPK [129].	Fewer angina pectoris attacks and ST segment changes during the balloon dilation stage of PCA compared to the conventional group.	
		Reduced blood levels of vWF and ET-1, while increasing NO levels, compared to the conventional group [130].	
Sevoflurane	Not yet clear [131].	No reduction in the incidence of cardiac and non-cardiac events during the 6 months following cardiac surgery involving ex-	
Severalane	The yet view [137].	tracorporeal circulation. Trend towards reduced treatment needs and fewer hospital admissions, when experiencing any events [131,132].	
Propofol	Akt/p53 [133].	After a CPB, compared with SEV, propofol reduced incidence of AKI, decreased inflammatory markers such as IL-6, CRP and segmented neutrophil counts [134].	
Deferoxamine	Chelates non-transferrin bound iron (free iron), iron in	Reduced post-PCI serum iron. No differences in serum ferritin, soluble transferrin receptor, F2-isoprostane levels, CRP levels,	
	transit between transferrin and ferritin (labile chelating iron pool), hemosiderin, and ferritin [135].	infarct size, creatine kinase, cTnI, the mean ST-segment resolution, the degree of wall motion abnormality, LVEF [136].	
		No significant difference between preischemia and reperfusion in the indirect measure of ROS activity. Better preservation of	
		myocardial cells. Reduction in the number of severely damaged mitochondria [137].	
		Reduction in thiobarbituric reactive substances (TBARS) levels, LVEF improvement, better myocardial recovery reflected by wall motion score index improvement [138].	

AA, ascorbic acid; AMPK, AMP-activated protein kinase; AKI, acute kidney injury; ARE, antioxidant response element; ATP, adenosine triphosphate; AUC, area under curve; BH4, tetrahydrobiopterin; cTnI, cardiac troponin I; CABG, coronary artery bypass graft; CPB, cardiopulmonary bypass; CK-MB, creatine kinase-MB; CRP, C-reactive protein; ECG, electrocardiogram; ET-1, endothelin-1; GSK-3β, glycogen synthase kinase 3 beta; HR, heart rate; ICU, intensive care unit; IL, interleukin; iNOS, inducible nitric oxide synthase; IV, intravenous; JAK2/STAT3, janus kinase 2/signal transducer and activator of transcription 3; LOX-1, lectin-like oxidized low-density lipoprotein receptor 1; LVEF, left ventricular ejection fraction; MACE, major adverse cardiaovascular events; MAPK, mitogen-activated protein kinase; MDA, malondialdehyde; MMP-2, metalloproteinase-2; MyD88, myeloid differentiation primary response 88; NADPH, nicotinamide adenine dinucleotide phosphate, NF-κB, nuclear factor kappa-light-chain-enhancer of activated B cells; NO, nitric oxide; Nrf2, nuclear factor erythroid 2-related factor 2; PCA, percutaneous coronary angioplasty; PCI, percutaneous coronary intervention; ROS, reactive oxygen species; SIRT1, sirtuin 1; SEV, sevoflurane; SOD, superoxide dismutase; STEMI, ST-segment elevation myocardial infarction; TBARS, thiobarbituric acid reactive substances; TIMI, thrombolysis in myocardial infarction; TLR4, toll-like receptor 4; TNF-α, tumor necrosis factor-alpha; vWF, von Willebrand factor; XO, xanthine oxidase.



A meta-analysis of several trials concluded that melatonin did not improve cardiac function or reduce infarct size [143]. The variability in trial methodologies, including differences in melatonin dosages (most studies used 50 mg intravenous (IV), nonetheless, dosages other treatment regimens 2 mg intracoronary (IC), and 3 mg per os (PO) routes of administration (PO, IV, and IC) could have explained the heterogeneity in infarct size reduction measured by troponin levels, since there is no consensus of the optimal clinical plasma concentration and mode of administration [144], and timing of treatment, may have influenced these findings. Notably, early administration, whether IC or IV, was associated with reductions in infarct size and improvements in LVEF, while delayed administration showed the opposite effect [143]. Regarding the measurement of the MRI, few trials have used cardiac magnetic resonance imaging (CMRI) to measure infarct size, while most of them rely on troponin levels [88,89]. From a safety perspective, melatonin administration has generally been well tolerated, with no significant adverse effects reported [145].

So far, the combination therapies with melatonin in the heart is not well studied. A recent study evaluated the synergistic effects of melatonin and captopril in a rat model of chronic heart failure. This combination improved systolic blood pressure, left ventricular function, while reducing myocardial injury markers and oxidative stress. The study demonstrated that the treatments effectively inhibited apoptosis by reducing caspase-3 activity and alleviated fibrosis by lowering the levels of transforming growth factor-beta 1 (TGF- β 1) and collagen-I, with the combination therapy of melatonin and captopril showing the most pronounced effects. Additionally, mitochondrial dynamics were significantly improved, as evidenced by enhanced mitophagy, boosted biogenesis, and improved mitochondrial fusion. Simultaneously, mitochondrial fission was reduced, contributing to the restoration of mitochondrial homeostasis and overall cellular function. Despite these promising results, its applicability to AMI remains limited [146].

Future research should focus on optimizing the timing and mode of melatonin administration to maximize its cardioprotective effects. Additionally, more extensive clinical trials are needed to account for patient-related factors such as age, comorbidities, and gender. Finally, further investigation is required to elucidate the precise mechanisms underlying melatonin's therapeutic actions, as many remain incompletely understood.

4.2 N-Acetylcysteine

N-acetylcysteine (NAC) is a compound derived from the amino acid L-cysteine, which has antioxidant properties and therapeutic uses in different clinical settings. It replenishes glutathione, a crucial antioxidant that helps protect cells from oxidative stress and damage [147]; it is also a reductant of disulfide bonds, a scavenger of reactive oxygen species, and its mechanism involves the inhibition of NF- κ B activation and neurokinin A production, which together result in reduced interleukin-6 production, thereby alleviating inflammation and oxidative stress [148].

Moreover, NAC has shown potential benefits in treating AMI. A study showed that oral supplementation of 600 mg NAC every eight hours for 72 hours can reduce high-sensitivity C-reactive protein, myeloperoxidase and Galectin-3 levels in AMI patients receiving fibrinolytic therapy [149]. Another study in patients who received fibrinolytic therapy plus NAC demonstrated that, compared to controls, plasma malondialdehyde (MDA) levels decreased, and LVEF was higher. However, there was no difference in infarct size according to CK-MB levels [150]. Furthermore, NAC may help prevent early cardiac remodeling by reducing levels of metalloproteinase (MMP)-2 and MMP-9. Additionally, it has been shown to shorten hospital stays in patients after AMI and also reduce the incidence of major adverse cardiovascular events (MACE) [151].

Despite these positive findings, evidence on the effect of NAC in reducing MRI in humans remains inconclusive [152,153]. One possible explanation is that the antioxidant effects of other agents used in reperfusion interventions, such as halogenated anesthetics or propofol, may diminish the observable benefit of NAC. Additionally, variability in NAC dosage (ranging from 4 to 300 mg/kg), treatment duration (1 hour to 5 days), and administration routes (PO, IV, or a combination of both) may have influenced the results. Also, clinical trials have been performed in different populations. Although they present MRI, differences between patients undergoing CABG, PCI, or thrombolysis make it difficult to draw broadly applicable conclusions.

In addition, NAC has been studied in combination with other treatments, such as nitroglycerin, to improve blood flow in patients experiencing AMI. Some trials have shown that when administered intravenously, high-dose NAC can enhance the effects of low-dose intravenous nitroglycerin, leading to better blood vessel dilation and increased oxygen delivery to heart tissue. This is related to a reduced infarct size in patients with STEMI undergoing PCI [92]. However, there are no recent studies in humans and the high incidence of side effects limits the clinical applicability of this therapeutic strategy [154]. In rats, a treatment combining NAC and allopurinol synergistically enhanced cardiac adiponectin (APN) content, an adipokine anti-ischemic properties, and reduced IRI. NAC alone increased cardiac APN levels and AdipoR2 expression, while allopurinol amplified NAC's effects, restoring key signaling pathways. Both NAC and allopurinol independently reduced myocardial infarct size and CK-MB release, with their combination demonstrating synergistic cardioprotective effects [93].

Regarding biomarkers, comparisons between levels of TGF- β and TNF- α after 24 and 72 hours within the NAC or placebo groups in patients with AMI revealed that there was not any significant difference in TNF- α levels. However,



NAC could prevent TGF- β levels from increasing after 72 hours, and this biomarker had strong correlations with the LVEF, suggesting it to be important in the prevention of remodeling [94].

While research is promising, more large-scale studies are needed to fully confirm the benefits of this safe drug in reducing damage and improving recovery in patients with AMI.

4.3 Allopurinol

Allopurinol is a medication primarily used to treat gout, some kidney stones, and cardiovascular diseases [94]. It is a purine base analog and works by inhibiting the enzyme xanthine oxidoreductase (XOR), which exists in two isoforms *in vivo*: xanthine dehydrogenase (XDH) and XO. XDH produces NADH and uric acid, the latter acting as a free radical scavenger, while XO catalyzes the formation of both uric acid and superoxide anion. Under conditions of ischemia and hypoxia, ATP depletion induces a reversible conversion of XDH to XO through the oxidation of sulfhydryl groups. Upon reperfusion, increased oxygen availability and elevated XO activity result in excessive ROS generation, exacerbating oxidative stress. In this context, allopurinol effectively inhibits XO activity and mitigates ROS production [155].

In the context of MRI in patients with STEMI, there is methodological heterogeneity. Further supporting its antioxidant role, when given before reperfusion in patients with AMI, the group treated with allopurinol showed reduced urinary isoprostane levels, a biomarker produced when free radicals cause lipid peroxidation in cell membranes. Also, there was a recovery of LVEF [156]. In a different RCT, allopurinol pre-treatment in 140 patients undergoing fibrinolysis reported improved ST-segment resolution at 90 minutes, reduced infarct size (as measured enzymatically), and a lower incidence of in-hospital MACE [95]. It has also shown an improvement in thrombolysis in myocardial infarction (TIMI) flow following PCI. This effect was expected considering the localization of ROS in capillary endothelial cells, along with cell swelling, activated neutrophils, and aggregated platelets, all of which contribute to the no-reflow phenomenon in STEMI patients undergoing emergency PCI. Nevertheless, other outcomes were not improved, such as cardiac adverse events, troponin levels, and electrocardiogram (ECG) ST-elevation regression [96]. Other RCT demonstrated that allopurinol resulted in a more effective ST elevation recovery and lower cardiac troponin I (cTnI), CK, and CK-MB peak values. In this same study, after a 1-month follow-up period, patients had a lower incidence of MACE, but no differences in LVEF were detected [97].

These findings suggest that while allopurinol may benefit coronary blood flow and specific inflammatory markers, its overall impact on cardiovascular outcomes in AMI remains limited.

4.4 Edaravone

Edara Edaravone is a lipophilic drug with potent free radical scavenging activity by quenching •OH and inhibiting both •OH-dependent and •OH-independent lipid peroxidation [98]. It was initially studied for its neuroprotective effects, particularly in the context of stroke and, more recently, neurodegenerative conditions such as amyotrophic lateral sclerosis (ALS) [98,157]. In recent decades, edaravone has also been recognized for its cardioprotective potential, mainly due to its capacity to attenuate cardiomyocyte apoptosis via activation of the JAK2/STAT3 signaling pathway [99].

Clinically, it has demonstrated efficacy in reducing IRI in AMI. When administered prior to reperfusion, it reduces infarct size, lowers the incidence of reperfusion-induced arrhythmias, and improves LVEF by decreasing oxidative stress and free radical generation [100]. Additionally, edaravone has been shown to lower plasma levels of monocyte chemoattractant protein-1 (MCP-1), which is associated with improved long-term cardiac recovery and a reduced incidence of heart failure [101].

Furthermore, reductions in biomarkers of tissue damage and oxidative stress, such as thioredoxin, have been observed, suggesting that edaravone may positively influence long-term survival and prognosis in AMI patients treated with this antioxidant [100]. Its ability to inhibit lipid peroxidation and prevent endothelial damage, combined with its high tissue accessibility due to its low molecular weight, further strengthens its potential as a cardioprotective agent against reperfusion injury [100].

Edaravone, in combination with obeticholic acid, has shown promising cardioprotective effects in an animal model of cardiotoxicity. The treatment significantly reduced levels of alkaline phosphatase (ALP), aspartate aminotransferase (AST), lactate dehydrogenase (LDH), CK-MB, and cTnI, while ameliorating histopathological cardiac abnormalities. It also decreased oxidative stress markers such as MDA, increased antioxidant defenses (superoxide dismutase [SOD] and GSH), and upregulated Nrf2, peroxisome proliferator-activated receptor gamma (PPAR- γ), and SIRT1 expression. Additionally, it lowered cardiac inflammatory markers (TNF- α , IL-1 β , IL-6) by inhibiting NF- κ B activity. While these findings highlight potential mechanisms of protection, their applicability to AMI remains uncertain and requires further investigation [158].

Despite these promising findings, the clinical evaluation of edaravone in this context remains limited, with few studies assessing its long-term benefits. Additional challenges include its rapid elimination from the body and the need for early, controlled administration to achieve optimal therapeutic effects [159]. Moreover, the absence of large-scale RCTs further limits the generalizability of these results.



4.5 Sodium Thiosulphate

Sodium thiosulphate (STS) is a water-soluble compound derived from hydrogen sulfide, employed for treating acute cyanide poisoning and carbon monoxide toxicity and to mitigate cisplatin-related toxicity in chemotherapy. Recently, STS's anti-inflammatory and antihypertensive antioxidant effects have been identified [160].

The antioxidant activity of STS is linked to its capacity to inhibit mitochondrial complex IV, which reduces the production of mitochondrial free radicals. Moreover, STS allosterically binds to the enzyme SOD and positively regulates Nrf2 enhancing its antioxidant activity [102,103].

The clinical application of STS in the context of AMI is relatively novel, with only one clinical trial conducted in 2022. This trial did not demonstrate a clinical benefit from STS administration, as treatment during reperfusion failed to reduce infarct size or improve LVEF [104,161]. Nevertheless, this study establishes a foundation for future research to realize the therapeutic benefits observed in preclinical models.

4.6 Nicorandil

Nicorandil is an antianginal drug distinguished by its dual mechanism of action. It functions as a nitric oxide donor, inducing vasodilation and as an agonist of ATP-sensitive potassium channels. These properties make it a potent vasodilator, reducing both ventricular preload and afterload and improving oxygen delivery under ischemic conditions [162]. However, nicorandil's potential extends beyond these effects. It attenuates oxidative stress by stimulating the opening of mitochondrial ATP-sensitive potassium channels, which optimizes oxidative phosphorylation, reduces free radical generation, and decreases the opening of the mitochondrial permeability transition pore [105]. Furthermore, preclinical studies suggest that nicorandil may positively regulate eNOS activity and reduce cardiomyocyte pyroptosis [106,163].

Given these properties, nicorandil has been proposed as a promising cardioprotective agent. A meta-analysis of 18 RCTs demonstrated that nicorandil administration improved coronary no-reflow phenomenon and ST-segment resolution after PCI, and reduced MACE [107]. The cardioprotective effects were more pronounced when nicorandil was administered IC or IV. Another meta-analysis analyzed 14 studies with 1762 patients to assess the effectiveness of nicorandil in reducing periprocedural myocardial injury (PMI) and MACE during PCI. The results showed that it significantly reduced both PMI and MACE, likely due to its dual action as a nitrate and potassium channel opener, which improves coronary blood flow and myocardial preconditioning. Specifically, it also reduced myocardial injury, oxidative stress, and ferroptosis markers. LV enddiastolic diameter and the incidence of unstable angina and heart failure 12 weeks post-PCI was also reduced although LV function parameters remained similar between groups

[164]. However, limitations such as variability in administration methods and lack of subgroup data indicate the need for further research on optimal dosing and comparative effectiveness.

Despite these promising results, the benefits of nicorandil have not been consistently observed. For example, a recent RCT with 83 patients found no significant improvement in infarct size, cardiac function, or the incidence of MACE [108]. One RCT reported that nicorandil treatment significantly reduced infarct size and edema compared to nitrate therapy, although the study's small sample size limited its generalizability [165]. Additionally, Ilyas *et al.* [166] reported that nicorandil enhanced ST-segment resolution, lowered cTnI levels 6 hours post-PCI, and reduced the incidence of MACE, although CK-MB levels were not significantly affected.

Therefore, while nicorandil shows potential cardioprotective effects, the evidence remains inconsistent across studies. Larger, well-designed RCTs are necessary to establish its efficacy and clarify nicorandil's efficacy in preventing IRI.

4.7 Quercetin

Quercetin is a natural polyphenol found in plants and is recognized for its broad spectrum of biological activities [167]. Its antioxidant capacity and potential as a cardioprotectant in the context of IRI have garnered attention. The primary mechanism proposed for these cardioprotective effects is the positive modulation of SIRT1 [109]. Activation of SIRT1 reduces oxidative stress by enhancing antioxidant defenses, promoting transcription factors Nrf2 and FOXO1, both of which are critical for cellular protection against oxidative damage [168]. Moreover, SIRT1 has been shown to attenuate inflammation and cardiomyocyte apoptosis by inhibiting the NF- κ B and p53 pathways [168].

Despite its promising cardioprotective profile in preclinical studies, the clinical use of quercetin in patients with AMI has been hindered by its unfavorable pharmacokinetics. Quercetin is poorly soluble in water, exhibits low bioavailability, and is rapidly metabolized [167]. However, the recent development of nanoformulations has helped to mitigate some of these limitations. In this context, an RCT recently investigated a nanoformulation of quercetin in patients with AMI to evaluate its ability to attenuate IRI. The study showed a reduction in infarct size and intramyocardial hemorrhage, although no significant effect on LVEF was observed [110].

This trial faced limitations, including using the area under the CK-MB curve to estimate infarct size. Additionally, the small sample size reduced the study's statistical power. Despite these constraints, the trial provides a foundation for future RCTs with larger sample sizes to better assess quercetin nanoformulations' efficacy, optimal dosage, and duration to maximize their cardioprotective potential.



4.8 Statins

Statins are a class of medications primarily known for inhibiting the enzyme 3-hydroxy-3-methylglutaryl-coenzyme A reductase (HMG-CoA reductase), a key regulator of cholesterol synthesis. By lowering cholesterol levels, statins address a significant risk factor for the development of cardiovascular disease. However, their cardioprotective properties extend beyond cholesterol reduction, as they have been shown to exert protective effects at the molecular level through various mechanisms. These include the interruption of RhoA-mediated ventricular remodeling pathways, elevation of adenosine levels, reduction of mPTP opening, enhancement of mitophagy via AMP-activated protein kinase (AMPK) pathway activation, and increased antioxidant activity [169,170].

Preclinical studies indicate that statins may also decrease the activity of NADPH oxidase, enhance the activation of the Nrf2/antioxidant response element (ARE) pathway, and reduce the expression of the lectin-like oxidized low-density lipoprotein receptor 1 (LOX-1), which is upregulated in cardiomyocytes under oxidative stress [111–115]. In this context, *in vivo* studies using swine models have demonstrated that early intravenous administration of atorvastatin prior to revascularization can significantly reduce IRI, leading to a decrease in infarct size [116,171].

Translating these findings to clinical practice has proven challenging, as results observed in preclinical animal models are not consistently replicated in human subjects. The prospective ROSEMARY study exemplifies this issue; it compared high-dose rosuvastatin administration (40 mg before primary PCI and 40 mg/day for 7 days thereafter) with low-dose administration (placebo before primary PCI and 10 mg/day for 7 days post-PCI) [116]. The results indicated that infarct size and clinical outcomes were comparable between both groups, with no significant differences observed.

A potential explanation for the inconclusive results of this study may be related to the use of specific antiplatelet agents commonly included in standard therapy, such as aspirin. These agents may diminish the cardioprotective effects of statins by inhibiting cyclooxygenase-2, leading to reduced levels of adenosine, which mediates some of the cardioprotective benefits [172,173]. In light of this, research has explored the use of alternative antiplatelet agents, such as cilostazol, which appear to enhance the effects of statins. However, human studies evaluating this combination remain limited [174–176].

4.9 Multitherapy Approaches in Myocardial Ischemia-Reperfusion Injury

There is substantial evidence supporting the role of oxidative stress in IRI following AMI. Given its complexity, the development of multi-targeted, antioxidant-based combination therapies presents a promising avenue for mitigating such damage. Multitherapy has been extensively stud-

ied as a strategy to combat IRI across various organs, including the liver, kidneys, brain, and myocardium. In this section, we will focus on the multitherapy approaches that have proven to be most effective in the context of MRI (Table 2, Ref. [177–189]).

4.10 Glucose, Insulin, and Potassium Therapy

GIK therapy has been studied since the 1960s as a potential strategy to mitigate IRI [190]. The rationale behind GIK therapy is to improve myocardial metabolism while leveraging the cardioprotective properties of insulin. Glucose and potassium are included primarily to counteract adverse effects, such as hypoglycemia and hypokalemia. Insulin plays a pivotal role by reducing glucose toxicity, acting as a positive inotrope, reducing oxidative stress, and activating anti-apoptotic and anti-inflammatory cell survival pathways [177]. Furthermore, insulin promotes glucose utilization as the primary energy source, thereby decreasing free fatty acid oxidation, reducing oxygen consumption, and attenuating oxidative stress [178]. However, the evidence remains contradictory, with some studies showing no change in post-infusion oxidative stress parameters [191], while others have reported alterations in SOD enzyme activity [192].

Since its introduction, numerous clinical trials have investigated GIK therapy, mostly yielding unfavorable outcomes. A 2010 meta-analysis, which included nine RCTs with a total of 28,000 patients, demonstrated no significant reduction in mortality with GIK therapy in cases of STEMI [193]. Among the most notable studies, the CREATE-ECLA trial, which enrolled over 20,000 patients, found no significant differences in in-hospital mortality or cardio-vascular complications following GIK infusion. This was likely influenced by the average administration time, which exceeded three hours after symptom onset [194].

However, the 2012 IMMEDIATE trial examined the early administration of GIK by emergency medical services, with an average time of 90 minutes from symptom onset. The primary outcomes (myocardial infarction biomarkers) were neutral, but secondary analyses revealed a significant reduction in infarct size at 30 days in 80% of patients with STEMI compared to placebo [195]. This finding has spurred further investigation, and the IMMEDIATE-2 trial is currently underway to explore these hypotheses [196].

More recent meta-analyses have sought to clarify the controversies surrounding GIK therapy. Yang *et al.* [197] reported that high-dose GIK reduces oxidative stress, improves LVEF, and decreases the risk of MACE. Similarly, Liu *et al.* [198] found that GIK improves coronary blood flow and cardiac function but is associated with an increased incidence of adverse events, such as phlebitis and hypoglycemia. Both meta-analyses underscore the need for large-scale clinical trials with long-term follow-up to more definitively assess the efficacy and safety of this therapy.





Table 2. Multitherapy approaches in myocardial ischemia-reperfusion injury.

Compound	Pathway/Mechanism involved	Results in humans
GIK	Insulin inhibits NF-κB (PI3K/Akt). Upregulates eNOS	A meta-analysis indicates that high-dose GIK in ACS patients undergoing reperfusion may enhance myocar-
	expression (PI3k/Akt). Inhibits the opening of mPTP	dial protection and long-term cardiac function, especially in STEMI cases. However, it also shows increased
	(PI3k/Akt/hexokinase) [177].	risks of phlebitis, hyperglycaemia, and hypoglycaemia, warranting cautious use in high-risk patients [180].
	Counteracts glucose toxicity [178].	High-dose GIK therapy in ACS patients receiving reperfusion therapy reduced MACEs and exhibited good oxidative stress-lowering efficacy, but did not improve survival outcomes [179].
		GIK therapy in ACS patients generally resulted in lower serious endpoint rates compared to placebo, with
		significant benefits observed in STEMI patients regarding mortality and cardiac arrest within 1 year [181].
AA + Vit E	Vitamin E (alpha-tocopherol) prevents lipid peroxidation of	Included 800 patients randomly assigned to receive either antioxidants or a matching placebo for 30 days.
	PUFAs [185].	Patients treated with antioxidants had a lower incidence of in-hospital cardiac mortality and non-fatal my-ocardial infarction (14% vs. 19%, OR 0.82) [186].
		The LVEF of the high ascorbate (HA) group was significantly higher than that of the low ascorbate (LA)
		group, with values on day 84 in the HA group being 33% higher. 95% of HA patients achieved a Throm-
		bolysis In Myocardial Infarction (TIMI) myocardial perfusion grade of 2–3, while only 79% of LA patients
		reached this grade [184].
AA + Vit E + Vit A + beta-carotene	Unknown.	Mean infarct size (measured by CK and CK-MB) was significantly lower in the antioxidant group compared
		to the placebo group.
		Lactate dehydrogenase levels increased slightly in the antioxidant group (88.6 IU/dL) compared to the placebo group (166.5 IU/dL).
		Angina pectoris, total arrhythmias, and poor left ventricular function occurred less frequently in the antioxidant group.
		Cardiac events were significantly lower in the antioxidant group (20.6% vs 30.6%) [185].
Curcumin + Piperine	Curcumin acts in the upregulation of PPAR γ , PGC1 α , and	In patients with acute myocardial infarction, daily 500 mg of curcumin with piperine for 8 weeks improved
•	UCP1 [187].	lipid profiles by lowering LDL and raising HDL, enhanced glycemic control (reduced HbA1C), and de-
		creased liver enzymes. No significant effects were noted on ejection fraction, cTnI, renal function, or elec-
		trolytes [183].
	Piperine acts in PPAR γ /Akt and activates the PI3K/Akt signal-	In post-CABG patients, a 5-day treatment with 500 mg of curcumin and 5 mg of piperine reduced C-reactive
	ing [188,189].	protein (CRP) and increased antioxidant capacity. There was a slight decrease in CK-MB, but no significant
		changes in cTnI, LDH, ejection fraction, atrial fibrillation incidence, or renal markers [182].

CK, creatine kinase; GIK, glucose-insulin-potassium; NF- κ B, nuclear factor kappa B; PI3K, phosphoinositide 3-kinase; Akt, protein kinase B; eNOS, endothelial Nitric Oxide Synthase; mPTP, mitochondrial permeability transition pore; ACS, acute coronary syndrome; STEMI, ST-elevation myocardial infarction; MACEs, major adverse cardiac events; AA, ascorbic acid; Vit E, Vitamin E; PUFA, polyunsaturated fatty acid; LVEF, left ventricular ejection fraction; TIMI, thrombolysis in myocardial infarction; Vit A, Vitamin A; LDL, low-density lipoprotein; HDL, high-density lipoprotein; HbA1C, glycated hemoglobin; CABG, coronary artery bypass graft; CK-MB, creatine kinase-MB; LDH, lactate dehydrogenase; PPAR γ , peroxisome proliferator-activated receptor gamma; PGC1 α , peroxisome proliferator-activated receptor gamma coactivator 1-alpha; UCP1, uncoupling protein 1; CRP, C-reactive protein; OR, odds ratio.

4.11 Curcumin + Piperine

Curcumin is the active compound found in turmeric (Curcuma longa), a bright yellow spice commonly used in cooking, especially in Asian cuisine. It is a natural polyphenol known for its anti-inflammatory, antioxidant, and anticancer properties [199]. One of curcumin's primary mechanisms of action is its ability to enhance the expression, stability, and nuclear translocation of Nrf2 [179]. It also has a direct role in IRI. In vitro studies have demonstrated its capacity to modulate apoptosis, autophagy, and, more recently, ferroptosis in cardiomyocytes [180,200]. Some proposed mechanisms are through positive modulation of the Hes1 protein, an intermediary in these cell death pathways. Specifically, curcumin enhances Bcl-2 protein activity to inhibit apoptosis, reduces beclin-1 and SIRT1 activity to mitigate autophagy, and increases GPX4 activity to suppress ferroptosis. However, its low bioavailability remains a significant limitation, hindering its clinical application.

On the other hand, piperine is an active compound found in black pepper (*Piper nigrum*), which also possesses anti-inflammatory, antihypertensive, anti-cancer, and antioxidant properties [181]. These effects are primarily attributed to the activation of the PPAR- γ , and its intrinsic antioxidant activity as a free radical scavenger [201]. Notably, one of piperine's most significant characteristics is its ability to enhance the bioavailability of various drugs. This is achieved by increasing gastrointestinal absorption, inhibiting efflux pumps, and reducing the activity of drugmetabolizing enzymes [202].

Despite the promising cardioprotective effects observed in preclinical studies, clinical evidence for this combination remains limited [203]. For instance, a study involving patients undergoing CABG surgery reported reduced in inflammatory parameters and increased total antioxidant capacity. However, no changes were observed in levels of cTnI, LDH, LVEF, or the incidence of atrial fibrillation [182]. Another study assessed the effects of the curcumin-piperine combination following AMI. The results demonstrated significant improvements in glycemic control and lipid profile. However, the treatment did not impact on LVEF or cTnI [183]. A recent review suggested that the curcumin doses used in clinical studies may have been too low to achieve meaningful cardioprotective effects. It proposed that higher doses, which have demonstrated safety, could more closely replicate the efficacy observed in animal models [184].

4.12 Antioxidant Vitamins

One of the most extensively studied approaches in this field is the combination of vitamins C (ascorbic acid, L-ascorbic acid or L-ascorbate) and E (alpha-tocopherol) [7]. Ascorbic acid (AA), a hydrophilic vitamin, functions primarily as a potent reducing agent, directly scavenging ROS such as superoxide anion [117]. Furthermore, it inhibits NOX synthesis and prevents the oxidation of tetrahydro-

biopterin (BH4), thus avoiding the uncoupling of eNOS [118,119]. Pharmacokinetic and pharmacodynamic studies have indicated that a minimum concentration of 10 mM is required to observe these effects in tissues under oxidative stress, a critical consideration for future studies evaluating its antioxidant efficacy [117].

Vitamin E is a fat-soluble micronutrient that accumulates in cell membranes, where it serves as both a membrane stabilizer and an antioxidant [204]. Its antioxidant capacity is primarily focused on preventing lipid peroxidation of PUFAs, thereby reducing damage to cell membranes [205]. The rationale behind combining vitamins C and E lies in their complementary actions: vitamin C operates in aqueous environments, while vitamin E functions in lipid environments [7]. Moreover, vitamin C plays a critical role in recycling alpha-tocopherol by reducing its oxidized form, thus enhancing its antioxidant activity [206].

Several clinical trials have been conducted to assess the efficacy of this combination therapy. One of the earliest studies, the MIVIT trial in 2005, investigated the clinical outcomes of patients treated with the combined therapy and observed a reduction in adverse events, although no specific clinical parameters were measured to explain these results [207]. The PREVEC trial demonstrated that patients who achieved elevated blood AA levels prior to reperfusion (via PCI) and continued oral supplementation with vitamins C and E showed improved ventricular function, as evidenced by increased LVEF and enhanced microvascular flow [208,209]. However, a key limitation of this study was the lack of assessment of the therapy's effects on infarct size and myocardial remodeling, both of which are critical factors in reperfusion injury [209]. A recent meta-analysis including eight RCTs that evaluated AA in PCI reported a reduction in myocardial damage biomarkers, without an improvement in myocardial function parameters [209]. As in other studies, there was considerable variability among trials, including differences in patient populations (AMI vs. stable angina) and timing of administration. Some studies implemented a continuation strategy extending weeks or months post-reperfusion. Additionally, most trials included relatively young patient cohorts, which may limit the generalizability of the findings [210–212].

Prior to the exclusive use of AA and vitamin E, combinations that included vitamin A and beta-carotene were tested and yielded favorable results in treated groups [185]. However, over time, the antioxidant and cardioprotective roles of vitamin A and beta-carotene have been questioned, leading to a decline in further studies on this combination [186,213,214].

4.13 Current Research in Combination Therapy

Several authors have proposed combination therapy as a promising strategy for MRI [215]. However, most of the evidence supporting the synergistic effects of multi-target antioxidant therapies comes from preclinical studies [216].



A combination of AA, NAC, and deferoxamine (DFO) has shown encouraging results and is currently being evaluated in early-stage clinical trials. AA acts as a potent antioxidant with cardioprotective properties, while NAC replenishes GSH stores, chelates metal ions, and modulates inflammation via NF-κB inhibition [217–219]. Meanwhile, DFO reduces labile iron availability, thereby limiting ROS production and preventing ferroptosis [15]. In isolated rat heart models, this combination significantly reduced infarct size and preserved myocardial tissue, leading to a human safety trial where intravenous administration of AA, NAC, and DFO achieved therapeutic plasma levels without severe adverse effects [220]. These findings have laid the foundation for an ongoing phase II clinical trial to assess its efficacy in patients with AMI.

Several clinical trials are currently investigating novel therapeutic approaches for AMI. One of the most studied is supersaturated oxygen (SSO₂) therapy, which has demonstrated a significant reduction in infarct size and improved microvascular function. The AMIHOT I and II trials reported a 26% reduction in infarct size in patients treated within six hours of symptom onset [221], prompting further evaluation in the AMIHOT III trial (NCT04743245). Other pharmacological strategies under investigation include colchicine (NCT05734612), which mitigates inflammation by inhibiting NLRP3 inflammasome activation, and nicorandil (NCT04665648), a nitric oxide donor with cardioprotective effects. Additionally, left ventricular unloading prior to reperfusion has been proposed as a strategy to minimise myocardial damage, with its clinical impact currently under investigation in the STEMI-DTU trial (NCT03947619). Despite promising findings in phase II studies, many phase III trials have struggled to demonstrate significant clinical benefits, particularly in terms of reducing mortality and heart failure incidence, likely due to patient heterogeneity and methodological limitations [215].

One of the major challenges in translating preclinical findings into clinical practice lies in the multifaceted nature of MRI, which involves oxidative stress, inflammation, mitochondrial dysfunction, and various cell death pathways. While several cardioprotective interventions have failed to demonstrate a substantial clinical impact, recent studies have identified novel therapeutic targets. Among these, advancements in paracrine signaling modulation have shown promise, with microRNAs, peptides, and extracellular vesicles exhibiting regenerative potential in experimental models [222]. Additionally, extracellular matrix modulation, particularly through agrin, has emerged as a potential strategy for myocardial repair [222].

Given the multifactorial nature of MRI, the most effective therapeutic approaches are likely to involve a combination of antioxidant therapy, pharmacological interventions, mechanical strategies, and regenerative medicine. The integration of these modalities may represent the most viable approach to improving clinical outcomes in AMI pa-

tients. Ongoing research will be critical in overcoming current barriers and facilitating the implementation of more effective combination therapies in routine clinical practice.

5. Discussion

MRI is a complex process characterized by oxidative stress, metabolic dysregulation, calcium overload, mitochondrial dysfunction, and alterations in cell signaling pathways, ultimately leading to membrane and/or DNA damage and inflammation. Oxidative stress plays a central role in driving cell death through these mechanisms.

Recent advances in understanding programmed cell death, including autophagy, apoptosis, necroptosis, ferroptosis, and pyroptosis, have revealed novel therapeutic targets for MRI. Strategies targeting oxidative stress, either through standalone antioxidants or combination therapies addressing multiple pathways, have shown promise *in vitro* and *in vivo*. For instance, iron chelators help regulate iron homeostasis, while insulin mitigates glucose toxicity. Compounds such as NAC, edaravone, melatonin, and allopurinol have demonstrated potential in reducing oxidative stress and infarct size (Fig. 2). However, their clinical efficacy may improve with multitarget approaches. Notably, the combination of AA, NAC, and DFO has reduced infarct size and provided myocardial protection in preclinical models, with initial clinical studies confirming its safety.

Despite significant advances in the clarification of the mechanisms of MRI, translating cardioprotective therapies from preclinical research to clinical practice remains challenging. While antioxidants have shown efficacy in animal models and some human trials, most studies are small-scale RCTs with limitations affecting generalizability. Emerging therapies, such as supersaturated oxygen (SSO₂), have demonstrated a 26% infarct size reduction in phase II trials, warranting further investigation in phase III studies. Additionally, colchicine, which targets the NLRP3 inflammasome, and nicorandil, a nitric oxide donor, are being explored for their cardioprotective effects in AMI patients. Left ventricular unloading before reperfusion has also been proposed to enhance myocardial salvage, with ongoing trials assessing its clinical impact. However, heterogeneity in study populations, disease severity, comorbidities, and concomitant medications remains a barrier to clinical translation [215,222].

Variability in therapeutic approaches further complicates implementation. Factors such as timing, dosage, and administration route (e.g., intracoronary vs. intravenous) vary across studies. Mechanistically, antioxidants are expected to be most effective when administered before and during reperfusion, a hypothesis supported by some trials, though rigorous evaluations remain limited. Outcome measures also differ, with infarct size widely regarded as the most robust indicator of myocardial injury, while functional outcomes such as LVEF, heart failure-related hospitalizations, and recurrent ischemic events provide additional in-



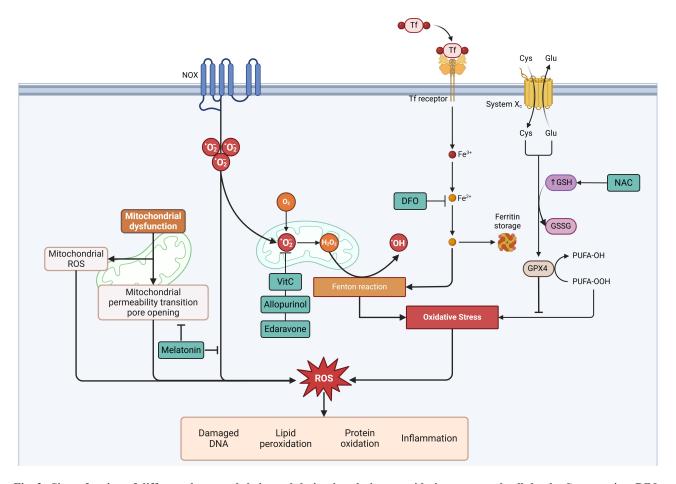


Fig. 2. Sites of action of different drugs and their modulation in relation to oxidative stress and cell death. Cys, cysteine; DFO, deferoxamine; Fe³⁺, ferric iron; Fe²⁺, ferrous iron; Glu, glutamate; GPX4, glutathione peroxidase 4; GSH, reduced glutathione; GSSG, oxidized glutathione; H₂O₂, hydrogen peroxide; NAC, N-acetylcysteine; NOX, NADPH oxidase; •OH, hydroxyl radical; •O₂⁻, superoxide anion; PUFA-OH, hydroxylated polyunsaturated fatty acid; PUFA-OOH, hydroperoxylated polyunsaturated fatty acid; System Xc⁻, cystine/glutamate antiporter; ROS, reactive oxygen species; Tf, transferrin; VitC, Vitamin C. Created with BioRender.com.

sights into long-term therapeutic impact. Given the limited clinical data and methodological constraints, larger, wellpowered, and mechanistically driven clinical trials are essential for a comprehensive evaluation of these strategies.

6. Conclusions

MRI remains a major therapeutic challenge due to its multifactorial nature and the complex interplay between oxidative stress, inflammation, and mitochondrial dysfunction. While significant progress has been made in identifying potential therapeutic targets, the translation of these findings into clinical practice has been hindered by limitations in study design, patient heterogeneity, and variability in outcome measures. Emerging multitarget approaches, such as combination antioxidant therapy, SSO₂ administration, ischemic conditioning, and NLRP3 inhibition, represent promising strategies, but their efficacy needs to be validated in well-designed, large-scale clinical trials.

Future research should focus on refining experimental models that better replicate real-world patient populations,

optimizing timing and dosing strategies for interventions, and standardizing clinical endpoints to facilitate meaningful comparisons across studies. Ultimately, a combination of pharmacological, mechanical, and regenerative strategies may be necessary to achieve significant improvements in MRI outcomes, underscoring the need for continued innovation and multidisciplinary collaboration in this field.

Author Contributions

EV and RR led the conceptualization of the study. EV, CRS, and VPG were responsible for writing the manuscript and for the creation of the figures. EV, CRS, VPG, LS, SC, JCP, AIJG, and RA made substantial contributions in the acquisition and analysis of literature, provided supervision throughout the study and critically reviewed and edited the manuscript. All authors contributed to editorial changes in the manuscript. All authors read and approved the final manuscript. All authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.



Ethics Approval and Consent to Participate

Not applicable.

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Conflict of Interest

The authors declare no conflict of interest. Luciano Saso is serving as Guest Editor of this journal. We declare that Luciano Saso had no involvement in the peer review of this article and has no access to information regarding its peer review. Full responsibility for the editorial process for this article was delegated to Ioanna-Katerina Aggeli.

References

- [1] Martin SS, Aday AW, Almarzooq ZI, Anderson CAM, Arora P, Avery CL, *et al.* 2024 Heart Disease and Stroke Statistics: A Report of US and Global Data From the American Heart Association. Circulation. 2024; 149: e347–e913. https://doi.org/10.1161/CIR.0000000000001209.
- [2] Stone GW, Selker HP, Thiele H, Patel MR, Udelson JE, Ohman EM, et al. Relationship Between Infarct Size and Outcomes Following Primary PCI: Patient-Level Analysis From 10 Randomized Trials. Journal of the American College of Cardiology. 2016; 67: 1674–1683. https://doi.org/10.1016/j.jacc.2016.01.069.
- [3] Hausenloy DJ, Yellon DM. Myocardial ischemia-reperfusion injury: a neglected therapeutic target. The Journal of Clinical Investigation. 2013; 123: 92–100. https://doi.org/10.1172/JC 162874.
- [4] Bell RM, Bøtker HE, Carr RD, Davidson SM, Downey JM, Dutka DP, et al. 9th Hatter Biannual Meeting: position document on ischaemia/reperfusion injury, conditioning and the ten commandments of cardioprotection. Basic Research in Cardiology. 2016; 111: 41. https://doi.org/10.1007/s00395-016-0558-1.
- [5] Davidson SM, Ferdinandy P, Andreadou I, Bøtker HE, Heusch G, Ibáñez B, et al. Multitarget Strategies to Reduce Myocardial Ischemia/Reperfusion Injury: JACC Review Topic of the Week. Journal of the American College of Cardiology. 2019; 73: 89–99. https://doi.org/10.1016/j.jacc.2018.09.086.
- [6] Chouchani ET, Pell VR, Gaude E, Aksentijević D, Sundier SY, Robb EL, et al. Ischaemic accumulation of succinate controls reperfusion injury through mitochondrial ROS. Nature. 2014; 515: 431–435. https://doi.org/10.1038/nature13909.
- [7] Del Re DP, Amgalan D, Linkermann A, Liu Q, Kitsis RN. Fundamental Mechanisms of Regulated Cell Death and Implications for Heart Disease. Physiological Reviews. 2019; 99: 1765–1817. https://doi.org/10.1152/physrev.00022.2018.
- [8] Rodrigo R, Libuy M, Feliú F, Hasson D. Molecular basis of cardioprotective effect of antioxidant vitamins in myocardial infarction. BioMed Research International. 2013; 2013: 437613. https://doi.org/10.1155/2013/437613.
- [9] Heusch G, Andreadou I, Bell R, Bertero E, Botker HE, Davidson SM, et al. Health position paper and redox perspectives on re-

- active oxygen species as signals and targets of cardioprotection. Redox Biology. 2023; 67: 102894. https://doi.org/10.1016/j.redox.2023.102894.
- [10] Brookes PS, Yoon Y, Robotham JL, Anders MW, Sheu SS. Calcium, ATP, and ROS: a mitochondrial love-hate triangle. American Journal of Physiology. Cell Physiology. 2004; 287: C817–33. https://doi.org/10.1152/ajpcell.00139.2004.
- [11] Yellon DM, Hausenloy DJ. Myocardial reperfusion injury. The New England Journal of Medicine. 2007; 357: 1121–1135. http s://doi.org/10.1056/NEJMra071667.
- [12] Tabata Fukushima C, Dancil IS, Clary H, Shah N, Nadtochiy SM, Brookes PS. Reactive oxygen species generation by reverse electron transfer at mitochondrial complex I under simulated early reperfusion conditions. Redox Biology. 2024; 70: 103047. https://doi.org/10.1016/j.redox.2024.103047.
- [13] Cadenas S. ROS and redox signaling in myocardial ischemiareperfusion injury and cardioprotection. Free Radical Biology & Medicine. 2018; 117: 76–89. https://doi.org/10.1016/j.freera dbiomed.2018.01.024.
- [14] Granger DN, Kvietys PR. Reperfusion injury and reactive oxygen species: The evolution of a concept. Redox Biology. 2015; 6: 524–551. https://doi.org/10.1016/j.redox.2015.08.020.
- [15] Fang X, Wang H, Han D, Xie E, Yang X, Wei J, et al. Ferroptosis as a target for protection against cardiomyopathy. Proceedings of the National Academy of Sciences of the United States of America. 2019; 116: 2672–2680. https://doi.org/10.1073/pnas .1821022116.
- [16] Zhang M, Shah AM. ROS signalling between endothelial cells and cardiac cells. Cardiovascular Research. 2014; 102: 249– 257. https://doi.org/10.1093/cvr/cvu050.
- [17] Radi R, Beckman JS, Bush KM, Freeman BA. Peroxynitrite oxidation of sulfhydryls. The cytotoxic potential of superoxide and nitric oxide. The Journal of Biological Chemistry. 1991; 266: 4244–4250.
- [18] Yakovlev VA, Mikkelsen RB. Protein tyrosine nitration in cellular signal transduction pathways. Journal of Receptor and Signal Transduction Research. 2010; 30: 420–429. https://doi.org/10.3109/10799893.2010.513991.
- [19] Iwase H, Robin E, Guzy RD, Mungai PT, Vanden Hoek TL, Chandel NS, et al. Nitric oxide during ischemia attenuates oxidant stress and cell death during ischemia and reperfusion in cardiomyocytes. Free Radical Biology & Medicine. 2007; 43: 590–599. https://doi.org/10.1016/j.freeradbiomed.2007.05.017.
- [20] Yu X, Ge L, Niu L, Lian X, Ma H, Pang L. The Dual Role of Inducible Nitric Oxide Synthase in Myocardial Ischemia/Reperfusion Injury: Friend or Foe? Oxidative Medicine and Cellular Longevity. 2018; 2018: 8364848. https://doi.org/ 10.1155/2018/8364848.
- [21] Webster KA. Mitochondrial membrane permeabilization and cell death during myocardial infarction: roles of calcium and reactive oxygen species. Future Cardiology. 2012; 8: 863–884. https://doi.org/10.2217/fca.12.58.
- [22] Naito Z, Kudo M, Xu G, Nishigaki R, Yokoyama M, Yamada N, et al. Immunohistochemical localization of mitogen-activated protein kinase (MAPK) family and morphological changes in rat heart after ischemia-reperfusion injury. Medical Electron Microscopy: Official Journal of the Clinical Electron Microscopy Society of Japan. 2000; 33: 74–81. https://doi.org/10.1007/s007950070005.
- [23] Gumina RJ, Buerger E, Eickmeier C, Moore J, Daemmgen J, Gross GJ. Inhibition of the Na(+)/H(+) exchanger confers greater cardioprotection against 90 minutes of myocardial ischemia than ischemic preconditioning in dogs. Circulation. 1999; 100: 2519–2526; discussion 2469–72. https://doi.org/10.1161/01.cir.100.25.2519.
- [24] Avkiran M, Marber MS. Na(+)/H(+) exchange inhibitors for cardioprotective therapy: progress, problems and prospects. Jour-



- nal of the American College of Cardiology. 2002; 39: 747–753. https://doi.org/10.1016/s0735-1097(02)01693-5.
- [25] Zeymer U, Suryapranata H, Monassier JP, Opolski G, Davies J, Rasmanis G, et al. The Na(+)/H(+) exchange inhibitor eniporide as an adjunct to early reperfusion therapy for acute myocardial infarction. Results of the evaluation of the safety and cardioprotective effects of eniporide in acute myocardial infarction (ESCAMI) trial. Journal of the American College of Cardiology. 2001; 38: 1644–1650. https://doi.org/10.1016/s0735-1097(01) 01608-4.
- [26] Boden WE, van Gilst WH, Scheldewaert RG, Starkey IR, Carlier MF, Julian DG, et al. Diltiazem in acute myocardial infarction treated with thrombolytic agents: a randomised placebocontrolled trial. Incomplete Infarction Trial of European Research Collaborators Evaluating Prognosis post-Thrombolysis (INTERCEPT). Lancet (London, England). 2000; 355: 1751–1756. https://doi.org/10.1016/s0140-6736(00)02262-5.
- [27] Lopaschuk G. Regulation of carbohydrate metabolism in ischemia and reperfusion. American Heart Journal. 2000; 139: S115–9. https://doi.org/10.1067/mhj.2000.103919.
- [28] Kulek AR, Anzell A, Wider JM, Sanderson TH, Przyklenk K. Mitochondrial Quality Control: Role in Cardiac Models of Lethal Ischemia-Reperfusion Injury. Cells. 2020; 9: 214. https://doi.org/10.3390/cells9010214.
- [29] Ziegler M, Wang X, Peter K. Platelets in cardiac ischaemia/reperfusion injury: a promising therapeutic target. Cardiovascular Research. 2019; 115: 1178–1188. https://doi.org/10. 1093/cvr/cvz070.
- [30] Vakeva AP, Agah A, Rollins SA, Matis LA, Li L, Stahl GL. Myocardial infarction and apoptosis after myocardial ischemia and reperfusion: role of the terminal complement components and inhibition by anti-C5 therapy. Circulation. 1998; 97: 2259– 2267. https://doi.org/10.1161/01.cir.97.22.2259.
- [31] Zhao ZQ, Lefer DJ, Sato H, Hart KK, Jefforda PR, Vinten-Johansen J. Monoclonal antibody to ICAM-1 preserves postischemic blood flow and reduces infarct size after ischemia-reperfusion in rabbit. Journal of Leukocyte Biology. 1997; 62: 292–300. https://doi.org/10.1002/jlb.62.3.292.
- [32] Hayward R, Campbell B, Shin YK, Scalia R, Lefer AM. Recombinant soluble P-selectin glycoprotein ligand-1 protects against myocardial ischemic reperfusion injury in cats. Cardiovascular Research. 1999; 41: 65–76. https://doi.org/10.1016/s0008-6363(98)00266-1.
- [33] APEX AMI Investigators, Armstrong PW, Granger CB, Adams PX, Hamm C, Holmes D, Jr, *et al.* Pexelizumab for acute ST-elevation myocardial infarction in patients undergoing primary percutaneous coronary intervention: a randomized controlled trial. JAMA. 2007; 297: 43–51. https://doi.org/10.1001/jama.297.1.43.
- [34] Atar D, Petzelbauer P, Schwitter J, Huber K, Rensing B, Kasprzak JD, *et al.* Effect of intravenous FX06 as an adjunct to primary percutaneous coronary intervention for acute ST-segment elevation myocardial infarction results of the F.I.R.E. (Efficacy of FX06 in the Prevention of Myocardial Reperfusion Injury) trial. Journal of the American College of Cardiology. 2009; 53: 720–729. https://doi.org/10.1016/j.jacc.2008.12.017.
- [35] Kowaltowski AJ, de Souza-Pinto NC, Castilho RF, Vercesi AE. Mitochondria and reactive oxygen species. Free Radical Biology & Medicine. 2009; 47: 333–343. https://doi.org/10.1016/j.free radbiomed.2009.05.004.
- [36] Zhang J, Bian HJ, Li XX, Liu XB, Sun JP, Li N, et al. ERK-MAPK signaling opposes rho-kinase to reduce cardiomyocyte apoptosis in heart ischemic preconditioning. Molecular Medicine (Cambridge, Mass.). 2010; 16: 307–315. https://doi. org/10.2119/molmed.2009.00121.
- [37] Ghigo A, Laffargue M, Li M, Hirsch E. PI3K and Calcium Signaling in Cardiovascular Disease. Circulation Research.

- 2017; 121: 282–292. https://doi.org/10.1161/CIRCRESAHA .117.310183.
- [38] Wei YR, Hou YL, Yin YJ, Li Z, Liu Y, Han NX, et al. Tongxinluo Activates PI3K/AKT Signaling Pathway to Inhibit Endothelial Mesenchymal Transition and Attenuate Myocardial Fibrosis after Ischemia-Reperfusion in Mice. Chinese Journal of Integrative Medicine. 2024; 30: 608–615. https://doi.org/10.1007/ s11655-024-3652-5.
- [39] Wang D, Zhang X, Li D, Hao W, Meng F, Wang B, et al. Kaempferide Protects against Myocardial Ischemia/Reperfusion Injury through Activation of the PI3K/Akt/GSK-3β Pathway. Mediators of Inflammation. 2017; 2017: 5278218. https://doi. org/10.1155/2017/5278218.
- [40] Tucka J, Yu H, Gray K, Figg N, Maguire J, Lam B, et al. Akt1 regulates vascular smooth muscle cell apoptosis through FoxO3a and Apaf1 and protects against arterial remodeling and atherosclerosis. Arteriosclerosis, Thrombosis, and Vascular Biology. 2014; 34: 2421–2428. https://doi.org/10.1161/ATVBAH A.114.304284.
- [41] Stephanou A. Role of STAT-1 and STAT-3 in ischaemia/reperfusion injury. Journal of Cellular and Molecular Medicine. 2004; 8: 519–525. https: //doi.org/10.1111/j.1582-4934.2004.tb00476.x.
- [42] Sciarretta S, Forte M, Frati G, Sadoshima J. New Insights Into the Role of mTOR Signaling in the Cardiovascular System. Circulation Research. 2018; 122: 489–505. https://doi.org/10.1161/ CIRCRESAHA.117.311147.
- [43] Galluzzi L, Baehrecke EH, Ballabio A, Boya P, Bravo-San Pedro JM, Cecconi F, *et al.* Molecular definitions of autophagy and related processes. The EMBO Journal. 2017; 36: 1811–1836. https://doi.org/10.15252/embj.201796697.
- [44] Wang Y, Zhou L, Su W, Huang F, Zhang Y, Xia ZY, et al. Selective Inhibition of PKCβ2 Restores Ischemic Postconditioning-Mediated Cardioprotection by Modulating Autophagy in Diabetic Rats. Journal of Diabetes Research. 2020; 2020: 2408240. https://doi.org/10.1155/2020/2408240.
- [45] Matsui Y, Takagi H, Qu X, Abdellatif M, Sakoda H, Asano T, et al. Distinct roles of autophagy in the heart during ischemia and reperfusion: roles of AMP-activated protein kinase and Beclin 1 in mediating autophagy. Circulation Research. 2007; 100: 914–922. https://doi.org/10.1161/01.RES.0000261924.76669.36.
- [46] Nakai A, Yamaguchi O, Takeda T, Higuchi Y, Hikoso S, Taniike M, *et al.* The role of autophagy in cardiomyocytes in the basal state and in response to hemodynamic stress. Nature Medicine. 2007; 13: 619–624. https://doi.org/10.1038/nm1574.
- [47] Robichaux DJ, Harata M, Murphy E, Karch J. Mitochondrial permeability transition pore-dependent necrosis. Journal of Molecular and Cellular Cardiology. 2023; 174: 47–55. https://doi.org/10.1016/j.yjmcc.2022.11.003.
- [48] Shimizu S, Narita M, Tsujimoto Y. Bcl-2 family proteins regulate the release of apoptogenic cytochrome c by the mitochondrial channel VDAC. Nature. 1999; 399: 483–487. https://doi.org/10.1038/20959.
- [49] Mocanu MM, Baxter GF, Yellon DM. Caspase inhibition and limitation of myocardial infarct size: protection against lethal reperfusion injury. British Journal of Pharmacology. 2000; 130: 197–200. https://doi.org/10.1038/sj.bjp.0703336.
- [50] McKimpson WM, Weinberger J, Czerski L, Zheng M, Crow MT, Pessin JE, et al. The apoptosis inhibitor ARC alleviates the ER stress response to promote β-cell survival. Diabetes. 2013; 62: 183–193. https://doi.org/10.2337/db12-0504.
- [51] Donath S, Li P, Willenbockel C, Al-Saadi N, Gross V, Willnow T, et al. Apoptosis repressor with caspase recruitment domain is required for cardioprotection in response to biomechanical and ischemic stress. Circulation. 2006; 113: 1203–1212. https://doi.org/10.1161/CIRCULATIONAHA.105.576785.



- [52] Inserte J, Cardona M, Poncelas-Nozal M, Hernando V, Vilardosa Ú, Aluja D, et al. Studies on the role of apoptosis after transient myocardial ischemia: genetic deletion of the executioner caspases-3 and -7 does not limit infarct size and ventricular remodeling. Basic Research in Cardiology. 2016; 111: 18. https://doi.org/10.1007/s00395-016-0537-6.
- [53] Seo J, Nam YW, Kim S, Oh DB, Song J. Necroptosis molecular mechanisms: Recent findings regarding novel necroptosis regulators. Experimental & Molecular Medicine. 2021; 53: 1007– 1017. https://doi.org/10.1038/s12276-021-00634-7.
- [54] Zhang W, Zhang J, Wang Z, Li T, Liu C, Kang X, et al. Extracellular RIPK3 Acts as a Damage-Associated Molecular Pattern to Exaggerate Cardiac Ischemia/Reperfusion Injury. Circulation. 2024; 150: 1791–1811. https://doi.org/10.1161/CIRCULATIONAHA.123.068595.
- [55] Oerlemans MIFJ, Liu J, Arslan F, den Ouden K, van Middelaar BJ, Doevendans PA, et al. Inhibition of RIP1-dependent necrosis prevents adverse cardiac remodeling after myocardial ischemiareperfusion in vivo. Basic Research in Cardiology. 2012; 107: 270. https://doi.org/10.1007/s00395-012-0270-8.
- [56] Wang C, Yuan W, Hu A, Lin J, Xia Z, Yang CF, et al. Dexmedetomidine alleviated sepsis induced myocardial ferroptosis and septic heart injury. Molecular Medicine Reports. 2020; 22: 175–184. https://doi.org/10.3892/mmr.2020.11114.
- [57] Zhang K, Tian XM, Li W, Hao LY. Ferroptosis in cardiac hypertrophy and heart failure. Biomedicine & Pharmacotherapy. 2023; 168: 115765. https://doi.org/10.1016/j.biopha.2023. 115765.
- [58] Liu G, Xie X, Liao W, Chen S, Zhong R, Qin J, et al. Ferroptosis in cardiovascular disease. Biomedicine & Pharmacotherapy. 2024; 170: 116057. https://doi.org/10.1016/j.biopha.2023. 116057
- [59] Zhao WK, Zhou Y, Xu TT, Wu Q. Ferroptosis: Opportunities and Challenges in Myocardial Ischemia-Reperfusion Injury. Oxidative Medicine and Cellular Longevity. 2021; 2021: 9929687. https://doi.org/10.1155/2021/9929687.
- [60] Feng H, Schorpp K, Jin J, Yozwiak CE, Hoffstrom BG, Decker AM, et al. Transferrin Receptor Is a Specific Ferroptosis Marker. Cell Reports. 2020; 30: 3411–3423.e7. https://doi.org/10.1016/j.celrep.2020.02.049.
- [61] Qian W, Liu D, Han Y, Liu M, Liu B, Ji Q, et al. Cyclosporine A-loaded apoferritin alleviates myocardial ischemia-reperfusion injury by simultaneously blocking ferroptosis and apoptosis of cardiomyocytes. Acta Biomaterialia. 2023; 160: 265–280. https://doi.org/10.1016/j.actbio.2023.02.025.
- [62] Wallace DF. The Regulation of Iron Absorption and Homeostasis. The Clinical Biochemist. Reviews. 2016; 37: 51–62.
- [63] Shesh BP, Connor JR. A novel view of ferritin in cancer. Biochimica et Biophysica Acta. Reviews on Cancer. 2023; 1878: 188917. https://doi.org/10.1016/j.bbcan.2023.188917.
- [64] Masaldan S, Clatworthy SAS, Gamell C, Meggyesy PM, Rigopoulos AT, Haupt S, et al. Iron accumulation in senescent cells is coupled with impaired ferritinophagy and inhibition of ferroptosis. Redox Biology. 2018; 14: 100–115. https: //doi.org/10.1016/j.redox.2017.08.015.
- [65] Santana-Codina N, Mancias JD. The Role of NCOA4-Mediated Ferritinophagy in Health and Disease. Pharmaceuticals (Basel, Switzerland). 2018; 11: 114. https://doi.org/10. 3390/ph11040114.
- [66] Shan X, Lv ZY, Yin MJ, Chen J, Wang J, Wu QN. The Protective Effect of Cyanidin-3-Glucoside on Myocardial Ischemia-Reperfusion Injury through Ferroptosis. Oxidative Medicine and Cellular Longevity. 2021; 2021: 8880141. https://doi.org/10.1155/2021/8880141.
- [67] Gao M, Monian P, Quadri N, Ramasamy R, Jiang X. Glutaminolysis and Transferrin Regulate Ferroptosis. Molecular Cell. 2015; 59: 298–308. https://doi.org/10.1016/j.molcel.2015.

- 06 011
- [68] Dawi J, Affa S, Gonzalez E, Misakyan Y, Nikoghosyan D, Haijar K, et al. Ferroptosis in Cardiovascular Disease and Cardiomyopathies: Therapeutic Implications of Glutathione and Iron Chelating Agents. Biomedicines. 2024; 12: 558. https://doi.org/10.3390/biomedicines12030558.
- [69] DeBoer DA, Clark RE. Iron chelation in myocardial preservation after ischemia-reperfusion injury: the importance of pretreatment and toxicity. The Annals of Thoracic Surgery. 1992; 53: 412–418. https://doi.org/10.1016/0003-4975(92)90260-b.
- [70] Kagan VE, Mao G, Qu F, Angeli JPF, Doll S, Croix CS, et al. Oxidized arachidonic and adrenic PEs navigate cells to ferroptosis. Nature Chemical Biology. 2017; 13: 81–90. https://doi.org/10.1038/nchembio.2238.
- [71] Piccolo M, Ferraro MG, Iazzetti F, Santamaria R, Irace C. Insight into Iron, Oxidative Stress and Ferroptosis: Therapy Targets for Approaching Anticancer Strategies. Cancers. 2024; 16: 1220. https://doi.org/10.3390/cancers16061220.
- [72] Tang LJ, Luo XJ, Tu H, Chen H, Xiong XM, Li NS, et al. Ferroptosis occurs in phase of reperfusion but not ischemia in rat heart following ischemia or ischemia/reperfusion. Naunyn-Schmiedeberg's Archives of Pharmacology. 2021; 394: 401– 410. https://doi.org/10.1007/s00210-020-01932-z.
- [73] Kobayashi M, Suhara T, Baba Y, Kawasaki NK, Higa JK, Matsui T. Pathological Roles of Iron in Cardiovascular Disease. Current Drug Targets. 2018; 19: 1068–1076. https://doi.org/10.2174/1389450119666180605112235.
- [74] Bulluck H, Rosmini S, Abdel-Gadir A, White SK, Bhuva AN, Treibel TA, et al. Residual Myocardial Iron Following Intramy-ocardial Hemorrhage During the Convalescent Phase of Reperfused ST-Segment-Elevation Myocardial Infarction and Adverse Left Ventricular Remodeling. Circulation. Cardiovascular Imaging. 2016; 9: e004940. https://doi.org/10.1161/CIRCIMAGIN G.116.004940.
- [75] Ding K, Liu C, Li L, Yang M, Jiang N, Luo S, et al. Acyl-CoA synthase ACSL4: an essential target in ferroptosis and fatty acid metabolism. Chinese Medical Journal. 2023; 136: 2521–2537. https://doi.org/10.1097/CM9.000000000002533.
- [76] Fan Z, Cai L, Wang S, Wang J, Chen B. Baicalin Prevents Myocardial Ischemia/Reperfusion Injury Through Inhibiting ACSL4 Mediated Ferroptosis. Frontiers in Pharmacology. 2021; 12: 628988. https://doi.org/10.3389/fphar.2021.628988.
- [77] Lv Y, Liang C, Sun Q, Zhu J, Xu H, Li X, et al. Structural insights into FSP1 catalysis and ferroptosis inhibition. Nature Communications. 2023; 14: 5933. https://doi.org/10.1038/s41467-023-41626-7.
- [78] Bersuker K, Hendricks JM, Li Z, Magtanong L, Ford B, Tang PH, et al. The CoQ oxidoreductase FSP1 acts parallel to GPX4 to inhibit ferroptosis. Nature. 2019; 575: 688–692. https://doi.or g/10.1038/s41586-019-1705-2.
- [79] Jiang X, Stockwell BR, Conrad M. Ferroptosis: mechanisms, biology and role in disease. Nature Reviews. Molecular Cell Biology. 2021; 22: 266–282. https://doi.org/10.1038/s41580-020-00324-8.
- [80] Sui X, Zhang R, Liu S, Duan T, Zhai L, Zhang M, et al. RSL3 Drives Ferroptosis Through GPX4 Inactivation and ROS Production in Colorectal Cancer. Frontiers in Pharmacology. 2018; 9: 1371. https://doi.org/10.3389/fphar.2018.01371.
- [81] Yu P, Zhang X, Liu N, Tang L, Peng C, Chen X. Pyroptosis: mechanisms and diseases. Signal Transduction and Targeted Therapy. 2021; 6: 128. https://doi.org/10.1038/s41392-021-00507-5.
- [82] Sun W, Wang C, Cui S, Wang Y, Zhao S, Lu M, et al. Association of GSDMD with microvascular-ischemia reperfusion injury after ST-elevation myocardial infarction. Frontiers in Cardiovascular Medicine. 2023; 10: 1138352. https://doi.org/10.3389/fc



- vm.2023.1138352.
- [83] Åkerblom A, James SK, Lakic TG, Becker RC, Cannon CP, Steg PG, et al. Interleukin-18 in patients with acute coronary syndromes. Clinical Cardiology. 2019; 42: 1202–1209. https://doi.org/10.1002/clc.23274.
- [84] Boutin JA, Kennaway DJ, Jockers R. Melatonin: Facts, Extrapolations and Clinical Trials. Biomolecules. 2023; 13: 943. https://doi.org/10.3390/biom13060943.
- [85] Galano A, Tan DX, Reiter RJ. On the free radical scavenging activities of melatonin's metabolites, AFMK and AMK. Journal of Pineal Research. 2013; 54: 245–257. https://doi.org/10.1111/ jpi.12010.
- [86] Wang X, Wang W, Zhang R, Ma B, Ni L, Feng H, *et al.* Melatonin attenuates high glucose induced endothelial cell pyroptosis by activating the Nrf2 pathway to inhibit NLRP3 inflammasome activation. Molecular Medicine Reports. 2023; 27: 71.
- [87] Yang Y, Sun Y, Yi W, Li Y, Fan C, Xin Z, et al. A review of melatonin as a suitable antioxidant against myocardial ischemia-reperfusion injury and clinical heart diseases. Journal of Pineal Research. 2014; 57: 357–366. https://doi.org/10.1111/jpi.12175.
- [88] Ekeloef S, Halladin N, Fonnes S, Jensen SE, Zaremba T, Rosenberg J, et al. Effect of Intracoronary and Intravenous Melatonin on Myocardial Salvage Index in Patients with ST-Elevation Myocardial Infarction: a Randomized Placebo Controlled Trial. Journal of Cardiovascular Translational Research. 2017; 10: 470–479. https://doi.org/10.1007/s12265-017-9768-7.
- [89] Dominguez-Rodriguez A, Abreu-Gonzalez P, de la Torre-Hernandez JM, Consuegra-Sanchez L, Piccolo R, Gonzalez-Gonzalez J, et al. Usefulness of Early Treatment With Melatonin to Reduce Infarct Size in Patients With ST-Segment Elevation Myocardial Infarction Receiving Percutaneous Coronary Intervention (From the Melatonin Adjunct in the Acute Myocardial Infarction Treated With Angioplasty Trial). The American Journal of Cardiology. 2017; 120: 522–526. https://doi.org/10.1016/j.amjcard.2017.05.018.
- [90] Dwaich KH, Al-Amran FGY, Al-Sheibani BIM, Al-Aubaidy HA. Melatonin effects on myocardial ischemia-reperfusion injury: Impact on the outcome in patients undergoing coronary artery bypass grafting surgery. International Journal of Cardiology. 2016; 221: 977–986. https://doi.org/10.1016/j.ijcard.2016. 07.108.
- [91] Nasseh N, Khezri MB, Farzam S, Shiravandi S, Shafikhani AA. The effect of melatonin on cardiac biomarkers after coronary artery bypass graft surgery: A double-blind, randomized pilot study. Journal of Cardiothoracic and Vascular Anesthesia. 2022; 36: 3800–3805. https://doi.org/10.1053/j.jvca.2022.06.003.
- [92] Pasupathy S, Tavella R, Grover S, Raman B, Procter NEK, Du YT, et al. Early Use of N-acetylcysteine With Nitrate Therapy in Patients Undergoing Primary Percutaneous Coronary Intervention for ST-Segment-Elevation Myocardial Infarction Reduces Myocardial Infarct Size (the NACIAM Trial [Nacetylcysteine in Acute Myocardial Infarction]). Circulation. 2017; 136: 894–903. https://doi.org/10.1161/CIRCULATIONA HA.117.027575.
- [93] Wang T, Qiao S, Lei S, Liu Y, Ng KFJ, Xu A, et al. N-acetylcysteine and allopurinol synergistically enhance cardiac adiponectin content and reduce myocardial reperfusion injury in diabetic rats. PloS One. 2011; 6: e23967. https://doi.org/10.1371/journal.pone.0023967.
- [94] Weisman A, Tomlinson GA, Lipscombe LL, Perkins BA, Hawker GA. Association between allopurinol and cardiovascular outcomes and all-cause mortality in diabetes: A retrospective, population-based cohort study. Diabetes, Obesity & Metabolism. 2019; 21: 1322–1329.
- [95] Separham A, Ghaffari S, Najafi H, Ghaffari R, Ziaee M, Babaei H. The Impact of Allopurinol on Patients With Acute ST Eleva-

- tion Myocardial Infarction Undergoing Thrombolytic Therapy. Journal of Cardiovascular Pharmacology. 2016; 68: 265–268. https://doi.org/10.1097/FJC.000000000000000409.
- [96] Kermani-Alghoraishi M, Sanei H, Heshmat-Ghahdarijani K, Ghahramani R, Honarvar M, Sadeghi M. Impact of Allopurinol Pretreatment on Coronary Blood Flow and Revascularization Outcomes after Percutaneous Coronary Intervention in Acute STEMI Patients: A Randomized Double Blind Clinical Trial. ARYA Atherosclerosis. 2023; 19: 1–9. https://doi.org/10.48305/ arya.2023.11577.2121.
- [97] Rentoukas E, Tsarouhas K, Tsitsimpikou C, Lazaros G, Deftereos S, Vavetsi S. The prognostic impact of allopurinol in patients with acute myocardial infarction undergoing primary percutaneous coronary intervention. International Journal of Cardiology. 2010; 145: 257–258. https://doi.org/10.1016/j.ij card.2009.08.037.
- [98] Yamashita T, Abe K. Update on Antioxidant Therapy with Edaravone: Expanding Applications in Neurodegenerative Diseases. International Journal of Molecular Sciences. 2024; 25: 2945. https://doi.org/10.3390/ijms25052945.
- [99] Chen H, Chen Y, Wang X, Yang J, Huang C. Edaravone attenuates myocyte apoptosis through the JAK2/STAT3 pathway in acute myocardial infarction. Free Radical Research. 2020; 54: 351–359. https://doi.org/10.1080/10715762.2020.1772469.
- [100] Tsujita K, Shimomura H, Kaikita K, Kawano H, Hokamaki J, Nagayoshi Y, et al. Long-term efficacy of edaravone in patients with acute myocardial infarction. Circulation Journal: Official Journal of the Japanese Circulation Society. 2006; 70: 832–837. https://doi.org/10.1253/circj.70.832.
- [101] Nakamura Y, Yamada Y, Shimomura H, Nagayoshi Y, Tsu-jita K, Yamashita T, et al. Effect of edaravone on plasma monocyte chemoattractant protein-1 levels in patients with acute my-ocardial infarction. Journal of Cardiology. 2009; 54: 416–424. https://doi.org/10.1016/j.jjcc.2009.07.001.
- [102] Sun WH, Liu F, Chen Y, Zhu YC. Hydrogen sulfide decreases the levels of ROS by inhibiting mitochondrial complex IV and increasing SOD activities in cardiomyocytes under ischemia/reperfusion. Biochemical and Biophysical Research Communications. 2012; 421: 164–169. https://doi.org/10.1016/j.bbrc.2012.03.121.
- [103] Calvert JW, Jha S, Gundewar S, Elrod JW, Ramachandran A, Pattillo CB, et al. Hydrogen sulfide mediates cardioprotection through Nrf2 signaling. Circulation Research. 2009; 105: 365– 374. https://doi.org/10.1161/CIRCRESAHA.109.199919.
- [104] de Koning MSL, van Dorp P, Assa S, Hartman MH, Voskuil M, Anthonio RL, et al. Rationale and Design of the Groningen Intervention Study for the Preservation of Cardiac Function with Sodium Thiosulfate after St-segment Elevation Myocardial Infarction (GIPS-IV) trial. American Heart Journal. 2022; 243: 167–176. https://doi.org/10.1016/j.ahj.2021.08.012.
- [105] Ozcan C, Bienengraeber M, Dzeja PP, Terzic A. Potassium channel openers protect cardiac mitochondria by attenuating oxidant stress at reoxygenation. American Journal of Physiology. Heart and Circulatory Physiology. 2002; 282: H531–H539. https://doi.org/10.1152/ajpheart.00552.2001.
- [106] Chen F, Chen ZQ, Zhong GL, Zhu JJ. Nicorandil inhibits TLR4/MyD88/NF-κB/NLRP3 signaling pathway to reduce pyroptosis in rats with myocardial infarction. Experimental Biology and Medicine (Maywood, N.J.). 2021; 246: 1938–1947. https://doi.org/10.1177/15353702211013444.
- [107] Geng N, Ren L, Xu L, Zou D, Pang W. Clinical outcomes of nicorandil administration in patients with acute ST-segment elevation myocardial infarction undergoing primary percutaneous coronary intervention: a systematic review and meta-analysis of randomized controlled trials. BMC Cardiovascular Disorders. 2021; 21: 488. https://doi.org/10.1186/s12872-021-02301-1.



- [108] Choe JC, Oh JH, Lee HC, Lee JW, Park TS, Park JH, et al. The effect of nicorandil on cardiac function and clinical outcomes in ST-segment elevation myocardial infarction patients undergoing primary percutaneous coronary intervention: a randomised trial. Acta Cardiologica. 2023; 78: 880–888. https://doi.org/10.1080/ 00015385.2022.2129592.
- [109] Chang X, Zhang T, Meng Q, ShiyuanWang, Yan P, Wang X, et al. Quercetin Improves Cardiomyocyte Vulnerability to Hypoxia by Regulating SIRT1/TMBIM6-Related Mitophagy and Endoplasmic Reticulum Stress. Oxidative Medicine and Cellular Longevity. 2021; 2021: 5529913. https://doi.org/10.1155/2021/5529913.
- [110] Kozhukhov S, Parkhomenko A, Lutay Y, Dovganych N, Study Investigators. Impact of quercetin in patients with myocardial infarction. A multicenter, randomized, and open-label pilot study. Hellenic Journal of Cardiology: HJC = Hellenike Kardiologike Epitheorese. 2024; 76: 68–74. https://doi.org/10.1016/ j.hic.2023.08.004.
- [111] Zhang L, Cheng L, Wang Q, Zhou D, Wu Z, Shen L, et al. Atorvastatin protects cardiomyocytes from oxidative stress by inhibiting LOX-1 expression and cardiomyocyte apoptosis. Acta Biochimica et Biophysica Sinica. 2015; 47: 174–182. https://doi.org/10.1093/abbs/gmu131.
- [112] Akhmedov A, Bonetti NR, Reiner MF, Spescha RD, Amstalden H, Merlini M, et al. Deleterious role of endothelial lectin-like oxidized low-density lipoprotein receptor-1 in ischaemia/reperfusion cerebral injury. Journal of Cerebral Blood Flow and Metabolism: Official Journal of the International Society of Cerebral Blood Flow and Metabolism. 2019; 39: 2233–2245. https://doi.org/10.1177/0271678X18793266.
- [113] Kang BY, Mehta JL. Rosuvastatin attenuates Ang II-mediated cardiomyocyte hypertrophy via inhibition of LOX-1. Journal of Cardiovascular Pharmacology and Therapeutics. 2009; 14: 283– 291. https://doi.org/10.1177/1074248409344329.
- [114] Pignatelli P, Carnevale R, Di Santo S, Bartimoccia S, Nocella C, Vicario T, et al. Rosuvastatin reduces platelet recruitment by inhibiting NADPH oxidase activation. Biochemical Pharmacology. 2012; 84: 1635–1642. https://doi.org/10.1016/j.bcp.2012.09.011.
- [115] Liu Z, Zhang F, Zhao L, Zhang X, Li Y, Liu L. Protective Effect of Pravastatin on Myocardial Ischemia Reperfusion Injury by Regulation of the miR-93/Nrf2/ARE Signal Pathway. Drug Design, Development and Therapy. 2020; 14: 3853–3864. https://doi.org/10.2147/DDDT.S251726.
- [116] Ko YG, Won H, Shin DH, Kim JS, Kim BK, Choi D, *et al.* Efficacy of early intensive rosuvastatin therapy in patients with ST-segment elevation myocardial infarction undergoing primary percutaneous coronary intervention (ROSEMARY Study). The American Journal of Cardiology. 2014; 114: 29–35. https://doi.org/10.1016/j.amjcard.2014.03.059.
- [117] Jackson TS, Xu A, Vita JA, Keaney JF, Jr. Ascorbate prevents the interaction of superoxide and nitric oxide only at very high physiological concentrations. Circulation Research. 1998; 83: 916–922. https://doi.org/10.1161/01.res.83.9.916.
- [118] Ulker S, McKeown PP, Bayraktutan U. Vitamins reverse endothelial dysfunction through regulation of eNOS and NAD(P)H oxidase activities. Hypertension (Dallas, Tex.: 1979). 2003; 41: 534–539. https://doi.org/10.1161/01.HYP.0000057421.28533. 37.
- [119] Newaz MA, Yousefipour Z, Nawal NNA. Modulation of nitric oxide synthase activity in brain, liver, and blood vessels of spontaneously hypertensive rats by ascorbic acid: protection from free radical injury. Clinical and Experimental Hypertension (New York, N.Y.: 1993). 2005; 27: 497–508. https://doi.org/10.1081/CEH-200067681.
- [120] Rozemeijer S, Hemilä H, van Baaren M, de Man AME. Vitamin C may reduce troponin and CKMB levels after PCI and CABG:

- a meta-analysis. BMC Cardiovascular Disorders. 2023; 23: 475. https://doi.org/10.1186/s12872-023-03459-6.
- [121] Wang ZJ, Hu WK, Liu YY, Shi DM, Cheng WJ, Guo YH, et al. The effect of intravenous vitamin C infusion on periprocedural myocardial injury for patients undergoing elective percutaneous coronary intervention. The Canadian Journal of Cardiology. 2014; 30: 96–101. https://doi.org/10.1016/j.cjca.2013.08.018.
- [122] Hahn JY, Kim HJ, Choi YJ, Jo SH, Kim HJ, Lee S, et al. Effects of atorvastatin pretreatment on infarct size in patients with ST-segment elevation myocardial infarction undergoing primary percutaneous coronary intervention. American Heart Journal. 2011; 162: 1026–1033. https://doi.org/10.1016/j.ahj.2011.08.
- [123] Wang Z, Yao M, Jiang L, Wang L, Yang Y, Wang Q, et al. Dexmedetomidine attenuates myocardial ischemia/reperfusioninduced ferroptosis via AMPK/GSK-3β/Nrf2 axis. Biomedicine & Pharmacotherapy. 2022; 154: 113572. https://doi.org/10. 1016/j.biopha.2022.113572.
- [124] Zhou H, Zhou D, Lu J, Wu C, Zhu Z. Effects of Pre-Cardiopulmonary Bypass Administration of Dexmedetomidine on Cardiac Injuries and the Inflammatory Response in Valve Replacement Surgery With a Sevoflurane Postconditioning Protocol: A Pilot Study. Journal of Cardiovascular Pharmacology. 2019; 74: 91–97. https://doi.org/10.1097/FJC. 0000000000000000698.
- [125] Chen W, Zhang Y, Wang Z, Tan M, Lin J, Qian X, et al. Dapagliflozin alleviates myocardial ischemia/reperfusion injury by reducing ferroptosis via MAPK signaling inhibition. Frontiers in Pharmacology. 2023; 14: 1078205. https://doi.org/10.3389/fphar.2023.1078205.
- [126] James S, Erlinge D, Storey RF, McGuire DK, de Belder M, Eriksson N, et al. Dapagliflozin in Myocardial Infarction without Diabetes or Heart Failure. NEJM Evidence. 2024; 3: EVI-Doa2300286. https://doi.org/10.1056/EVIDoa2300286.
- [127] Lu Q, Yang L, Xiao JJ, Liu Q, Ni L, Hu JW, et al. Empagliflozin attenuates the renal tubular ferroptosis in diabetic kidney disease through AMPK/NRF2 pathway. Free Radical Biology & Medicine. 2023; 195: 89–102. https://doi.org/10.1016/j.freera dbiomed.2022.12.088.
- [128] Butler J, Jones WS, Udell JA, Anker SD, Petrie MC, Harrington J, et al. Empagliflozin after Acute Myocardial Infarction. The New England Journal of Medicine. 2024; 390: 1455–1466. https://doi.org/10.1056/NEJMoa2314051.
- [129] Zhou B, Zhang J, Chen Y, Liu Y, Tang X, Xia P, *et al.* Puerarin protects against sepsis-induced myocardial injury through AMPK-mediated ferroptosis signaling. Aging. 2022; 14: 3617–3632. https://doi.org/10.18632/aging.204033.
- [130] Xie R, Du J, Hao Y. Myocardial protection and mechanism of Puerarin Injection on patients of coronary heart disease with ischemia/reperfusion. Zhongguo Zhong Xi Yi Jie He Za Zhi Zhongguo Zhongxiyi Jiehe Zazhi= Chinese Journal of Integrated Traditional and Western Medicine. 2003; 23: 895–897. (In Chinese)
- [131] Sheng H, Xiong J, Yang D. Protective Effect of Sevoflurane Preconditioning on Cardiomyocytes Against Hypoxia/Reoxygenation Injury by Modulating Iron Homeostasis and Ferroptosis. Cardiovascular Toxicology. 2023; 23: 86–92. https://doi.org/10.1007/s12012-023-09782-w.
- [132] Bonvini JM, Beck-Schimmer B, Kuhn SJ, Graber SM, Neff TA, Schläpfer M. Late Post-Conditioning with Sevoflurane after Cardiac Surgery—Are Surrogate Markers Associated with Clinical Outcome? PloS One. 2015; 10: e0132165. https://doi.org/ 10.1371/journal.pone.0132165.
- [133] Li S, Lei Z, Yang X, Zhao M, Hou Y, Wang D, et al. Propofol Protects Myocardium From Ischemia/Reperfusion Injury by Inhibiting Ferroptosis Through the AKT/p53 Signaling Pathway.



- Frontiers in Pharmacology. 2022; 13: 841410. https://doi.org/10.3389/fphar.2022.841410.
- [134] Yoo YC, Shim JK, Song Y, Yang SY, Kwak YL. Anesthetics influence the incidence of acute kidney injury following valvular heart surgery. Kidney International. 2014; 86: 414–422. https://doi.org/10.1038/ki.2013.532.
- [135] Hershko C, Konijn AM, Nick HP, Breuer W, Cabantchik ZI, Link G. ICL670A: a new synthetic oral chelator: evaluation in hypertransfused rats with selective radioiron probes of hepatocellular and reticuloendothelial iron stores and in iron-loaded rat heart cells in culture. Blood. 2001; 97: 1115–1122. https://doi.org/10.1182/blood.v97.4.1115.
- [136] Chan W, Taylor AJ, Ellims AH, Lefkovits L, Wong C, Kingwell BA, et al. Effect of iron chelation on myocardial infarct size and oxidative stress in ST-elevation-myocardial infarction. Circulation. Cardiovascular Interventions. 2012; 5: 270–278. https://doi.org/10.1161/CIRCINTERVENTIONS.111.966226.
- [137] Ferreira R, Burgos M, Milei J, Llesuy S, Molteni L, Hourquebie H, et al. Effect of supplementing cardioplegic solution with deferoxamine on reperfused human myocardium. The Journal of Thoracic and Cardiovascular Surgery. 1990; 100: 708–714.
- [138] Paraskevaidis IA, Iliodromitis EK, Vlahakos D, Tsiapras DP, Nikolaidis A, Marathias A, *et al.* Deferoxamine infusion during coronary artery bypass grafting ameliorates lipid peroxidation and protects the myocardium against reperfusion injury: immediate and long-term significance. European Heart Journal. 2005; 26: 263–270. https://doi.org/10.1093/eurheartj/ehi028.
- [139] Zhang W, Wang X, Tang Y, Huang C. Melatonin alleviates doxorubicin-induced cardiotoxicity via inhibiting oxidative stress, pyroptosis and apoptosis by activating Sirt1/Nrf2 pathway. Biomedicine & Pharmacotherapy. 2023; 162: 114591. https://doi.org/10.1016/j.biopha.2023.114591. https://doi.org/10.3892/mmr.2023.12958.
- [140] Han D, Huang W, Li X, Gao L, Su T, Li X, et al. Melatonin facilitates adipose-derived mesenchymal stem cells to repair the murine infarcted heart via the SIRT1 signaling pathway. Journal of Pineal Research. 2016; 60: 178–192. https://doi.org/10.1111/jpi.12299.
- [141] Rahim I, Sayed RK, Fernández-Ortiz M, Aranda-Martínez P, Guerra-Librero A, Fernández-Martínez J, et al. Melatonin alleviates sepsis-induced heart injury through activating the Nrf2 pathway and inhibiting the NLRP3 inflammasome. Naunyn-Schmiedeberg's Archives of Pharmacology. 2021; 394: 261–277. https://doi.org/10.1007/s00210-020-01972-5.
- [142] Niu YJ, Zhou W, Nie ZW, Shin KT, Cui XS. Melatonin enhances mitochondrial biogenesis and protects against rotenone-induced mitochondrial deficiency in early porcine embryos. Journal of Pineal Research. 2020; 68: e12627. https://doi.org/10.1111/jpi.12627.
- [143] Lv T, Yan J, Lou Y, Zhang Z, Ye M, Zhou J, *et al.* Evaluation of Melatonin Therapy in Patients with Myocardial Ischemia-Reperfusion Injury: A Systematic Review and Meta-Analysis. Oxidative Medicine and Cellular Longevity. 2022; 2022: 4610522. https://doi.org/10.1155/2022/4610522.
- [144] Domínguez-Rodríguez A, Abreu-González P, Báez-Ferrer N, Reiter RJ, Avanzas P, Hernández-Vaquero D. Melatonin and Cardioprotection in Humans: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. Frontiers in Cardiovascular Medicine. 2021; 8: 635083. https://doi.org/10.3389/fc vm.2021.635083.
- [145] Andersen LPH, Gögenur I, Rosenberg J, Reiter RJ. The Safety of Melatonin in Humans. Clinical Drug Investigation. 2016; 36: 169–175. https://doi.org/10.1007/s40261-015-0368-5.
- [146] El-Sayed SF, Abdelhamid AM, ZeinElabdeen SG, El-Wafaey DI, Moursi SMM. Melatonin enhances captopril mediated cardioprotective effects and improves mitochondrial dynamics in male Wistar rats with chronic heart failure. Scientific Reports.

- 2024; 14: 575. https://doi.org/10.1038/s41598-023-50730-z.
- [147] Sahasrabudhe SA, Terluk MR, Kartha RV. N-acetylcysteine Pharmacology and Applications in Rare Diseases-Repurposing an Old Antioxidant. Antioxidants (Basel, Switzerland). 2023; 12: 1316. https://doi.org/10.3390/antiox12071316.
- [148] Santus P, Signorello JC, Danzo F, Lazzaroni G, Saad M, Radovanovic D. Anti-Inflammatory and Anti-Oxidant Properties of N-Acetylcysteine: A Fresh Perspective. Journal of Clinical Medicine. 2024; 13: 4127. https://doi.org/10.3390/jc m13144127.
- [149] Wasyanto T, Yasa' A, Jalaludinsyah A. Effect of Oral N-Acetylcysteine Supplementation on the Immunity System in Patients with Acute Myocardial Infarction. Acta Medica Indonesiana. 2019; 51: 311–317.
- [150] Yesilbursa D, Serdar A, Senturk T, Serdar Z, Sağ S, Cordan J. Effect of N-acetylcysteine on oxidative stress and ventricular function in patients with myocardial infarction. Heart and Vessels. 2006; 21: 33–37. https://doi.org/10.1007/s00380-005-0854-4.
- [151] Talasaz AH, Khalili H, Jenab Y, Salarifar M, Broumand MA, Darabi F. N-Acetylcysteine effects on transforming growth factor-β and tumor necrosis factor-α serum levels as pro-fibrotic and inflammatory biomarkers in patients following ST-segment elevation myocardial infarction. Drugs in R&D. 2013; 13: 199– 205. https://doi.org/10.1007/s40268-013-0025-5.
- [152] Pereira JEG, El Dib R, Braz LG, Escudero J, Hayes J, Johnston BC. N-acetylcysteine use among patients undergoing cardiac surgery: A systematic review and meta-analysis of randomized trials. PloS One. 2019; 14: e0213862. https://doi.org/10.1371/journal.pone.0213862.
- [153] Khan SA, Campbell AM, Lu Y, An L, Alpert JS, Chen QM. N-Acetylcysteine for Cardiac Protection During Coronary Artery Reperfusion: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. Frontiers in Cardiovascular Medicine. 2021; 8: 752939. https://doi.org/10.3389/fcvm.2021. 752939.
- [154] Ardissino D, Merlini PA, Savonitto S, Demicheli G, Zanini P, Bertocchi F, et al. Effect of Transdermal Nitroglycerin or N-Acetylcysteine, or Both, in the Long-Term Treatment of Unstable Angina Pectoris. Journal of the American College of Cardiology. 1997; 29: 941–947. https://doi.org/10.1016/S0735-1097(97)00005-3. https://doi.org/10.1111/dom.13656.
- [155] Zhang YS, Lu LQ, Jiang YQ, Li NS, Luo XJ, Peng JW, et al. Allopurinol attenuates oxidative injury in rat hearts suffered ischemia/reperfusion via suppressing the xanthine oxidase/vascular peroxidase 1 pathway. European Journal of Pharmacology. 2021; 908: 174368. https://doi.org/10.1016/j.ejphar.2021.174368.
- [156] Guan W, Osanai T, Kamada T, Hanada H, Ishizaka H, Onodera H, et al. Effect of allopurinol pretreatment on free radical generation after primary coronary angioplasty for acute myocardial infarction. Journal of Cardiovascular Pharmacology. 2003; 41: 699–705. https://doi.org/10.1097/00005344-200305000-00005.
- [157] Kawai H, Nakai H, Suga M, Yuki S, Watanabe T, Saito KI. Effects of a novel free radical scavenger, MCl-186, on ischemic brain damage in the rat distal middle cerebral artery occlusion model. The Journal of Pharmacology and Experimental Therapeutics. 1997; 281: 921–927.
- [158] El-Shoura EAM, Hassanein EHM, Taha HH, Shalkami AGS, Hassanein MMH, Ali FEM, et al. Edaravone and obeticholic acid protect against cisplatin-induced heart toxicity by suppressing oxidative stress and inflammation and modulating Nrf2, TLR4/p38MAPK, and JAK1/STAT3/NF-κB signals. Naunyn-Schmiedeberg's Archives of Pharmacology. 2024; 397: 5649– 5662. https://doi.org/10.1007/s00210-024-02956-5.
- [159] Kikuchi K, Tancharoen S, Takeshige N, Yoshitomi M, Morioka



- M, Murai Y, *et al*. The efficacy of edaravone (radicut), a free radical scavenger, for cardiovascular disease. International Journal of Molecular Sciences. 2013; 14: 13909–13930. https://doi.org/10.3390/ijms140713909.
- [160] Zhang MY, Dugbartey GJ, Juriasingani S, Sener A. Hydrogen Sulfide Metabolite, Sodium Thiosulfate: Clinical Applications and Underlying Molecular Mechanisms. International Journal of Molecular Sciences. 2021; 22: 6452. https://doi.org/10.3390/ij ms22126452.
- [161] de Koning MSLY, van Dorp P, Assa S, Pundziute-Do Prado G, Voskuil M, Anthonio RL, et al. Sodium Thiosulfate in Acute Myocardial Infarction: A Randomized Clinical Trial. JACC. Basic to Translational Science. 2023; 8: 1285–1294. https://doi.org/10.1016/j.jacbts.2023.06.001.
- [162] Goel H, Carey M, Elshaikh A, Krinock M, Goyal D, Nadar SK. Cardioprotective and Antianginal Efficacy of Nicorandil: A Comprehensive Review. Journal of Cardiovascular Pharmacology. 2023; 82: 69–85. https://doi.org/10.1097/FJC. 00000000000001436.
- [163] Refaie MMM, Shehata S, El-Hussieny M, Abdelraheem WM, Bayoumi AMA. Role of ATP-Sensitive Potassium Channel (K_{ATP}) and eNOS in Mediating the Protective Effect of Nicorandil in Cyclophosphamide-Induced Cardiotoxicity. Cardiovascular Toxicology. 2020; 20: 71–81. https://doi.org/10.1007/ s12012-019-09535-8.
- [164] Tariq H, Ahmed S, Ahmed S, Hanif N, Anwar E, Kumari A, et al. Efficacy of Nicorandil in Preventing Myocardial Injury and Cardiovascular Outcomes in Patients Undergoing Percutaneous Coronary Intervention (PCI): A Systematic Review and Meta-Analysis. Cureus. 2024; 16: e66938. https://doi.org/10.7759/cureus.66938.
- [165] Yamada K, Isobe S, Ishii H, Yokouchi K, Iwata H, Sawada K, et al. Impacts of nicorandil on infarct myocardium in comparison with nitrate: assessed by cardiac magnetic resonance imaging. Heart and Vessels. 2016; 31: 1430–1437. https://doi.org/10.1007/s00380-015-0752-3.
- [166] Ilyas M, Noor M, Haroon S, Farhat K, Ali S, Wahid M. Assessment of cardiac parameters after the administration of nicorandil before primary percutaneous coronary intervention. JPMA. the Journal of the Pakistan Medical Association. 2024; 74: 917–921. https://doi.org/10.47391/JPMA.9981.
- [167] Alizadeh SR, Ebrahimzadeh MA. Quercetin derivatives: Drug design, development, and biological activities, a review. European Journal of Medicinal Chemistry. 2022; 229: 114068. https://doi.org/10.1016/j.ejmech.2021.114068.
- [168] Ding X, Zhu C, Wang W, Li M, Ma C, Gao B. SIRT1 is a regulator of autophagy: Implications for the progression and treatment of myocardial ischemia-reperfusion. Pharmacological Research. 2024; 199: 106957. https://doi.org/10.1016/j.phrs.2023. 106957.
- [169] Mendieta G, Ben-Aicha S, Casani L, Badimon L, Sabate M, Vilahur G. Molecular pathways involved in the cardioprotective effects of intravenous statin administration during ischemia. Basic Research in Cardiology. 2019; 115: 2. https://doi.org/10.1007/s00395-019-0760-z.
- [170] Bland AR, Payne FM, Ashton JC, Jamialahmadi T, Sahebkar A. The cardioprotective actions of statins in targeting mitochondrial dysfunction associated with myocardial ischaemia-reperfusion injury. Pharmacological Research. 2022; 175: 105986. https:// doi.org/10.1016/j.phrs.2021.105986.
- [171] Mendieta G, Ben-Aicha S, Gutiérrez M, Casani L, Aržanauskaitė M, Carreras F, *et al.* Intravenous Statin Administration During Myocardial Infarction Compared With Oral Post-Infarct Administration. Journal of the American College of Cardiology. 2020; 75: 1386–1402. https://doi.org/10.1016/j.jacc.2020.01.042.
- [172] Atar S, Ye Y, Lin Y, Freeberg SY, Nishi SP, Rosanio S, et al.

- Atorvastatin-induced cardioprotection is mediated by increasing inducible nitric oxide synthase and consequent S-nitrosylation of cyclooxygenase-2. American Journal of Physiology. Heart and Circulatory Physiology. 2006; 290: H1960–H1968. https://doi.org/10.1152/ajpheart.01137.2005.
- [173] Birnbaum Y, Lin Y, Ye Y, Martinez JD, Huang MH, Lui CY, et al. Aspirin before reperfusion blunts the infarct size limiting effect of atorvastatin. American Journal of Physiology. Heart and Circulatory Physiology. 2007; 292: H2891–H2897. https://doi.org/10.1152/ajpheart.01269.2006.
- [174] Ye Y, Lin Y, Perez-Polo R, Huang MH, Hughes MG, McAdoo DJ, et al. Enhanced cardioprotection against ischemiareperfusion injury with a dipyridamole and low-dose atorvastatin combination. American Journal of Physiology. Heart and Circulatory Physiology. 2007; 293: H813–H818. https://doi.or g/10.1152/ajpheart.00210.2007.
- [175] Ye Y, Long B, Qian J, Perez-Polo JR, Birnbaum Y. Dipyridamole with low-dose aspirin augments the infarct size-limiting effects of simvastatin. Cardiovascular Drugs and Therapy. 2010; 24: 391–399. https://doi.org/10.1007/s10557-010-6252-x.
- [176] Manickavasagam S, Ye Y, Lin Y, Perez-Polo RJ, Huang MH, Lui CY, et al. The cardioprotective effect of a statin and cilostazol combination: relationship to Akt and endothelial nitric oxide synthase activation. Cardiovascular Drugs and Therapy. 2007; 21: 321–330. https://doi.org/10.1007/s10557-007-6036-0.
- [177] Ng KW, Allen ML, Desai A, Macrae D, Pathan N. Cardioprotective effects of insulin: how intensive insulin therapy may benefit cardiac surgery patients. Circulation. 2012; 125: 721–728. https://doi.org/10.1161/CIRCULATIONAHA.111.063784.
- [178] Lazzeri C, Valente S, Chiostri M, Attanà P, Picariello C, Gensini GF. The glucose dysmetabolism in the acute phase of non-diabetic ST-elevation myocardial infarction: from insulin resistance to hyperglycemia. Acta Diabetologica. 2013; 50: 293–300. https://doi.org/10.1007/s00592-011-0325-6.
- [179] Shannar A, Chou PJ, Peter R, Dave PD, Patel K, Pan Y, et al. Pharmacodynamics (PD), Pharmacokinetics (PK) and PK-PD Modeling of NRF2 Activating Dietary Phytochemicals in Cancer Prevention and in Health. Current Pharmacology Reports. 2025; 11: 6. https://doi.org/10.1007/s40495-024-00388-6.
- [180] Huang Z, Ye B, Dai Z, Wu X, Lu Z, Shan P, et al. Curcumin inhibits autophagy and apoptosis in hypoxia/reoxygenationinduced myocytes. Molecular Medicine Reports. 2015; 11: 4678–4684. https://doi.org/10.3892/mmr.2015.3322.
- [181] Dludla PV, Cirilli I, Marcheggiani F, Silvestri S, Orlando P, Muvhulawa N, et al. Bioactive Properties, Bioavailability Profiles, and Clinical Evidence of the Potential Benefits of Black Pepper (Piper nigrum) and Red Pepper (Capsicum annum) against Diverse Metabolic Complications. Molecules (Basel, Switzerland). 2023; 28: 6569. https://doi.org/10.3390/molecule s28186569.
- [182] Tehrani SD, Hosseini A, Shahzamani M, Heidari Z, Askari G, Majeed M, et al. Evaluation of the effectiveness of curcumin and piperine co-supplementation on inflammatory factors, cardiac biomarkers, atrial fibrillation, and clinical outcomes after coronary artery bypass graft surgery. Clinical Nutrition ESPEN. 2024; 62: 57–65. https://doi.org/10.1016/j.clnesp.2024.05.003.
- [183] Tabaee S, Sahebkar A, Aghamohammadi T, Pakdel M, Dehabeh M, Sobhani R, et al. The Effects of Curcumin Plus Piperine Supplementation in Patients with Acute Myocardial Infarction: A Randomized, Double-Blind, and Placebo-Controlled Trial. Advances in Experimental Medicine and Biology. 2021; 1328: 199–211. https://doi.org/10.1007/978-3-030-73234-9_13.
- [184] Li T, Jin J, Pu F, Bai Y, Chen Y, Li Y, et al. Cardioprotective effects of curcumin against myocardial I/R injury: A systematic review and meta-analysis of preclinical and clinical



- studies. Frontiers in Pharmacology. 2023; 14: 1111459. https://doi.org/10.3389/fphar.2023.1111459.
- [185] Singh RB, Niaz MA, Rastogi SS, Rastogi S. Usefulness of antioxidant vitamins in suspected acute myocardial infarction (the Indian experiment of infarct survival-3). The American Journal of Cardiology. 1996; 77: 232–236. https://doi.org/10.1016/s0002-9149(97)89384-8.
- [186] McNulty H, Jacob RF, Mason RP. Biologic activity of carotenoids related to distinct membrane physicochemical interactions. The American Journal of Cardiology. 2008; 101: 20D– 29D. https://doi.org/10.1016/j.amjcard.2008.02.004.
- [187] Cox FF, Misiou A, Vierkant A, Ale-Agha N, Grandoch M, Haendeler J, et al. Protective Effects of Curcumin in Cardiovascular Diseases-Impact on Oxidative Stress and Mitochondria. Cells. 2022; 11: 342. https://doi.org/10.3390/cells11030342.
- [188] Ma ZG, Yuan YP, Zhang X, Xu SC, Wang SS, Tang QZ. Piperine Attenuates Pathological Cardiac Fibrosis Via PPAR-γ/AKT Pathways. EBioMedicine. 2017; 18: 179–187. https://doi.org/ 10.1016/j.ebiom.2017.03.021.
- [189] Li YP, Chen Z, Cai YH. Piperine protects against myocardial ischemia/reperfusion injury by activating the PI3K/AKT signaling pathway. Experimental and Therapeutic Medicine. 2021; 21: 374. https://doi.org/10.3892/etm.2021.9805.
- [190] Shipp JC, Opie LH, Challoner D. Fatty Acid and Glucose Metabolism in the Perfused Heart. Nature. 1961; 189: 1018– 1019. https://doi.org/10.1038/1891018a0.
- [191] Díaz-Araya G, Nettle D, Castro P, Miranda F, Greig D, Campos X, et al. Oxidative stress after reperfusion with primary coronary angioplasty: lack of effect of glucose-insulin-potassium infusion. Critical Care Medicine. 2002; 30: 417–421. https://doi.org/10.1097/00003246-200202000-00025.
- [192] Demircan S, Yazici M, Diraman E, Demircan G, Kilicaslan F, Durna K, et al. The effect of glucose-insulin-potassium treatment on myocardial oxidative stress in patients with acute coronary syndromes undergoing percutaneous coronary intervention. Coronary Artery Disease. 2008; 19: 99–104. https://doi.org/10.1097/MCA.0b013e3282f27c34.
- [193] Zhao YT, Weng CL, Chen ML, Li KB, Ge YG, Lin XM, et al. Comparison of glucose-insulin-potassium and insulin-glucose as adjunctive therapy in acute myocardial infarction: a contemporary meta-analysis of randomised controlled trials. Heart (British Cardiac Society). 2010; 96: 1622–1626. https://doi.org/ 10.1136/hrt.2010.194563.
- [194] Yusuf S, Mehta SR, Díaz R, Paolasso E, Pais P, Xavier D, *et al.* Challenges in the conduct of large simple trials of important generic questions in resource-poor settings: the CREATE and ECLA trial program evaluating GIK (glucose, insulin and potassium) and low-molecular-weight heparin in acute myocardial infarction. American Heart Journal. 2004; 148: 1068–1078. https://doi.org/10.1016/j.ahj.2004.08.033.
- [195] Selker HP, Beshansky JR, Griffith JL, D'Agostino RB, Massaro JM, Udelson JE, *et al.* Study design for the Immediate Myocardial Metabolic Enhancement During Initial Assessment and Treatment in Emergency Care (IMMEDIATE) Trial: A double-blind randomized controlled trial of intravenous glucose, insulin, and potassium for acute coronary syndromes in emergency medical services. American Heart Journal. 2012; 163: 315–322. https://doi.org/10.1016/j.ahj.2012.02.002.
- [196] Udelson JE, Selker HP, Braunwald E. Glucose-Insulin-Potassium Therapy for Acute Myocardial Infarction: 50 Years On and Time for a Relook. Circulation. 2022; 146: 503–505. https://doi.org/10.1161/CIRCULATIONAHA.121.058740.
- [197] Yang Z, Liu H, Lu D, Cao S, Xu F, Li C. Effects of high-dose glucose-insulin-potassium on acute coronary syndrome patients receiving reperfusion therapy: a meta-analysis. World Journal of Emergency Medicine. 2024; 15: 181–189. https://doi.org/10.5847/wjem.j.1920-8642.2024.048.

- [198] Liu H, Liu R, Yang Z, Xu F, Li C. Effect of preinitiated glucose-insulin-potassium strategy for patients with undergoing planned percutaneous coronary intervention: a systematic review and meta-analysis. BMJ Open. 2023; 13: e073557. https://doi.org/10.1136/bmjopen-2023-073557.
- [199] El-Saadony MT, Yang T, Korma SA, Sitohy M, Abd El-Mageed TA, Selim S, et al. Impacts of turmeric and its principal bioactive curcumin on human health: Pharmaceutical, medicinal, and food applications: A comprehensive review. Frontiers in Nutrition. 2023; 9: 1040259. https://doi.org/10.3389/fn ut.2022.1040259.
- [200] Yuan Y, Huang H, Hu T, Zou C, Qiao Y, Fang M, et al. Curcumin pretreatment attenuates myocardial ischemia/reperfusion injury by inhibiting ferroptosis, autophagy and apoptosis via HES1. International Journal of Molecular Medicine. 2024; 54: 110. https://doi.org/10.3892/ijmm.2024.5434.
- [201] Yan J, Xu SC, Kong CY, Zhou XY, Bian ZY, Yan L, et al. Piperine Alleviates Doxorubicin-Induced Cardiotoxicity via Activating PPAR-γ in Mice. PPAR Research. 2019; 2019: 2601408. https://doi.org/10.1155/2019/2601408.
- [202] Wiraswati HL, Ma'ruf IF, Sharifi-Rad J, Calina D. Piperine: an emerging biofactor with anticancer efficacy and therapeutic potential. BioFactors. 2025; 51: e2134. https://doi.org/10.1002/ biof.2134.
- [203] Chakraborty M, Bhattacharjee A, Kamath JV. Cardioprotective effect of curcumin and piperine combination against cyclophosphamide-induced cardiotoxicity. Indian Journal of Pharmacology. 2017; 49: 65–70. https://doi.org/10.4103/0253-7613.201015.
- [204] Pinilla-González V, Rojas-Solé C, Gómez-Hevia F, González-Fernández T, Cereceda-Cornejo A, Chichiarelli S, et al. Tapping into Nature's Arsenal: Harnessing the Potential of Natural Antioxidants for Human Health and Disease Prevention. Foods (Basel, Switzerland). 2024; 13: 1999. https://doi.org/10.3390/foods13131999.
- [205] Shi H, Noguchi N, Niki E. Comparative study on dynamics of antioxidative action of alpha-tocopheryl hydroquinone, ubiquinol, and alpha-tocopherol against lipid peroxidation. Free Radical Biology & Medicine. 1999; 27: 334–346. https://doi.org/10.1016/s0891-5849(99)00053-2.
- [206] Packer JE, Slater TF, Willson RL. Direct observation of a free radical interaction between vitamin E and vitamin C. Nature. 1979; 278: 737–738. https://doi.org/10.1038/278737a0.
- [207] Jaxa-Chamiec T, Bednarz B, Drozdowska D, Gessek J, Gniot J, Janik K, et al. Antioxidant effects of combined vitamins C and E in acute myocardial infarction. The randomized, double-blind, placebo controlled, multicenter pilot Myocardial Infarction and VITamins (MIVIT) trial. Kardiologia Polska. 2005; 62: 344–350.
- [208] Rodrigo R, Hasson D, Prieto JC, Dussaillant G, Ramos C, León L, et al. The effectiveness of antioxidant vitamins C and E in reducing myocardial infarct size in patients subjected to percutaneous coronary angioplasty (PREVEC Trial): study protocol for a pilot randomized double-blind controlled trial. Trials. 2014; 15: 192. https://doi.org/10.1186/1745-6215-15-192.
- [209] Valls N, Gormaz JG, Aguayo R, González J, Brito R, Hasson D, et al. Amelioration of persistent left ventricular function impairment through increased plasma ascorbate levels following myocardial infarction. Redox Report: Communications in Free Radical Research. 2016; 21: 75–83. https://doi.org/10.1179/1351000215Y.0000000018.
- [210] Ramos C, Brito R, González-Montero J, Valls N, Gormaz JG, Prieto JC, et al. Effects of a novel ascorbate-based protocol on infarct size and ventricle function in acute myocardial infarction patients undergoing percutaneous coronary angioplasty. Archives of Medical Science: AMS. 2017; 13: 558–567. https:



- //doi.org/10.5114/aoms.2016.59713.
- [211] Basili S, Tanzilli G, Mangieri E, Raparelli V, Di Santo S, Pignatelli P, et al. Intravenous ascorbic acid infusion improves myocardial perfusion grade during elective percutaneous coronary intervention: relationship with oxidative stress markers. JACC. Cardiovascular Interventions. 2010; 3: 221–229. https://doi.org/10.1016/j.jcin.2009.10.025.
- [212] Wang ZJ, Hu WK, Liu YY, Shi DM, Cheng WJ, Guo YH, et al. The effect of intravenous vitamin C infusion on periprocedural myocardial injury for patients undergoing elective percutaneous coronary intervention. The Canadian Journal of Cardiology. 2014; 30: 96–101. https://doi.org/10.1016/j.cjca.2013.08.018.
- [213] Manolis AA, Manolis T, Melita H, Manolis AS. Role of Vitamins in Cardiovascular Health: Know Your Facts Part 1. Current Vascular Pharmacology. 2023; 21: 378–398. https://doi.org/10.2174/1570161121666230912155548.
- [214] Sesso HD. Carotenoids and cardiovascular disease: what research gaps remain? Current Opinion in Lipidology. 2006; 17: 11–16. https://doi.org/10.1097/01.mol.0000203888.42514.27.
- [215] Welt FGP, Batchelor W, Spears JR, Penna C, Pagliaro P, Ibanez B, et al. Reperfusion Injury in Patients With Acute Myocardial Infarction: JACC Scientific Statement. Journal of the American College of Cardiology. 2024; 83: 2196–2213. https://doi.org/10.1016/j.jacc.2024.02.056.
- [216] Zhang S, Yan F, Luan F, Chai Y, Li N, Wang YW, et al. The pathological mechanisms and potential therapeutic drugs for myocardial ischemia reperfusion injury. Phytomedicine: International Journal of Phytotherapy and Phytopharmacology. 2024; 129: 155649. https://doi.org/10.1016/j.phymed.2024.155649.
- [217] Dodd S, Dean O, Copolov DL, Malhi GS, Berk M. N-

- acetylcysteine for antioxidant therapy: pharmacology and clinical utility. Expert Opinion on Biological Therapy. 2008; 8: 1955–1962. https://doi.org/10.1517/14728220802517901.
- [218] Lu Y, Qin W, Shen T, Dou L, Man Y, Wang S, *et al.* The antioxidant N-acetylcysteine promotes atherosclerotic plaque stabilization through suppression of RAGE, MMPs and NF-κB in ApoE-deficient mice. Journal of Atherosclerosis and Thrombosis. 2011; 18: 998–1008. https://doi.org/10.5551/jat.8870.
- [219] Joshi D, Mittal DK, Shrivastava S, Shukla S. Protective role of thiol chelators against dimethylmercury induced toxicity in male rats. Bulletin of Environmental Contamination and Toxicology. 2010; 84: 613–617. https://doi.org/10.1007/ s00128-010-9982-3.
- [220] Gajardo Cortez AIJ, Lillo-Moya J, San-Martín-Martinez D, Pozo-Martinez J, Morales P, Prieto JC, et al. Safety and Pharmacokinetics of a Combined Antioxidant Therapy against Myocardial Reperfusion Injury: A Phase 1 Randomized Clinical Trial in Healthy Humans. Clinical Pharmacology in Drug Development. 2024; 13: 1051–1060. https://doi.org/10.1002/cpdd.1443.
- [221] Stone GW, Martin JL, de Boer MJ, Margheri M, Bramucci E, Blankenship JC, et al. Effect of supersaturated oxygen delivery on infarct size after percutaneous coronary intervention in acute myocardial infarction. Circulation. Cardiovascular Interventions. 2009; 2: 366–375. https://doi.org/10.1161/CIRCINTE RVENTIONS.108.840066.
- [222] Heusch G. Myocardial ischemia/reperfusion: Translational pathophysiology of ischemic heart disease. Med (New York, N.Y.). 2024; 5: 10–31. https://doi.org/10.1016/j.medj.2023.12. 007.

