Review

Pulmonary Arterial Hypertension: Emerging Principles of Precision Medicine across Basic Science to Clinical Practice

Neil J. Kelly¹, Stephen Y. Chan^{1,*}

¹Center for Pulmonary Vascular Biology and Medicine and Pittsburgh Heart, Lung, and Blood Vascular Medicine Institute; Division of Cardiology; Department of Medicine, University of Pittsburgh School of Medicine and University of Pittsburgh Medical Center, Pittsburgh, PA 15213, USA *Correspondence: chansy@pitt.edu (Stephen Y. Chan)

Academic Editor: Harald Kaemmerer

Submitted: 16 June 2022 Revised: 20 August 2022 Accepted: 9 September 2022 Published: 9 November 2022

Abstract

Pulmonary arterial hypertension (PAH) is an enigmatic and deadly vascular disease with no known cure. Recent years have seen rapid advances in our understanding of the molecular underpinnings of PAH, with an expanding knowledge of the molecular, cellular, and systems-level drivers of disease that are being translated into novel therapeutic modalities. Simultaneous advances in clinical technology have led to a growing list of tools with potential application to diagnosis and phenotyping. Guided by fundamental biology, these developments hold the potential to usher in a new era of personalized medicine in PAH with broad implications for patient management and great promise for improved outcomes.

Keywords: pulmonary hypertension; disease mechanism; systems biology; translational biology; endothelium; precision medicine

1. Introduction

Pulmonary hypertension (PH) is a complex and progressive disease involving elevated pressures in the pulmonary arteries due to one or multiple varied etiologies. Left untreated, PH is associated with right ventricular (RV) hypertrophy and failure resulting in markedly reduced life expectancy. Insights and mechanistic discoveries over the past two decades have begun to untangle this enigmatic and, so far, incurable disease. As a result, novel approaches to PH management are emerging which set the stage for introducing an era of precision medicine—holding the potential to bring earlier diagnoses, more effective treatments, and improved patient outcomes. This review will discuss key principles of the scientific basis of PH and its clinical management while highlighting emerging and potentially practice-changing concepts and technologies.

2. Clinical Definitions

Prior to our current understanding of disease mechanisms, PH was classified as either "primary" (idiopathic) or "secondary" to any of a variety of diverse clinical states. In 1998, a working group of the World Society of Pulmonary Hypertension (WSPH), sponsored by the World Health Organization (WHO), devised the current schema of clinical groupings which aims to categorize PH according to its etiology [1]. In the most recent iteration drafted by the 6th WSPH [2], PH is defined by a resting mean pulmonary artery pressure (mPAP) greater than 20 mmHg by right heart catheterization and categorized according to additional hemodynamic and clinical factors: Pulmonary arterial hypertension (PAH – WSPH Group 1), PH secondary to left heart disease (PH-LHD – WSPH Group 2),

PH secondary to chronic lung disease or hypoxia (PH-CLD – WSPH Group 3), chronic thromboembolic pulmonary hypertension (CTEPH – WSPH Group 4), and PH due to multifactorial or miscellaneous causes (WSPH Group 5). To be classified as WSPH Group 1 PAH, precapillary hemodynamics must be observed, with mPAP >20 mmHg accompanied by an elevated pulmonary vascular resistance (PVR) of greater than or equal to 3 Wood units (mmHg/L/min) and pulmonary artery wedge pressure (PAWP) less than or equal to 15 mmHg. Importantly, expert clinical assessment must rule out predominant contributions by left heart disease, hypoxic lung disease, and chronic pulmonary emboli. On the other hand, PH secondary to left heart disease (PH-LHD - WSPH Group 2) is defined by a PAWP greater than 15 mmHg regardless of PVR. In clinical practice, patients often fall into more than one category of PH [2].

While epidemiological data comparing across PH groups are less available and often limited to diagnoses inferred from echocardiography, it is clear that PAH constitutes a minority of total global PH burden, but the exact prevalence of PAH is not known. In total, PAH is diagnosed in an estimated 2.4–7.6 million individuals annually with a prevalence of 15 to 50 million cases and a strong female predominance [3,4]. This may differ, depending upon geography or epidemiologic techniques. For example, in a large population-based cohort study from Ontario, Canada, the annual prevalence of any form of PH was 127.3/100,000 of which PAH accounted for 15.6% [5], while an Australian cohort identified the proportion of overall PH prevalence due to PAH at 2.7% [6]. In recent years, the foundational and clinical understanding of PAH has advanced dramatically, and this review will focus on recent progress made in

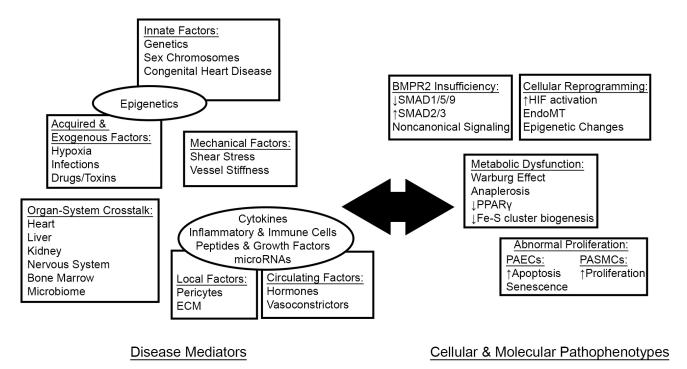


Fig. 1. PAH as a systemic disease. A growing body of literature reveals the interplay between systemic disease mediators and cellular pathophenotypes in driving pathologic vascular pathology in PAH. ECM, extracellular matrix.

the field of PAH and possible clinical advancements in the near future.

3. Etiology

PAH describes a clinical syndrome in which primary remodeling of the small muscularized arterioles and precapillary vessels (50–500 μ m diameter) promotes pathologic increases in pulmonary vascular resistance, culminating in elevated pulmonary artery pressures, RV hypertrophy, and symptomatology and death from RV failure [7]. A minority of PAH cases (2-3%) can be directly attributed to heritable causes (HPAH) [8], while roughly 50% of PAH cases are classified as idiopathic (IPAH). However, it is increasingly recognized that a substantial proportion— 20-30%—of cases labeled as idiopathic are likely to be hereditary [9]. Much of the remaining PAH burden is derived from connective tissue diseases (systemic sclerosis, lupus, rheumatoid arthritis, and others). Rare causes of PAH have been linked to associated triggers including portopulmonary hypertension, drugs/toxins (with recent increasing cases of methamphetamine use), infections (human immunodeficiency virus [HIV], schistosomiasis), congenital heart disease, pulmonary venoocclusive disease (PVOD)/pulmonary capillary hemangiomatosis (PCH), and persistent pulmonary hypertension of the newborn (PPHN)

With the exceptions of the rare entities of PVOD/PCH [10] and PPHN [11], the various causes of PAH share similar histopathological features. PAH is characterized by resistive changes to the small precapillary arterioles including

medial hypertrophy and hyperplasia, intimal and adventitial fibrosis, and thrombotic and plexiform lesions [12]. Over variable time frames, the progression and accrual of vascular remodeling manifests as clinical disease.

4. Disease Mechanisms (Fig. 1)

4.1 Disrupted Homeostasis of Vascular Effectors

Prior to our current understanding of genetic disease influences, it was apparent that the vasoconstrictive phenotype of PAH was provoked in part by endothelial dysfunction and disrupted homeostasis between various mediators of vascular tone. Among the best studied are the vasodilatory arachidonic acid metabolite prostacyclin and free radical nitric oxide (NO), as well as the vasoconstrictive peptide hormone endothelin-1 (ET-1, also known as EDN1); manipulation of these vasoactive mediators forms the basis of current pharmacotherapy in PAH [13].

Prostacyclin (PGI2) is a potent vasodilator and inhibitor of platelet activation derived from the arachidonic acid metabolite prostaglandin H2 (PGH2) [14]. Examination of the urine of PAH patients has shown that PGI2 breakdown products are decreased while those of the vasoconstrictive and platelet-activating PGH2 derivate thromboxane A2 (TXA2) are increased [15]. Additionally, prostacyclin synthase, which catalyzes the conversion of PGH2 to PGI2, is decreased in the lungs of IPAH patients [16], favoring increased flux of PGH2 towards TXA2.

NO is synthesized from L-arginine through the actions of the nitric oxide synthase (NOS) isoenzymes in concert with multiple cofactors [17]. Among its many effects, NO



generally causes vasodilation while inhibiting pulmonary artery endothelial cell (PAEC) apoptosis, PA smooth muscle cell (PASMC) proliferation, and platelet aggregation all key pathologic features of PAH. Numerous mechanisms contribute to a reduction in bioavailable NO in the setting of PAH, including decreased NOS expression, decreased substrate availability by upregulation of arginases, cofactor oxidation, and rapid scavenging by local reactive oxygen species (ROS) (reviewed in [18]). While endothelial NOS (NOS3, also known as eNOS) expression is decreased in lung sections from PAH patients [19], it is paradoxically increased in plexiform lesions [20]; however, eNOS is unlikely to contribute significantly to NO synthesis in this environment where it probably exists in an uncoupled state and catalyzes the formation of the superoxide radical, promoting oxidative stress and pulmonary vascular remodeling [21].

ET-1 is a peptide hormone primarily synthesized in the endothelium where it is translated as a prepropeptide and undergoes two stages of proteolytic activation to reach its mature form. ET-1 exerts its effect through the actions of two G-protein coupled receptors, ET-A and ET-B, localized on the smooth muscle cells. Expression of ET-1—as well as its associated activating proteases and receptors—is increased in PAH [22], where it directs a program of vasoconstriction and PASMC proliferation (reviewed in [23]).

4.2 Genetics

Over two decades ago, the discovery of causative heterozygous bone morphogenetic protein 2 (BMPR2) mutations within familial cases of PAH [24,25] marked a foundational moment in our understanding of the disease. BMPR2 encodes a membrane-bound type 2 receptor of the transforming growth factor beta (TGF β) receptor superfamily which heterodimerizes with type 1 receptors and, upon ligand binding, canonically transduces cytosolic and transcriptional signals through the "mothers against decapentaplegic" SMAD1/5/9 signaling pathway [26]. It is estimated that BMPR2 mutations account for roughly 75% of HPAH and perhaps as much as 25% of IPAH [27]. Inherited in an autosomal dominant pattern, the penetrance of HPAH due to BMPR2 mutations is estimated to be 14% in males and 42% in females [28], indicating that sex and other factors play a strong role in disease manifestation.

Mutations in other BMPR2-related genes have been linked to PAH albeit at lower frequencies, including type I receptors (ACVRL1, ENG), SMAD family members (SMAD4, SMAD9), and BMPR2 ligands (GDF2 which encodes BMP9) [29–33]. Pathogenic BMPR2 mutations result in BMPR2 haploinsufficiency with a decrease in pulmonary vascular BMPR2 protein expression; notably, BMPR2 expression is also decreased in the pulmonary vasculature of patients with other forms of pulmonary hypertension, suggesting mechanistic parallels between the various subgroups [34]. More recently, rare PAH-associated

mutations have been identified in genes and/or neighboring chromosomal loci without a clear link to BMPR2 signaling, including caveolin 1 (*CAVI*) [35], potassium channel subfamily K, member 3 (*KCNK3*), probable cation-transporting ATPase 13A3 (*ATP13A3*) [36], aquaporin 1 (*AQP1*) [36], SRY-box 17 (*SOX17*) [36,37], and major histocompatibility complex, class II, DP alpha 1 and beta 1 (*HLA-DPA1/HLA-DPB1*) [37]. It is hoped that greater clarity into the functional consequences of these rare disease-associated mutations will bring a fuller picture of the mechanistic underpinnings of PAH.

4.3 BMPR2 Insufficiency

The exact molecular mechanisms that explain the link of BMPR2 haploinsufficiency to vascular remodeling and PAH remain incompletely defined; however, significant progress has been achieved in recent years in understanding this complex paradigm. In vitro studies have demonstrated that under normal circumstances, ligand binding to BMPR2 protects PAECs from apoptosis [38] while suppressing the proliferation of PASMCs [39,40]. Phenotypically, BMPR2-deficient pulmonary artery endothelial cells display increased apoptosis [38], disrupted vasodilator/vasoconstrictor homeostasis [41], endothelial-tomesenchymal transition (EndoMT) [42], and dysregulated metabolism [43–45]—alterations which mirror those seen in cultured PAECs from PAH patients in general. In models of BMPR2 insufficiency, rodents with heterozygous knockout of *Bmpr2* are phenotypically similar to their wild-type counterparts at baseline but develop PAH under inflammatory stress [46,47]. In this respect, animal models suggest the requirement for a second hit as an explanation for the low penetrance of PAH in BMPR2 haploinsufficiency. In PAH subjects, disruption of BMPR2 signaling and the downstream SMAD1/5/9 cascade is accompanied by increased pathologic TGF β signaling through SMAD2/3 [48]. Recently, Yung and colleagues [49] found that the activin and growth and differentiation factor (GDF) ligands activin A, GDF8 and GDF11 were upregulated in the lungs of PAH subjects and activated a SMAD2/3-mediated proproliferative, antiapoptotic phenotype in PAECs and PASMCs. When these effectors were antagonized with a ligand trap, SMAD2/3 signaling was attenuated and experimental PH was reversed, suggesting restoration of balance to amongst BMP versus $TGF\beta/Activin/GDF$ as a therapeutic target in PAH. In addition to canonical SMAD signaling, BMPR2 effector functions may vary depending on context and the engagement of noncanonical pathways (reviewed in [50]) and cell types beyond endothelium and smooth muscle cells, adding further complexity to its pleiotropic roles.

4.4 Acquired & Environmental Factors

Data from the Registry to Evaluate Early and Long-Term Pulmonary Arterial Hypertension Disease Management (REVEAL) suggest that drugs and toxins are respon-



sible for approximately 1 in 20 total PAH cases [8]. Among the earliest drugs associated with PAH was the anorexigen aminorex (2-amino-5-phenyl-2-oxazoline) [51], whose association with an epidemic of PAH in the 1960s led to heightened awareness of PAH in general [52]. Since then, additional stimulant anorexigens—notably fenfluramine/dexfenfluramine [53,54]—as well as recreational amphetamines and other drugs have been linked to the development of PAH [2,55]. The known serotonergic properties of these toxins sparked interest in serotonin (5hydroxytryptamine, 5HT) as a molecular mediator of disease [56]. 5HT promotes smooth muscle cell proliferation and pulmonary vasoconstriction [57,58], and the active metabolite of dexfenfluramine is a potent activator of the 5HT2B receptor [59]. Additionally, genetic deletion or pharmacologic blockade of 5HT1B [60] or 5HT2B [59] is protective against hypoxia-induced rodent PH. In human IPAH, plasma 5HT is increased [61] suggesting a broader role for 5HT beyond anorexigen-induced PAH. Interestingly, 5HT exacerbates hypoxia-induced PH in BMPR2deficient mice while inhibiting BMP signaling via SMAD proteins, suggesting mechanistic overlap with HPAH [62].

The cellular response to hypoxia has long been recognized to play a crucial role in the pathogenesis of PAH. While chronic exposure to hypoxia and high altitude retains a separate designation within Group 3 PH, hypoxic signaling pathways are intimately involved in PAH and other WHO groups [63]. In the pulmonary vasculature, acute hypoxia leads to vasoconstriction through transcriptional reprogramming to promote the synthesis of vasoconstrictors such as ET-1 [64] over vasodilators including NO [65]; with sustained hypoxia, remodeling leads to alterations in pulmonary vascular architecture and pulmonary hypertension [66]. The hypoxia inducible factors (HIFs) are master transcriptional regulators of hypoxic cellular responses composed of an oxygen-sensitive alpha subunit (HIF1 α , $HIF2\alpha$, $HIF3\alpha$) and an oxygen-insensitive beta subunit (aryl hydrocarbon receptor nuclear translocator ARNT1, ARNT2, ARNT3) shared with the aryl hydrocarbon receptor (AhR) [67]. In normoxic conditions, the alpha subunit is marked for ubiquitination and degradation by prolyl hydroxylase domain (PHD) mediated hydroxylation leading to binding of the von Hippel Lindau (VHL) E3 ubiquitin ligase complex and subsequent degradation. In hypoxic conditions, the alpha subunit heterodimerizes with the beta subunit and translocates to the nucleus where it initiates transcriptional events through binding to HIF response elements (HREs). It is increasingly appreciated that HIF complexes can be stabilized by alternative means in normoxic conditions, including the conditions of inflammation, mechanical stretch, and metabolic stress characteristic of PAH (reviewed in [63]). Additionally, a variant in the EPASI gene encoding HIF2 α has been associated with the development of RV failure in cattle living at high altitude, known as Brisket Disease, adding evidence of genetic influences

on aberrant HIF activation in PH [68].

More recently, it has been demonstrated by independent laboratories that the aryl hydrocarbon receptor (AhR), a master regulator of xenobiotic responses which shares a heterodimerization partner with the HIF α subunits, is of critical importance in experimental PH [69,70]. In rodent models of PH, the phenotype triggered by Sugen (SU5416) as a model of PAH was attributed to its activation of AhR rather than inhibition of VEGFR2 as previously accepted [71]. AhR activation provoked inflammation and PH in animal models, while plasma agonistic AhR activity was higher in PAH patients than in healthy controls [70]. If this finding is upheld, it would suggest a role for countless additional environmental xenobiotics in the development of PAH. For example, a recent epidemiological study based in the United Kingdom found that air pollution may be linked to PAH outcomes [72], although the mechanism of this association has not yet been explored.

4.5 Sex Differences

The female predominance of PAH is well-established and extends to nearly all subgroups of the disease [4,8]. While females outnumber males by more than 2:1 in national registries [3], prevalent males have a significantly higher mortality. This observation that females have better outcomes in the context of higher disease burden has been termed the "estrogen paradox" [73]; these findings have spurred investigation of the effects of sex hormonespredominantly the major female estrogen, estradiol (17 β estradiol, E2)—and their metabolites in PAH. As with humans, model organisms also exhibit sexual dimorphism whereby males experience more severe disease than females [74,75], while exogenous estradiol administration prevents and reverses rodent PH [76,77]. Estradiol modulates transcriptional programming through binding to alpha (ER α , also known as nuclear receptor subfamily 3, group A, member 2 [NR3A1]) or beta (ER β , also known as NR3A2) hormone receptors as well as non-genomic effects on binding to G-protein coupled estrogen receptors [78]. In pulmonary vascular cells, estradiol generally yields an antimitogenic, anti-proliferative phenotype in PASMCs while inducing the synthesis of vasodilatory mediators [73].

However, the actions of estradiol are complex and context-dependent across its metabolites and target cell types. The first step in estradiol metabolism is mediated by the cytochrome P450 (CYP450) superfamily of hemecontaining monooxygenases and predominantly involves hydroxylation at the C2 and C4 positions, although hydroxylation at other carbons including C16 also occurs [79]. Broadly speaking, the anti-mitogenic, anti-proliferative 2-estrogens are thought to be protective in PAH, while the proinflammatory, pro-proliferative 16-estrogens are believed to be pathogenic (reviewed in [73]). CYP1B1, which hydroxylates estrogen at the C4 and, to a lesser degree, C2 and C16 positions, is of particular interest as it is highly ex-



pressed in PASMCs isolated from PAH patients [80], and genetic polymorphisms affecting CYP1B1 protein function have been associated with decreased ratios of "good" 2-estrogens metabolite to "bad" 16-estrogen metabolites as well as PAH penetrance in females with *BMPR2* mutations [81]. In rodent models of PAH secondary to serotonergic excess—including after administration of the anorexigen dexfenfluramine implicated in drug-induced PAH—CYP1B1 and 16-estrogen levels are increased, while genetic knockout or pharmacologic inhibition of CYP1B1 prevents disease [82,83]. CYP1A1, meanwhile, is also involved in estrogen metabolism and is upregulated in experimental PAH through activation of AhR, although further work is needed to define its relationship to estrogen signaling in PAH [69].

Upstream of its metabolism, estradiol is synthesized from androgens through the action of aromatase (CYP19A1) [79]. Implicating a pathogenic role for estrogens, a polymorphism in CYP19A1 was associated with increased estradiol levels and the presence of portopulmonary hypertension (PoPH)—a subgroup of PAH—in patients with advanced liver disease, while PoPH was also associated with increased levels of 16-estrogens [84]. Interestingly, roughly one-third of PAH patients in the USbased REVEAL registry were classified as obese at the time of enrollment [85], and visceral adipose tissue is known to be a major site of aromatase expression and estrogen biosynthesis [86]. In leptin-deficient obese mice, which spontaneously develop PH and pulmonary vascular remodeling, these pathophenotypes were attenuated by aromatase inhibition with anastrozole or CYP1B1 inhibition with 2,2',4,6'-tetramethoxystilbene, suggesting a mechanistic role for estrogen in this disease model [87]. In a distinct rodent model of PH, the anti-diabetic drug metformin was shown to reverse PH and vascular remodeling through transcriptional repression of aromatase, again suggesting a therapeutic effect of estrogen inhibition [88]. In a small randomized trial comparing anastrozole to placebo in PAH patients on background therapy, anastrozole was associated with a modest improvement in 6-minute walk distance after 3 months while having no effect on an echocardiographic metric of RV systolic function [89]. Taken together, these findings indicate important influences including estrogen metabolism, cell type, and disease model in defining the effects of estrogens on pulmonary vascular remodeling.

In contrast to its variable effects on the pulmonary vasculature, the beneficial effects of estradiol on RV structure and function are well-established and thought to explain in large part the estrogen paradox [90]. In human cohorts, the estrogen-diminished state of menopause has been associated with onset of CTD-PAH [91], while post-menopausal women receiving hormone replacement therapy have evidence of improved RV systolic function on cardiac imaging [92]. Meanwhile, females have evidence of improved RV adaptation to PAH by invasive [93] and non-invasive [94] methods as compared to males. In animal models of PH, females develop less severe RV hypertrophy (RVH) [74], while estradiol administration attenuates RV remodeling [76,95,96]. Historical studies have demonstrated that estradiol inhibits cardiac fibrosis in models of left ventricular (LV) failure via an ER β -dependent mechanism [97]. Recently, however, experimental models have shown a critical role for ER α in orchestrating adaptive RV remodeling, at least in part through a BMPR2-dependent mechanism [98,99]. The success of pharmacologic manipulation of estrogen signaling in PAH will likely depend on the ability to balance its opposing effects which define the estrogen paradox.

There remain significant gaps in our understanding of PAH during the unique state of pregnancy. Although estrogen levels increase throughout pregnancy [100], PAH in pregnancy poses serious maternal and fetal risks (reviewed in [101]) such that current guidelines recommend against conception [102]. These risks are thought to be mediated by hemodynamic and hemostatic changes of pregnancy, yet the precise molecular mediators and roles of sex hormones are understudied and may hold novel insights into the effects of pregnancy-specific estrogen derivatives.

Finally, in contrast to the pleiotropic effects of female sex hormones, recent evidence has indicated a protective effect of the Y chromosome in experimental hypoxic PH [103,104]. A subsequent study found that the transcription factor SRY, encoded on the Y chromosome, is a direct positive regulator of *BMPR2* expression at the transcriptional level [105], providing an additional mechanistic explanation for sex disparities in PAH.

4.6 Dysregulated Metabolism

The transition from oxidative phosphorylation to aerobic glycolysis in normoxic conditions, known as the Warburg effect (named the Pasteur effect in hypoxia), is a hallmark of PAH [106]. Central to this shift is the inhibition of pyruvate dehydrogenase, which catalyzes conversion of pyruvate to acetyl-CoA allowing progression from glycolysis to the tricarboxylic acid (TCA) cycle. In PAH-relevant pathways including hypoxia and tyrosine kinase signaling [107,108], this intermediate step is blocked via upregulation of the inhibitory enzyme pyruvate dehydrogenase kinase (PDK) thereby shunting pyruvate to glycolysis. In experimental PH, the PDK inhibitor dichloroacetate (DCA) prevents and reverses pulmonary hypertension and causes apoptosis of PASMCs [109-111]. Interestingly, an openlabel study of DCA in IPAH patients on baseline therapy led to variable reductions in mean PA pressure, with lack of response predicted by genotypic variants in key mitochondrial genes [112]. Additional factors, including inflammation and BMPR2 deficiency, have also been suggested to contribute to the Warburg effect in PAH. Recent studies have also shown that anaerobic glycolysis is favored in PAH by alternative splicing of pyruvate kinase muscle (PKM)



isoforms in response to downregulation of microRNA-124 (miR-124), a process linked to BMPR2 deficiency [44,113]. Additional PAH-related mediators, including the inflammatory cytokine TNF- α [114] as well as HIV infection [115] have been proposed to contribute to Warburg physiology, demonstrating the overlap between various pathophysiologic influences in PAH.

As seen in the Warburg effect, the metabolic shift to anaerobic glycolysis facilitates the use of cellular carbons to generate biomass and meet the anabolic demands of rapid proliferation [116]. In a process known as anaplerosis, the TCA cycle intermediate oxaloacetate is replenished to maintain the pool of biosynthetic and bioenergetic precursors either through the actions of pyruvate carboxylase on pyruvate or deamidation of glutamine ("glutaminolysis") by glutaminase [117]. In experimental PAH, glutaminase (GLS1) is upregulated in a yes-associated protein (YAP1)dependent fashion in order to generate macromolecular precursors and sustain proliferation, with inhibitors of these proteins leading to prevention and reversal of rodent PH [118-120]. Interestingly, NO has been shown to promote Warburg-type physiology and glutaminolysis in ovarian cancer cells, although it is unknown whether comparable mechanisms translate to pulmonary vascular cell types [121].

Electron transport is a critical feature of mitochondrial metabolism and requires the presence of evolutionarily ancient iron-sulfur (Fe-S) clusters, bioinorganic prosthetic groups which facilitate cellular redox processes. Fe-S cluster biogenesis requires more than 30 cytosolic and mitochondrial proteins, and synthetic dysfunction attenuates oxidative phosphorylation as well as other critical metabolic and cellular events including DNA repair [122, 123]. Our group previously identified a hypoxia-inducible microRNA, microRNA-210 (miR-210), which translationally represses the Fe-S cluster assembly enzymes ISCU1/2 and is upregulated in PAH [124,125]. Importantly, both forced overexpression of miR-210 or pharmacologic silencing of ISCU promoted experimental PAH [124]. Mutations in Fe-S biogenesis proteins are also associated with several Mendelian disorders, including Friedreich's Ataxia (Frataxin, FXN) and the multiple mitochondrial dysfunction syndromes (MMDS) 1 (NFU1 iron-sulfur cluster scaffold, NFUI) and 2 (BolA family member 3, BOLA3) [126, 127]. PAH is frequently associated with the clinical syndrome of MMDS1, and rats harboring a human NFU1 mutation develop spontaneous PH [128-130]. Additionally, we have shown that deficiency in BOLA3 or FXN causes experimental PH through multiple mechanisms related to Fe-S biology including attenuation of oxidative phosphorylation, accumulation metabolic intermediates, and induction of cellular senescence [131,132]. Collectively, these findings indicate a critical role for Fe-S clusters in the maintenance of metabolic integrity and normal cellular proliferation processes which, when perturbed, contribute to pulmonary

vascular remodeling.

It is increasingly appreciated that pathologic metabolic abnormalities in PAH extend well beyond mitochondrial flux. Recent evidence has pointed to metabolic dysfunction and aberrant insulin signaling and lipid handling in multiple forms of PH, including PAH [133] and with BMPR2 deficiency [134]. Our group and others have shown a critical role for the nuclear receptor peroxisome proliferator-activated receptor gamma (PPAR γ), a master regulator of glucose and fatty acid utilization, in PAH pathogenesis [119,135,136]. Upregulation of the microRNA miR-130/301 in PH family led to translational repression of PPAR γ with cell-type and context-dependent effects on PAEC and PASMC proliferation resulting in pathologic vascular remodeling [119]. PPAR γ has also been identified as a downstream mediator of non-canonical SMAD1/5/9-independent BMP2/BMPR2 signaling where it functions to repress PASMC proliferation [137]. The protective actions of PPAR γ extend to the failing right ventricle, where it exerts therapeutic effects by restoring homeostasis to glucose utilization and fatty acid oxidation [136]. In sum, the diverse and growing list of metabolic perturbations in PAH is reflective of the multidimensional links between metabolism and pulmonary vascular homeostasis.

4.7 Inflammation & Immune Activation

Perivascular inflammation involving macrophages, dendritic cells, T and B lymphocytes, and mast cells is a characteristic feature PAH and correlates with vascular remodeling [138,139], suggesting a causal relationship between the two. Myeloid cell recruitment from bone marrow and blood has been highlighted as a key process in the development of vascular inflammation in PAH [140]. Furthermore, targeting right ventricular inflammation via the NLRP3 inflammasome has recently been described [141]. Correspondingly, circulating cytokine levels, including interleukin- 1β (IL- 1β), interleukin-6 (IL-6), and tumor necrosis factor- α (TNF- α) are increased in PAH and correlate with mortality [142-144]. In mouse models, transgenic overexpression of the proinflammatory cytokine interleukin-6 (IL-6) is sufficient to cause PH, while its deficiency exerts protective effects against disease development [145]. Interestingly, cultured PASMCs treated with silencing RNA to BMPR2 overexpress IL-6, and IL-6 is upregulated in mice harboring a dominant negative Bmpr2 transgene [146], thus linking proinflammatory cytokine expression to BMPR2 deficiency. Inflammatory cytokines can directly induce PAH-relevant phenotypes including proliferation in cultured PASMCs via induction of mitogenic stimuli [147,148]. Moreover, pharmacologic antagonism of IL-1 β and IL-6 receptors is protective in experimental PH [149,150], although clinical trials of IL-6 receptor blockade in PAH have questioned its translatability [151]. Taken together, these points underscore a complex



and unresolved interplay between inflammatory mediators and PAH.

The CTD-PAH subgroup is characterized by systemic immune dysregulation and autoimmunity. Studies of T lymphocyte populations in PAH have suggested that regulatory T cells play an integral role in the maintenance of vascular integrity and protect against the development of PAH [152]. Autoantibodies to endothelial cell antigens have been described in scleroderma-associated PAH and induce apoptosis in cell culture [153], although further study of the exact role of anti-endothelial antibodies and B cell depletion in CTD-PAH requires further study [55].

PAH secondary to infection is also thought to be at least partly related to particular inflammatory signatures. While incompletely understood, the pathogenesis of HIV-PAH is likely multifactorial with contributions from the direct actions of viral proteins, inflammatory mediators, and other factors leading to pulmonary vascular remodeling (reviewed in [154]). Expanding on this concept, Saito and colleagues [155] offered recent evidence that endogenous human retroviruses contribute to PAH pathogenesis, as well. In their study, they showed that transcripts of human endogenous retrovirus K (HERV-K) are upregulated in the lungs of PAH patients and that HERV-K proteins can drive pathogenic vascular changes in rodent models of PH, suggesting that both exogenous and endogenous viruses can modulate inflammatory signatures to promote PAH. Recently, observational data showed that, while the incidence of Coronavirus Disease 2019 (COVID19) in PAH is similar to that of the general population, outcomes are significantly worse [156,157]. There is currently insufficient evidence to suggest a mechanistic link between the causative severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection and PAH, and the poor outcomes may result, at least in part, from the inherent fragility of the PAH patient population.

Schistosomiasis-associated PAH (Sch-PAH), caused by infection with the helminth *Schistosoma mansoni*, is a common cause of PAH globally, though its mechanisms remain ill-defined (reviewed in [158]). In murine models of Sch-PAH, codeletion of the T_H2 cytokines IL-4 and IL-13 protects against the development of experimental PH, which is thought to be related to IL-13-mediated upregulation of TGFb with consequent SMAD2/3 activation and PASMC proliferation [159,160]. Interestingly, IL-13 overexpression induces experiment rodent PH [161], and plasma IL-13 is elevated in scleroderma-associated PAH as compared to scleroderma without PAH [162], suggesting a plausible role for type 2 inflammation in PAH more generally.

As evidenced by the myriad inflammatory mediators associated with PAH, their precise role in disease progression is complex and incompletely understood (reviewed in [163]). Recently, the application of high-throughput techniques has been helpful in defining PAH-associated inflam-

matory and immune signatures [164,165], and longitudinal studies will add additional clarity given the dynamic nature of tissue inflammation [166].

4.8 Epigenetics

Epigenetic modifications describe heritable changes in gene expression that do not alter DNA sequence and mainly comprise DNA methylation, histone modifications, and changes in non-coding RNA expression [167]. As alluded to above, non-coding RNAs (ncRNAs) have been implicated in multiple pathways related to PAH pathogenesis. ncRNAs fall into several categories, including microRNAs (miRNAs), long non-coding RNAs (lncRNAs), and many others [168] where contributions to PAH are only beginning to be appreciated. miRNAs exert their effects through sequence-dependent binding and posttranscriptional repression of target mRNAs in order to orchestrate the downregulation of a wide range of targets [169]. The importance of miRNAs in PAH is well-established, as they have been shown to affect a number of PAH-relevant pathways related to BMPR2, hypoxia, estrogen, PPAR γ , inflammation, and more (reviewed in [170]). lncRNAs are single-stranded RNAs with multiple potential functions, most notably as facilitators of chromatin modification, although additional roles in miRNA antagonism and scaffolding have been suggested [171]. Recent discoveries have shed light onto the involvement of lncRNAs in PAH pathobiology; notably, tyrosine kinase receptor-inducing lncRNA (TYKRIL) was recently shown to be upregulated in PASMCs and pericytes from PAH patients and promotes cellular proliferation by interfering with p53-mediated transcriptional repression of platelet-derived growth factor receptor beta (PDGFR β) [172]. Given the abundant and growing understanding of ncRNAs in PAH pathology, it is expected that some will emerge as promising therapeutic targets in the future.

DNA methylation is generally associated with gene silencing through the covalent addition of methyl groups to cytosine residues—interfering with the binding of cofactors to DNA—and has been well-described in PAH (reviewed in [173]). Recently, RNA methylation has also been described in PAH [174]. Histone modifications take many forms, the best studied of which are acetylation and methylation of histone tails with subsequent implications for gene regulation [175] and for PAH-relevant phenotypes [176]. Histone methylation has been observed in controlling pathogenic processes in PAH [177]. The discovery of histone acetylation signatures in PAH has catalyzed an intent toward pharmacologic targeting. Bromodomain containing protein 4 (BRD4), a member of the bromodomain and extraterminal domain (BET) family of proteins which bind to acetylated histones, modulates cell cycle progression and inflammation, among others, and has been studied extensively in cancer biology [178]. BRD4 is also upregulated in PAH, and its pharmacologic inhibition ameliorates disease in preclinical models [179]. More recently, inobrodib (CCS1477),



a specific bromodomain inhibitor targeting the paralogous histone acetyl transferases p300 and CREB binding protein (CBP) [180], has shown therapeutic efficacy in experimental models [181].

4.9 DNA Damage & Senescence

In addition to epigenetic modifications, the accumulation of DNA damage and impaired DNA repair have been described in PAH [182], including in connection with methamphetamine use [183]. In the setting of accumulated damage, as with aging, cells adopt a senescent phenotype with limited proliferative potential yet apoptosis resistance that is accompanied by a pro-inflammatory senescenceassociated secretory phenotype (SASP) [184]. An emerging hypothesis positions PAEC senescence as a unifying feature of PAH, based partly on its observation in multiple diverse disease models [132,185]. Interestingly, BRD4 inhibition has been shown to modulate the SASP in cancer cells, potentially contributing to its efficacy in preclinical PH [186]. Additional modulators of senescence, so-called "senolytics" which have been extensively studied in cancer, are ripe for further examination in PAH [184].

4.10 Non-Canonical Cell Types & Circulating Bodies

Beyond the well-recognized roles of endothelial, smooth muscle, fibroblast, and immune cells in promoting PAH pathogenesis, there is increasing appreciation for the contributions from non-canonical cell types and circulating bodies in the disease process. It is now recognized that pericytes, subintimal support cells which assist with the maintenance of normal vascular homeostasis (reviewed in [187]), are dysfunctional in PAH and play a role in the pathogenic loss of distal arteriolar beds [188-190]. Recent research has also identified non-canonical functions of well-studied proteins; for example, keratin-1 (KRT1), which is mainly found in hair follicles, has been shown to be regulated by hypoxia and is a negative modulator of PASMC migration and proliferation in experimental PH [191]. Additionally, the role of peripheral nervous system innervation of the pulmonary vasculature is increasingly appreciated [192], and pulmonary artery denervation has shown beneficial signals for the treatment of PAH in uncontrolled studies [193,194]. Stem cell and endothelial progenitor cell biology has been implicated in PAH pathogenesis [195], and endothelial progenitor cell therapy is under clinical study (ClinicalTrials.gov identifier NCT03001414). Mesenchymal stem cells and secreted of circulating microvesicles have displayed therapeutic properties in experimental models. Recent administration of conditioned media from such stem cells resulted in clinical and hemodynamic improvement of severe PAH in a single pediatric patient [196]. Yet, given the extreme pleiotropy of these stem cells and their microvesicle content, identification of the exact causative components of this biology has been challenging. Prior studies have established that miRNAs can be packaged into exosomes to

transmit intercellular signals [197], and recent work in our lab demonstrating endocrine delivery of miR-210 during hypoxia in mice with conjoined circulatory systems [198] provides a plausible framework for the effects of circulating microvesicles on pulmonary vascular biology.

4.11 Mechanobiology in PAH

Mechanical forces contribute to PAH at the cellular, tissue, and organ levels. The effects of deranged flow patterns are best exemplified in the setting of CHD with systemic-to-pulmonary shunting, although they likely contributed to all subgroups of PAH (reviewed in [199]). Mechanoreceptors on the surface of endothelial cells respond to perturbations in flow [200], with physiologic increases in laminar shear stress leading to activation of NO and PGI2 biosynthetic pathways, downregulation of ET-1, and decreased ROS generation. In this manner, the pulmonary vasculature is able to accommodate increased cardiac output. However, supraphysiologic shear stress and cyclic strain, as seen in the setting of left to right shunting, are accompanied by increases in ET-1, thromboxane A2, ROS production, and pathologic vascular remodeling [201,202].

At the tissue level, vascular stiffness is increased in PAH and correlates with survival [203]. Our group and others have shown that vascular stiffness promotes the activation of that mechanoeffectors YAP and transcriptional co-activator with PDZ-binding motif (TAZ) [204]. The resultant signaling cascades lead to miRNA dysregulation [205], metabolic reprogramming and glutaminolysis [120], downregulation of cyclic oxygenase-2 (COX2) and prostaglandin synthesis [206], and other YAP/TAZ-associated disease mechanisms [207]. In addition to fibroblast and smooth muscle function, endothelial cell production of collagen may also contribute to pulmonary vascular stiffening [208].

The organ-level response of the RV to increased afterload drives morbidity and mortality in PAH [209]. In the setting of pressure overload, the RV undergoes adaptive concentric hypertrophy which results in decreased wall stress and increased contracticility allowing RV stroke volume to remain "coupled" with its load. At a certain point, cardiac output can only be maintained through maladaptive eccentric hypertrophy (dilation) and tachycardia, eventually leading to RV-PA "uncoupling" with a drop in cardiac output (reviewed in [210]). The detailed molecular underpinnings of RV failure in PAH are incompletely understood, although fibrosis [211], cytoskeletal and sarcomeric remodeling [212], and altered bioenergetics and glutaminolysis [213] are known to play important roles. Interestingly, recent studies have shown that inhibition of IL6 signaling by pharmacologic blockade of its coreceptor, glycoprotein 130 (gp130, also known as IL6ST), attenuates pathologic RV remodeling without impacting the degree of pulmonary vascular remodeling [141,214]. Clinically, morbidity and



mortality follow RV dysfunction, which may progress regardless of the use of PVR-lowering therapy [215]. The experimental finding of dissociated vascular and ventricular pathologies adds to growing mechanistic rationale for the development of therapeutics specifically targeting the RV

4.12 Systemic Connections to PAH

It is now clear that a multitude of circulating factors contribute to PAH pathogenesis, including neurohormonal mediators of the renin-angiotensin-aldosterone (reviewed in [216]) and sympathetic nervous systems [217], immune cells and cytokines [163], growth factors, and others. It is also understood that primary disorders of solid organs including the liver, LV, and kidney can result in result in PoPH, Group 2 PH, and Group 5 PH, respectively. The observation that circulating BMP9, a hepatically-synthesized BMPR2 ligand, is decreased in PoPH compared to cirrhosis without PH suggests a direct mechanistic link between PoPH and BMPR2 insufficiency [218,219]. The LV relates to PH in large part due to its interdependence with the RV: in LV dysfunction, elevated filling pressures are experienced as increased afterload by the RV. Meanwhile, RV failure in advanced PAH has significant implications for left ventricular (LV) function, as well—leftward bowing of the interventricular septum and decreased RV stroke volume both necessarily result in decreased LV diastolic filling [220]. Chronic kidney disease (CKD) also coexists frequently with PH [221,222]; while a number of mechanisms have been proposed, including hemodynamic factors observed in cardiorenal syndrome, endothelial dysfunction, and arteriovenous shunting [223], the precise events connecting PH and CKD are unknown. Recently, novel links from pulmonary vascular disease to the gut microbiome [224] and the central nervous system [225] have been proposed, reinforcing the idea of PAH as a systemic disease.

5. Presentation & Prognosis

PAH classically presents with nonspecific symptoms of exertional dyspnea and fatigue due to an inadequate increase in cardiac output during activity. Later in the course of the disease, symptoms of RV failure manifest, including leg edema, abdominal distension, early satiety, and near-syncope or syncope [226]. A substantial minority of patients—greater than one third in early registries—will have symptoms of RV failure by the time the diagnosis is established [227]. PAH is associated with significant morbidity and mortality. While limited to 2.8 years prior to the advent of modern therapies [228], median survival is estimated at ~7 years from the time of diagnosis in the current treatment era [229].

Historically, clinical severity and risk of mortality had been categorized primarily by WHO functional class (WHO-FC) [227,230,231], modeled after the New York Heart Association (NYHA) functional classes in heart fail-

ure. In the modern era, the synthesis of information across multiple clinical indices and demographics has yielded a more sophisticated algorithm to prognosticate risk of future morbid or mortal events and thus guide therapy. Specifically, once a diagnosis of PAH has been confirmed, an initial risk assessment is performed to gauge prognosis and to guide therapy. Risk stratification is based upon scoring tools derived from PAH registries, including the USbased REVEAL/REVEAL 2.0 [230,232] and the Europeanbased Swedish Pulmonary Arterial Hypertension Register (SPAHR) [233], Comparative, Prospective Registry of Newly Initiated Therapies for Pulmonary Hypertension (COMPERA) [234], and French Pulmonary Hypertension Registry (FPHR) [235]. While varying in their precise formulations, all tools use a combination of clinical, functional, exercise, hemodynamic, and biochemical inputs to assign a risk category (low, intermediate, or high) and inform initial management strategies [236]. Importantly, current guidelines endorse the use of serial risk assessments at 3-6 months with a goal of maintaining or achieving a lowrisk profile through escalation of therapy [102].

6. In Pursuit of Early Diagnosis

Given the apparent efficacy of early therapy at reducing mortality [229,237], a major focus of PAH management has been on the early diagnosis of disease [238]. The initial symptoms of PAH are nonspecific and require a degree of clinical suspicion in order to pursue a thorough diagnostic evaluation. This workup is burdensome and typically begins with transthoracic echocardiography (TTE). When TTE shows features consistent with pulmonary hypertension, or if uncertainty remains, an invasive hemodynamic assessment with right heart catheterization should be performed. If right-sided hemodynamics are consistent with a diagnosis of PH, additional imaging and serologic studies must be performed to rule out more common causes of PH and establish a diagnosis of PAH [102].

In the REVEAL registry, more than 20% of patients experienced a delay of greater than 2 years between the onset of symptoms and the diagnosis of PAH [239]. As PAH is a progressive disease and more advanced disease is associated with poor outcomes, it is unsurprising that early diagnosis and treatment of PAH is critical to improving survival [237]. Detection of early or pre-symptomatic PAH is difficult, due in part to the very nature of the disease [238]. Recent studies have found that hemodynamic values of mPAP [240] and PVR [241] previously considered as "borderline" in fact portend worse clinical outcomes, prompting alterations to the hemodynamic definitions of clinically significant disease [2]. Even in familial PAH, the low penetrance of disease-associated mutations [242] means that genetic testing alone is insufficient to identify individuals who will develop clinical disease. In addition, physiologic adaptation of the right ventricle to increased afterload may delay clinical symptoms until disease is already present. Further-



more, intrinsic reserves constituting greater than 60% of the pulmonary vascular cross-sectional area allow for normal resting hemodynamics even when pathogenic remodeling is well underway [243].

In the face of these challenges, the development of improved screening and risk-stratification tools has become an area of intense interest. One area that has received prolonged attention but has yet to see widespread adoption is invasive cardiopulmonary exercise testing (iCPET) [244]. In theory, iCPET can serve as a "stress test" for the pulmonary circulation and unmask latent PAH: with the obliteration of pulmonary vascular beds and reduction in vascular distensibility, early PAH would be expected to be accompanied by a disproportionate rise in mPAP during exercise. However, the lack of clear data on what constitutes an abnormally elevated mPAP during exercise, and the difficulty in distinguishing between pre- and post-capillary causes of such elevations, have made it challenging to include exercise PH in current guidelines [2]. In addition, technical requirements and limited access are hurdles to the widespread adoption of iCPET. Nonetheless, interest in iCPET as a screening modality has continued, with some investigators advocating its use in the identification of affected carriers in familial forms of disease [245] or those with known risk factors of PAH.

6.1 Novel Imaging Platforms

While invasive hemodynamic measurements remain the gold standard in assessing the presence and severity of PAH [2], they ultimately reveal phenotypic rather than histopathological insights. As a result, they are a lagging indicator of disease progression and, as mentioned, remain normal until severe pathologic alterations have already taken place. Similarly, widely used imaging modalities such as echocardiography and cardiac magnetic resonance imaging (MRI) are useful in assessing and monitoring phenotypic consequences—including elevations in pulmonary artery pressures and declines in right ventricular systolic function—of pathologic events [246]. 4D flow MRI is a recent advancement which combines threedimensional spatial encoding with three-directional velocities to allow for improved hemodynamic assessment, although its application to PAH is in its early stages [247]. Magnetic resonance spectroscopy (MRS) is an older technology with the ability to provide add molecular quantitation to traditional imaging data, and its experimental use to quantify metabolites in the failing RV suggests that it may have clinical application, as well [248,249]. More recently, novel molecular imaging modalities have been developed which, if translatable to the clinical realm, may be able to identify PAH before disease is clinically evident. ¹²⁹Xe MRI is an emerging pulmonary imaging technique which utilizes the stable xenon isotope ¹²⁹Xe to generate three dimensional maps of lung uptake, interstitial diffusion, and erythrocyte transfer of gaseous or soluble ¹²⁹Xe

[250]. Applied to animal models and two patients with PAH [251,252], ¹²⁹Xe MRI revealed a signature impairment in erythrocyte transfer that was distinct from other studied lung pathologies and preceded the onset of severe disease in rodents. A second emerging technology, positron emission tomography (PET) imaging utilizing a macrophage-targeting tracer identified rodent disease prior to hemodynamic derangements and was able to distinguish PAH from PH-LHD in a small cohort of human subjects [253]. Such molecular imaging techniques have the potential to fundamentally alter the diagnostic evaluation of PAH, shifting the process from procedural assessments of late phenotypic sequelae to noninvasive measurements of early pathologic derangements.

6.2 Biomarkers

A perhaps simpler means of disease detection would be to through the use of a diagnostic blood test. Although biomarkers have seen robust interest, there has been limited success in their application to PAH [254]. B-type natriuretic peptide (BNP) is perhaps the most widely-used biomarker in PAH, a preformed peptide release from the ventricle during periods of increased wall tension that correlates with hemodynamic derangements [255], RV systolic function [256], and mortality [257]. However, it does not distinguish between right- and left-sided heart disease; even after controlling specificity in a high-risk scleroderma population, NT-proBNP performed poorly (56% sensitivity) in the detection of early disease [258]. Therefore, the identification of circulating factors that are both specific to PAH and sensitive to early pathology is essential to the development of clinically useful biomarkers.

In order to improve biomarker specificity, investigators have examined mechanistic biomarkers that may better reflect underlying pathologic processes in retrospective analyses. In one such recent study, the novel biomarker NEDD9 was found to be increased in PAH [208]. In another small study, our group proposed Signal peptide, CUB domain and EGF like domain containing 1 (SCUBE1) as a mechanistic biomarker of PAH based on its differential expression in induced pluripotent stem cell endothelial cells (iPSC-Ecs) derived from affected and carrier BMPR2 mutant heterozygotes. Plasma SCUBE1 levels were able to distinguish PAH from controls and the other more common WSPH Groups 2 and 3 PH [259]. In addition to peptides, microRNAs are known to mediate crucial pathogenic processes in PAH, and circulating disease-relevant microR-NAs have been proposed as biomarkers of early disease (reviewed in [170]). While these assays are far from clinical deployment, it is clear that similar mechanistic approaches will be essential to bringing a useful biomarker into clinical practice.

Of course, PAH, while hemodynamically defined as a single disease, can arise from several distinct etiologies. Nikolic and colleagues recently showed that circulating lev-



els BMP9, a ligand for BMPR2 synthesized in the liver, are significantly reduced in PoPH but not in other forms of PAH [219]. This heterogeneity within various subtypes of PAH suggests that multiple biomarkers or molecular panels may be necessary to provide early and accurate diagnoses.

7. Current & Future Therapies

Pulmonary vasodilators, which predate our current understanding of disease mechanisms, form the backbone of pharmacotherapy in PH. These drugs fall into three categories depending on the targeted pathway—prostanoids, nitric oxide potentiators (phosphodiesterase 5 [PDE5] inhibitors and soluble guanylate cyclase [sGC] activators), and endothelin receptor antagonists (ERA)—and have been extensively reviewed previously [13]. Additionally, highdose calcium channel blockers (CCBs) are indicated in a small subset of PAH patients who respond to invasive vasoreactivity testing [102]. Current recommendations indicate that, in low and intermediate-risk patients, initial combination therapy with a PDE5 inhibitor and ERA is appropriate. Meanwhile, high-risk patients should be started on combination therapy which includes an intravenous prostanoid. On sequential assessment, patients at low risk may be continued on their current regimens, while those at intermediate or high risk should advance to triple combination therapy including a PDE5 inhibitor, ERA, and intravenous prostanoid [236]. The era of vasodilator therapy has been accompanied by improvements in quality and quantity of life [229], although vasodilators do not reverse the pathological features of PAH. When medical therapy fails, lung or heart-lung transplantation is the only option [260], highlighting the need for effective and targeted therapeutics.

7.1 Drugs Targeting BMPR2 Signaling

With greater understanding of disease mechanisms, drug-development efforts have shifted from nonspecific vasodilators to targeted therapeutics. Chief among these targeting strategies are drugs that aim to restore balance between BMPR2 signaling—which is diminished in hereditary and other forms of PAH—and TGF β signaling, which is increased. Sotatercept, initially developed to treat osteoporosis, is a fusion protein consisting of the extracellular domain of human activin receptor type IIA and the Fc domain of IgG1 which serves as a ligand trap for members of the TGF- β superfamily thereby decreasing pro-growth SMAD2/3 signaling to restore balance with the growthinhibiting SMAD1/5/9 signaling diminished by BMPR2 insufficiency [48]. In a recent randomized controlled trial, sotatercept treatment resulted in a significant decrease in pulmonary vascular resistance among patients on maximum tolerated background PAH therapy [261]. Alternatively, the augmentation of BMPR2 can also rebalance the BMPR2/TGF β scale; the BMPR2 ligand BMP9 has been proposed as a means of restoring balanced SMAD signaling in the pulmonary vasculature and has shown efficacy at reversing PAH in preclinical studies [262]. Similarly, the immunosuppressive drug tacrolimus (FK506) used in transplant recipients was identified from a screen of more than 3500 compounds as harboring potent BMPR2 agonism [263]. Tacrolimus prevented and reversed pulmonary hypertension in multiple rodent disease models, and clinical trials are planned [264].

7.2 Repurposing of Cancer Therapies

As illustrated by the application of tacrolimus to PAH, repurposing of existing drugs to the treatment of PAH has emerged as a strategy to overcome the costs of de novo drug development and the inherent difficulty of conducting clinical trials in rare diseases [265]. Cancer therapies have attracted significant interest in PAH given the substantial mechanistic overlap between cancer and PH [266]. As with cancer, tyrosine kinase receptors (TKRs) play crucial roles in transmitting mitogenic signals to the pulmonary arterial smooth muscle resulting in pathogenic hypertrophy and hyperplasia [267]. This knowledge spurred interest in the study of the tyrosine kinase inhibitor (TKI) imatinib, a partially selective inhibitor of the platelet-derived growth factor receptor approved for the treatment of chronic myelogenous leukemia, for the treatment of PAH. While imatinib was efficacious at improving symptoms and functional class—as well as reversing disease in preclinical models—the high rate of severe adverse events, notably subdural hematomas, precluded its clinical use [267,268]. In order to minimize off-target effects, inhaled TKIs have been developed, including aerosolized imatinib (AV-101) and seralutinib (GB002) which are currently in clinical trials for the treatment of patients with PAH on background vasodilator therapy (ClinicalTrials.gov identifiers NCT05036135, NCT04816604) [269,270]. Paradoxically, the TKI dasatinib—and potentially others—has been linked to the development of PAH [271,272]. While the precise mechanisms of these divergent effects are unclear, they may be a consequence of variable TKR specificity profiles, including Src inhibition, as well as other mechanisms [273].

The case of TKIs shows the challenges of predicting cumulative drug effects based on mechanism alone. One strategy to address this concern is to infer net effects based on predictive algorithms. Our group recently analyzed transcriptomic differential dependency networks of a library of cancer drugs [274] to identify compounds leading to the rewiring of PH gene clusters. This approach led to the identification of a bromodomain-containing protein BRD2/4 inhibitor and a piperlongumine-like GSTP1 inhibitor, both of which ameliorated experimental PH [275]. Correspondingly, the BRD4 inhibitor JQ1 has previously been shown to reverse experimental PH in rodent models [179], and the BRD4 inhibitor apabetalone (RVX208) is currently under Phase 2 clinical investigation in PAH (ClinicalTrials.gov identifier NCT04915300) [276,277].



Several additional cancer therapeutics have garnered interest in PAH, including anastrazole and tamoxifen targeting estrogen signaling [89,278]; palbociclib-mediated cyclin-dependent kinase 4/9 (CDK4/9) inhibition [279]; and modulation DNA damage/repair with the poly-ADP ribose polymerase inhibitor olaparib [182], highlighting the overlapping pathophenotypes between PAH and cancer as well as the hope that these drugs can be successfully translated to the clinical management of pulmonary vascular disease.

7.3 Drugs Targeting Metabolic Dysregulation

Similar to cancer, metabolic reprogramming from oxidative phosphorylation to glycolysis under aerobic conditions—known as the Warburg effect—is a core feature of PAH associated with aberrant activation of proliferative pathways and adverse RV remodeling [106]. In addition to the aforementioned trial of DCA in PAH, other metabolic drugs are under investigation. As discussed earlier, cells must maintain adequate biomass to sustain proliferation through anaplerosis. ECM stiffening characteristic of PAH stimulates glutaminolytic generation of TCA carbon intermediates through the activation of a YAP-GLS1 molecular axis to sustain pulmonary vascular cell proliferation through YAP1-dependent upregulation of GLS1 [120,207]. Both the YAP inhibitor verteporfin, used in the treatment of macular degeneration [280], and GLS1 inhibitor CB-839 ameliorated cellular proliferation and PH in multiple rodent and primate disease models [120]. Given the ubiquitous expression of YAP1 and GLS1 and in order to minimize systemic toxicities, an inhaled delivery system was developed that a showed a synergistic benefit of combined verteporfin and CB-839 therapy in the treatment of experimental PH [118], establishing these drugs and drug targets, singly or in combination, as promising candidates for further development.

The distressed right ventricle also undergoes metabolic rewiring in advanced PAH whereby the normal balance between glucose and fatty acid utilization, established through substrate competition in a process known as the Randle Cycle, is disrupted in favor of increased fatty acid oxidation [213,281]. By inhibiting fatty acid oxidation, it has been shown that fatty acid oxidase (FAO) inhibitors can shift metabolic substrates toward glucose oxidation and thereby improve right ventricular function [106,282]. Ranolazine and trimetazidine, two FAO inhibitors used clinically to treat refractory angina pectoris [283], increased RV cardiomyocyte glucose oxidation, reversed RV hypertrophy, and improved exercise capacity in a PA-banding model of RV pressure-overload failure [213]. In independent small human pilot studies, ranolazine was found to improve various clinical aspects of right ventricular function and size in PAH [284,285]. Trimetazidine is likewise the subject of active clinical trials investigating its impact on RV function and metabolism in

PAH.

The repurposing of medications used in diabetes mellitus, specifically the PPAR γ agonist thiazolidinediones (TZDs) and the AMP-activated protein kinase (AMPK) stimulator metformin, has also been of interest in PAH, in part based on findings that insulin resistance is common in the disease [133,135,286]. However, the beneficial mechanisms of these medications in PAH are believed to extend beyond their antihyperglycemic effects. PPAR γ is a transcriptional regulator of key enzymes involving glucose and fatty acid utilization which is suppressed in experimental PH and linked to BMPR2 signaling [137,287]. Pharmacologic activation of PPAR γ with the TZDs rosiglitazone or pioglitazone has been consistently shown to prevent and reverse PH in preclinical models [119,135,136]. In light of the beneficial effects of FAO inhibitors on RV performance, it is counterintuitive that TZDs have shown a beneficial effect on RV function attributed to increased fatty acid utilization [136], perhaps best explained by the restoration of glucose/fatty acid homeostasis rather than an intrinsic preference for a particular fuel source. Despite strong evidence of benefit in animal models, concerns about the cardiac risk profile of TZDs—namely, their association with heart failure exacerbations [288,289]—have thus far prevented their advancement to clinical trials in PAH. Metformin, meanwhile, a well-tolerated first-line anti-diabetic agent, ameliorates vascular cell proliferation and RV dysfunction in multiple animal models as well as in a small human cohort [290,291]. Metformin has also shown to be therapeutic in preclinical models of Group 2 PH, and debate persists over whether the observed benefits of the drug are limited to metabolic syndrome-associated diastolic heart failure or truly extend to Group 1 PAH [292]. In fact, momentum appears to be shifting away from the study of metformin and TZDs. However, given the preclinical successes of the newer antihyperglycemic agents of the sodium glucose cotransporter-2 (SGLT2) inhibitor [293] and glucagon-like peptide-1 (GLP1) receptor agonist [294] on the treatment of experimental PH and in human trials of heart failure with PH in general, it is expected that these drugs will soon advance to clinical trials in PAH.

7.4 Drugs Targeting Inflammation & Immunity

As discussed, inflammatory factors and immune mediators are tightly linked to the signature pathogenic changes in PAH [163]. They have received attention as potential therapeutic targets but with less robust results. IL-6, a central inflammatory cytokine produced by vascular and non-vascular cells, is quantitatively associated with PAH outcomes [295], and forced overexpression of its receptor IL6R causes vascular remodeling in animal models of PH [150]. Tocilizumab, a humanized monoclonal antibody targeting IL6R and approved for use in certain diseases such as cytokine release syndrome, has shown efficacy at reversing disease pathology in preclinical mod-



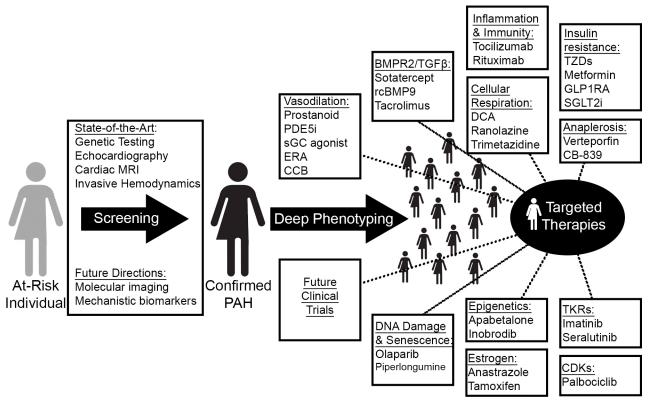


Fig. 2. Precision medicine and novel treatment paradigms in PAH. Guided by improved diagnostic technologies and omics-level deep phenotyping, therapeutic targeting of novel PAH-relevant processes will match a potentially new and molecularly-guided catalog of disease clusters to tailored regimens. MRI, magnetic resonance imaging; PDE5i, phosphodiesterase 5 inhibitor; sGC, soluble guany-late cyclase; ERA, endothelin receptor antagonist; CCB, calcium channel blocker; rcBMP9, recombinant bone morphogenic protein 9; GLP1RA, glucagon-like peptide 1 receptor agonist; SGLT2i, sodium-glucose cotransporter-2 inhibitor; TKRs, tyrosine kinase receptors; CDKs, cyclin-dependent kinases.

els. However, human data have so far been less compelling [296], with a small 6-month phase 2 study showing a decrease in serum inflammatory markers but no change in pulmonary vascular resistance or functional outcomes [151]. Interestingly, a modest reduction in PVR was noted in four of six patients with CTD-PAH which, interpreted cautiously, may suggest that particular subsets will respond favorably to tocilizumab therapy. In a similar fashion, a small randomized-controlled pilot study of B-cell depletion therapy in SSc-PAH produced mixed results, with low levels of rheumatoid factor (RF), IL-12, and IL-17 predictive of improvements in 6-minute walk distance after rituximab therapy [297]. Collectively, these results indicate that enhanced strategies to align patients with individualized anti-inflammatory regimens may improve therapeutic responses.

8. Precision Medicine (Fig. 2)

PAH is a heterogeneous disorder with a multitude of causes as outlined in this review. It is already well-established that subsets of PAH patients—notably those with PVOD/PCH [10]—may not respond favorably to existing vasodilator therapies. As the pharmacologic arma-

mentarium of PAH expands, it is unlikely that all patients will derive equal benefit from targeted therapies. For example, it has already been suggested that individuals with CTD-PAH may be more likely to benefit from antiinflammatory biologics [151], while polymorphisms in certain endothelin-related genes may predict the clinical response to ERAs [298]. Hence, matching the appropriate therapy to the proper patient will become paramount, particularly if more drugs are to be tested appropriately in the limited global number of PAH patients available for recruitment. The National Research Council defines precision medicine as the "tailoring of medical treatment to the individual characteristics of each patient" [299]. Given the diversity of pathologic insults resulting in PAH, it is reasonable to expect that individualized care will yield benefits in patient outcomes.

One could consider an early observation in PAH therapy as an example of precision medicine before it was known as such. It has long been recognized that calcium channel blockers (CCBs) cause an acute vasodilator response in a small subset (less than 10%) of patients with IPAH [300]. In clinical studies, responders have been observed in idiopathic, heritable, and anorexigen-induced



PAH [301,302] and are identified by an acute vasodilator response to nitric oxide, epoprostenol, or, less commonly, adenosine during invasive hemodynamic testing [302]. When treated with long-term CCB therapy, such patients have markedly improved survival compared to non-responders [302,303]. More recently, transcriptomic signatures in peripheral blood samples have shown the ability to differentiate vasoreactive and non-vasoreactive patients with high sensitivity and specificity [304], suggesting a unique molecular phenotype of CCB responders. The distinct clinical and molecular profile of CCB responders led to their inclusion as a separate subset of PAH in the most recent clinical classification guidelines [2]. One goal of precision medicine is to identify the contours of additional subgroups so that they may be targeted with specific therapies.

Early attempts to apply deep omics-level phenotyping to PAH have already begun, including genomic [37], transcriptomic [305], proteomic [306], metabolic [307] and immune [164] profiling of PAH subjects. As a proof of concept, Sweatt and colleagues [164] recently utilized a machine learning approach to identify 4 immune clusters in PAH based on cross-sectional levels of 48 circulating cytokines, chemokines, and growth factors. Despite the inclusion of numerous PAH subgroups, the identified clusters did not correlate with clinical classifications but were strongly predictive of survival. Hence, the these clusters may represent a surrogate of disease severity rather than distinct molecular phenotypes, a possibility that will be addressed by future studies with longitudinal data. Such an effort is currently underway—the Pulmonary Vascular Disease Phenomics Program (PVDOMICS)—that seeks to redefine PH subgroups in place since 1998 based on clusters identified through deep phenotyping [308].

With vast quantities of population and patient-specific information spanning the molecular, genomic, radiographic, demographic, and clinical realms, novel computational methods employing multiscale modeling and machine learning will be required to integrate these data into clinically meaningful tools. If employed successfully, such algorithms have the potential to provide improved diagnostic and risk assessment platforms, inform research directions and drug development, and guide patients toward tailored therapies and clinical trials.

9. Conclusions

The past 30 years have brought multiple gains to the management and prognosis of PAH. However, the clinical application of fundamental discoveries and technological advances developed in this time frame promises to accelerate this trajectory. Pulmonary hypertension is a field where basic science and clinical care are rapidly evolving together, and it will benefit our patients to have clinicians who are well-versed in the two. With improved diagnostic capabilities and expanded treatment options tailored to well-defined molecular phenotypes, the future of precision PAH manage-

ment is promising.

Author Contributions

NJK and SYC wrote the manuscript, contributed to editorial changes in the manuscript, and approved the final manuscript.

Ethics Approval and Consent to Participate

Not applicable.

Acknowledgment

Not applicable.

Funding

This work was supported by NIH grants T32 HL129964 (N.J.K.), R01 HL124021 (S.Y.C.), and R01 HL 122596 (S.Y.C.) as well as by the AHA grant 18EIA33900027 (S.Y.C.) and the McKamish Family Foundation, the Hemophilia Center of Western Pennsylvania, and the Institute for Transfusion Medicine (N.J.K.).

Conflict of Interest

S.Y.C. has served as a consultant for Acceleron Pharma and United Therapeutics. S.Y.C. is a director, officer, and shareholder in Synhale Therapeutics. S.Y.C. has held research grants from Actelion, Bayer, and Pfizer. S.Y.C. has filed patent applications regarding the targeting of metabolism in pulmonary hypertension.

References

- [1] World Health Organization. Primary Pulmonary Hypertension: Executive Summary. In Rich S (ed.) World Symposium - Primary Pulmonary Hypertension. Evian: France. 1998.
- [2] Simonneau G, Montani D, Celermajer DS, Denton CP, Gatzoulis MA, Krowka M, et al. Haemodynamic definitions and updated clinical classification of pulmonary hypertension. European Respiratory Journal. 2019; 53: 1801913.
- [3] McGoon MD, Benza RL, Escribano-Subias P, Jiang X, Miller DP, Peacock AJ, et al. Pulmonary Arterial Hypertension: epidemiology and registries. Journal of the American College of Cardiology. 2013; 62: D51–D59.
- [4] Humbert M, Sitbon O, Chaouat A, Bertocchi M, Habib G, Gressin V, *et al.* Pulmonary Arterial Hypertension in France: results from a national registry. American Journal of Respiratory and Critical Care Medicine. 2006; 173: 1023–1030.
- [5] Wijeratne DT, Lajkosz K, Brogly SB, Lougheed MD, Jiang L, Housin A, et al. Increasing Incidence and Prevalence of World Health Organization Groups 1 to 4 Pulmonary Hypertension: A Population-Based Cohort Study in Ontario, Canada. Circulation: Cardiovascular Quality and Outcomes. 2018; 11: e003973.
- [6] Strange G, Playford D, Stewart S, Deague JA, Nelson H, Kent A, et al. Pulmonary hypertension: prevalence and mortality in the Armadale echocardiography cohort. Heart. 2012; 98: 1805– 1811
- [7] Tuder RM, Marecki JC, Richter A, Fijalkowska I, Flores S. Pathology of Pulmonary Hypertension. Clinics in Chest Medicine. 2007; 28: 23–42.
- [8] Badesch DB, Raskob GE, Elliott CG, Krichman AM, Farber HW, Frost AE, et al. Pulmonary Arterial Hypertension: baseline



- characteristics from the REVEAL Registry. Chest. 2010; 137: 376-387.
- [9] Morrell NW, Aldred MA, Chung WK, Elliott CG, Nichols WC, Soubrier F, et al. Genetics and genomics of pulmonary arterial hypertension. European Respiratory Journal. 2019; 53: 1801899.
- [10] Montani D, Lau EM, Dorfmüller P, Girerd B, Jaïs X, Savale L, et al. Pulmonary veno-occlusive disease. European Respiratory Journal. 2016; 47: 1518–1534.
- [11] Mandell E, Kinsella JP, Abman SH. Persistent pulmonary hypertension of the newborn. Pediatric Pulmonology. 2021; 56: 661–669.
- [12] Humbert M, Guignabert C, Bonnet S, Dorfmüller P, Klinger JR, Nicolls MR, et al. Pathology and pathobiology of pulmonary hypertension: state of the art and research perspectives. European Respiratory Journal. 2019; 53: 1801887.
- [13] Bisserier M, Pradhan N, Hadri L. Current and emerging therapeutic approaches to pulmonary hypertension. Reviews in Cardiovascular Medicine. 2020; 21: 163–179.
- [14] Gerber JG, Voelkel N, Nies AS, McMurtry IF, Reeves JT. Moderation of hypoxic vasoconstriction by infused arachidonic acid: role of PGI2. Journal of Applied Physiology: Respiratory, Environmental and Exercise Physiology. 1980; 49: 107–112.
- [15] Christman BW, McPherson CD, Newman JH, King GA, Bernard GR, Groves BM, et al. An Imbalance between the Excretion of Thromboxane and Prostacyclin Metabolites in Pulmonary Hypertension. New England Journal of Medicine. 1992; 327: 70–75.
- [16] Tuder R, Cool C, Geraci M, Wang J, Abman S, Wright L, et al. Prostacyclin Synthase Expression is Decreased in Lungs from Patients with Severe Pulmonary Hypertension. American Journal of Respiratory and Critical Care Medicine. 1999; 159: 1925– 1932.
- [17] Alderton WK, Cooper CE, Knowles RG. Nitric oxide synthases: structure, function and inhibition. Biochemical Journal. 2001; 357: 593–615.
- [18] Klinger JR, Abman SH, Gladwin MT. Nitric Oxide Deficiency and Endothelial Dysfunction in Pulmonary Arterial Hypertension. American Journal of Respiratory and Critical Care Medicine. 2013; 188: 639–646.
- [19] Giaid A, Saleh D. Reduced Expression of Endothelial Nitric Oxide Synthase in the Lungs of Patients with Pulmonary Hypertension. New England Journal of Medicine. 1995; 333: 214–221.
- [20] Mason NA, Springall DR, Burke M, Pollock J, Mikhail G, Yacoub MH, *et al.* High expression of endothelial nitric oxide synthase in plexiform lesions of pulmonary hypertension. The Journal of Pathology. 1998; 185: 313–318.
- [21] Alp NJ, Channon KM. Regulation of Endothelial Nitric Oxide Synthase by Tetrahydrobiopterin in Vascular Disease. Arteriosclerosis, Thrombosis, and Vascular Biology. 2004; 24: 413– 420.
- [22] Giaid A, Yanagisawa M, Langleben D, Michel RP, Levy R, Shennib H, et al. Expression of Endothelin-1 in the Lungs of Patients with Pulmonary Hypertension. New England Journal of Medicine. 1993; 328: 1732–1739.
- [23] Galie N. The endothelin system in pulmonary arterial hypertension. Cardiovascular Research. 2004; 61: 227–237.
- [24] Deng Z, Morse JH, Slager SL, Cuervo N, Moore KJ, Venetos G, et al. Familial Primary Pulmonary Hypertension (Gene PPH1) is Caused by Mutations in the Bone Morphogenetic Protein Receptor–II Gene. The American Journal of Human Genetics. 2000; 67: 737–744.
- [25] International PPHC, Lane KB, Machado RD, Pauciulo MW, Thomson JR, Phillips JA, et al. Heterozygous germline mutations in BMPR2, encoding a TGF-beta receptor, cause familial primary pulmonary hypertension. Nature Genetics. 2000; 26:

- 81_84
- [26] Moustakas A, Souchelnytskyi S, Heldin CH. Smad regulation in TGF-beta signal transduction. Journal of Cell Science. 2001; 114: 4359–4369.
- [27] Soubrier F, Chung WK, Machado R, Grünig E, Aldred M, Geraci M, et al. Genetics and Genomics of Pulmonary Arterial Hypertension. Journal of the American College of Cardiology. 2013; 62: D13–D21.
- [28] Cogan J, Austin E, Hedges L, Womack B, West J, Loyd J, et al. Role of BMPR2 alternative splicing in heritable pulmonary arterial hypertension penetrance. Circulation. 2012; 126: 1907–1916
- [29] McAllister KA, Grogg KM, Johnson DW, Gallione CJ, Baldwin MA, Jackson CE, et al. Endoglin, a TGF-beta binding protein of endothelial cells, is the gene for hereditary haemorrhagic telangiectasia type 1. Nature Genetics. 1994; 8: 345–351.
- [30] Johnson DW, Berg JN, Baldwin MA, Gallione CJ, Marondel I, Yoon SJ, et al. Mutations in the activin receptor–like kinase 1 gene in hereditary haemorrhagic telangiectasia type 2. Nature Genetics. 1996; 13: 189–195.
- [31] Nasim MT, Ogo T, Ahmed M, Randall R, Chowdhury HM, Snape KM, et al. Molecular genetic characterization of SMAD signaling molecules in pulmonary arterial hypertension. Human Mutation. 2011; 32: 1385–1389.
- [32] Shintani M, Yagi H, Nakayama T, Saji T, Matsuoka R. A new nonsense mutation of SMAD8 associated with pulmonary arterial hypertension. Journal of Medical Genetics. 2009; 46: 331– 337.
- [33] Wang XJ, Lian TY, Jiang X, Liu SF, Li SQ, Jiang R, *et al.* Germline *BMP9* mutation causes idiopathic pulmonary arterial hypertension. The European Respiratory Journal. 2019; 53: 1801609
- [34] Atkinson C, Stewart S, Upton PD, Machado R, Thomson JR, Trembath RC, et al. Primary Pulmonary Hypertension is Associated with Reduced Pulmonary Vascular Expression of Type II Bone Morphogenetic Protein Receptor. Circulation. 2002; 105: 1672–1678.
- [35] Austin ED, Ma L, LeDuc C, Berman Rosenzweig E, Borczuk A, Phillips JA, *et al.* Whole Exome Sequencing to Identify a Novel Gene (Caveolin-1) Associated with Human Pulmonary Arterial Hypertension. Circulation: Cardiovascular Genetics. 2012; 5: 336–343.
- [36] Graf S, Haimel M, Bleda M, Hadinnapola C, Southgate L, Li W, et al. Identification of rare sequence variation underlying heritable pulmonary arterial hypertension. Nature Communications. 2018; 9: 1416.
- [37] Rhodes CJ, Batai K, Bleda M, Haimel M, Southgate L, Germain M, et al. Genetic determinants of risk in pulmonary arterial hypertension: international genome-wide association studies and meta-analysis. The Lancet. Respiratory Medicine. 2019; 7: 227–238.
- [38] Teichert-Kuliszewska K, Kutryk MJB, Kuliszewski MA, Karoubi G, Courtman DW, Zucco L, et al. Bone Morphogenetic Protein Receptor-2 Signaling Promotes Pulmonary Arterial Endothelial Cell Survival: implications for loss-of-function mutations in the pathogenesis of pulmonary hypertension. Circulation Research. 2006; 98: 209–217.
- [39] Zhang S, Fantozzi I, Tigno DD, Yi ES, Platoshyn O, Thistleth-waite PA, et al. Bone morphogenetic proteins induce apoptosis in human pulmonary vascular smooth muscle cells. American Journal of Physiology-Lung Cellular and Molecular Physiology. 2003; 285: L740–L754.
- [40] Zakrzewicz A, Hecker M, Marsh LM, Kwapiszewska G, Nejman B, Long L, et al. Receptor for Activated C-Kinase 1, a Novel Interaction Partner of Type II Bone Morphogenetic Protein Receptor, Regulates Smooth Muscle Cell Proliferation



- in Pulmonary Arterial Hypertension. Circulation. 2007; 115: 2957-2968
- [41] Gangopahyay A, Oran M, Bauer EM, Wertz JW, Comhair SA, Erzurum SC, et al. Bone Morphogenetic Protein Receptor II is a Novel Mediator of Endothelial Nitric-oxide Synthase Activation. Journal of Biological Chemistry. 2011; 286: 33134–33140.
- [42] Ranchoux B, Antigny F, Rucker-Martin C, Hautefort A, Péchoux C, Bogaard HJ, *et al.* Endothelial-to-Mesenchymal Transition in Pulmonary Hypertension. Circulation. 2015; 131: 1006–1018.
- [43] Diebold I, Hennigs J, Miyagawa K, Li C, Nickel N, Kaschwich M, et al. BMPR2 Preserves Mitochondrial Function and DNA during Reoxygenation to Promote Endothelial Cell Survival and Reverse Pulmonary Hypertension. Cell Metabolism. 2015; 21: 596–608.
- [44] Caruso P, Dunmore BJ, Schlosser K, Schoors S, Dos Santos C, Perez-Iratxeta C, et al. Identification of MicroRNA-124 as a Major Regulator of Enhanced Endothelial Cell Glycolysis in Pulmonary Arterial Hypertension via PTBP1 (Polypyrimidine Tract Binding Protein) and Pyruvate Kinase M2. Circulation. 2017; 136: 2451–2467.
- [45] Egnatchik RA, Brittain EL, Shah AT, Fares WH, Ford HJ, Monahan K, *et al.* Dysfunctional BMPR2 signaling drives an abnormal endothelial requirement for glutamine in pulmonary arterial hypertension. Pulmonary Circulation. 2017; 7: 186–199.
- [46] Song Y, Jones JE, Beppu H, Keaney JF, Loscalzo J, Zhang Y. Increased Susceptibility to Pulmonary Hypertension in Heterozygous BMPR2-Mutant Mice. Circulation. 2005; 112: 553–562.
- [47] Tian W, Jiang X, Sung YK, Shuffle E, Wu T, Kao PN, et al. Phenotypically Silent Bone Morphogenetic Protein Receptor 2 Mutations Predispose Rats to Inflammation-Induced Pulmonary Arterial Hypertension by Enhancing the Risk for Neointimal Transformation. Circulation. 2019; 140: 1409–1425.
- [48] Yung LM, Nikolic I, Paskin-Flerlage SD, Pearsall RS, Kumar R, Yu PB. A Selective Transforming Growth Factor-beta Ligand Trap Attenuates Pulmonary Hypertension. American Journal of Respiratory and Critical Care Medicine. 2016; 194: 1140–1151.
- [49] Yung L, Yang P, Joshi S, Augur ZM, Kim SSJ, Bocobo GA, et al. ACTRIIA-Fc rebalances activin/GDF versus BMP signaling in pulmonary hypertension. Science Translational Medicine. 2020; 12: eaar5660.
- [50] Andruska A, Spiekerkoetter E. Consequences of BMPR2 Deficiency in the Pulmonary Vasculature and Beyond: Contributions to Pulmonary Arterial Hypertension. International Journal of Molecular Sciences. 2018; 19: 2499.
- [51] Kay JM, Smith P, Heath D. Aminorex and the pulmonary circulation. Thorax. 1971; 26: 262–270.
- [52] Fishman AP. Primary pulmonary arterial hypertension: a look back. Journal of the American College of Cardiology. 2004; 43: \$2_\$64
- [53] Abenhaim L, Moride Y, Brenot F, Rich S, Benichou J, Kurz X, et al. Appetite-Suppressant Drugs and the Risk of Primary Pulmonary Hypertension. International Primary Pulmonary Hypertension Study Group. New England Journal of Medicine. 1996; 335: 609–616.
- [54] Brenot F, Herve P, Petitpretz P, Parent F, Duroux P, Simonneau G. Primary pulmonary hypertension and fenfluramine use. Heart. 1993; 70: 537–541.
- [55] Zamanian RT, Hedlin H, Greuenwald P, Wilson DM, Segal JI, Jorden M, et al. Features and Outcomes of Methamphetamineassociated Pulmonary Arterial Hypertension. American Journal of Respiratory and Critical Care Medicine. 2018; 197: 788–800.
- [56] Angevine LS, Mehendale HM. Effect of chlorphentermine on the pulmonary disposition of 5-hydroxytryptamine in the isolated perfused rabbit lung. The American Review of Respiratory Disease. 1980; 122: 891–898.
- [57] Morecroft I, Heeley RP, Prentice HM, Kirk A, MacLean MR.

- 5-hydroxytryptamine receptors mediating contraction in human small muscular pulmonary arteries: importance of the 5-HT1B receptor. British Journal of Pharmacology. 1999; 128: 730–734.
- [58] Lee SL, Wang WW, Lanzillo JJ, Fanburg BL. Serotonin produces both hyperplasia and hypertrophy of bovine pulmonary artery smooth muscle cells in culture. American Journal of Physiology-Lung Cellular and Molecular Physiology. 1994; 266: L46–L52.
- [59] Launay JM, Hervé P, Peoc'h K, Tournois C, Callebert J, Nebigil CG, et al. Function of the serotonin 5-hydroxytryptamine 2B receptor in pulmonary hypertension. Nature Medicine. 2002; 8: 1129–1135.
- [60] Keegan A, Morecroft I, Smillie D, Hicks MN, MacLean MR. Contribution of the 5-HT(1B) receptor to hypoxia-induced pulmonary hypertension: converging evidence using 5-HT(1B)receptor knockout mice and the 5-HT(1B/1D)-receptor antagonist GR127935. Circulation Research. 2001; 89: 1231–1239.
- [61] Hervé P, Launay J, Scrobohaci M, Brenot F, Simonneau G, Petitpretz P, et al. Increased plasma serotonin in primary pulmonary hypertension. The American Journal of Medicine. 1995; 99: 249–254
- [62] Long L, MacLean MR, Jeffery TK, Morecroft I, Yang X, Rudarakanchana N, et al. Serotonin increases susceptibility to pulmonary hypertension in BMPR2-deficient mice. Circulation Research. 2006; 98: 818–827.
- [63] Pullamsetti SS, Mamazhakypov A, Weissmann N, Seeger W, Savai R. Hypoxia-inducible factor signaling in pulmonary hypertension. Journal of Clinical Investigation. 2020; 130: 5638– 5651.
- [64] Kourembanas S, Marsden PA, McQuillan LP, Faller DV. Hypoxia induces endothelin gene expression and secretion in cultured human endothelium. Journal of Clinical Investigation. 1991; 88: 1054–1057.
- [65] McQuillan LP, Leung GK, Marsden PA, Kostyk SK, Kourembanas S. Hypoxia inhibits expression of eNOS via transcriptional and posttranscriptional mechanisms. American Journal of Physiology-Heart and Circulatory Physiology. 1994; 267: H1921–H1927.
- [66] Dunham-Snary KJ, Wu D, Sykes EA, Thakrar A, Parlow LRG, Mewburn JD, et al. Hypoxic Pulmonary Vasoconstriction: From Molecular Mechanisms to Medicine. Chest. 2017; 151: 181– 192.
- [67] Prabhakar NR, Semenza GL. Adaptive and Maladaptive Cardiorespiratory Responses to Continuous and Intermittent Hypoxia Mediated by Hypoxia-Inducible Factors 1 and 2. Physiological Reviews. 2012; 92: 967–1003.
- [68] Newman JH, Holt TN, Cogan JD, Womack B, Phillips JA, Li C, et al. Increased prevalence of EPAS1 variant in cattle with high-altitude pulmonary hypertension. Nature Communications. 2015; 6: 6863.
- [69] Dean A, Gregorc T, Docherty CK, Harvey KY, Nilsen M, Morrell NW, et al. Role of the Aryl Hydrocarbon Receptor in Sugen 5416—induced Experimental Pulmonary Hypertension. American Journal of Respiratory Cell and Molecular Biology. 2018; 58: 320–330.
- [70] Masaki T, Okazawa M, Asano R, Inagaki T, Ishibashi T, Yamagishi A, et al. Aryl hydrocarbon receptor is essential for the pathogenesis of pulmonary arterial hypertension. Proceedings of the National Academy of Sciences of the United States of America. 2021; 118: e2023899118.
- [71] Taraseviciene-Stewart L, Kasahara Y, Alger L, Hirth P, Mahon GM, Waltenberger J, et al. Inhibition of the VEGF receptor 2 combined with chronic hypoxia causes cell death-dependent pulmonary endothelial cell proliferation and severe pulmonary hypertension. The FASEB Journal. 2001; 15: 427–438.
- [72] Sofianopoulou E, Kaptoge S, Gräf S, Hadinnapola C, Treacy



- CM, Church C, *et al.* Traffic exposures, air pollution and outcomes in pulmonary arterial hypertension: a UK cohort study analysis. European Respiratory Journal. 2019; 53: 1801429.
- [73] Tofovic SP. Estrogens and Development of Pulmonary Hypertension: Interaction of Estradiol Metabolism and Pulmonary Vascular Disease. Journal of Cardiovascular Pharmacology. 2010; 56: 696–708.
- [74] Rabinovitch M, Gamble WJ, Miettinen OS, Reid L. Age and sex influence on pulmonary hypertension of chronic hypoxia and on recovery. American Journal of Physiology-Heart and Circulatory Physiology. 1981; 240: H62–H72.
- [75] McMurtry IF, Frith CH, Will DH. Cardiopulmonary responses of male and female swine to simulated high altitude. Journal of Applied Physiology. 1973; 35: 459–462.
- [76] Farhat MY, Chen M, Bhatti T, Iqbal A, Cathapermal S, Ramwell PW. Protection by oestradiol against the development of cardiovascular changes associated with monocrotaline pulmonary hypertension in rats. British Journal of Pharmacology. 1993; 110: 719–723.
- [77] Umar S, Iorga A, Matori H, Nadadur RD, Li J, Maltese F, et al. Estrogen Rescues Preexisting Severe Pulmonary Hypertension in Rats. American Journal of Respiratory and Critical Care Medicine. 2011; 184: 715–723.
- [78] Mendelsohn ME. Genomic and nongenomic effects of estrogen in the vasculature. The American Journal of Cardiology. 2002; 90: F3–F6.
- [79] Tsuchiya Y, Nakajima M, Yokoi T. Cytochrome P450-mediated metabolism of estrogens and its regulation in human. Cancer Letters. 2005; 227: 115–124.
- [80] White K, Loughlin L, Maqbool Z, Nilsen M, McClure J, Dempsie Y, et al. Serotonin transporter, sex, and hypoxia: microarray analysis in the pulmonary arteries of mice identifies genes with relevance to human PAH. Physiological Genomics. 2011; 43: 417–437
- [81] Austin ED, Cogan JD, West JD, Hedges LK, Hamid R, Dawson EP, et al. Alterations in oestrogen metabolism: implications for higher penetrance of familial pulmonary arterial hypertension in females. European Respiratory Journal. 2009; 34: 1093–1099.
- [82] Dempsie Y, MacRitchie NA, White K, Morecroft I, Wright AF, Nilsen M, et al. Dexfenfluramine and the oestrogenmetabolizing enzyme CYP1B1 in the development of pulmonary arterial hypertension. Cardiovascular Research. 2013; 99: 24–34
- [83] Johansen AKZ, Dean A, Morecroft I, Hood K, Nilsen M, Loughlin L, et al. The Serotonin Transporter Promotes a Pathological Estrogen Metabolic Pathway in Pulmonary Hypertension via Cytochrome P450 1B1. Pulmonary Circulation. 2016; 6: 82–92.
- [84] Al-Naamani N, Krowka MJ, Forde KA, Krok KL, Feng R, Heresi GA, *et al.* Estrogen Signaling and Portopulmonary Hypertension: the Pulmonary Vascular Complications of Liver Disease Study (PVCLD2). Hepatology. 2021; 73: 726–737.
- [85] Farber HW, Miller DP, Poms AD, Badesch DB, Frost AE, Rouzic EM, et al. Five-Year Outcomes of Patients Enrolled in the REVEAL Registry. Chest. 2015; 148: 1043–1054.
- [86] Bhardwaj P, Au CC, Benito-Martin A, Ladumor H, Oshchep-kova S, Moges R, et al. Estrogens and breast cancer: Mechanisms involved in obesity-related development, growth and progression. The Journal of Steroid Biochemistry and Molecular Biology. 2019; 189: 161–170.
- [87] Mair KM, Harvey KY, Henry AD, Hillyard DZ, Nilsen M, MacLean MR. Obesity alters oestrogen metabolism and contributes to pulmonary arterial hypertension. European Respiratory Journal. 2019; 53: 1801524.
- [88] Dean A, Nilsen M, Loughlin L, Salt IP, MacLean MR. Metformin Reverses Development of Pulmonary Hypertension via Aromatase Inhibition. Hypertension. 2016; 68: 446–454.

- [89] Kawut SM, Archer-Chicko CL, DeMichele A, Fritz JS, Klinger JR, Ky B, et al. Anastrozole in Pulmonary Arterial Hypertension. a Randomized, Double-Blind, Placebo-controlled Trial. American Journal of Respiratory and Critical Care Medicine. 2017; 195; 360–368.
- [90] Jacobs W, van de Veerdonk MC, Trip P, de Man F, Heymans MW, Marcus JT, et al. The Right Ventricle Explains Sex Differences in Survival in Idiopathic Pulmonary Arterial Hypertension. Chest. 2014; 145: 1230–1236.
- [91] Scorza R, Caronni M, Bazzi S, Nador F, Beretta L, Antonioli R, et al. Post-Menopause is the Main Risk Factor for Developing Isolated Pulmonary Hypertension in Systemic Sclerosis. Annals of the New York Academy of Sciences. 2002; 966: 238–246.
- [92] Ventetuolo CE, Ouyang P, Bluemke DA, Tandri H, Barr RG, Bagiella E, et al. Sex Hormones are Associated with Right Ventricular Structure and Function: The MESA-right ventricle study. American Journal of Respiratory and Critical Care Medicine. 2011; 183: 659–667.
- [93] Tello K, Richter MJ, Yogeswaran A, Ghofrani HA, Naeije R, Vanderpool R, et al. Sex Differences in Right Ventricular– Pulmonary Arterial Coupling in Pulmonary Arterial Hypertension. American Journal of Respiratory and Critical Care Medicine. 2020; 202: 1042–1046.
- [94] Swift AJ, Capener D, Hammerton C, Thomas SM, Elliot C, Condliffe R, et al. Right ventricular sex differences in patients with idiopathic pulmonary arterial hypertension characterised by magnetic resonance imaging: pair-matched case controlled study. PLoS ONE. 2015; 10: e0127415.
- [95] Ahn BH, Park HK, Cho HG, Lee HA, Lee YM, Yang EK, et al. Estrogen and Enalapril attenuate the Development of Right Ventricular Hypertrophy induced by Monocrotaline in Ovariectomized Rats. Journal of Korean Medical Science. 2003; 18: 641–648.
- [96] Frump AL, Goss KN, Vayl A, Albrecht M, Fisher A, Tursunova R, et al. Estradiol improves right ventricular function in rats with severe angioproliferative pulmonary hypertension: effects of endogenous and exogenous sex hormones. American Journal of Physiology-Lung Cellular and Molecular Physiology. 2015; 308: L873–L890.
- [97] Pedram A, Razandi M, O'Mahony F, Lubahn D, Levin ER. Estrogen receptor-beta prevents cardiac fibrosis. Molecular Endocrinology. 2010; 24: 2152–2165.
- [98] Frump AL, Albrecht M, Yakubov B, Breuils-Bonnet S, Nadeau V, Tremblay E, *et al.* 17beta-Estradiol and estrogen receptor alpha protect right ventricular function in pulmonary hypertension via BMPR2 and apelin. The Journal of Clinical Investigation. 2021; 131: e129433.
- [99] Cheng TC, Philip JL, Tabima DM, Kumari S, Yakubov B, Frump AL, et al. Estrogen receptor-alpha prevents right ventricular diastolic dysfunction and fibrosis in female rats. American Journal of Physiology. Heart and Circulatory Physiolog. 2020; 319: H1459–H1473.
- [100] Schock H, Zeleniuch-Jacquotte A, Lundin E, Grankvist K, Lakso H, Idahl A, et al. Hormone concentrations throughout uncomplicated pregnancies: a longitudinal study. BMC Pregnancy and Childbirth. 2016; 16: 146.
- [101] Olsson KM, Channick R. Pregnancy in pulmonary arterial hypertension. European Respiratory Review. 2016; 25: 431–437.
- [102] Galiè N, Humbert M, Vachiery J, Gibbs S, Lang I, Torbicki A, et al. 2015 ESC/ERS Guidelines for the diagnosis and treatment of pulmonary hypertension: The Joint Task Force for the Diagnosis and Treatment of Pulmonary Hypertension of the European Society of Cardiology (ESC) and the European Respiratory Society (ERS): Endorsed by: Association for European Paediatric and Congenital Cardiology (AEPC), International Society for Heart and Lung Transplantation (ISHLT). European Heart



- Journal. 2016; 37: 67-119.
- [103] Umar S, Cunningham CM, Itoh Y, Moazeni S, Vaillancourt M, Sarji S, et al. The Y Chromosome Plays a Protective Role in Experimental Hypoxic Pulmonary Hypertension. American Journal of Respiratory and Critical Care Medicine. 2018; 197: 952– 955.
- [104] Cunningham CM, Li M, Ruffenach G, Doshi M, Aryan L, Hong J, et al. Y-Chromosome Gene, Uty, Protects Against Pulmonary Hypertension by Reducing Proinflammatory Chemokines. American Journal of Respiratory and Critical Care Medicine. 2022; 206: 186–196.
- [105] Yan L, Cogan JD, Hedges LK, Nunley B, Hamid R, Austin ED. The Y Chromosome Regulates BMPR2 Expression via SRY: a Possible Reason "why" Fewer Males Develop Pulmonary Arterial Hypertension. American Journal of Respiratory and Critical Care Medicine. 2018; 198: 1581–1583.
- [106] Chan SY, Rubin LJ. Metabolic dysfunction in pulmonary hypertension: from basic science to clinical practice. European Respiratory Review. 2017; 26: 170094.
- [107] Kim J, Tchernyshyov I, Semenza GL, Dang CV. HIF-1-mediated expression of pyruvate dehydrogenase kinase: a metabolic switch required for cellular adaptation to hypoxia. Cell Metabolism. 2006; 3: 177–185.
- [108] Hitosugi T, Fan J, Chung T, Lythgoe K, Wang X, Xie J, et al. Tyrosine Phosphorylation of Mitochondrial Pyruvate Dehydrogenase Kinase 1 is Important for Cancer Metabolism. Molecular Cell. 2011; 44: 864–877.
- [109] Michelakis ED, McMurtry MS, Wu XC, Dyck JR, Moudgil R, Hopkins TA, et al. Dichloroacetate, a metabolic modulator, prevents and reverses chronic hypoxic pulmonary hypertension in rats: role of increased expression and activity of voltage-gated potassium channels. Circulation. 2002; 105: 244–250.
- [110] McMurtry MS, Bonnet S, Wu X, Dyck JRB, Haromy A, Hashimoto K, et al. Dichloroacetate Prevents and Reverses Pulmonary Hypertension by Inducing Pulmonary Artery Smooth Muscle Cell Apoptosis. Circulation Research. 2004; 95: 830– 840.
- [111] Bonnet S, Michelakis ED, Porter CJ, Andrade-Navarro MA, Thebaud B, Bonnet S, *et al*. An abnormal mitochondrial-hypoxia inducible factor-1alpha-Kv channel pathway disrupts oxygen sensing and triggers pulmonary arterial hypertension in fawn hooded rats: similarities to human pulmonary arterial hypertension. Circulation. 2006; 113: 2630–2641.
- [112] Michelakis ED, Gurtu V, Webster L, Barnes G, Watson G, Howard L, et al. Inhibition of pyruvate dehydrogenase kinase improves pulmonary arterial hypertension in genetically susceptible patients. Science Translational Medicine. 2017; 9: eaao4583.
- [113] Zhang H, Wang D, Li M, Plecitá-Hlavatá L, D'Alessandro A, Tauber J, *et al.* Metabolic and Proliferative State of Vascular Adventitial Fibroblasts in Pulmonary Hypertension is Regulated through a MicroRNA-124/PTBP1 (Polypyrimidine Tract Binding Protein 1)/Pyruvate Kinase Muscle Axis. Circulation. 2017; 136: 2468–2485.
- [114] Sutendra G, Dromparis P, Bonnet S, Haromy A, McMurtry MS, Bleackley RC, et al. Pyruvate dehydrogenase inhibition by the inflammatory cytokine TNFalpha contributes to the pathogenesis of pulmonary arterial hypertension. Journal of Molecular Medicine. 2011; 89: 771–783.
- [115] Aounallah M, Dagenais-Lussier X, El-Far M, Mehraj V, Jenabian M, Routy J, et al. Current topics in HIV pathogenesis, part 2: Inflammation drives a Warburg-like effect on the metabolism of HIV-infected subjects. Cytokine and Growth Factor Reviews. 2016; 28: 1–10.
- [116] Vander Heiden MG, Cantley LC, Thompson CB. Understanding the Warburg Effect: the Metabolic Requirements of Cell Pro-

- liferation. Science. 2009; 324: 1029-1033.
- [117] DeBerardinis RJ, Lum JJ, Hatzivassiliou G, Thompson CB. The biology of cancer: metabolic reprogramming fuels cell growth and proliferation. Cell Metabolism. 2008; 7: 11–20.
- [118] Acharya AP, Tang Y, Bertero T, Tai Y, Harvey LD, Woodcock CC, et al. Simultaneous Pharmacologic Inhibition of yes-Associated Protein 1 and Glutaminase 1 via Inhaled Poly(Lactic-co-Glycolic) Acid–Encapsulated Microparticles Improves Pulmonary Hypertension. Journal of the American Heart Association. 2021; 10: e019091.
- [119] Bertero T, Lu Y, Annis S, Hale A, Bhat B, Saggar R, et al. Systems-level regulation of microRNA networks by miR-130/301 promotes pulmonary hypertension. Journal of Clinical Investigation. 2014; 124: 3514–3528.
- [120] Bertero T, Oldham WM, Cottrill KA, Pisano S, Vanderpool RR, Yu Q, et al. Vascular stiffness mechanoactivates YAP/TAZdependent glutaminolysis to drive pulmonary hypertension. Journal of Clinical Investigation. 2016; 126: 3313–3335.
- [121] Caneba CA, Yang L, Baddour J, Curtis R, Win J, Hartig S, et al. Nitric oxide is a positive regulator of the Warburg effect in ovarian cancer cells. Cell Death and Disease. 2014; 5: e1302.
- [122] Rouault TA, Maio N. Biogenesis and functions of mammalian iron-sulfur proteins in the regulation of iron homeostasis and pivotal metabolic pathways. Journal of Biological Chemistry. 2017; 292: 12744–12753.
- [123] Fuss JO, Tsai C, Ishida JP, Tainer JA. Emerging critical roles of Fe–S clusters in DNA replication and repair. Biochimica Et Biophysica Acta (BBA) - Molecular Cell Research. 2015; 1853: 1253–1271.
- [124] White K, Lu Y, Annis S, Hale AE, Chau BN, Dahlman JE, et al. Genetic and hypoxic alterations of the microRNA-210-ISCU1/2 axis promote iron-sulfur deficiency and pulmonary hypertension. EMBO Molecular Medicine. 2015; 7: 695–713.
- [125] Chan SY, Zhang Y, Hemann C, Mahoney CE, Zweier JL, Loscalzo J. MicroRNA-210 Controls Mitochondrial Metabolism during Hypoxia by Repressing the Iron-Sulfur Cluster Assembly Proteins ISCU1/2. Cell Metabolism. 2009; 10: 273–284.
- [126] Cameron J, Janer A, Levandovskiy V, MacKay N, Rouault T, Tong W, et al. Mutations in Iron-Sulfur Cluster Scaffold Genes NFU1 and BOLA3 Cause a Fatal Deficiency of Multiple Respiratory Chain and 2-Oxoacid Dehydrogenase Enzymes. The American Journal of Human Genetics. 2011; 89: 486–495.
- [127] Colin F, Martelli A, Clemancey M, Latour JM, Gambarelli S, Zeppieri L, *et al.* Mammalian frataxin controls sulfur production and iron entry during de novo Fe4S4 cluster assembly. Journal of the American Chemical Society. 2013; 135: 733–740.
- [128] Navarro-Sastre A, Tort F, Stehling O, Uzarska M, Arranz J, del Toro M, et al. A Fatal Mitochondrial Disease is Associated with Defective NFU1 Function in the Maturation of a Subset of Mitochondrial Fe-S Proteins. The American Journal of Human Genetics. 2011; 89: 656–667.
- [129] Ahting U, Mayr JA, Vanlander AV, Hardy SA, Santra S, Makowski C, et al. Clinical, biochemical, and genetic spectrum of seven patients with NFU1 deficiency. Frontiers in Genetics. 2015; 6: 123.
- [130] Niihori M, Eccles CA, Kurdyukov S, Zemskova M, Varghese MV, Stepanova AA, et al. Rats with a Human Mutation of NFU1 Develop Pulmonary Hypertension. American Journal of Respiratory Cell and Molecular Biology. 2020; 62: 231–242.
- [131] Yu Q, Tai Y, Tang Y, Zhao J, Negi V, Culley MK, et al. BOLA (BolA Family Member 3) Deficiency Controls Endothelial Metabolism and Glycine Homeostasis in Pulmonary Hypertension. Circulation. 2019; 139: 2238–2255.
- [132] Culley MK, Zhao J, Tai YY, Tang Y, Perk D, Negi V, et al. Frataxin deficiency promotes endothelial senescence in pul-



- monary hypertension. The Journal of Clinical Investigation. 2021; 131: e136459.
- [133] Hemnes AR, Luther JM, Rhodes CJ, Burgess JP, Carlson J, Fan R, *et al.* Human PAH is characterized by a pattern of lipid-related insulin resistance. JCI Insight. 2019; 4: e123611.
- [134] Hemnes AR, Fessel JP, Chen X, Zhu S, Fortune NL, Jetter C, et al. BMPR2 dysfunction impairs insulin signaling and glucose homeostasis in cardiomyocytes. American Journal of Physiology. Lung Cellular and Molecular Physiology. 2020; 318: L429–L441.
- [135] Hansmann G, Wagner RA, Schellong S, Perez VA, Urashima T, Wang L, *et al.* Pulmonary arterial hypertension is linked to insulin resistance and reversed by peroxisome proliferator-activated receptor-gamma activation. Circulation. 2007; 115: 1275–1284.
- [136] Legchenko E, Chouvarine P, Borchert P, Fernandez-Gonzalez A, Snay E, Meier M, et al. PPARgamma agonist pioglitazone reverses pulmonary hypertension and prevents right heart failure via fatty acid oxidation. Science Translational Medicine. 2018; 10: eaao0303.
- [137] Hansmann G, de Jesus Perez VA, Alastalo TP, Alvira CM, Guignabert C, Bekker JM, et al. An antiproliferative BMP-2/PPARgamma/apoE axis in human and murine SMCs and its role in pulmonary hypertension. The Journal of Clinical Investigation. 2008; 118: 1846–1857.
- [138] Stacher E, Graham BB, Hunt JM, Gandjeva A, Groshong SD, McLaughlin VV, et al. Modern Age Pathology of Pulmonary Arterial Hypertension. American Journal of Respiratory and Critical Care Medicine. 2012; 186: 261–272.
- [139] Price LC, Wort SJ, Perros F, Dorfmüller P, Huertas A, Montani D, et al. Inflammation in Pulmonary Arterial Hypertension. Chest. 2012; 141: 210–221.
- [140] Florentin J, Coppin E, Vasamsetti SB, Zhao J, Tai Y, Tang Y, *et al.* Inflammatory Macrophage Expansion in Pulmonary Hypertension Depends upon Mobilization of Blood-Borne Monocytes. The Journal of Immunology. 2018; 200: 3612–3625.
- [141] Al-Qazazi R, Lima PDA, Prisco SZ, Potus F, Dasgupta A, Chen K, et al. Macrophage–NLRP3 Activation Promotes Right Ventricle Failure in Pulmonary Arterial Hypertension. American Journal of Respiratory and Critical Care Medicine. 2022; 206: 608–624.
- [142] Soon E, Holmes AM, Treacy CM, Doughty NJ, Southgate L, Machado RD, *et al.* Elevated levels of inflammatory cytokines predict survival in idiopathic and familial pulmonary arterial hypertension. Circulation. 2010; 122: 920–927.
- [143] Cracowski J, Chabot F, Labarere J, Faure P, Degano B, Schwebel C, *et al.* Proinflammatory cytokine levels are linked to death in pulmonary arterial hypertension. European Respiratory Journal. 2014; 43: 915–917.
- [144] Humbert M, Monti G, Brenot F, Sitbon O, Portier A, Grangeot-Keros L, et al. Increased interleukin-1 and interleukin-6 serum concentrations in severe primary pulmonary hypertension. American Journal of Respiratory and Critical Care Medicine. 1995; 151: 1628–1631.
- [145] Steiner MK, Syrkina OL, Kolliputi N, Mark EJ, Hales CA, Waxman AB. Interleukin-6 Overexpression Induces Pulmonary Hypertension. Circulation Research. 2009; 104: 236–244.
- [146] Hagen M, Fagan K, Steudel W, Carr M, Lane K, Rodman DM, et al. Interaction of interleukin-6 and the BMP pathway in pulmonary smooth muscle. American Journal of Physiology-Lung Cellular and Molecular Physiology. 2007; 292: L1473–L1479.
- [147] Houssaini A, Abid S, Mouraret N, Wan F, Rideau D, Saker M, et al. Rapamycin Reverses Pulmonary Artery Smooth Muscle Cell Proliferation in Pulmonary Hypertension. American Journal of Respiratory Cell and Molecular Biology. 2013; 48: 568– 577.

- [148] Courboulin A, Tremblay VL, Barrier M, Meloche J, Jacob MH, Chapolard M, et al. Kruppel-like factor 5 contributes to pulmonary artery smooth muscle proliferation and resistance to apoptosis in human pulmonary arterial hypertension. Respiratory Research. 2011; 12: 128.
- [149] Voelkel NF, Tuder RM, Bridges J, Arend WP. Interleukin-1 receptor antagonist treatment reduces pulmonary hypertension generated in rats by monocrotaline. American Journal of Respiratory Cell and Molecular Biology. 1994; 11: 664–675.
- [150] Tamura Y, Phan C, Tu L, Le Hiress M, Thuillet R, Jutant E, et al. Ectopic upregulation of membrane-bound IL6R drives vascular remodeling in pulmonary arterial hypertension. Journal of Clinical Investigation. 2018; 128: 1956–1970.
- [151] Toshner M, Church C, Harbaum L, Rhodes C, Villar Moreschi SS, Liley J, et al. Mendelian randomisation and experimental medicine approaches to interleukin-6 as a drug target in pulmonary arterial hypertension. The European Respiratory Journal. 2022; 59: 2002463.
- [152] Tamosiuniene R, Tian W, Dhillon G, Wang L, Sung YK, Gera L, et al. Regulatory T Cells Limit Vascular Endothelial Injury and Prevent Pulmonary Hypertension. Circulation Research. 2011: 109: 867–879.
- [153] Arends SJ, Damoiseaux JGMC, Duijvestijn AM, Debrus-Palmans L, Vroomen M, Boomars KA, et al. Immunoglobulin G anti-endothelial cell antibodies: inducers of endothelial cell apoptosis in pulmonary arterial hypertension? Clinical and Experimental Immunology. 2013; 174: 433–440.
- [154] Basyal B, Jarrett H, Barnett CF. Pulmonary Hypertension in HIV. Canadian Journal of Cardiology. 2019; 35: 288–298.
- [155] Saito T, Miyagawa K, Chen S, Tamosiuniene R, Wang L, Sharpe O, et al. Upregulation of Human Endogenous Retrovirus-K is Linked to Immunity and Inflammation in Pulmonary Arterial Hypertension. Circulation. 2017; 136: 1920–1935.
- [156] Lee JD, Burger CD, Delossantos GB, Grinnan D, Ralph DD, Rayner SG, et al. A Survey-based Estimate of COVID-19 Incidence and Outcomes among Patients with Pulmonary Arterial Hypertension or Chronic Thromboembolic Pulmonary Hypertension and Impact on the Process of Care. Annals of the American Thoracic Society. 2020; 17: 1576–1582.
- [157] Montani D, Certain MC, Weatherald J, Jais X, Bulifon S, Noel-Savina E, et al. COVID-19 in Patients with Pulmonary Hypertension: A National Prospective Cohort Study. American Journal of Respiratory and Critical Care Medicine. 2022; 206: 573–583.
- [158] Knafl D, Gerges C, King CH, Humbert M, Bustinduy AL. Schistosomiasis-associated pulmonary arterial hypertension: a systematic review. European Respiratory Review. 2020; 29: 190089.
- [159] Graham BB, Chabon J, Gebreab L, Poole J, Debella E, Davis L, et al. Transforming growth factor-beta signaling promotes pulmonary hypertension caused by Schistosoma mansoni. Circulation. 2013; 128: 1354–1364.
- [160] Kumar R, Mickael C, Chabon J, Gebreab L, Rutebemberwa A, Garcia AR, et al. The Causal Role of IL-4 and IL-13 in Schistosoma mansoni Pulmonary Hypertension. American Journal of Respiratory and Critical Care Medicine. 2015; 192: 998–1008.
- [161] Cho WK, Lee CM, Kang MJ, Huang Y, Giordano FJ, Lee PJ, et al. IL-13 receptor alpha2-arginase 2 pathway mediates IL-13-induced pulmonary hypertension. American Journal of Physiology. Lung Cellular and Molecular Physiology. 2013; 304: L112–L124.
- [162] Christmann RB, Hayes E, Pendergrass S, Padilla C, Farina G, Affandi AJ, et al. Interferon and alternative activation of monocyte/macrophages in systemic sclerosis-associated pulmonary arterial hypertension. Arthritis and Rheumatism. 2011; 63: 1718–1728.



- [163] Rabinovitch M, Guignabert C, Humbert M, Nicolls MR. Inflammation and Immunity in the Pathogenesis of Pulmonary Arterial Hypertension. Circulation Research. 2014; 115: 165–175.
- [164] Sweatt AJ, Hedlin HK, Balasubramanian V, Hsi A, Blum LK, Robinson WH, et al. Discovery of Distinct Immune Phenotypes Using Machine Learning in Pulmonary Arterial Hypertension. Circulation Research. 2019; 124: 904–919.
- [165] Marsh LM, Jandl K, Grünig G, Foris V, Bashir M, Ghanim B, et al. The inflammatory cell landscape in the lungs of patients with idiopathic pulmonary arterial hypertension. European Respiratory Journal. 2018; 51: 1701214.
- [166] Kumar R, Graham B. How does inflammation contribute to pulmonary hypertension? European Respiratory Journal. 2018; 51: 1702403.
- [167] Loscalzo J, Handy DE. Epigenetic Modifications: Basic Mechanisms and Role in Cardiovascular Disease (2013 Grover Conference Series). Pulmonary Circulation. 2014; 4: 169–174.
- [168] Cech T, Steitz J. The Noncoding RNA Revolution—Trashing Old Rules to Forge New Ones. Cell. 2014; 157: 77–94.
- [169] Bartel DP. MicroRNAs: Target Recognition and Regulatory Functions. Cell. 2009; 136: 215–233.
- [170] Negi V, Chan SY. Discerning functional hierarchies of microR-NAs in pulmonary hypertension. JCI Insight. 2017; 2: e91327.
- [171] Ulitsky I, Bartel D. LincRNAs: Genomics, Evolution, and Mechanisms. Cell. 2013; 154: 26–46.
- [172] Zehendner CM, Valasarajan C, Werner A, Boeckel J, Bischoff FC, John D, et al. Long Noncoding RNA TYKRIL Plays a Role in Pulmonary Hypertension via the p53-mediated Regulation of PDGFRβ. American Journal of Respiratory and Critical Care Medicine. 2020; 202: 1445–1457.
- [173] Gamen E, Seeger W, Pullamsetti SS. The emerging role of epigenetics in pulmonary hypertension. European Respiratory Journal. 2016; 48: 903–917.
- [174] Hu L, Wang J, Huang H, Yu Y, Ding J, Yu Y, et al. YTHDF1 Regulates Pulmonary Hypertension through Translational Control of MAGED1. American Journal of Respiratory and Critical Care Medicine. 2021; 203: 1158–1172.
- [175] Bannister AJ, Kouzarides T. Regulation of chromatin by histone modifications. Cell Research. 2011; 21: 381–395.
- [176] Napoli C, Benincasa G, Loscalzo J. Epigenetic Inheritance Underlying Pulmonary Arterial Hypertension. Arteriosclerosis, Thrombosis, and Vascular Biology. 2019; 39: 653–664.
- [177] Bisserier M, Mathiyalagan P, Zhang S, Elmastour F, Dorfmuller P, Humbert M, *et al.* Regulation of the Methylation and Expression Levels of the BMPR2 Gene by SIN3a as a Novel Therapeutic Mechanism in Pulmonary Arterial Hypertension. Circulation. 2021; 144: 52–73.
- [178] Belkina AC, Denis GV. BET domain co-regulators in obesity, inflammation and cancer. Nature Reviews Cancer. 2012; 12: 465–477.
- [179] Meloche J, Potus F, Vaillancourt M, Bourgeois A, Johnson I, Deschamps L, et al. Bromodomain-Containing Protein 4: The Epigenetic Origin of Pulmonary Arterial Hypertension. Circulation Research. 2015; 117: 525–535.
- [180] Welti J, Sharp A, Brooks N, Yuan W, McNair C, Chand SN, et al. Targeting the p300/CBP Axis in Lethal Prostate Cancer. Cancer Discovery. 2021; 11: 1118–1137.
- [181] Chelladurai P, Kuenne C, Bourgeois A, Günther S, Valasarajan C, Cherian AV, *et al.* Epigenetic reactivation of transcriptional programs orchestrating fetal lung development in human pulmonary hypertension. Science Translational Medicine. 2022; 14: eabe5407.
- [182] Meloche J, Pflieger A, Vaillancourt M, Paulin R, Potus F, Zervopoulos S, et al. Role for DNA Damage Signaling in Pulmonary Arterial Hypertension. Circulation. 2014; 129: 786–797.

- [183] Chen P, Cao A, Miyagawa K, Tojais NF, Hennigs JK, Li CG, et al. Amphetamines promote mitochondrial dysfunction and DNA damage in pulmonary hypertension. JCI Insight. 2017; 2: e90427
- [184] Culley MK, Chan SY. Endothelial Senescence: a New Age in Pulmonary Hypertension. Circulation Research. 2022; 130: 928–941.
- [185] van der Feen DE, Bossers GPL, Hagdorn QAJ, Moonen J, Kurakula K, Szulcek R, et al. Cellular senescence impairs the reversibility of pulmonary arterial hypertension. Science Translational Medicine. 2020; 12: eaaw4974.
- [186] Tasdemir N, Banito A, Roe J, Alonso-Curbelo D, Camiolo M, Tschaharganeh DF, et al. BRD4 Connects Enhancer Remodeling to Senescence Immune Surveillance. Cancer Discovery. 2016; 6: 612–629.
- [187] Yuan K, Agarwal S, Chakraborty A, Condon DF, Patel H, Zhang S, et al. Lung Pericytes in Pulmonary Vascular Physiology and Pathophysiology. Comprehensive Physiology. 2021; 11: 2227–2247.
- [188] Yuan K, Shamskhou EA, Orcholski ME, Nathan A, Reddy S, Honda H, et al. Loss of Endothelium-Derived Wnt5a is Associated with Reduced Pericyte Recruitment and Small Vessel Loss in Pulmonary Arterial Hypertension. Circulation. 2019; 139: 1710–1724.
- [189] Yuan K, Shao N, Hennigs JK, Discipulo M, Orcholski ME, Shamskhou E, et al. Increased Pyruvate Dehydrogenase Kinase 4 Expression in Lung Pericytes is Associated with Reduced Endothelial-Pericyte Interactions and Small Vessel Loss in Pulmonary Arterial Hypertension. The American Journal of Pathology. 2016; 186: 2500–2514.
- [190] Ricard N, Tu L, Le Hiress M, Huertas A, Phan C, Thuillet R, et al. Increased Pericyte Coverage Mediated by Endothelial-Derived Fibroblast Growth Factor-2 and Interleukin-6 is a Source of Smooth Muscle-Like Cells in Pulmonary Hypertension. Circulation. 2014; 129: 1586–1597.
- [191] Zhang L, Zeng X, Li Y, Chen S, Tang L, Wang N, et al. Keratin 1 attenuates hypoxic pulmonary artery hypertension by suppressing pulmonary artery media smooth muscle expansion. Acta Physiologica. 2021; 231: e13558.
- [192] Kummer W. Pulmonary Vascular Innervation and its Role in Responses to Hypoxia: Size Matters! Proceedings of the American Thoracic Society. 2011; 8: 471–476.
- [193] Chen SL, Zhang FF, Xu J, Xie DJ, Zhou L, Nguyen T, et al. Pulmonary artery denervation to treat pulmonary arterial hypertension: the single-center, prospective, first-in-man PADN-1 study (first-in-man pulmonary artery denervation for treatment of pulmonary artery hypertension). Journal of the American College of Cardiology. 2013; 62: 1092–1100.
- [194] Zhang H, Zhang J, Chen M, Xie D, Kan J, Yu W, et al. Pulmonary Artery Denervation Significantly Increases 6-Min Walk Distance for Patients with Combined Pre- and Post-Capillary Pulmonary Hypertension Associated with Left Heart Failure. JACC: Cardiovascular Interventions. 2019; 12: 274–284.
- [195] Granton J, Langleben D, Kutryk MB, Camack N, Galipeau J, Courtman DW, et al. Endothelial NO-Synthase Gene-Enhanced Progenitor Cell Therapy for Pulmonary Arterial Hypertension. Circulation Research. 2015; 117: 645–654.
- [196] Hansmann G, Chouvarine P, Diekmann F, Giera M, Ralser M, Mülleder M, et al. Human umbilical cord mesenchymal stem cell-derived treatment of severe pulmonary arterial hypertension. Nature Cardiovascular Research. 2022; 1: 568–576.
- [197] Valadi H, Ekström K, Bossios A, Sjöstrand M, Lee JJ, Lötvall JO. Exosome-mediated transfer of mRNAs and microRNAs is a novel mechanism of genetic exchange between cells. Nature Cell Biology. 2007; 9: 654–659.
- [198] Zhao J, Florentin J, Tai Y, Torrino S, Ohayon L, Brzoska T,



- *et al.* Long Range Endocrine Delivery of Circulating miR-210 to Endothelium Promotes Pulmonary Hypertension. Circulation Research. 2020; 127: 677–692.
- [199] Fineman JR, Black SM. Pressure vs Flow-Induced Pulmonary Hypertension. Advances in Pulmonary Hypertension. 2019; 18: 19–24.
- [200] Tzima E, Irani-Tehrani M, Kiosses WB, Dejana E, Schultz DA, Engelhardt B, et al. A mechanosensory complex that mediates the endothelial cell response to fluid shear stress. Nature. 2005; 437: 426–431.
- [201] Li M, Stenmark KR, Shandas R, Tan W. Effects of Pathological Flow on Pulmonary Artery Endothelial Production of Vasoactive Mediators and Growth Factors. Journal of Vascular Research. 2009; 46: 561–571.
- [202] Steinhorn RH, Russell JA, Lakshminrusimha S, Gugino SF, Black SM, Fineman JR. Altered endothelium-dependent relaxations in lambs with high pulmonary blood flow and pulmonary hypertension. American Journal of Physiology-Heart and Circulatory Physiology. 2001; 280: H311–H317.
- [203] Gan CT, Lankhaar J, Westerhof N, Marcus JT, Becker A, Twisk JWR, et al. Noninvasively Assessed Pulmonary Artery Stiffness Predicts Mortality in Pulmonary Arterial Hypertension. Chest. 2007; 132: 1906–1912.
- [204] Halder G, Dupont S, Piccolo S. Transduction of mechanical and cytoskeletal cues by YAP and TAZ. Nature Reviews Molecular Cell Biology. 2012; 13: 591–600.
- [205] Bertero T, Cottrill K, Lu Y, Haeger C, Dieffenbach P, Annis S, et al. Matrix Remodeling Promotes Pulmonary Hypertension through Feedback Mechanoactivation of the YAP/TAZ-miR-130/301 Circuit. Cell Reports. 2015; 13: 1016–1032.
- [206] Dieffenbach PB, Haeger CM, Coronata AMF, Choi KM, Varelas X, Tschumperlin DJ, et al. Arterial stiffness induces remodeling phenotypes in pulmonary artery smooth muscle cells via YAP/TAZ-mediated repression of cyclooxygenase-2. American Journal of Physiology-Lung Cellular and Molecular Physiology. 2017; 313: L628–L647.
- [207] Kudryashova TV, Goncharov DA, Pena A, Kelly N, Vanderpool R, Baust J, et al. HIPPO-Integrin-linked Kinase Cross-Talk Controls Self-Sustaining Proliferation and Survival in Pulmonary Hypertension. American Journal of Respiratory and Critical Care Medicine. 2016; 194: 866–877.
- [208] Samokhin AO, Stephens T, Wertheim BM, Wang RS, Vargas SO, Yung LM, et al. NEDD9 targets COL3A1 to promote endothelial fibrosis and pulmonary arterial hypertension. Science Translational Medicine. 2018; 10: eaap7294.
- [209] Bogaard HJ, Natarajan R, Henderson SC, Long CS, Kraskauskas D, Smithson L, et al. Chronic Pulmonary Artery Pressure Elevation is Insufficient to Explain Right Heart Failure. Circulation. 2009; 120: 1951–1960.
- [210] Sharifi Kia D, Kim K, Simon MA. Current Understanding of the Right Ventricle Structure and Function in Pulmonary Arterial Hypertension. Frontiers in Physiology. 2021; 12: 641310.
- [211] Andersen S, Nielsen-Kudsk JE, Vonk Noordegraaf A, de Man FS. Right Ventricular Fibrosis. Circulation. 2019; 139: 269–285.
- [212] Hill MR, Simon MA, Valdez-Jasso D, Zhang W, Champion HC, Sacks MS. Structural and Mechanical Adaptations of Right Ventricle Free Wall Myocardium to Pressure Overload. Annals of Biomedical Engineering. 2014; 42: 2451–2465.
- [213] Fang Y, Piao L, Hong Z, Toth PT, Marsboom G, Bache-Wiig P, et al. Therapeutic inhibition of fatty acid oxidation in right ventricular hypertrophy: exploiting Randle's cycle. Journal of Molecular Medicine. 2012; 90: 31–43.
- [214] Prisco SZ, Hartweck LM, Rose L, Lima PDA, Thenappan T, Archer SL, et al. Inflammatory Glycoprotein 130 Signaling Links Changes in Microtubules and Junctophilin-2 to Altered Mitochondrial Metabolism and Right Ventricular Contractility.

- Circulation: Heart Failure. 2022; 15: e008574.
- [215] van de Veerdonk MC, Kind T, Marcus JT, Mauritz G, Heymans MW, Bogaard H, et al. Progressive Right Ventricular Dysfunction in Patients with Pulmonary Arterial Hypertension Responding to Therapy. Journal of the American College of Cardiology. 2011; 58: 2511–2519.
- [216] Maron BA, Leopold JA. The Role of the Renin-Angiotensin-Aldosterone System in the Pathobiology of Pulmonary Arterial Hypertension (2013 Grover Conference Series). Pulmonary Circulation. 2014; 4: 200–210.
- [217] Velez-Roa S, Ciarka A, Najem B, Vachiery JL, Naeije R, van de Borne P. Increased sympathetic nerve activity in pulmonary artery hypertension. Circulation. 2004; 110: 1308–1312.
- [218] Rochon ER, Krowka MJ, Bartolome S, Heresi GA, Bull T, Roberts K, et al. BMP9/10 in Pulmonary Vascular Complications of Liver Disease. American Journal of Respiratory and Critical Care Medicine. 2020; 201: 1575–1578.
- [219] Nikolic I, Yung L, Yang P, Malhotra R, Paskin-Flerlage SD, Dinter T, et al. Bone Morphogenetic Protein 9 is a Mechanistic Biomarker of Portopulmonary Hypertension. American Journal of Respiratory and Critical Care Medicine. 2019; 199: 891–902.
- [220] Vonk Noordegraaf A, Westerhof BE, Westerhof N. The Relationship between the Right Ventricle and its Load in Pulmonary Hypertension. Journal of the American College of Cardiology. 2017; 69: 236–243.
- [221] Nickel NP, O'Leary JM, Brittain EL, Fessel JP, Zamanian RT, West JD, et al. Kidney dysfunction in patients with pulmonary arterial hypertension. Pulmonary Circulation. 2017; 7: 38–54.
- [222] O'Leary JM, Assad TR, Xu M, Birdwell KA, Farber-Eger E, Wells QS, et al. Pulmonary hypertension in patients with chronic kidney disease: invasive hemodynamic etiology and outcomes. Pulmonary Circulation. 2017; 7: 674–683.
- [223] Kawar B, Ellam T, Jackson C, Kiely DG. Pulmonary Hypertension in Renal Disease: Epidemiology, Potential Mechanisms and Implications. American Journal of Nephrology. 2013; 37: 281–290.
- [224] Kim S, Rigatto K, Gazzana MB, Knorst MM, Richards EM, Pepine CJ, et al. Altered Gut Microbiome Profile in Patients with Pulmonary Arterial Hypertension. Hypertension. 2020; 75: 1063–1071.
- [225] Vaillancourt M, Chia P, Medzikovic L, Cao N, Ruffenach G, Younessi D, et al. Experimental Pulmonary Hypertension Is Associated With Neuroinflammation in the Spinal Cord. Frontiers in Physiology. 2019; 10: 1186.
- [226] Rich JD, Rich S. Clinical Diagnosis of Pulmonary Hypertension. Circulation. 2014; 130: 1820–1830.
- [227] Rich S, Dantzker DR, Ayres SM, Bergofsky EH, Brundage BH, Detre KM, et al. Primary Pulmonary Hypertension. Annals of Internal Medicine. 1987; 107: 216–223.
- [228] D'Alonzo GE, Barst RJ, Ayres SM, Bergofsky EH, Brundage BH, Detre KM, et al. Survival in Patients with Primary Pulmonary Hypertension. Results from a national prospective registry. Annals of Internal Medicine. 1991; 115: 343–349.
- [229] Benza RL, Miller DP, Barst RJ, Badesch DB, Frost AE, Mc-Goon MD. An Evaluation of Long-term Survival from Time of Diagnosis in Pulmonary Arterial Hypertension from the RE-VEAL Registry. Chest. 2012; 142: 448–456.
- [230] Benza RL, Miller DP, Gomberg-Maitland M, Frantz RP, Foreman AJ, Coffey CS, et al. Predicting Survival in Pulmonary Arterial Hypertension: insights from the Registry to Evaluate Early and Long-Term Pulmonary Arterial Hypertension Disease Management (REVEAL). Circulation. 2010; 122: 164–172.
- [231] Humbert M, Sitbon O, Chaouat A, Bertocchi M, Habib G, Gressin V, et al. Survival in Patients with Idiopathic, Familial, and Anorexigen-Associated Pulmonary Arterial Hypertension in the Modern Management Era. Circulation. 2010; 122: 156–163.



- [232] Benza RL, Gomberg-Maitland M, Elliott CG, Farber HW, Foreman AJ, Frost AE, et al. Predicting Survival in Patients with Pulmonary Arterial Hypertension: The REVEAL Risk Score Calculator 2.0 and Comparison With ESC/ERS-Based Risk Assessment Strategies. Chest. 2019; 156: 323–337.
- [233] Kylhammar D, Kjellström B, Hjalmarsson C, Jansson K, Nisell M, Söderberg S, et al. A comprehensive risk stratification at early follow-up determines prognosis in pulmonary arterial hypertension. European Heart Journal. 2018; 39: 4175–4181.
- [234] Hoeper MM, Kramer T, Pan Z, Eichstaedt CA, Spiesshoefer J, Benjamin N, et al. Mortality in pulmonary arterial hypertension: prediction by the 2015 European pulmonary hypertension guidelines risk stratification model. European Respiratory Journal. 2017; 50: 1700740.
- [235] Boucly A, Weatherald J, Savale L, Jaïs X, Cottin V, Prevot G, et al. Risk assessment, prognosis and guideline implementation in pulmonary arterial hypertension. European Respiratory Journal. 2017; 50: 1700889.
- [236] Galie N, Channick RN, Frantz RP, Grunig E, Jing ZC, Moiseeva O, et al. Risk stratification and medical therapy of pulmonary arterial hypertension. The European Respiratory Journal. 2019; 53: 1801889.
- [237] Galiè N, Rubin L, Hoeper M, Jansa P, Al-Hiti H, Meyer G, et al. Treatment of patients with mildly symptomatic pulmonary arterial hypertension with bosentan (EARLY study): a double-blind, randomised controlled trial. The Lancet. 2008; 371: 2093–2100.
- [238] Lau EM, Humbert M, Celermajer DS. Early detection of pulmonary arterial hypertension. Nature Reviews Cardiology. 2015; 12: 143–155.
- [239] Brown LM, Chen H, Halpern S, Taichman D, McGoon MD, Farber HW, et al. Delay in recognition of pulmonary arterial hypertension: factors identified from the REVEAL Registry. Chest. 2011; 140: 19–26.
- [240] Assad TR, Maron BA, Robbins IM, Xu M, Huang S, Harrell FE, et al. Prognostic Effect and Longitudinal Hemodynamic Assessment of Borderline Pulmonary Hypertension. JAMA Cardiology. 2017; 2: 1361.
- [241] Maron BA, Brittain EL, Hess E, Waldo SW, Barón AE, Huang S, *et al.* Pulmonary vascular resistance and clinical outcomes in patients with pulmonary hypertension: a retrospective cohort study. The Lancet Respiratory Medicine. 2020; 8: 873–884.
- [242] Larkin EK, Newman JH, Austin ED, Hemnes AR, Wheeler L, Robbins IM, et al. Longitudinal Analysis Casts Doubt on the Presence of Genetic Anticipation in Heritable Pulmonary Arterial Hypertension. American Journal of Respiratory and Critical Care Medicine. 2012; 186: 892–896.
- [243] Elder JC, Brofman BL, Kohn PM, Charms BL, Lawrence J, Godfrey AB. Unilateral pulmonary artery absence or hypoplasia; radiographic and cardiopulmonary studies in five patients. Circulation. 1958; 17: 557–566.
- [244] Maron BA, Cockrill BA, Waxman AB, Systrom DM. The Invasive Cardiopulmonary Exercise Test. Circulation. 2013; 127: 1157–1164.
- [245] Trip P, Vonk-Noordegraaf A, Bogaard HJ. Cardiopulmonary Exercise Testing Reveals Onset of Disease and Response to Treatment in a Case of Heritable Pulmonary Arterial Hypertension. Pulmonary Circulation. 2012; 2: 387–389.
- [246] Remy-Jardin M, Ryerson CJ, Schiebler ML, Leung ANC, Wild JM, Hoeper MM, *et al.* Imaging of pulmonary hypertension in adults: a position paper from the Fleischner Society. European Respiratory Journal. 2021; 57: 2004455.
- [247] Ota H, Kamada H, Higuchi S, Takase K. Clinical Application of 4D Flow MR Imaging to Pulmonary Hypertension. Magnetic Resonance in Medical Sciences. 2022; 21: 309–318.
- [248] Brittain EL, Talati M, Fessel JP, Zhu H, Penner N, Calcutt MW, et al. Fatty Acid Metabolic Defects and Right Ventricular Lipo-

- toxicity in Human Pulmonary Arterial Hypertension. Circulation. 2016; 133: 1936–1944.
- [249] Spindler M, Schmidt M, Geier O, Sandstede J, Hahn D, Ertl G, et al. Functional and Metabolic Recovery of the Right Ventricle during Bosentan Therapy in Idiopathic Pulmonary Arterial Hypertension. Journal of Cardiovascular Magnetic Resonance. 2005; 7: 853–854.
- [250] Eddy RL, Parraga G. Pulmonary xenon-129 MRI: new opportunities to unravel enigmas in respiratory medicine. European Respiratory Journal. 2020; 55: 1901987.
- [251] Virgincar RS, Nouls JC, Wang Z, Degan S, Qi Y, Xiong X, et al. Quantitative 129Xe MRI detects early impairment of gasexchange in a rat model of pulmonary hypertension. Scientific Reports. 2020; 10: 7385.
- [252] Dahhan T, Kaushik SS, He M, Mammarappallil JG, Tapson VF, McAdams HP, et al. Abnormalities in hyperpolarized (129)Xe magnetic resonance imaging and spectroscopy in two patients with pulmonary vascular disease. Pulmonary Circulation. 2016; 6: 126–131.
- [253] Park JB, Suh M, Park JY, Park JK, Kim YI, Kim H, et al. Assessment of Inflammation in Pulmonary Artery Hypertension by (68)Ga-Mannosylated Human Serum Albumin. American Journal of Respiratory and Critical Care Medicine. 2020; 201: 95–106
- [254] Anwar A, Ruffenach G, Mahajan A, Eghbali M, Umar S. Novel biomarkers for pulmonary arterial hypertension. Respiratory Research. 2016; 17: 88.
- [255] Fijalkowska A, Kurzyna M, Torbicki A, Szewczyk G, Florczyk M, Pruszczyk P, et al. Serum N-Terminal Brain Natriuretic Peptide as a Prognostic Parameter in Patients with Pulmonary Hypertension. Chest. 2006; 129: 1313–1321.
- [256] Blyth KG, Groenning BA, Mark PB, Martin TN, Foster JE, Steedman T, et al. NT-proBNP can be used to detect right ventricular systolic dysfunction in pulmonary hypertension. The European Respiratory Journal. 2007; 29: 737–744.
- [257] Nagaya N, Nishikimi T, Uematsu M, Satoh T, Kyotani S, Sakamaki F, et al. Plasma Brain Natriuretic Peptide as a Prognostic Indicator in Patients with Primary Pulmonary Hypertension. Circulation. 2000; 102: 865–870.
- [258] Williams MH, Handler CE, Akram R, Smith CJ, Das C, Smee J, et al. Role of N-terminal brain natriuretic peptide (N-TproBNP) in scleroderma-associated pulmonary arterial hypertension. European Heart Journal. 2006; 27: 1485–1494.
- [259] Sun W, Tang Y, Tai YY, Handen A, Zhao J, Speyer G, et al. SCUBE1 Controls BMPR2-Relevant Pulmonary Endothelial Function: Implications for Diagnostic Marker Development in Pulmonary Arterial Hypertension. JACC: Basic to Translational Science. 2020; 5: 1073–1092.
- [260] Bartolome S, Hoeper MM, Klepetko W. Advanced pulmonary arterial hypertension: mechanical support and lung transplantation. European Respiratory Review. 2017; 26: 170089.
- [261] Humbert M, McLaughlin V, Gibbs JSR, Gomberg-Maitland M, Hoeper MM, Preston IR, et al. Sotatercept for the Treatment of Pulmonary Arterial Hypertension. New England Journal of Medicine. 2021; 384: 1204–1215.
- [262] Long L, Ormiston ML, Yang X, Southwood M, Gräf S, Machado RD, et al. Selective enhancement of endothelial BMPR-II with BMP9 reverses pulmonary arterial hypertension. Nature Medicine. 2015; 21: 777–785.
- [263] Spiekerkoetter E, Tian X, Cai J, Hopper RK, Sudheendra D, Li CG, et al. FK506 activates BMPR2, rescues endothelial dysfunction, and reverses pulmonary hypertension. Journal of Clinical Investigation. 2013; 123: 3600–3613.
- [264] Spiekerkoetter E, Sung YK, Sudheendra D, Scott V, Del Rosario P, Bill M, *et al.* Randomised placebo-controlled safety and tolerability trial of FK506 (tacrolimus) for pulmonary ar-



- terial hypertension. European Respiratory Journal. 2017; 50: 1602449.
- [265] Prins KW, Thenappan T, Weir EK, Kalra R, Pritzker M, Archer SL. Repurposing Medications for Treatment of Pulmonary Arterial Hypertension: what's Old is New again. Journal of the American Heart Association. 2019; 8: e011343.
- [266] Boucherat O, Vitry G, Trinh I, Paulin R, Provencher S, Bonnet S. The cancer theory of pulmonary arterial hypertension. Pulmonary Circulation. 2017; 7: 285–299.
- [267] Schermuly RT, Dony E, Ghofrani HA, Pullamsetti S, Savai R, Roth M, et al. Reversal of experimental pulmonary hypertension by PDGF inhibition. Journal of Clinical Investigation. 2005; 115: 2811–2821.
- [268] Ghofrani HA, Seeger W, Grimminger F. Imatinib for the treatment of pulmonary arterial hypertension. The New England Journal of Medicine. 2005; 353: 1412–1413.
- [269] Frantz RP, Benza RL, Channick RN, Chin K, Howard LS, McLaughlin VV, et al. TORREY, a Phase 2 study to evaluate the efficacy and safety of inhaled seralutinib for the treatment of pulmonary arterial hypertension. Pulmonary Circulation. 2021; 11: 20458940211057071.
- [270] Gillies H, Niven R, Dake B, Chakinala MM, Feldman JP, Hill NS, et al. A Phase 1 Single and Multiple Ascending Dose (SAD/MAD) Study to Evaluate the Safety, Tolerability, and Pharmacokinetics of AV-101, a Novel Inhaled Dry Powder Formulation of Imatinib in Healthy Subjects. B106 (pp. A3594–A3594). Observational Studies and Clinical Trials in Pulmonary Hypertension: Union Square. 2022.
- [271] Montani D, Bergot E, Günther S, Savale L, Bergeron A, Bourdin A, *et al.* Pulmonary Arterial Hypertension in Patients Treated by Dasatinib. Circulation. 2012; 125: 2128–2137.
- [272] Cornet L, Khouri C, Roustit M, Guignabert C, Chaumais M, Humbert M, et al. Pulmonary arterial hypertension associated with protein kinase inhibitors: a pharmacovigilance—pharmacodynamic study. European Respiratory Journal. 2019; 53: 1802472.
- [273] Guignabert C, Phan C, Seferian A, Huertas A, Tu L, Thuillet R, et al. Dasatinib induces lung vascular toxicity and predisposes to pulmonary hypertension. Journal of Clinical Investigation. 2016; 126: 3207–3218.
- [274] Basu A, Bodycombe N, Cheah J, Price E, Liu K, Schaefer G, et al. An Interactive Resource to Identify Cancer Genetic and Lineage Dependencies Targeted by Small Molecules. Cell. 2013; 154: 1151–1161.
- [275] Negi V, Yang J, Speyer G, Pulgarin A, Handen A, Zhao J, et al. Computational repurposing of therapeutic small molecules from cancer to pulmonary hypertension. Science Advances. 2021; 7: eabh3794.
- [276] Van der Feen DE, Kurakula K, Tremblay E, Boucherat O, Bossers GPL, Szulcek R, et al. Multicenter Preclinical Validation of BET Inhibition for the Treatment of Pulmonary Arterial Hypertension. American Journal of Respiratory and Critical Care Medicine. 2019; 200: 910–920.
- [277] Provencher S, Potus F, Blais-Lecours P, Bernard S, Martineau S, Breuils-Bonnet S, et al. BET Protein Inhibition for Pulmonary Arterial Hypertension: a Pilot Clinical Trial. American Journal of Respiratory and Critical Care Medicine. 2022; 205: 1357–1360.
- [278] Chen X, Austin ED, Talati M, Fessel JP, Farber-Eger E, Brittain EL, et al. Oestrogen inhibition reverses pulmonary arterial hypertension and associated metabolic defects. European Respiratory Journal. 2017; 50: 1602337.
- [279] Weiss A, Neubauer MC, Yerabolu D, Kojonazarov B, Schlueter BC, Neubert L, et al. Targeting cyclin-dependent kinases for the treatment of pulmonary arterial hypertension. Nature Communications. 2019; 10: 2204.

- [280] Felley-Bosco E, Stahel R. Hippo/YAP pathway for targeted therapy. Translational Lung Cancer Research. 2014; 3: 75–83.
- [281] Graham RM, Frazier DP, Thompson JW, Haliko S, Li H, Wasserlauf BJ, *et al.* A unique pathway of cardiac myocyte death caused by hypoxia–acidosis. Journal of Experimental Biology. 2004; 207: 3189–3200.
- [282] Kantor PF, Lucien A, Kozak R, Lopaschuk GD. The Antianginal Drug Trimetazidine Shifts Cardiac Energy Metabolism from Fatty Acid Oxidation to Glucose Oxidation by Inhibiting Mitochondrial Long-Chain 3-Ketoacyl Coenzyme A Thiolase. Circulation Research. 2000; 86: 580–588.
- [283] Knuuti J, Wijns W, Saraste A, Capodanno D, Barbato E, Funck-Brentano C, et al. 2019 ESC Guidelines for the diagnosis and management of chronic coronary syndromes. European Heart Journal. 2020; 41: 407–477.
- [284] Khan SS, Cuttica MJ, Beussink-Nelson L, Kozyleva A, Sanchez C, Mkrdichian H, et al. Effects of Ranolazine on Exercise Capacity, Right Ventricular Indices, and Hemodynamic Characteristics in Pulmonary Arterial Hypertension: a Pilot Study. Pulmonary Circulation. 2015; 5: 547–556.
- [285] Han Y, Forfia P, Vaidya A, Mazurek JA, Park MH, Ramani G, et al. Ranolazine Improves Right Ventricular Function in Patients with Precapillary Pulmonary Hypertension: Results from a Double-Blind, Randomized, Placebo-Controlled Trial. Journal of Cardiac Failure. 2021; 27: 253–257.
- [286] Zamanian RT, Hansmann G, Snook S, Lilienfeld D, Rappaport KM, Reaven GM, et al. Insulin resistance in pulmonary arterial hypertension. The European Respiratory Journal. 2009; 33: 318–324.
- [287] Calvier L, Chouvarine P, Legchenko E, Hoffmann N, Geldner J, Borchert P, et al. PPARgamma Links BMP2 and TGF-beta1 Pathways in Vascular Smooth Muscle Cells, Regulating Cell Proliferation and Glucose Metabolism. Cell Metabolism. 2017; 25: 1118–1134.e7.
- [288] Mannucci E, Monami M, Di Bari M, Lamanna C, Gori F, Gensini GF, et al. Cardiac safety profile of rosiglitazone: a comprehensive meta-analysis of randomized clinical trials. International Journal of Cardiology. 2010; 143: 135–140.
- [289] Lincoff AM, Wolski K, Nicholls SJ, Nissen SE. Pioglitazone and Risk of Cardiovascular Events in Patients with Type 2 Diabetes Mellitus: a meta-analysis of randomized trials. The Journal of the American Medical Association. 2007; 298: 1180–1188.
- [290] Lai Y, Tabima DM, Dube JJ, Hughan KS, Vanderpool RR, Goncharov DA, et al. SIRT3–AMP-Activated Protein Kinase Activation by Nitrite and Metformin Improves Hyperglycemia and Normalizes Pulmonary Hypertension Associated with Heart Failure with Preserved Ejection Fraction. Circulation. 2016; 133: 717–731.
- [291] Brittain EL, Niswender K, Agrawal V, Chen X, Fan R, Pugh ME, et al. Mechanistic Phase II Clinical Trial of Metformin in Pulmonary Arterial Hypertension. Journal of the American Heart Association. 2020; 9: e018349.
- [292] Goncharov DA, Goncharova EA, Tofovic SP, Hu J, Baust JJ, Pena AZ, et al. Metformin Therapy for Pulmonary Hypertension Associated with Heart Failure with Preserved Ejection Fraction versus Pulmonary Arterial Hypertension. American Journal of Respiratory and Critical Care Medicine. 2018; 198: 681–684.
- [293] Chowdhury B, Luu AZ, Luu VZ, Kabir MG, Pan Y, Teoh H, et al. The SGLT2 inhibitor empagliflozin reduces mortality and prevents progression in experimental pulmonary hypertension. Biochemical and Biophysical Research Communications. 2020; 524: 50–56.
- [294] Lee M, Tsai K, Hsu J, Shin S, Wu J, Yeh J. Liraglutide prevents and reverses monocrotaline-induced pulmonary arterial hypertension by suppressing ET-1 and enhancing eNOS/sGC/PKG pathways. Scientific Reports. 2016; 6: 31788.



- [295] Simpson CE, Chen JY, Damico RL, Hassoun PM, Martin LJ, Yang J, et al. Cellular sources of interleukin-6 and associations with clinical phenotypes and outcomes in pulmonary arterial hypertension. The European Respiratory Journal. 2020; 55: 1901761
- [296] Hernández-Sánchez J, Harlow L, Church C, Gaine S, Knight-bridge E, Bunclark K, et al. Clinical trial protocol for TRANS-FORM-UK: a therapeutic open-label study of tocilizumab in the treatment of pulmonary arterial hypertension. Pulmonary Circulation. 2018; 8: 2045893217735820.
- [297] Zamanian RT, Badesch D, Chung L, Domsic RT, Medsger T, Pinckney A, et al. Safety and Efficacy of B-Cell Depletion with Rituximab for the Treatment of Systemic Sclerosis—associated Pulmonary Arterial Hypertension: a Multicenter, Double-Blind, Randomized, Placebo-controlled Trial. American Journal of Respiratory and Critical Care Medicine. 2021; 204: 209–221.
- [298] Benza RL, Gomberg-Maitland M, Demarco T, Frost AE, Torbicki A, Langleben D, et al. Endothelin-1 Pathway Polymorphisms and Outcomes in Pulmonary Arterial Hypertension. American Journal of Respiratory and Critical Care Medicine. 2015; 192: 1345–1354.
- [299] National Research Council. Toward Precision Medicine: Building a Knowledge Network for Biomedical Research and a New Taxonomy of Disease. The National Academies Press: Washington (DC). 2011.
- [300] Rich S, Brundage BH. High-dose calcium channel-blocking therapy for primary pulmonary hypertension: evidence for longterm reduction in pulmonary arterial pressure and regression of right ventricular hypertrophy. Circulation. 1987; 76: 135–141.
- [301] Montani D, Savale L, Natali D, Jais X, Herve P, Garcia G, et al. Long-term response to calcium-channel blockers in

- non-idiopathic pulmonary arterial hypertension. European Heart Journal. 2010; 31: 1898–1907.
- [302] Sitbon O, Humbert M, Jaïs X, Ioos V, Hamid AM, Provencher S, et al. Long-Term Response to Calcium Channel Blockers in Idiopathic Pulmonary Arterial Hypertension. Circulation. 2005; 111: 3105–3111.
- [303] Rich S, Kaufmann E, Levy PS. The Effect of High Doses of Calcium-Channel Blockers on Survival in Primary Pulmonary Hypertension. New England Journal of Medicine. 1992; 327: 76–81.
- [304] Hemnes AR, Trammell AW, Archer SL, Rich S, Yu C, Nian H, et al. Peripheral blood signature of vasodilator-responsive pulmonary arterial hypertension. Circulation. 2015; 131: 401–409.
- [305] Rhodes CJ, Otero-Nunez P, Wharton J, Swietlik EM, Kariotis S, Harbaum L, et al. Whole-Blood RNA Profiles Associated with Pulmonary Arterial Hypertension and Clinical Outcome. American Journal of Respiratory and Critical Care Medicine. 2020; 202: 586–594.
- [306] Harbaum L, Rhodes CJ, Wharton J, Lawrie A, Karnes JH, Desai AA, et al. Mining the Plasma Proteome for Insights into the Molecular Pathology of Pulmonary Arterial Hypertension. American Journal of Respiratory and Critical Care Medicine. 2022; 205: 1449–1460.
- [307] Rhodes CJ, Ghataorhe P, Wharton J, Rue-Albrecht KC, Hadinnapola C, Watson G, et al. Plasma Metabolomics Implicates Modified Transfer RNAs and Altered Bioenergetics in the Outcomes of Pulmonary Arterial Hypertension. Circulation. 2017; 135: 460–475.
- [308] Hemnes AR, Beck GJ, Newman JH, Abidov A, Aldred MA, Barnard J, et al. PVDOMICS: A Multi-Center Study to Improve Understanding of Pulmonary Vascular Disease Through Phenomics. Circulation Research. 2017; 121: 1136–1139.

