Review

The Neglected Internodal Tract—A Cardiac Conduction System Structure Homologous to the Development and Regulation of the Sinoatrial Node

Yuanqin Zhao^{1,2,3}, Juyi Wan^{2,3}, Bin Liao^{2,3}, Man Qi^{1,4,5,6}, C

Academic Editor: Zoltán Papp

Submitted: 4 November 2024 Revised: 5 December 2024 Accepted: 12 December 2024 Published: 22 April 2025

Abstract

The existence of internodal tracts (ITs) is controversial. Indeed, ITs in the cardiac conduction system (CCS), connected to the sinoatrial node (SAN), transmit electrical signals quickly to the left atrium and the atrioventricular node (AVN). Interestingly, research has suggested that the ITs and the tail of the SAN may share developmental homology. Additionally, many studies indicate that ITs blockage can lead to atrial conduction block and is associated with atrial fibrillation (AF). However, few studies have been reported on the morphogenesis, development, and function of ITs. Therefore, this paper aims to review the morphogenesis, development, and function of ITs, focusing on the regulatory mechanisms of transcription factors (TFs), such as NK2 homeobox 5 (NKX2.5), SHOX homeobox 2 (SHOX2), hyperpolarization activated cyclic nucleotide gated potassium channel 4 (HCN4), and T-box transcription factor 3 (TBX3) in the development and morphogenesis of ITs. This review also explores the causes of arrhythmias, especially atrial block, in order to provide new insights into the pathogenesis of CCS disorders.

Keywords: cardiac conduction system; internodal tracts; sinoatrial node; arrhythmia; transcription factors

1. Introduction

According to the Centers for Disease Control and Prevention (CDC), more than 600,000 people in the United States die annually from heart failure or sudden cardiac death [1]. Approximately 50% of patients with heart diseases die from arrhythmias [2]. Studies also revealed that nearly 50% of patients undergoing congenital heart surgery developed arrhythmias after surgery [3]. Arrhythmia mechanisms include abnormalities in impulse formation and/or conduction, among which abnormalities in impulse conduction often present as various conduction blocks, including sinoatrial, atrial, atrioventricular, and ventricular blocks. The cardiac conduction system (CCS) generates and transmits impulses to maintain the heart's rhythmic beats. CCS also consists of specialized myocardial cells that coordinate the contractions of the atria and ventricles to establish the heart's rhythm. Thus, CCS is essential in the formation and normal function of the heart (Fig. 1). Consequently, the dysfunction of any component can lead to serious heart conditions, such as arrhythmias, decreased cardiac output, and even sudden death. Although noted as key components in CCS, the existence, morphogenesis, development and function of internodal tracts (ITs) have rarely been reported systematically. Actually, recent studies have increasingly confirmed the existence of ITs, although their specific anatomy still require further investigation [4,5]. Studies also indicate that functional block of ITs can cause clinical manifestations such as atrial conduction block or atrial fibrillation (AF) [6]. Therefore, this article aims to review the structural composition and development of ITs in CCS as well as transcription factors (TFs) playing regulatory roles in ITs' development for the purpose of laying groundwork for further research into the molecular mechanisms of atrial block and AF and also providing new directions for treating arrhythmias.

2. The Development of Internodal Tracts in Cardiac Conduction System

The CCS consists of ITs with fast conduction, sinoatrial node (SAN), atrioventricular node (AVN) with slow

¹ State Key Laboratory of Cardiovascular Disease, Fuwai Hospital, National Center for Cardiovascular Diseases, Chinese Academy of Medical Sciences and Peking Union Medical College, 100037 Beijing, China

²Department of Cardiovascular Surgery, The Affiliated Hospital, Southwest Medical University, Metabolic Vascular Diseases Key Laboratory of Sichuan Province, Key Laboratory of Cardiovascular Remodeling and Dysfunction, 646000 Luzhou, Sichuan, China

³Key Laboratory of Medical Electrophysiology, Ministry of Education & Medical Electrophysiological Key Laboratory of Sichuan Province (Collaborative Innovation Center for Prevention of Cardiovascular Diseases), Institute of Cardiovascular Research, Southwest Medical University, 646000 Luzhou, Sichuan, China

⁴Chinese People's Liberation Army (PLA) General Hospital, College of Pulmonary & Critical Care Medicine, 100091 Beijing, China

⁵Department of Cardiology, Chinese PLA General Hospital, 100853 Beijing, China

⁶State Key Laboratory of Cardiovascular Diseases and Medical Innovation Center, Shanghai East Hospital, School of Medicine, Tongji University, 200120 Shanghai, China

^{*}Correspondence: wanjuyi@swmu.edu.cn (Juyi Wan); liaobin@swmu.edu.cn (Bin Liao); qimandonkey@163.com (Man Qi)

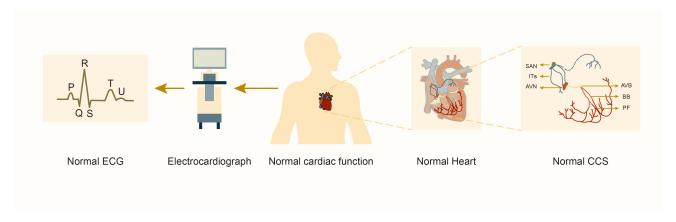


Fig. 1. Structure and function of the normal cardiac conduction system. Abbreviations: SAN, sinoatrial node; ITs, internodal tracts; AVN, atrioventricular node; AVB, atrioventricular bundle; BB, bundle branches; PF, Purkinje fibers; CCS, cardiac conduction system; ECG, electrocardiograph.

conduction and specialized myocardial cells such as the atrioventricular bundle (AVB, or "His bundle"), left and right bundle branches, and Purkinje fibers (Fig. 1). These cells coordinate the rhythmic contractions of the atria and ventricles to establish the heart's rhythmic beating. Studies of lineage tracking and electrophysiological characteristics of CCS cells reveal that these cells don't belong to the same type. Different cell types exhibit significant heterogeneity in morphology, function and molecular characterization [7]. SAN in the right atrium initially generates electrical pulse, depolarizing adjacent atrial myocardial cells before rapidly propagating through ITs to AVN and the left atrium. AVN delays impulse transmission, slowing the electrical pulse speed by approximately 0.04 seconds. This delay allows the atrium to contract and pump blood into the ventricle before it contracts, ensuring the sequential contraction of the atrial and ventricular muscles and adapting to intracardiac blood circulation. Subsequently, the electrical pulse rapidly travels through the atrioventricular bundle to the left and right bundle branches and the Purkinje fiber network, activating ventricular myocardial contractions [8].

In cardiac development, the first heart field (FHF) and the second heart field (SHF) are critical embryonic regions derived from distinct populations of cardiac progenitor cells. The temporal sequence of their involvement defines their contributions to the cardiac lineage. The FHF primarily contributes to the formation of the left ventricle (LV), whereas the SHF promotes the right ventricle (RV) and outflow tract (OFT). Both fields also play essential roles in developing the atria and septum [9]. Using specific markers such as Hcn4 (hyperpolarization-activated cyclic nucleotide gated K⁺ 4, in FHF), Isl1 (ISL1 transcription factor, LIM/homeodomain, in SHF) and Tbx18 (T-box18, in the venous pole of the posterior heart field), CCS lineage tracing in mice revealed that AVN, His bundle and Purkinje fibers express only Hcn4, but not Isl1 or Tbx18 [10-12]. This suggests that the AVN, His bundle, and Purkinje fibers primarily develop from the FHF. Further, experiments tracking Tbx18 expression revealed that cells in the SAN head express both Tbx18 and Isl1, while the SAN junction/tail and ITs express only Isl1 [12]. Results also indicate that cells in the SAN head may originate from the posterior heart field in the venous pole of the second heart field, while cells in the ITs and SAN junction originate from the SHF [10–12]. Studies using CCS–lacZ transgenic mice described three ITs connecting the SAN and AVN, as well as the embryonic origins of the SAN and ITs [13,14]. Subsequent studies on the expression patterns of *Hcn4* and *Tbx3* (CCS markers) in mouse embryonic heart development suggested that development of ITs may occur simultaneously with the SAN junction [15,16]. Morphogenesis might begin at embryonic day (E) 9.5 and be completed by E12.5, with lacZ and Tbx3 expression continuing at least until E17.5. Nakashima et al. [17] found that NK2 homeobox 5 (Nkx2-5, also named Nkx2.5) suppresses the proliferation of atrial working myocytes and confines SAN, AVN, and ITs. In addition, other laboratories have found that Shox2 is expressed as early as E8.5 in mouse embryos on the dorsal side of the primitive cardiac tube; on E9.5, Shox2 is expressed in the transition zone between the venous sinus and the total atrium; on E10.5, Shox2 is most strongly expressed in the venous sinus and venous valve; on E11.5, in situ hybridization of the entire heart showed that Shox2 expression expanded to include the SAN, venous valves and ITs [14,18-20]. This suggests that ITs and SAN junction development may share similar regulatory mechanisms (Fig. 2).

3. The Morphology of Internodal Tracts

There is ongoing controversy regarding the existence of specialized conduction tissues, known as ITs, between the SAN and AVNs in the CCS. Early theories suggested that these conduction bundles were crucial for rapidly transmitting electrical signals [21]. However, due to the low expression of gap junction proteins in these bundles, they were once thought to have little or degraded signal conduction function and were considered less significant in ma-



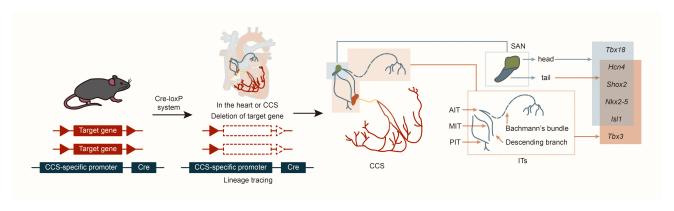


Fig. 2. Structure and lineage tracing of sinoatrial node and internodal tracts. Abbreviations: Cre, cyclization recombinase; AIT, anterior internodal tract; MIT, middle internodal tract; PIT, posterior internodal tract; *Tbx18*, T-box18; *Hcn4*, hyperpolarization activated cyclic nucleotide gated K⁺ 4; *Shox2*, SHOX homeobox2; *Nkx2-5*, NK2 homeobox 5; *Isl1*, ISL1 transcription factor, LIM/homeodomain; *Tbx3*, T-box 3.

ture hearts [16]. Some researchers argue that electrical signals are transmitted radially along the atrial muscle, relying on the arrangement of myocardial cells and gap junction proteins rather than on specialized conduction bundles. Clinical observations of dynamic electrocardiograms have shown that atrial contraction time is slightly prolonged during sinus excitation compared to ectopic excitation. This has led to skepticism about specialized atrial conduction pathways [22]. However, whether these conduction pathways were because of the presence of specialized conduction tissue or because of the anisotropic orientation of the muscle fibers remains controversial [6].

Nevertheless, numerous electrophysiological and biochemical studies have confirmed the existence of intraatrial conduction bundles. Certain myocardial cells in the atrium, which differ in the electrophysiological aspect, connect SAN and AVN and are termed ITs [23]. In 1963, Thomas N. James [24] identified three conduction tracts connecting the SAN and the AVN, named the anterior internodal tract (AIT), middle internodal tract (MIT), and posterior internodal tract (PIT), based on their anatomical locations. The AIT, which includes part of the Bachmann bundle (BB), is the most complex (Fig. 2) and originates from the anterior edge of the SAN, extending towards the left front, arching around the superior vena cava (SVC) and the anterior wall of the right atrium. The AIT splits into two bundles: one extends through the atrial sulcus into the left atrium, known as the BB, connecting the left and right atrial walls; the other descends diagonally behind the aortic root to the upper edge of the AVN, known as the descending branch. The MIT, or Wenckebach's bundle, originates from the posterior upper edge of the SAN, arches to the right, passes behind the superior vena cava, and descends along the right side of the atrial septum into the upper edge of the AVN. The PIT, or Thorel's bundle, extends from the posterior edge of the SAN along the terminal ridge to the inferior vena cava valve (Eustachian's ridge), then passes slightly above the coronary sinus to the posterior edge of the AVN before extending rapidly downward into it. During its course, the PIT branches out and spreads along the right atrial comb muscle to the back of the right atrium [25]. The anterior, middle, and posterior ITs converge near the AVN, dividing into two parts: a small portion of the AIT and MIT fibers, along with a large portion of the PIT fibers, enter the lower part of the AVN; the majority of AIT and MIT fibers and a small portion of the PIT fibers terminate at the posterior upper edge of the AVN [26]. The ITs consists of myocardial cells, Purkinje fibers, and a few transitional cells characterized by their compact structure, extending from the oval fossa to the AVN through Todaro's tendon. This cell population is confined to the left atrial septum [27].

4. Manifestations of Dysfunction in Internodal Tracts

The ITs play a crucial role in the CCS by rapidly transmitting electrical signals from SAN to left atrium and AVN [4]. Generally, electrical impulse generated by the SAN is transmitted through the AIT, MIT and PIT (including the BB) to the AVN and the left atrium. Thus, blockage or damage to the ITs can result in internodal or interatrial conduction block, which is associated with electromechanical dysfunction of the left atrium and initiation of various atrial arrhythmias [28-30]. In an experiment with dogs, Bachmann [31] first described damage to the apical muscle bundle in the atrial septum connecting the left and right atrial appendages, resulting in interatrial conduction block. Waldo et al. [32] produced surgical lesions in the left and right atria portions of the BB in the canine heart, and subsequently, delayed conduction of the BB led to partial interatrial block (IAB) according to electrocardiogram observations, while a complete block of the BB led to advanced IAB. Studies indicate that anatomic sites associated with the occurrence of atrial arrhythmia include the atrial septum (where the MIT is located) and crista terminalis (CT, where the PIT runs through) in adult hearts. These areas are critical for transmitting electrical signals



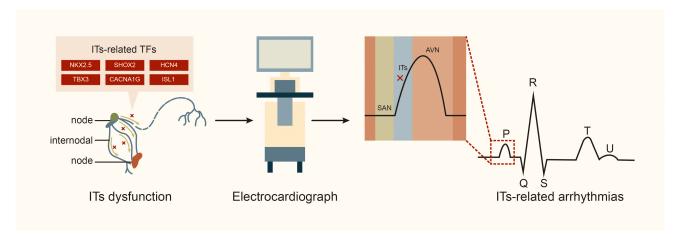


Fig. 3. Internodal tracts dysfunction and related transcription factors. Abbreviations: TFs, transcription factors; STBX3, T-box transcription factor 3; HCN4, hyperpolarization activated cyclic nucleotide gated potassium channel 4; SHOX2, SHOX homeobox 2; NKX2.5, NK2 homeobox 5; ISL1, ISL LIM homeobox 1; CACNA1G, calcium voltage-gated channel subunit alpha1 G.

from SAN to AIT in the left atrium [6,14]. A study of the Finnish national population, including 8028 individuals aged 30 and older (of whom 6354 underwent health examinations, including electrocardiograms, and were followed for 15 years) [33], found that approximately 10.7% of patients had symptoms of IAB; the high incidence of IAB in the population suggests that the number of people with ITs conduction disorder may be underestimated. Furthermore, interatrial conduction block is a risk factor for conditions such as supraventricular arrhythmia (including AF), stroke, thromboembolism, sinus node dysfunction, and transient ischemic attacks [33,34]. Additionally, about half of patients have a history of recurrent paroxysmal AF or atrial flutter, and 40% may experience atrial premature beats and atrial tachycardia [27,35,36]. The Wenckebach bundle is associated with the induction of AF, while the unique electrophysiological characteristics of the BB have been observed in patients with AF [14]. In patients with atrial septal defect, the maze procedure can lead to atrial conduction block and AF due to blockage of the AIT and MIT by the defect, while the right lower atrial incision and intermediate septal incision block the PIT [37]. In a study exploring the relationship between damage to the internodal conduction pathway and supraventricular arrhythmia in dogs, conduction time and synchronous activation sequences from the SAN to the vicinity of the AVN were measured using endocardial mapping. The study found that injury to the AIT during cardiopulmonary bypass surgery, regardless of location or extent, significantly prolonged the conduction time from the SAN to the vicinity of the AVN by approximately 15 milliseconds. Additionally, separation of the AIT and PIT, or all the three tracts, can lead to sinus arrhythmias. About 50% of dogs also experience bradycardia, including atrioventricular nodal arrhythmias [38], highlighting the crucial role of internodal tracts in maintaining normal cardiac conduction function (Fig. 3).

Studies have also found that atrial muscle contraction ceases in a high potassium ion environment, and the P wave on the electrocardiogram disappears; however, the ITs continue to transmit impulses from the SAN to the AVN and maintain contractions [25]. Except for the ITs, electrical signal transmission in other atrial cells is completely lost [39], indicating that ITs have greater resistance to high potassium ion environments than general atrial cardiomyocytes.

5. Transcription Factors and Internodal Tracts

Research has identified that TFs such as NKX2.5 [17], SHOX2 [40,41], HCN4 [42], and TBX3 [16] are expressed in the ITs. Additionally, calcium voltage-gated channel subunit alpha1 G (CACNA1G) and ISL1 are crucial in the development of CCS (Table 1, Ref. [17,41–46]).

5.1 NKX2-5 and Internodal Tracts

NKX2-5 is a member of the NK2 family of homeobox genes in humans and is homologous to the tinman gene found in Drosophila melanogaster. Furthermore, NKX2-5 plays a critical role in cardiac morphogenesis by regulating transcription of various genes involved in this process [47]. Clinically, NKX2-5 haploinsufficiency is associated with progressive heart defects, such as atrioventricular block, atrial septal defect, and ventricular septal defect [48]. Studies knocking out the Nkx2-5 gene in mice have shown that the loss of Nkx2-5 leads to severe cardiac developmental abnormalities, resulting in death around embryonic day 10.5 (E10.5). This underscores the essential role NKX2-5 has in cardiac structure development. However, the precise role of Nkx2-5 in the CCS remains poorly understood, partly due to the premature death of Nkx2-5 knockout mice, which hinders the analysis of tissue development in the SHF [49]. Research on atrial Nkx2-5 knockout mice [17] revealed three *Hcn4*-positive ITs between the



Table 1. The discovery of transcription factors possibly associated with ITs and their regulatory function.

Author	Year	Transcription factors of ITs	Coding gene	Function of transcription factors
Nakashima et al. [17]	2014	NKX2.5	Nkx2-5	Inhibits ITs morphogenesis and conductivity
Sun et al. [41]	2013	SHOX2	Shox2	Regulates the development and maintenance
				of ITs
Vicente-Steijn et al. [42]	2011	HCN4	Hcn4	Maintains normal rhythm and regulates ITs
				development
Soufan et al. [43]	2004	TBX3	Tbx3	Prevents ITs from developing into ventricular
				and atrial myocardium
Baudot et al. [44]	2020	CACNA1G	Cacna1g	Increases the speed of conduction from SAN
				to ITs
Cai et al. [45]	2003	ISL1	Isl1	Regulates the ITs development and function
Ye et al. [46]	2015	NKX2.5 and SHOX2	Nkx2-5 and Shox2	Co-regulates the cell fate of SAN and ITs

SAN and the AVN. Additionally, Nkx2-5 knockout led to a significant increase of the *Hcn4*-positive area and thickening of these tracts, whereas the Hcn4-positive ITs in the control group were very thin. In the Nkx2-5 conditional knockout and control groups, Connexin 40 (Cx40) in the ITs was negative, indicating that these tracts only maintained slow conductivity. Mice with atrial-specific deficiency of Nkx2-5 frequently exhibit major atrial abnormalities, such as atrial septal defects (ASDs), supraventricular bradvcardia, and atrial conduction block. These abnormalities are attributed to the proliferation of myocardial working cells and conduction cells. Genetic analysis indicated that Notch signaling promotes atrial myocardium proliferation in mouse embryos, while transcriptome analysis showed that Nkx2-5 inhibits Notch activity in the atrium [17]. These findings suggest that Nkx2-5 and Notch signaling work together to regulate the proliferation of atrial working and conduction cardiomyocytes. Thus, transcription factor NKX2.5 plays an inhibitory role in the morphogenesis and conductivity of ITs, and they help to prevent the development of these tracts into atrial myocardial tissue by inhibiting Notch signaling. Therefore, NKX2.5 is crucial for both cardiac structure formation and normal function of ITs.

5.2 SHOX2 and Internodal Tracts

SHOX2 is a homologous domain transcription factor that promotes the activation of *short stature homeobox* 2 (*Shox2*) gene, a member of the homeobox gene family known as *SHOX2* in humans. Mouse *Shox2* and human *SHOX2* share up to 99% homology in amino acid sequences [50]. SHOX2 is a key regulatory factor in the development of CCS, participating in heart rate regulation, and is genetically related to atrial arrhythmias. Its deficiency can lead to bradycardia [51]. Researchers also confirmed that the *SHOX2* gene in humans is a susceptibility gene for AF [52,53]. In humans, *SHOX2* is located on chromosome 3q25.32 and encodes three protein isoforms: isoform a (331 amino acids), isoform b (355 amino acids), and isoform c (319 amino acids). Previous research has established that SHOX2 regulates the expression of many car-

diac genes through transcription, including Hcn4, Nkx2-5, Cx40, Connexin 43 (Cx43), natriuretic precursor peptide A (Nppa), Isl1, and bone morphogenetic protein 4 (Bmp4) [54,55]. Meanwhile, findings by Wang et al. [56] show that paired-like homeodomain transcription factor 2 (Pitx2) upregulates the miR-17-92/106b-25 cluster, which inhibits Shox2, leading to restricted sinus node development. The absence of this miRNA cluster in mice results in sinus node dysfunction, prolonged PR interval, and increased susceptibility to AF. Studies have also found that AF patients carrying the SHOX2 3'-UTR c.*28T>C variant (where thymine at the 28th base in the 3' non-coding region of the SHOX2 gene is replaced by cytosine) have significantly longer PR intervals compared to patients with wild-type alleles [52]. Additionally, patients with the c.580C>T variant (where thymine replaces cytosine at the 580th base) may produce a p.R194X truncated protein (with only 193 amino acids), leading to sinus bradycardia, as well as a significantly prolonged PR interval, RR interval, and QRS wave [53]. These studies indicate that missense mutations in both coding and non-coding regions of the SHOX2 gene can significantly reduce or even eliminate SHOX2 expression, and the resulting arrhythmia symptoms can be stably inherited. *Isl1*, a specific marker for the second cardiac region, is activated by wild-type Shox2 during transcription. In addition, the Shox2 R194X mutant mice fail to activate Isl1, demonstrating a direct transcriptional connection between Shox2 and the ITs [53,57]. In mouse models, homozygous Shox2 deletion leads to increased mortality between E11.5 and E13.5 due to severe cardiovascular defects, including bradycardia, underdeveloped sinus node and valve, and abnormal expressions of Cx40, Cx43, and Nkx2-5. The mortality rate can be mitigated with closely related hypomorphic *Shox* allele replacing *Shox2*, however the arrhythmias persist [19,58]. Early in embryonic cardiac development, Shox2 is expressed in the SAN, right venous valve, and dorsal mesenchymal protrusion of the heart [19,40]. Sun et al. [40,41] have also observed Shox2 expression in the ITs, which share an origin with the right venous valve. Conditional knockout of Shox2 in mice results in incomplete



development of the SAN junction and the MIT and PIT, causing symptoms such as atrial block and bradycardia in adult mice. Further, *Hcn4* expression in the AIT disappears while the AIT morphology remains unaffected, suggesting potential alterations in ITs' conductivity. These findings indicate that the deletion of the *SHOX2* gene or a mutation may lead to defects in the morphology, development and function of the SAN or ITs. SHOX2 is thus a critical factor in regulating the development and maintenance of the entire CCS, especially the SAN and the ITs.

5.3 HCN4 and Internodal Tracts

HCN4 is a subtype of hyperpolarization-activated cyclic nucleotide-gated (HCN) channels, acting as a pacing channel that is highly expressed during sinus node development. Studies have shown that Hcn4 is a marker for the first cardiac region and cardiac pacing cells. During different stages of cardiac development, Hcn4 is dynamically expressed in various differentiated cardiomyocyte precursors, including those in the CCS that produce different components essential for cardiac function [10]. In studies on chicken embryonic CCS development, HCN4 expression was found to be restricted to the SAN, ITs, AVN, bundle branch, and atrioventricular annulus myocardium, making it as a CCS development marker in chickens [42]. Meanwhile, changes in HCN4 channel function are associated with sinus node dysfunction and arrhythmias such as AF, ventricular tachycardia, and atrioventricular block [59]. Complete or heart-specific ablation of *Hcn4* in mice impairs embryonic development, resulting in death between E9.5 and E11.5 [60]. Cardiac specific *Hcn4* knockout mice exhibit severe bradycardia and atrioventricular block, leading to cardiac arrest and death [61], highlighting the critical role of Hcn4 in maintaining normal rhythm. Thus, the expression pattern of HCN4 during development may indicate potential arrhythmia targets in hearts and has connections with the development and function of the ITs.

5.4 TBX3 and Internodal Tracts

The T-box factor is part of an ancient protein family consisting of evolutionarily conserved transcription factors crucial for embryonic development. T-box 3 (Tbx3), a member of this family, is expressed in various tissues and plays a key regulatory role in organs such as the heart, breast, and limbs [62]. Soufan et al. [43] have identified TBX3 as a transcription inhibitor in the internodal region. Tbx3 is continuously expressed throughout cardiac development from the SAN to the atrioventricular region, starting from E8.5. The expression regions coincide with regions showing higher levels of Nppa and Cx40, which are markers of faster conduction velocity [63-65]. The peripheral ventricular conduction network, marked by rapid conduction and expression of ventricular-specific myocardial markers, contrasts with the CCS, which features slower conduction and lower expression of these markers [20,65]. Tbx3 and Tbx2 share structural and functional similarities,

including inhibitory characteristics and DNA recognition elements [66]. From early embryonic stages, Tbx2 expression decreases while Tbx3 expression increases in the heart. Development of the CCS occurs within the Tbx2 expression regions, where TBX2 inhibits ventricular formation by suppressing ventricular type-specific myocardial markers Nppa and Cx40 [67,68]. This indicates that the CCS develops in regions where ventricular formation is inhibited. In another three-dimensional reconstruction study on mouse heart development, Tbx3 was selectively expressed throughout the CCS, including in the ITs [43]. TBX3 inhibits the promoter activity of Nppa and Cx40 and blocks the synergistic activation of the Nppa promoter by TBX5 and NKX2.5, thus contributing to the sustained inhibition of ventricular phenotype development in the CCS-derived myocardium [16]. Additionally, TBX3 has been shown to bind and inhibit atrial genes, such as the Cx43 gene, thereby suppressing the working atrial myocardial phenotype [69]. These findings suggest that the TBX3 transcription factor plays a crucial role in the development of ITs and may prevent their differentiation into the ventricular or atrial myocardium.

5.5 CACNAIG and Internodal Tracts

Recent studies have confirmed the presence of the CACNA1G transcription factor in the CCS. CACNA1G encodes the pore-forming subunit of the low-voltage activated T-type calcium channel (Ca_v3.1) channel, which is expressed in various regions of the central nervous system, particularly in Purkinje neurons and deep cerebellar nuclear. The Ca_v3.1 channel is characterized by low activation voltage, relatively small single-channel conductivity, and rapid deactivation [70]. Further, the Ca_v3.1 channel is believed to be associated with pacemaker activity, lowthreshold calcium spikes, neuronal oscillation, resonance, and rebound pulse discharge. Although Ca_v 3.1 is expressed in the embryonic hearts of various mammals, its expression decreases with the progress of development [71]. The functional role of Ca_v3.1 has been confirmed in rodents; mice lacking Ca_v3.1 channels exhibited reduced pacemaker activity and worsened atrioventricular conduction compared to wild-type mice [72]. Ca_v3.1 mRNA and protein have also been detected in the CCS of humans [73]. Baudot et al. [44] demonstrated that Cacnalg expression is linked to the autonomy of pacemaker cells, whereby increased Cacnalg expression correlated with increased conduction velocity from the SAN to the ITs while pacing conduction velocity decreased upon reaching the AVN. This suggests that the ITs are the dominant conduction pathway from the SAN to the AVN and may be associated with atrioventricular delay.

5.6 ISL1 and Internodal Tracts

The ISL1 transcription factor is crucial for the development of the CCS. The *Isl1* gene encodes a transcription factor with a LIM homeodomain. *Isl1* is expressed in adult mammalian cardiac stem cells and progenitor cells, primar-



ily in the outflow tract, right ventricle, and parts of the atrium. This area, known as the second cardiac region, is considered a convergence place for cardiac progenitor cells; therefore, deletion of this gene results in abnormal atrioventricular formation [45]. Homozygous *Isl1* mutants exhibit growth retardation around E9.5–10 and death around E10.5–11. Histological analysis showed severe cardiac developmental defects in these mutants, including the absence of the right ventricle and outflow tract and a common atrium [45]. Additionally, *Isl1* is a direct transcriptional target of *Shox2*, which can rescue *Shox2*-mediated bradycardia [57]. During SAN development, *Shox2* regulates *Isl1* expression, suggesting that *Isl1* is crucial for SAN development and may influence ITs' development and function, potentially impacting atrioventricular delay.

5.7 Potential Co-regulation of Internodal Tracts by SHOX2 and NKX2.5

Research by Ye et al. [46] on the SAN revealed that it can be divided into two distinct regions: the $Shox2^+/Hcn4^+$ SAN head and the $Shox2^+/Nkx2-5^+/Hcn4^+$ SAN junction. The specific knockout of *Shox2* at the SAN junction results in an underdeveloped SAN junction and ectopic expression of the working myocardial marker Cx40 in cells at this junction. This indicates that Shox2 knockout can cause a shift of cell fate from SAN cells to working myocardial cells. Additionally, an intriguing antagonistic interaction between Shox2 and Nkx2-5 was observed. Conditional knockout of the Shox2 gene at Nkx2-5 expression sites and subsequent reduction of Nkx2-5 expression partially reversed the SAN junction defects caused by Shox2 knockout, allowing for the re-expression of *Hcn4*, *Tbx3*, and *Isl1* at the SAN junction. This suggests a reversion of cell fate from working myocardial cells to $Shox2^+/Nkx2-5^+$ pacemaker cells. These findings highlight how the Shox2-Nkx2-5 antagonistic mechanism can alter cell fate in the SAN and regulate its morphology. Given that the morphogenesis of both the ITs and the SAN junction starts simultaneously from the second cardiac region [10,11,74], it is reasonable to hypothesize that the ITs are also co-regulated by *Shox2* and *Nkx2-5*.

Furthermore, mutations in the human Notch and Ras-MAPK (mitogen-activated protein kinase) signal pathways are crucial for myocardial cell proliferation and can lead to atrial block and atrial septal defect (ASD) [75]. Indeed, it was confirmed that *Shox2* is a target gene of the pituitary homeobox gene (*Pitx2*) [56]. Meanwhile, one *Pitx2* subtype, *Pitx2c*, directly binds to and inhibits *Shox2* expression. Haploid dysfunction of *Pitx2c* predisposes mice to atrial arrhythmias [69], suggesting that Notch, Ras-MAPK signal pathways and *Pitx2* may play roles in the developmental regulation of internodal tracts.

6. Expectations

As a part of the CCS, the ITs are extremely important for studying cardiovascular diseases, especially in exploring the pathogenic mechanisms of congenital or acquired heart conduction system-related diseases. However, there are still limitations in current understanding, with limited reports on the existence, morphogenesis, developmental process, and function of ITs. Firstly, Ye et al. [46] found a mutually antagonistic molecular mechanism between Nkx2-5 and Shox2, which has potential regulatory effects on the morphogenesis of SAN junction and the fates of working myocardial cells and pacemaker cells. As the morphogenesis process of the SAN junction and ITs begin simultaneously and both originate from the second cardiac region, it is reasonable to speculate that Nkx2-5 and Shox2 may jointly regulate ITs. However, more experiments are still needed for proof. Secondly, the regulatory mechanisms of transcription factors such as HCN4, TBX3, and PITX2 on the morphogenesis and function of ITs have not been fully elucidated, and the relationship between *Hcn4*, *Tbx3*, *Nkx2-5*, Shox2 and the regulation of ITs still needs further exploration. Thirdly, transcription factors such as CACNA1G and ISL1 are closely related to the development of CCS and may also be involved in the regulation of ITs' development and function. Lastly, other complex molecular mechanisms may exist in the regulation of ITs, which require further exploration.

7. Conclusions

Although significant progress has been made in the study of ITs, the developmental mechanisms, formation processes, and specific functions of these tracts still need to be better understood, particularly the regulatory mechanisms, where various research views exist. Research on ITs has reshaped our understanding of CCS and offers a theoretical foundation for the onset of certain arrhythmias. However, the exact role of ITs in CCS and their contributions to the pathogenesis of cardiovascular diseases, especially arrhythmias, remains to be fully elucidated. In conclusion, ITs warrant increased attention, and further research investment is essential to advance our understanding of this crucial apparatus, ultimately driving progress in cardiology and clinical medicine.

Abbreviations

AIT, anterior internodal tract; CCS, cardiac conduction system; ITs, internodal tracts; MIT, middle internodal tract; PIT, posterior internodal tract; SAN, sinoatrial node.

Author Contributions

YZ performed conceptualization and wrote original draft. BL and JW performed methodology, funding acquisition, supervision and revised the manuscript critically for important intellectual contents. MQ performed conceptualization, funding acquisition, review, and editing. All authors read and approved the final manuscript. All authors have participated sufficiently in the paper writing and agreed to be accountable for all aspects of the article.



Ethics Approval and Consent to Participate

Not applicable.

Acknowledgment

Not applicable.

Funding

This work was financially supported by The National Key Research and Development Program of China (grant 2021YFC2701700); the China Postdoctoral Science Foundation (grant 2024T170070), National Natural Science Foundation of China (82400381), Open Project Fund of State Key Laboratory of Cardiovascular Diseases, Shanghai East Hospital (2024SKL-TJ001), Chinese PLA General Hospital Sixth medical center Innovation and Culture Grant (CXPY202304) and Key Lab Construction Grant of PCCM college (XXB2024-1), Open Project Fund of National Clinical Research Center for Geriatric Diseases (NCRCG-PLAGH-2024008) to MQ; National Natural Science Foundation of China (82070277 to BL, 82170325 to JW), Science and Technology Strategic Cooperation Programs of Luzhou Municipal People's Government and Southwest Medical University (2024LZXNYDJ004 to BL, 2024LZXNYDJ028 to JW), the Central Government Funds of Guiding Local Scientific and Technological Development (2023ZYD0069) to BL; Natural Science Foundation of Sichuan Province of China (2024NSFSC2085), Sichuan Province science and technology projects (2024YFFK0042), Science and Technology Strategic Cooperation Programs of Luzhou Municipal People's Government and Southwest Medical University (2024LZXNYDJ028) to JW.

Conflict of Interest

The authors declare no conflict of interest.

References

- [1] Ahmad FB, Cisewski JA, Anderson RN. Mortality in the United States - Provisional Data, 2023. MMWR. Morbidity and Mortality Weekly Report. 2024; 73: 677–681. https://doi.org/10. 15585/mmwr.mm7331a1.
- [2] Evbayekha E, Antia A, Dixon B, Reiss C, LaRue S. Predictors of mortality and burden of arrhythmias in endstage heart failure. Current Problems in Cardiology. 2024; 49: 102541. https://doi. org/10.1016/j.cpcardiol.2024.102541.
- [3] Alp H, Narin C, Baysal T, Sarıgül A. Prevalence of and risk factors for early postoperative arrhythmia in children after cardiac surgery. Pediatrics International: Official Journal of the Japan Pediatric Society. 2014; 56: 19–23. https://doi.org/10.1111/ped.12209.
- [4] Anderson RH, Sánchez-Quintana D, Spicer DE, Farré J, Back Sternick E. How does the cardiac impulse pass from the sinus to the atrioventricular node? Heart Rhythm. 2022; 19: 1738–1746. https://doi.org/10.1016/j.hrthm.2022.05.032.
- [5] Cavero I, Holzgrefe H. Internodal conduction pathways: revisiting a century-long debate on their existence, morphology, and location in the context of 2023 best science. Advances in Physiology Education. 2023; 47: 838–850. https://doi.org/10.1152/advan.00029.2023.

- [6] van Campenhout MJH, Yaksh A, Kik C, de Jaegere PP, Ho SY, Allessie MA, et al. Bachmann's bundle: a key player in the development of atrial fibrillation? Circulation. Arrhythmia and Electrophysiology. 2013; 6: 1041–1046. https://doi.org/10.1161/CIRCEP.113.000758.
- [7] Goodyer WR, Beyersdorf BM, Paik DT, Tian L, Li G, Buikema JW, et al. Transcriptomic Profiling of the Developing Cardiac Conduction System at Single-Cell Resolution. Circulation Research. 2019; 125: 379–397. https://doi.org/10.1161/CIRCRE SAHA.118.314578.
- [8] Tan SY, Fritsch MK, White S, Arva NC. Dissecting the Cardiac Conduction System: Is It Worthwhile? Pediatric and Developmental Pathology: the Official Journal of the Society for Pediatric Pathology and the Paediatric Pathology Society. 2020; 23: 413–423. https://doi.org/10.1177/1093526620944756.
- [9] Miyamoto M, Gangrade H, Tampakakis E. Understanding Heart Field Progenitor Cells for Modeling Congenital Heart Diseases. Current Cardiology Reports. 2021; 23: 38. https://doi.org/10. 1007/s11886-021-01468-5.
- [10] Liang X, Wang G, Lin L, Lowe J, Zhang Q, Bu L, et al. HCN4 dynamically marks the first heart field and conduction system precursors. Circulation Research. 2013; 113: 399–407. https:// doi.org/10.1161/CIRCRESAHA.113.301588.
- [11] Kelly RG, Buckingham ME, Moorman AF. Heart fields and cardiac morphogenesis. Cold Spring Harbor Perspectives in Medicine. 2014; 4: a015750. https://doi.org/10.1101/cshpersp ect.a015750.
- [12] Meilhac SM, Lescroart F, Blanpain C, Buckingham ME. Cardiac cell lineages that form the heart. Cold Spring Harbor Perspectives in Medicine. 2014; 4: a013888. https://doi.org/10.1101/cs hperspect.a013888.
- [13] Blom NA, Gittenberger-de Groot AC, DeRuiter MC, Poelmann RE, Mentink MM, Ottenkamp J. Development of the cardiac conduction tissue in human embryos using HNK-1 antigen expression: possible relevance for understanding of abnormal atrial automaticity. Circulation. 1999; 99: 800–806. https://doi.org/10.1161/01.cir.99.6.800.
- [14] Jongbloed MRM, Schalij MJ, Poelmann RE, Blom NA, Fekkes ML, Wang Z, et al. Embryonic conduction tissue: a spatial correlation with adult arrhythmogenic areas. Journal of Cardiovascular Electrophysiology. 2004; 15: 349–355. https://doi.org/10.1046/j.1540-8167.2004.03487.x.
- [15] Wang Y, Lu P, Jiang L, Wu B, Zhou B. Control of sinus venous valve and sinoatrial node development by endocardial NOTCH1. Cardiovascular Research. 2020; 116: 1473–1486. https://doi.org/10.1093/cvr/cvz249.
- [16] Hoogaars WMH, Tessari A, Moorman AFM, de Boer PAJ, Hagoort J, Soufan AT, et al. The transcriptional repressor Tbx3 delineates the developing central conduction system of the heart. Cardiovascular Research. 2004; 62: 489–499. https://doi.org/10.1016/j.cardiores.2004.01.030.
- [17] Nakashima Y, Yanez DA, Touma M, Nakano H, Jaroszewicz A, Jordan MC, et al. Nkx2-5 suppresses the proliferation of atrial myocytes and conduction system. Circulation Research. 2014; 114: 1103–1113. https://doi.org/10.1161/CIRCRESAHA
- [18] Espinoza-Lewis RA, Yu L, He F, Liu H, Tang R, Shi J, *et al.* Shox2 is essential for the differentiation of cardiac pacemaker cells by repressing Nkx2-5. Developmental Biology. 2009; 327: 376–385. https://doi.org/10.1016/j.ydbio.2008.12.028.
- [19] Blaschke RJ, Hahurij ND, Kuijper S, Just S, Wisse LJ, Deissler K, et al. Targeted mutation reveals essential functions of the homeodomain transcription factor Shox2 in sinoatrial and pacemaking development. Circulation. 2007; 115: 1830–1838. https://doi.org/10.1161/CIRCULATIONAHA.106.637819.
- [20] Rentschler S, Vaidya DM, Tamaddon H, Degenhardt K, Sassoon D, Morley GE, et al. Visualization and functional charac-



- terization of the developing murine cardiac conduction system. Development (Cambridge, England). 2001; 128: 1785–1792. https://doi.org/10.1242/dev.128.10.1785.
- [21] Ho SY, Anderson RH, Sánchez-Quintana D. Atrial structure and fibres: morphologic bases of atrial conduction. Cardiovascular Research. 2002; 54: 325–336. https://doi.org/10.1016/ s0008-6363(02)00226-2.
- [22] Shaoxiang D. Discussion on the rationality of internodal bundle. National Medical Frontiers of China. 2009; 18: 9-9,33. (In Chinese) https://doi.org/10.3969/j.issn.1673-5552.2009.18.006.
- [23] Liebman J. Are there internodal tracts? Yes. International Journal of Cardiology. 1985; 7: 175–185. https://doi.org/10.1016/0167-5273(85)90362-6.
- [24] James TN. The connecting pathways between the sinus node and AV node and between the right and the left atrium in the human heart. American Heart Journal. 1963; 66: 498–508. https://doi.org/10.1016/0002-8703(63)90382-x.
- [25] Holsinger JW, Jr, Wallace AG, Sealy WC. The identification and surgical significance of the atrial internodal conduction tracts. Annals of Surgery. 1968; 167: 447–453. https://doi.org/10.1097/ 00000658-196804000-00001.
- [26] James TN. Anatomy of the A–V node of the dog. The Anatomical Record. 1964; 148: 15–27. https://doi.org/10.1002/ar.1091480103.
- [27] Jain S, Mirza S, Shukla G. Inadvertent atrial dissociation following catheter ablation: A demonstration of cardiac anisotropy and functional block. HeartRhythm Case Reports. 2019; 5: 201–204. https://doi.org/10.1016/j.hrcr.2018.12.010.
- [28] Ariyarajah V, Spodick DH. The Bachmann Bundle and interatrial conduction. Cardiology in Review. 2006; 14: 194–199. https://doi.org/10.1097/01.crd.0000195221.26979.2b.
- [29] Knol WG, Teuwen CP, Kleinrensink GJ, Bogers AJJC, de Groot NMS, Taverne YJHJ. The Bachmann bundle and interatrial conduction: comparing atrial morphology to electrical activity. Heart Rhythm. 2019; 16: 606–614. https://doi.org/10.1016/j.hr thm.2018.10.021.
- [30] Carmona Puerta R. Interatrial blocks: diagnosis and clinical significance. Medicina Clinica. 2020; 155: 207–214. https://doi.or g/10.1016/j.medcli.2020.03.011.
- [31] Bachmann G. The inter-auricular time interval. American Journal of Physiology-Legacy Content. 1916; 41: 309–320.
- [32] Waldo AL, Bush HL, Jr, Gelband H, Zorn GL, Jr, Vitikainen KJ, Hoffman BF. Effects on the canine P wave of discrete lesions in the specialized atrial tracts. Circulation Research. 1971; 29: 452–467. https://doi.org/10.1161/01.res.29.5.452.
- [33] Istolahti T, Eranti A, Huhtala H, Lyytikäinen LP, Kähönen M, Lehtimäki T, et al. The prevalence and prognostic significance of interatrial block in the general population. Annals of Medicine. 2020; 52: 63–73. https://doi.org/10.1080/07853890. 2020.1731759.
- [34] A Reiffel J. Intra-Atrial Block: Definition and Relationship to Atrial Fibrillation and Other Adverse Outcomes. Journal of Atrial Fibrillation. 2019; 12: 2234. https://doi.org/10.4022/jafi b.2234.
- [35] Xixue D, Haihong Y. Clinical analysis of 100 cases of internodal tracts block. Practical Electrocardiology and Clinical Treatment. 2006; 4: 294. (In Chinese) https://doi.org/10.3969/j.issn.1008-0740.2006.04.036.
- [36] Xiang W, Jianan W, Huiqiang C. Atrial block and atrial arrhythmia. Journal of Electrocardiology and Circulation. 2014; 3: 191–195. (In Chinese)
- [37] Nitta T, Yamauchi H, Ohkubo N, Ishii Y, Tanaka S, Hayashi M, et al. Modification of the radial procedure in a patient with partial atrioventricular septal defect. The Annals of Thoracic Surgery. 2002; 73: 661–663. https://doi.org/10.1016/s0003-4975(01)03009-0.
- [38] Kuge K. An experimental study on the influence of surgical injuries to the internodal conduction tracts on supraventricular

- arrhythmias associated with intraatrial operations. Nihon Geka Gakkai Zasshi. 1993; 94: 285–296. (In Japanese)
- [39] Racker DK. Sinoventricular transmission in 10 mM K+ by canine atrioventricular nodal inputs. Superior atrionodal bundle and proximal atrioventricular bundle. Circulation. 1991; 83: 1738–1753. https://doi.org/10.1161/01.cir.83.5.1738.
- [40] Sun C, Yu D, Ye W, Liu C, Gu S, Sinsheimer NR, et al. The short stature homeobox 2 (Shox2)-bone morphogenetic protein (BMP) pathway regulates dorsal mesenchymal protrusion development and its temporary function as a pacemaker during cardiogenesis. The Journal of Biological Chemistry. 2015; 290: 2007– 2023. https://doi.org/10.1074/jbc.M114.619007.
- [41] Sun C, Zhang T, Liu C, Gu S, Chen Y. Generation of Shox2-Cre allele for tissue specific manipulation of genes in the developing heart, palate, and limb. Genesis (New York, N.Y.: 2000). 2013; 51: 515–522. https://doi.org/10.1002/dvg.22397.
- [42] Vicente-Steijn R, Passier R, Wisse LJ, Schalij MJ, Poelmann RE, Gittenberger-de Groot AC, et al. Funny current channel HCN4 delineates the developing cardiac conduction system in chicken heart. Heart Rhythm. 2011; 8: 1254–1263. https://doi.org/10.1016/j.hrthm.2011.03.043.
- [43] Soufan AT, van den Hoff MJB, Ruijter JM, de Boer PAJ, Hagoort J, Webb S, et al. Reconstruction of the patterns of gene expression in the developing mouse heart reveals an architectural arrangement that facilitates the understanding of atrial malformations and arrhythmias. Circulation Research. 2004; 95: 1207–1215. https://doi.org/10.1161/01.RES.0000150852.04747.e1.
- [44] Baudot M, Torre E, Bidaud I, Louradour J, Torrente AG, Fossier L, et al. Concomitant genetic ablation of L-type Ca_v1.3 (α_{1D}) and T-type Ca_v3.1 (α_{1G}) Ca²⁺ channels disrupts heart automaticity. Scientific Reports. 2020; 10: 18906. https://doi.org/10.1038/s41598-020-76049-7.
- [45] Cai CL, Liang X, Shi Y, Chu PH, Pfaff SL, Chen J, et al. Isl1 identifies a cardiac progenitor population that proliferates prior to differentiation and contributes a majority of cells to the heart. Developmental Cell. 2003; 5: 877–889. https://doi.org/10.1016/s1534-5807(03)00363-0.
- [46] Ye W, Wang J, Song Y, Yu D, Sun C, Liu C, et al. A common Shox2-Nkx2-5 antagonistic mechanism primes the pacemaker cell fate in the pulmonary vein myocardium and sinoatrial node. Development (Cambridge, England). 2015; 142: 2521–2532. ht tps://doi.org/10.1242/dev.120220.
- [47] Chung IM, Rajakumar G. Genetics of Congenital Heart Defects: The NKX2-5 Gene, a Key Player. Genes. 2016; 7: 6. https://doi.org/10.3390/genes7020006.
- [48] Jay PY, Berul CI, Tanaka M, Ishii M, Kurachi Y, Izumo S. Cardiac conduction and arrhythmia: insights from Nkx2.5 mutations in mouse and humans. Novartis Foundation Symposium. 2003; 250: 227–238; discussion 238–241, 276–279.
- [49] Ye W, Song Y, Huang Z, Zhang Y, Chen Y. Genetic Regulation of Sinoatrial Node Development and Pacemaker Program in the Venous Pole. Journal of Cardiovascular Development and Disease. 2015; 2: 282–298. https://doi.org/10.3390/jcdd2040282.
- [50] Hu W, Xin Y, Zhao Y, Hu J. Shox2: The Role in Differentiation and Development of Cardiac Conduction System. The Tohoku Journal of Experimental Medicine. 2018; 244: 177–186. https://doi.org/10.1620/tjem.244.177.
- [51] Hoffmann S, Schmitteckert S, Raedecke K, Rheinert D, Diebold S, Roeth R, et al. Network-driven discovery yields new insight into Shox2-dependent cardiac rhythm control. Biochimica et Biophysica Acta. Gene Regulatory Mechanisms. 2021; 1864: 194702. https://doi.org/10.1016/j.bbagrm.2021.194702.
- [52] Hoffmann S, Clauss S, Berger IM, Weiß B, Montalbano A, Röth R, et al. Coding and non-coding variants in the SHOX2 gene in patients with early-onset atrial fibrillation. Basic Research in Cardiology. 2016; 111: 36. https://doi.org/10.1007/ s00395-016-0557-2.



- [53] Li N, Wang ZS, Wang XH, Xu YJ, Qiao Q, Li XM, et al. A SHOX2 loss-of-function mutation underlying familial atrial fibrillation. International Journal of Medical Sciences. 2018; 15: 1564–1572. https://doi.org/10.7150/ijms.27424.
- [54] Liu H, Chen CH, Ye W, Espinoza-Lewis RA, Hu X, Zhang Y, et al. Phosphorylation of Shox2 is required for its function to control sinoatrial node formation. Journal of the American Heart Association. 2014; 3: e000796. https://doi.org/10.1161/JAHA.114.000796.
- [55] Puskaric S, Schmitteckert S, Mori AD, Glaser A, Schneider KU, Bruneau BG, et al. Shox2 mediates Tbx5 activity by regulating Bmp4 in the pacemaker region of the developing heart. Human Molecular Genetics. 2010; 19: 4625–4633. https://doi.org/10. 1093/hmg/ddq393.
- [56] Wang J, Bai Y, Li N, Ye W, Zhang M, Greene SB, et al. Pitx2-microRNA pathway that delimits sinoatrial node development and inhibits predisposition to atrial fibrillation. Proceedings of the National Academy of Sciences of the United States of America. 2014; 111: 9181–9186. https://doi.org/10.1073/pnas.1405411111.
- [57] Hoffmann S, Berger IM, Glaser A, Bacon C, Li L, Gretz N, et al. Islet1 is a direct transcriptional target of the homeodomain transcription factor Shox2 and rescues the Shox2-mediated bradycardia. Basic Research in Cardiology. 2013; 108: 339. https://doi.org/10.1007/s00395-013-0339-z.
- [58] Liu H, Chen CH, Espinoza-Lewis RA, Jiao Z, Sheu I, Hu X, et al. Functional redundancy between human SHOX and mouse Shox2 genes in the regulation of sinoatrial node formation and pacemaking function. The Journal of Biological Chemistry. 2011; 286: 17029–17038. https://doi.org/10.1074/jbc.M111.234252.
- [59] Rivolta I, Binda A, Masi A, DiFrancesco JC. Cardiac and neuronal HCN channelopathies. Pflugers Archiv: European Journal of Physiology. 2020; 472: 931–951. https://doi.org/10.1007/s00424-020-02384-3.
- [60] Stieber J, Herrmann S, Feil S, Löster J, Feil R, Biel M, et al. The hyperpolarization-activated channel HCN4 is required for the generation of pacemaker action potentials in the embryonic heart. Proceedings of the National Academy of Sciences of the United States of America. 2003; 100: 15235–15240. https://doi. org/10.1073/pnas.2434235100.
- [61] Baruscotti M, Bucchi A, Viscomi C, Mandelli G, Consalez G, Gnecchi-Rusconi T, et al. Deep bradycardia and heart block caused by inducible cardiac-specific knockout of the pacemaker channel gene Hcn4. Proceedings of the National Academy of Sciences of the United States of America. 2011; 108: 1705–1710. https://doi.org/10.1073/pnas.1010122108.
- [62] Dong L, Lyu X, Faleti OD, He ML. The special stemness functions of Tbx3 in stem cells and cancer development. Seminars in Cancer Biology. 2019; 57: 105–110. https://doi.org/10.1016/j.semcancer.2018.09.010.
- [63] Van Kempen MJ, Vermeulen JL, Moorman AF, Gros D, Paul DL, Lamers WH. Developmental changes of connexin40 and connexin43 mRNA distribution patterns in the rat heart. Cardio-vascular Research. 1996; 32: 886–900. https://doi.org/10.1016/0008-6363(96)00131-9.

- [64] Delorme B, Dahl E, Jarry-Guichard T, Briand JP, Willecke K, Gros D, et al. Expression pattern of connexin gene products at the early developmental stages of the mouse cardiovascular system. Circulation Research. 1997; 81: 423–437. https://doi.org/10.1161/01.res.81.3.423.
- [65] Moorman AFM, Christoffels VM. Cardiac chamber formation: development, genes, and evolution. Physiological Reviews. 2003; 83: 1223–1267. https://doi.org/10.1152/physrev.00006. 2003.
- [66] Lingbeek ME, Jacobs JJL, van Lohuizen M. The T-box repressors TBX2 and TBX3 specifically regulate the tumor suppressor gene p14ARF via a variant T-site in the initiator. The Journal of Biological Chemistry. 2002; 277: 26120–26127. https://doi.org/10.1074/jbc.M200403200.
- [67] Habets PEMH, Moorman AFM, Clout DEW, van Roon MA, Lingbeek M, van Lohuizen M, et al. Cooperative action of Tbx2 and Nkx2.5 inhibits ANF expression in the atrioventricular canal: implications for cardiac chamber formation. Genes & Development. 2002; 16: 1234–1246. https://doi.org/10.1101/ gad.222902.
- [68] Christoffels VM, Hoogaars WMH, Tessari A, Clout DEW, Moorman AFM, Campione M. T-box transcription factor Tbx2 represses differentiation and formation of the cardiac chambers. Developmental Dynamics: an Official Publication of the American Association of Anatomists. 2004; 229: 763–770. https://doi.org/10.1002/dvdy.10487.
- [69] Wang J, Klysik E, Sood S, Johnson RL, Wehrens XHT, Martin JF. Pitx2 prevents susceptibility to atrial arrhythmias by inhibiting left-sided pacemaker specification. Proceedings of the National Academy of Sciences of the United States of America. 2010; 107: 9753–9758. https://doi.org/10.1073/pnas.0912585107.
- [70] Engbers JDT, Anderson D, Zamponi GW, Turner RW. Signal processing by T-type calcium channel interactions in the cerebellum. Frontiers in Cellular Neuroscience. 2013; 7: 230. https: //doi.org/10.3389/fncel.2013.00230.
- [71] Mesirca P, Torrente AG, Mangoni ME. Functional role of voltage gated Ca(2+) channels in heart automaticity. Frontiers in Physiology. 2015; 6: 19. https://doi.org/10.3389/fphys.2015. 00019.
- [72] Mangoni ME, Traboulsie A, Leoni AL, Couette B, Marger L, Le Quang K, et al. Bradycardia and slowing of the atrioventricular conduction in mice lacking Ca_v3.1/alpha1G T-type calcium channels. Circulation Research. 2006; 98: 1422–1430. https://doi.org/10.1161/01.RES.0000225862.14314.49.
- [73] Chandler NJ, Greener ID, Tellez JO, Inada S, Musa H, Molenaar P, et al. Molecular architecture of the human sinus node: insights into the function of the cardiac pacemaker. Circulation. 2009; 119: 1562–1575. https://doi.org/10.1161/CIRCULATIONAHA 108 804369
- [74] Zaffran S, Kelly RG. New developments in the second heart field. Differentiation; Research in Biological Diversity. 2012; 84: 17–24. https://doi.org/10.1016/j.diff.2012.03.003.
- [75] Gelb BD, Tartaglia M. Noonan syndrome and related disorders: dysregulated RAS-mitogen activated protein kinase signal transduction. Human Molecular Genetics. 2006; 15 Spec No 2: R220–R226. https://doi.org/10.1093/hmg/ddl197.

